

*Appendix A*

LIST OF STRAINS, VECTORS, AND PLASMIDS

TABLE OF CONTENTS

A.1 Strains for Cloning and Protein Expression.....	A-2
DH10B.....	A-2
BL21.....	A-3
A.2 Vectors for Cloning and Protein Expression.....	A-4
pUC19.....	A-4
pQE-80L.....	A-5
A.3 Plasmids Encoding Artificial Protein Genes.....	A-6
pUC19 P.....	A-6
pCU19 A.....	A-8
pUC19 P L37A.....	A-10
pUC19 P L37V.....	A-12
pUC19 P L37I.....	A-14
pUC19 P L44A.....	A-16
pUC19 P T40A.....	A-18
pUC19 P Q54A.....	A-20
pUC19 P I58A.....	A-22
pQE-80L EPE.....	A-24
pQE-80L ERE.....	A-26
pQE-80L ER <sub>c</sub> E.....	A-28
pQE-80L EAE.....	A-30
pQE-80L EPE L37A.....	A-32
pQE-80L EPE L37V.....	A-34
pQE-80L EPE L37I.....	A-36
pQE-80L EPE L44A.....	A-38
pQE-80L EPE T40A.....	A-40
pQE-80L EPE Q54A.....	A-42
pQE-80L EPE I58A.....	A-44

## A.1 Strains for Cloning and Protein Expression

### DH10B

- Genotype:** F- *mcrA*  $\Delta$ (*mrr-hsdRMS-mcrBC*)  $\phi$ 80*dlacZ* $\Delta$ M15 *lacX74 recA1 endA1 araD139*  $\Delta$ (*ara, leu*)7697 *galU galK*  $\lambda$ - *rpsL nupG tonA*
- Source:** Invitrogen Max Efficiency DH10B-T1<sup>R</sup> (from Kai Yuet)
- Use(s):** Cloning strain. Plasmid storage and propagation. Preparation of chemically competent bacteria using the Mix & Go *E. coli* Transformation kit (Zymo Research).
- Notes:** Resistant to phage T1 and T5 infection (*tonA*)
- Availability:**
- (1) 25 v/v % glycerol stock
  - (2) Invitrogen Cat. No. 12331-013

**BL21**

- Genotype:** F- *fhuA2 [lon] ompT gal [dcm] ΔhdsS*
- Source:** New England BioLabs
- Use(s):** Protein expression strain. Preparation of chemically competent bacteria using the Mix & Go *E. coli* Transformation kit (Zymo Research).
- Notes:** Resistant to phage T1 and T5 infection (*fhuA2*, synonym for *tonA*). This strain does not contain the DE3 lysogen and cannot be used to express proteins from the T7 promoter.
- Availability:**
- (1) 25 v/v % glycerol stock
  - (2) NEB Cat. No. C2530H

## A.2 Vectors for Cloning and Protein Expression

### pUC19

**Strain(s)/Plasmid:** DH10B/pUC19

**Antibiotic Resistance:** Ampicillin

**Size** 2686 bp

**Description:** Standard cloning strain developed by Messing and coworkers (*Gene*. 33, 103-119). Site-directed mutagenesis of the P coding region to generate P variants was carried out in pUC19 due to its small size as well as the lack of elastin domains present in pQE-80L EPE, which were found to complicate the mutagenesis protocol.

The multiple cloning site of pUC19 contains numerous restriction enzymes. *EcoRI* and *XbaI* were used to subclone P.

**Sequencing primers:** M13F, M13R available from most sequencing facilities.

**Available Sources:**

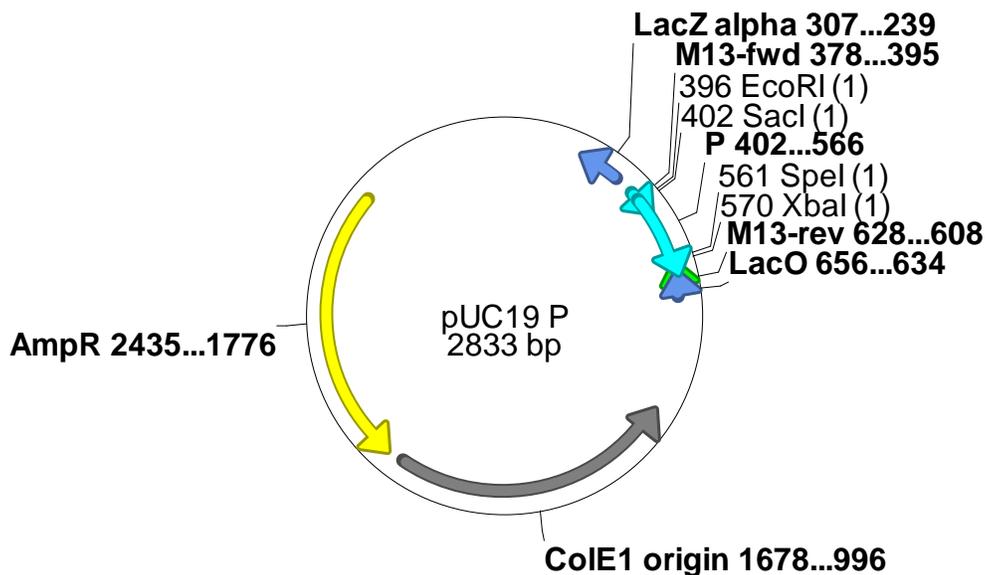
- (1) miniprep DNA (-20°C)
- (2) 25 v/v % glycerol stock in (a) DH10B
- (3) Addgene plasmid #50005

**pQE-80L**

<b><u>Strain(s)/Plasmid:</u></b>	None
<b><u>Antibiotic Resistance:</u></b>	Ampicillin
<b><u>Size</u></b>	4751 bp
<b><u>Description:</u></b>	Qiagen vector for protein expression in <i>E. coli</i> . T5 promoter recruits endogenous RNA polymerase (ie. DE3 lysogen is not required for T7 RNAP mediated transcription). Contains two copies of lac operator (lacO) in promoter region to regulate transcription. Contains a copy of lac repressor gene (lacI) (ie. pREP4 plasmid is not required as it is for other <i>trans</i> -repressed pQE vectors). The ribosomal binding site (RBS) is located several bp upstream of the start codon. The vector contains a sequence encoding an N-terminal 6xHis tag for IMAC purification. This can be removed by cloning at the <i>EcoRI</i> restriction enzyme site, however this site is upstream of the RBS and start codon so these elements must be included in the sequence subcloned into this site.
<b><u>Note:</u></b>	The pQE-80L vectors used in this work were all modified to remove the XhoI restriction site at position 1.
<b><u>Sequencing primers:</u></b>	pQEfor 5' CCC GAA AAG TGC CAC CTG pQErev 5' GTT CTG AGG TCA TTA CTG G
<b><u>Available Sources:</u></b>	(1) Qiagen Cat. No. 32943

**A.3 Plasmids Encoding Artificial Protein Genes****pUC19 P**

<b><u>Submitted by:</u></b>	Larry Dooling
<b><u>Strain(s)/Plasmid:</u></b>	DH10B/pUC19 P
<b><u>Vector:</u></b>	pUC19
<b><u>Insert:</u></b>	P
<b><u>Antibiotic Resistance:</u></b>	Ampicillin
<b><u>Description:</u></b>	P coiled coil domain in the pUC19 vector for mutagenesis. Does not contain start/stop codons. This plasmid is not for protein expression.
<b><u>Construction:</u></b>	The P domain was amplified from pQE-80L EPE with <i>EcoRI</i> and <i>XbaI</i> overhangs, digested and subcloned into pUC19 at the same sites.
<b><u>Available Sources:</u></b>	(1) miniprep DNA (-20°C) (2) 25 v/v % glycerol stock in DH10B (-80°C)

**Plasmid Map:****Coding sequence (*EcoRI* to *XbaI*):**

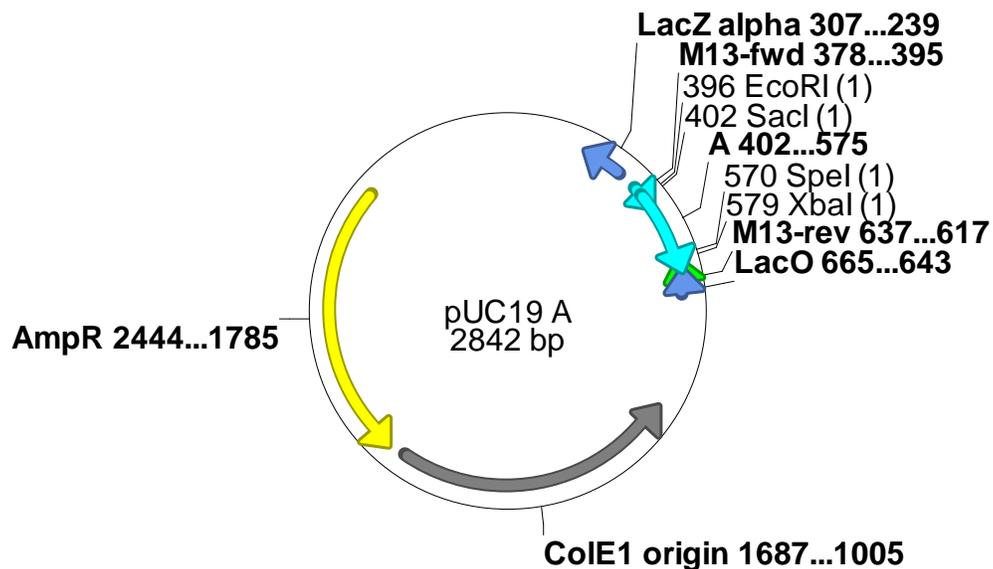
GAATTCGAGCTCGGATCAGGACTTGGATCCGCGCCGCAAATGCTGCGTGAAGTCA  
 GGAAACCAATGCCGCGCTTCAGGATGTGCGGGAATTGCTTCGTCAACAGGTCAAGG  
 AGATAACGTTCTTGAAGAACACCGTCATGGAGTCGGATGCGTCCAAGCTTAATACTA  
 GTGTGTCTAGA

**Translation (*EcoRI* to *XbaI*):**

EFELGSLGSAPQMLRELQETNAALQDVRELLRQQVKEITFLKNTVMESDASKLNTSVS  
 R

**pUC19 A**

<b><u>Submitted by:</u></b>	Larry Dooling
<b><u>Strain(s)/Plasmid:</u></b>	DH10B/pUC19 A
<b><u>Vector:</u></b>	pUC19
<b><u>Insert:</u></b>	A
<b><u>Antibiotic Resistance:</u></b>	Ampicillin
<b><u>Description:</u></b>	A coiled coil domain in the pUC19 vector. Does not contain start/stop codons.
<b><u>Construction:</u></b>	The A domain was amplified by PCR from pQE-9 PC10A with <i>SacI</i> and <i>SpeI</i> overhangs, digested and subcloned into pUC19 at the same sites.
<b><u>Available Sources:</u></b>	(1) miniprep DNA (-20°C) (2) 25 v/v % glycerol stock in DH10B (-80°C)

**Plasmid Map:****Coding sequence (*EcoRI* to *XbaI*):**

GAATTCGAGCTCATGCCGACTAGCGGTGACCTGGAAAACGAAGTGGCCCAGCTGGA  
 AAGGGAAGTTAGATCTCTGGAAGATGAAGCGGCTGAACTGGAACAAAAAGTCTCGA  
 GACTGAAAAATGAAATCGAAGACCTGAAAGCCGAAATTGGTGACCATGTGGCGCCT  
 CGAGACACTAGTGTGTCTAGA

**Translation (*EcoRI* to *XbaI*):**

EFELMPTSGDLENEVAQLEREVRSLEDEAAELEQKVSRLKNEIEDLKAEIGDHVAPRDTS  
 VSR

**pUC19 P L37A**

**Submitted by:** Larry Dooling

**Strain(s)/Plasmid:** DH10B/pUC19 P L37A

**Vector:** pUC19

**Insert:** P L37A

**Antibiotic Resistance:** Ampicillin

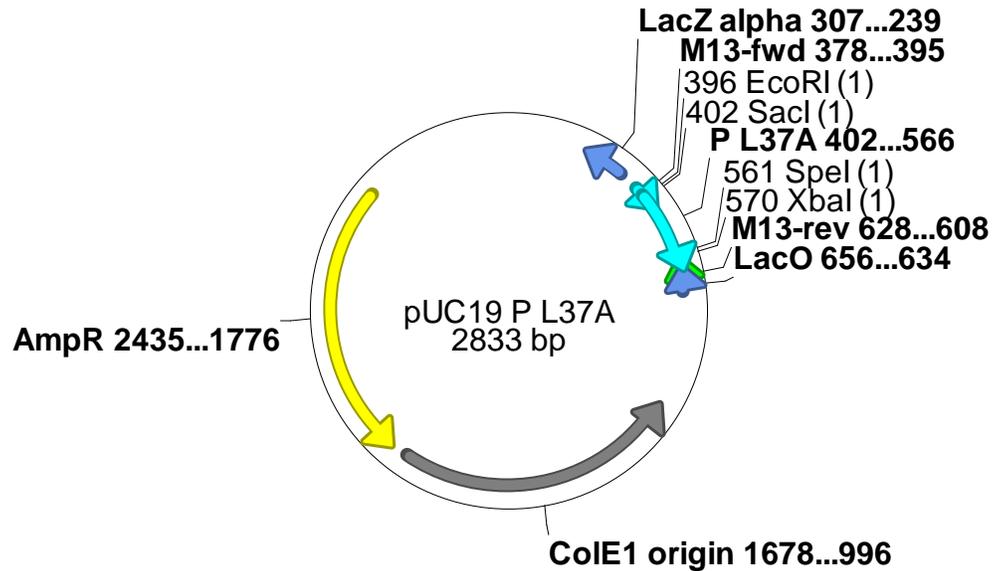
**Description:** P coiled coil domain with L37A mutation based on Gunasekar *et al.*, *Biochemistry* 2009.

**Construction:** The P domain in pUC19 P was mutated by Quick Change with the following primers:

(+) CAA ATG CTG CGT GAA **GCG** CAG GAA ACC AAT GCC  
(-) GGC ATT GGT TTC CTG **CGC** TTC ACG CAG CAT TTG

**Available Sources:**

- (1) miniprep DNA (-20°C)
- (2) 25 v/v % glycerol stock in DH10B (-80°C)

**Plasmid Map:****Coding sequence (*EcoRI* to *XbaI*):**

GAATTCGAGCTCGGATCAGGACTTGGATCCGCGCCGCAAATGCTGCGTGAAGCGCA  
 GGAAACCAATGCCGCGCTTCAGGATGTGCGGGAATTGCTTCGTCAACAGGTCAAGG  
 AGATAACGTTCTTGAAGAACACCGTCATGGAGTCGGATGCGTCCAAGCTTAATACTA  
 GTGTGTCTAGA

**Translation (*EcoRI* to *XbaI*):**

EFELGSLGSAPQMLREAQETNAALQDVRELLRQQVKEITFLKNTVMESDASKLNLSVS  
 R

**pUC19 P L37V**

**Submitted by:** Larry Dooling

**Strain(s)/Plasmid:** DH10B/pUC19 P L37V

**Vector:** pUC19

**Insert:** P L37V

**Antibiotic Resistance:** Ampicillin

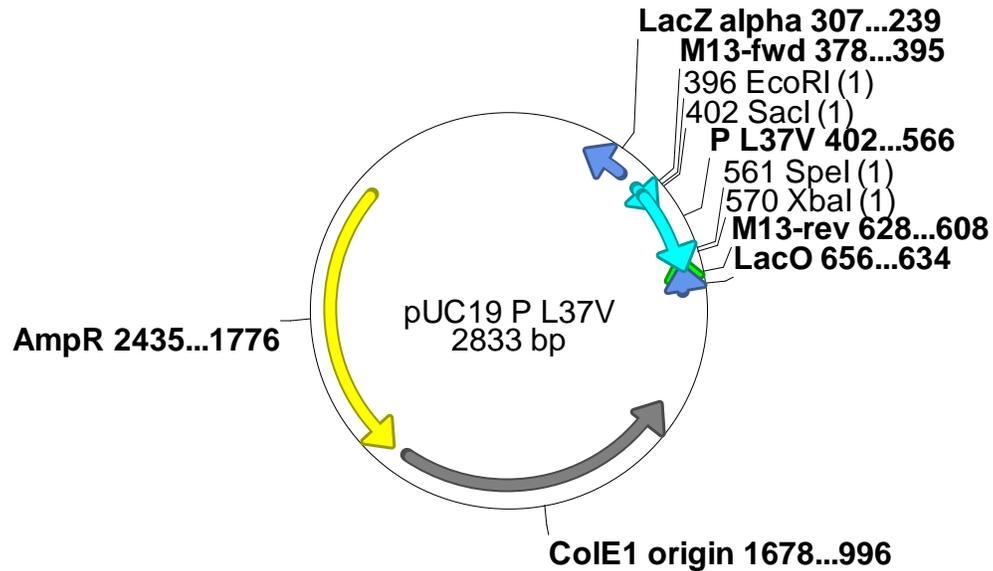
**Description:** P coiled coil domain with L37V mutation. This mutation was not made in Gunasekar *et al.*, *Biochemistry* 2009, but is at the same position as the L37A mutation in that work.

**Construction:** The P domain in pUC19 P was mutated by Quick Change with the following primers:

(+) CAA ATG CTG CGT GAA **GTG** CAG GAA ACC AAT GCC  
(-) GGC ATT GGT TTC CTG **CAC** TTC ACG CAG CAT TTG

**Available Sources:**

- (1) miniprep DNA (-20°C)
- (2) 25 v/v % glycerol stock in DH10B (-80°C)

**Plasmid Map:****Coding sequence (*EcoRI* to *XbaI*):**

GAATTCGAGCTCGGATCAGGACTTGGATCCGCGCCGCAAATGCTGCGTGAAGTGCA  
 GGAAACCAATGCCGCGCTTCAGGATGTGCGGGAATTGCTTCGTCAACAGGTCAAGG  
 AGATAACGTTCTTGAAGAACACCGTCATGGAGTCGGATGCGTCCAAGCTTAATACTA  
 GTGTGTCTAGA

**Translation (*EcoRI* to *XbaI*):**

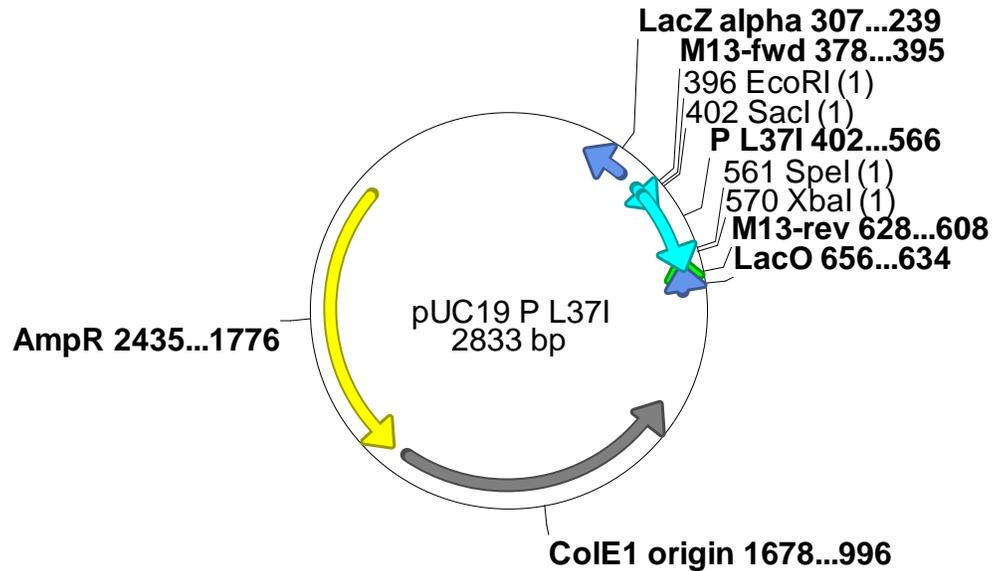
EFELGSLGSAPQMLREVQETNAALQDVRELLRQQVKEITFLKNTVMESDASKLNLSVS  
 R

**pUC19 P L37I**

<b><u>Submitted by:</u></b>	Larry Dooling
<b><u>Strain(s)/Plasmid:</u></b>	DH10B/pUC19 P L37I
<b><u>Vector:</u></b>	pUC19
<b><u>Insert:</u></b>	P L37I
<b><u>Antibiotic Resistance:</u></b>	Ampicillin
<b><u>Description:</u></b>	P coiled coil domain with L37I mutation. This mutation was not made in Gunasekar <i>et al.</i> , <i>Biochemistry</i> 2009, but is at the same position as the L37A mutation in that work.
<b><u>Construction:</u></b>	The P domain in pUC19 P was mutated by Quick Change with the following primers:

(+) CAA ATG CTG CGT GAA **ATT** CAG GAA ACC AAT GCC  
(-) GGC ATT GGT TTC CTG **CAC** TTC ACG CAG CAT TTG

<b><u>Available Sources:</u></b>	(1) miniprep DNA (-20°C)
	(2) 25 v/v % glycerol stock in DH10B (-80°C)

**Plasmid Map:****Coding sequence (*EcoRI* to *XbaI*):**

GAATTCGAGCTCGGATCAGGACTTGGATCCGCGCCGCAAATGCTGCGTGAAATTCA  
 GGAAACCAATGCCGCGCTTCAGGATGTGCGGGAATTGCTTCGTCAACAGGTCAAGG  
 AGATAACGTTCTTGAAGAACACCGTCATGGAGTCGGATGCGTCCAAGCTTAATACTA  
 GTGTGTCTAGA

**Translation (*EcoRI* to *XbaI*):**

EFELGSGGLGSAPQMLREIQETNAALQDVRELLRQQVKEITFLKNTVMESDASKLNTSVS  
 R

**pUC19 P L44A**

**Submitted by:** Larry Dooling

**Strain(s)/Plasmid:** DH10B /pUC19 P L44A

**Vector:** pUC19

**Insert:** P L44A

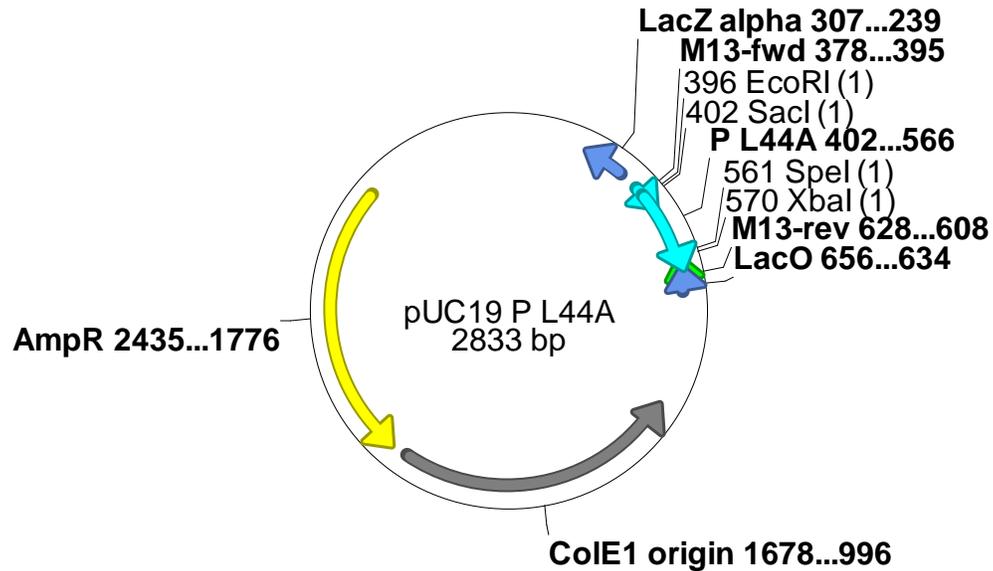
**Antibiotic Resistance:** Ampicillin

**Description:** P coiled coil domain with L44A mutation based on Gunasekar *et al.*, *Biochemistry* 2009.

**Construction:** The P domain in pUC19 P was mutated by Quick Change with the following primers:

(+) CAG GAA ACC AAT GCC GCG **GCT** CAG GAT GTG CGG GAA TTG C  
(-) GCA ATT CCC GCA CAT CCT **GAG** **CCG** CGG CAT TGG TTT CCT G

**Available Sources:** (1) miniprep DNA (-20°C)  
(2) 25 v/v % glycerol stock in DH10B (-80°C)

**Plasmid Map:****Coding sequence (*EcoRI* to *XbaI*):**

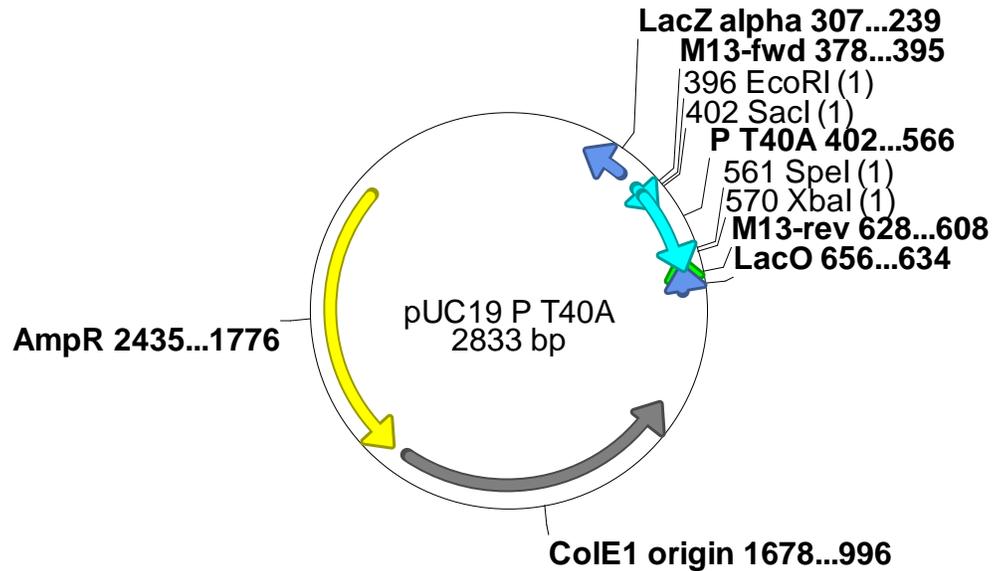
GAATTCGAGCTCGGATCAGGACTTGGATCCGCGCCGCAAATGCTGCGTGAAGTCA  
 GGAAACCAATGCCGCGGCTCAGGATGTGCGGGAATTGCTTCGTCAACAGGTCAAGG  
 AGATAACGTTCTTGAAGAACACCGTCATGGAGTCGGATGCGTCCAAGCTTAATACTA  
 GTGTGTCTAGA

**Translation (*EcoRI* to *XbaI*):**

EFELGSLGSAPQMLRELQETNAAAQDVRELLRQQVKEITFLKNTVMESDASKLNLSVS  
 R

**pUC19 P T40A**

<b><u>Submitted by:</u></b>	Larry Dooling
<b><u>Strain(s)/Plasmid:</u></b>	DH10B/pUC19 P T40A
<b><u>Vector:</u></b>	pUC19
<b><u>Insert:</u></b>	P T40A
<b><u>Antibiotic Resistance:</u></b>	Ampicillin
<b><u>Description:</u></b>	P coiled coil domain with T40A mutation based on Gunasekar <i>et al.</i> , <i>Biochemistry</i> 2009.
<b><u>Construction:</u></b>	The P domain in pUC19 P was mutated by Quick Change with the following primers:  (+) GAA CTG CAG GAA <u>G</u> CC AAT GCC GCG C (-) G CGC GGC ATT <u>G</u> GC TTC CTG CAG TTC
<b><u>Available Sources:</u></b>	(1) miniprep DNA (-20°C)  (2) 25 v/v % glycerol stock in DH10B (-80°C)

**Plasmid Map:****Coding sequence (*EcoRI* to *XbaI*):**

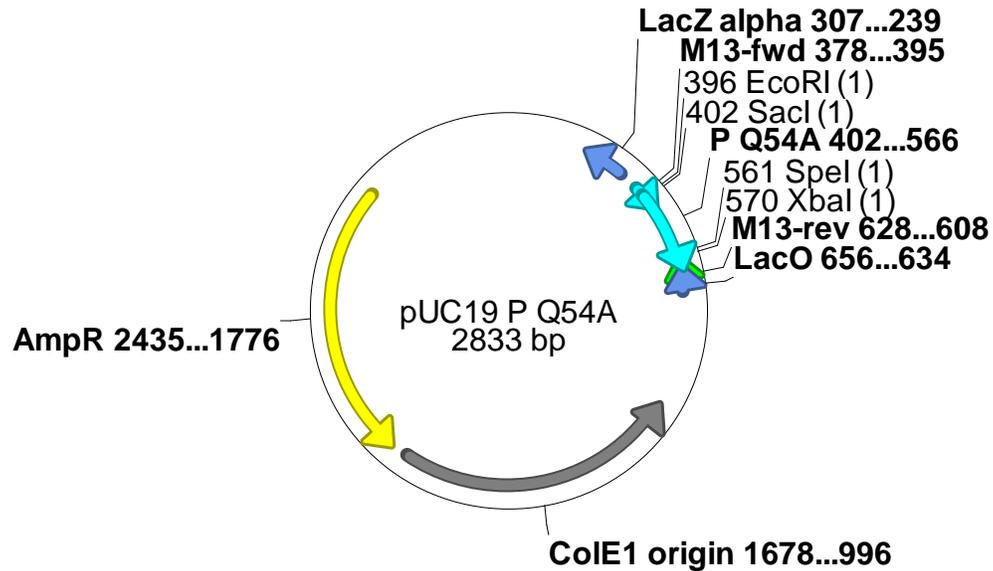
GAATTCGAGCTCGGATCAGGACTTGGATCCGCGCCGCAAATGCTGCGTGAAGTCA  
GGAAGCCAATGCCGCGCTTCAGGATGTGCGGGAATTGCTTCGTCAACAGGTCAAGG  
AGATAACGTTCTTGAAGAACACCGTCATGGAGTCGGATGCGTCCAAGCTTAATACTA  
GTGTGTCTAGA

**Translation (*EcoRI* to *XbaI*):**

EFELGSGLGSA PQMLRELQEANAALQDVRELLRQQVKEITFLKNTVMESDASKLNLSVS  
R

**pUC19 P Q54A**

<b><u>Submitted by:</u></b>	Larry Dooling
<b><u>Strain(s)/Plasmid:</u></b>	DH10B/pUC19 P Q54A
<b><u>Vector:</u></b>	pUC19
<b><u>Insert:</u></b>	P Q54A
<b><u>Antibiotic Resistance:</u></b>	Ampicillin
<b><u>Description:</u></b>	P coiled coil domain with Q54A mutation based on Gunasekar <i>et al.</i> , <i>Biochemistry</i> 2009.
<b><u>Construction:</u></b>	The P domain in pUC19 P was mutated by Quick Change with the following primers:  (+) GAA TTG CTT CGT CAA <b><u>GCG</u></b> GTC AAG GAG ATA AC (-) GT TAT CTC CTT GAC <b><u>CGC</u></b> TTG ACG AAG CAA TTC
<b><u>Available Sources:</u></b>	(1) miniprep DNA (-20°C)  (2) 25 v/v % glycerol stock in DH10B (-80°C)

**Plasmid Map:****Coding sequence (*EcoRI* to *XbaI*):**

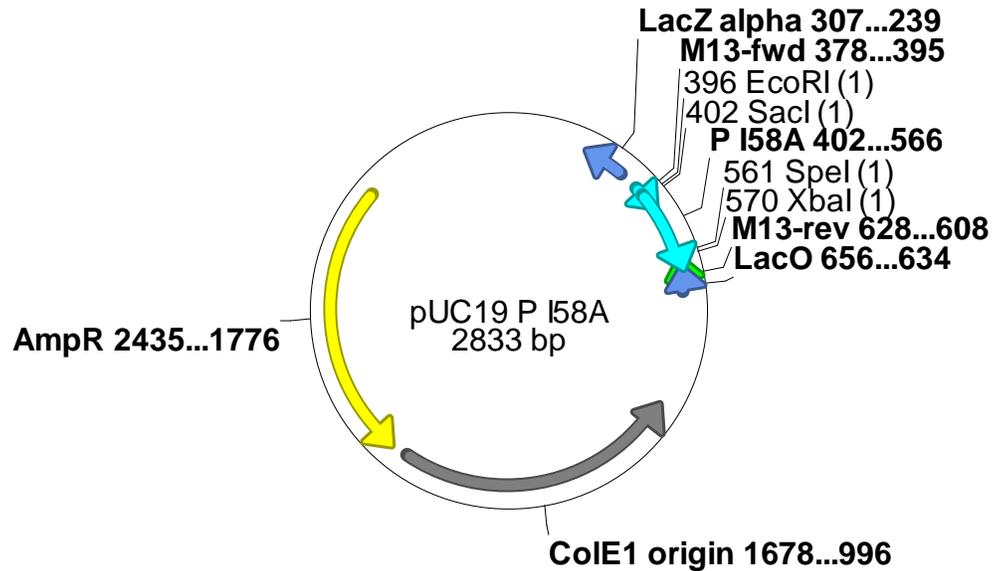
GAATTCGAGCTCGGATCAGGACTTGGATCCGCGCCGCAAATGCTGCGTGAAGTCA  
 GGAAACCAATGCCGCGCTTCAGGATGTGCGGGAATTGCTTCGTCAAGCGGTCAAGG  
 AGATAACGTTCTTGAAGAACACCGTCATGGAGTCGGATGCGTCCAAGCTTAATACTA  
 GTGTGTCTAGA

**Translation (*EcoRI* to *XbaI*):**

EFELGSLGSA PQMLRELQETNAALQDVRELLRQAVKEITFLKNTVMESDASKLNLSVS  
 R

**pUC19 P I58A**

<b><u>Submitted by:</u></b>	Larry Dooling
<b><u>Strain(s)/Plasmid:</u></b>	DH10B/pUC19 P I58A
<b><u>Vector:</u></b>	pUC19
<b><u>Insert:</u></b>	P I58A
<b><u>Antibiotic Resistance:</u></b>	Ampicillin
<b><u>Description:</u></b>	P coiled coil domain with I58A mutation based on Gunasekar <i>et al.</i> , <i>Biochemistry</i> 2009.
<b><u>Construction:</u></b>	The P domain in pUC19 P was mutated by Quick Change with the following primers:  (+) CAG GTC AAG GAG <b><u>GCA</u></b> ACG TTC TTG AAG (-) CTT CAA GAA CGT <b><u>TGC</u></b> CTC CTT GAC CTG
<b><u>Available Sources:</u></b>	(1) miniprep DNA (-20°C)  (2) 25 v/v % glycerol stock in DH10B (-80°C)

**Plasmid Map:****Coding sequence (*EcoRI* to *XbaI*):**

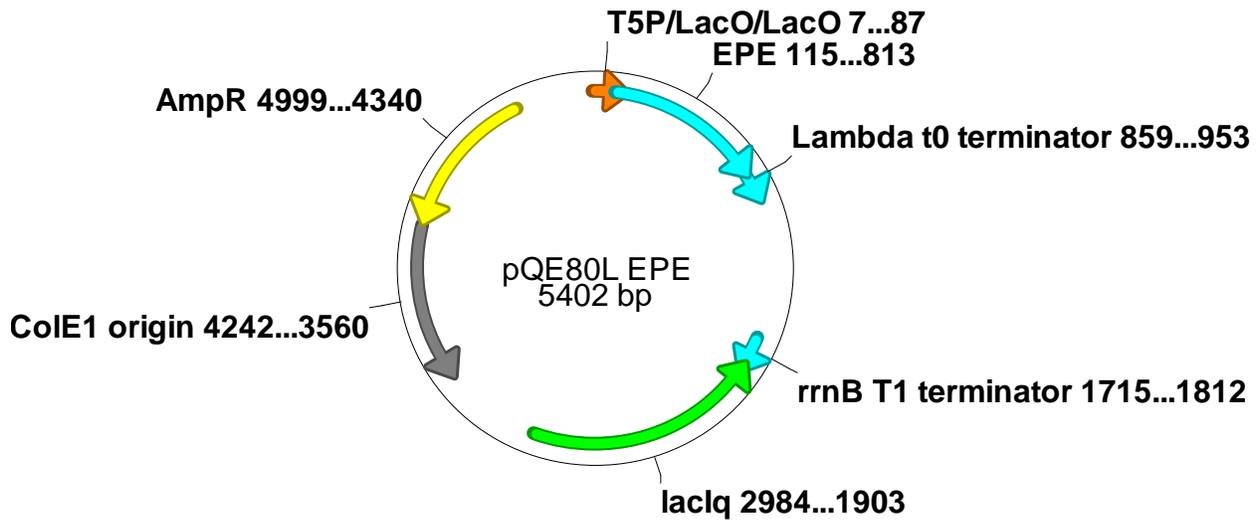
GAATTCGAGCTCGGATCAGGACTTGGATCCGCGCCGCAAATGCTGCGTGAAGTCA  
 GGAAACCAATGCCGCGCTTCAGGATGTGCGGGAATTGCTTCGTCAACAGGTCAAGG  
 AGGCAACGTTCTTGAAGAACACCGTCATGGAGTCGGATGCGTCCAAGCTTAATACTA  
 GTGTGTCTAGA

**Translation (*EcoRI* to *XbaI*):**

EFELGSGLGSA PQMLRELQETNAALQDVRELLRQQVKEATFLKNTVMESDASKLNTSV  
 SR

**pQE-80L EPE**

<b><u>Submitted by:</u></b>	Wen-Bin Zhang/Larry Dooling
<b><u>Strain(s)/Plasmid:</u></b>	DH10B/pQE-80L EPE BL21/pQE-80L EPE
<b><u>Vector:</u></b>	pQE-80L
<b><u>Insert:</u></b>	EPE
<b><u>Antibiotic Resistance:</u></b>	Ampicillin
<b><u>Description:</u></b>	Hydrophilic elastin-like repeats [(VPGVG) <sub>2</sub> VPGE(VPGVG) <sub>2</sub> ] <sub>3</sub> flanking a P coiled coil motif. N- and C-terminal Cys residues facilitate cross-linking with 4-arm PEG maleimide, vinyl sulfone, etc.
<b><u>Construction:</u></b>	Prepared by Wen-Bin Zhang.
<b><u>Available Sources:</u></b>	(1) miniprep DNA (-20°C) (2) 25 v/v % glycerol stock in DH10B (-80°C) (3) 25 v/v % glycerol stock in BL21 (-80°C)

**Plasmid Map:****Coding sequence:**

```

ATGAGATGCAGCAGCCATCATCATCATCACGTCGACGGCCACGGCGTGGGTGTT
CCGGGCGTCCGGTGTGCCGGGTGTGGGTGTGCCGGGCGAAGGTGTGCCGGGCGTCCG
TGTGCCGGGTGTTGGTGTTCGGGGCGTTGGTGTGCCGGGCGTTGGCGTGCCGGGCGA
GGGTGTGCCGGGCGTTGGTGTTCGGGGCGTGGGTGTGCCGGGCGTGGGCGTGCCGG
GCGTCCGTGTTCCGGGCGAGGGTGTGCCGGGCGTAGGTGTGCCGGGCGTTGGTGAG
CTCGGATCAGGACTTGGATCCGCGCCGCAAATGCTGCGTGAAGTGCAGGAAACCAA
TGCCGCGCTTCAGGATGTGCCGGAATTGCTTCGTCAACAGGTCAAGGAGATAACGTT
CTTGAAGAACACCGTTCATGGAGTCGGATGCGTCCAAGCTTAATACTAGTGTGCCGGG
CGTCGGGCGTGCCGGGCGTAGGTGTTCCGGGCGAGGGTGTTCGGGCGTTGGTGTGCC
GGGCGTCGGCGTGCCGGGCGTGGGTGTTCCGGGCGTAGGTGTGCCGGGCGAGGGTG
TGCCGGGCGTGGGCGTGCCGGGCGTAGGTGTTCCGGGCGTAGGTGTTCCGGGCGTA
GGTGTTCGGGTGAAGGCGTGCCGGGCGTTGGTGTGCCGGGTGTGGGCGTGCCGGG
CGGGCTGCTCGAGTGCATGTAA

```

**Translation:**

```

MRCSSHHHHHVDGHGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPG
EGVPGVGVPGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPGVGVPG
AALQDVRELLRQQVKEITFLKNTVMESDASKLNTSVPGVGVPGVGVPGEGVPGVGVPG
VGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPGVGVPGVGVPGVGVPGG
LLECM*

```

**pQE-80L ERE**

<b><u>Submitted by:</u></b>	Wen-Bin Zhang/Larry Dooling
<b><u>Strain(s)/Plasmid:</u></b>	DH10B/pQE-80L ERE BL21/pQE-80L ERE
<b><u>Vector:</u></b>	pQE-80L
<b><u>Insert:</u></b>	ERE
<b><u>Antibiotic Resistance:</u></b>	Ampicillin
<b><u>Description:</u></b>	Hydrophilic elastin-like repeats [(VPGVG) <sub>2</sub> VPGEG(VPGVG) <sub>2</sub> ] <sub>3</sub> flanking a 17mer RGD motif. N- and C-terminal Cys residues facilitate cross-linking with 4-arm PEG maleimide, vinyl sulfone, etc. The MMP1 recognition sequence (GPQGIWGQ) is included before the C-terminal Cys. Also originally denoted as CEC for Cysteine-Elastin-Cysteine.
<b><u>Construction:</u></b>	Prepared by Wen-Bin Zhang.
<b><u>Available Sources:</u></b>	(1) miniprep DNA (-20°C) (2) 25 v/v % glycerol stock in DH10B (-80°C) (3) 25 v/v % glycerol stock in BL21 (-80°C)



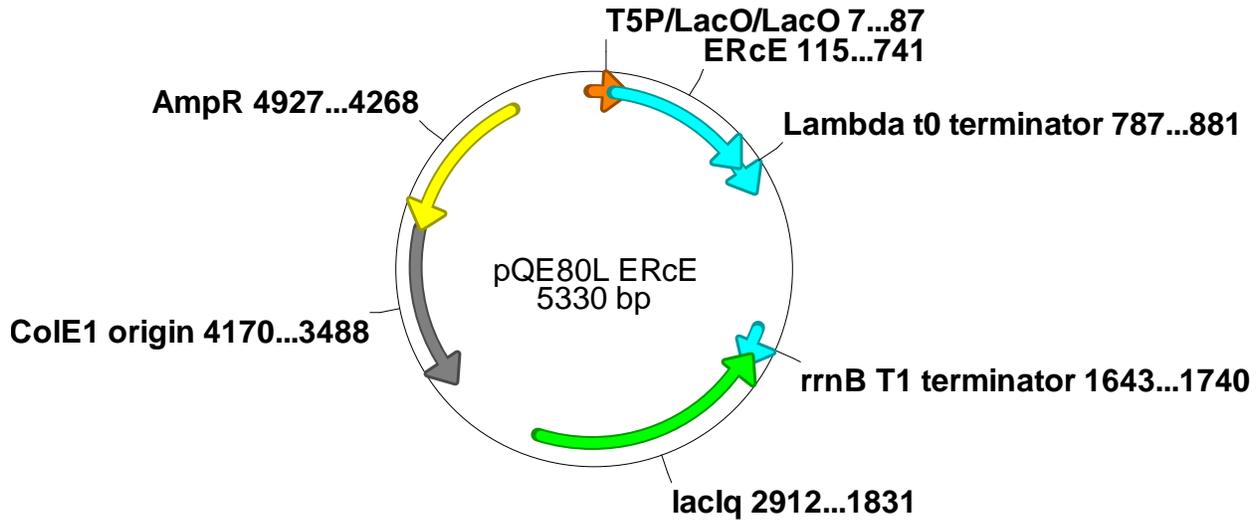
**pQE-80L ER<sub>C</sub>E****Submitted by:** Larry Dooling**Strain(s)/Plasmid:** DH10B/pQE-80L ER<sub>C</sub>E  
BL21/pQE-80L ER<sub>C</sub>E**Vector:** pQE-80L**Insert:** ER<sub>C</sub>E or ERE S104C**Antibiotic Resistance:** Ampicillin**Description:** Hydrophilic elastin-like repeats [(VPGVG)<sub>2</sub>VPGEG(VPGVG)<sub>2</sub>]<sub>3</sub> flanking a 17mer RGD motif. N- and C-terminal Cys residues facilitate cross-linking with 4-arm PEG maleimide, vinyl sulfone, etc. The MMP1 recognition sequence (GPQGIWGQ) is included before the C-terminal Cys. The RGD domains contains a Ser to Cys mutation (RGDS → RGDC) to introduce a cross-linking site in the middle of the protein. This new RGD domain is abbreviated R<sub>C</sub>.**Construction:** Serine 104 was mutated to cysteine by Quick Change of pQE-80L ERE with the following primers:

(+) GTT ACC GGC CGT GGT GAT **TGT** CCG GCC AGC TCT GCC  
 (-) GGC AGA GCT GGC CGG **ACA** ATC ACC ACG GCC GGT AAC

This resulted in the correct mutation but was accompanied by the deletion of the C-terminal E domain. To generate the correct construct, the *EcoRI-SpeI* fragment of this plasmid was isolated by restriction enzyme digestion and subcloned into pQE-80L ERE digested with the same enzymes (replacing ER with ER<sub>C</sub>).

**Available Sources:**

- (1) miniprep DNA (-20°C)
- (2) 25 v/v % glycerol stock in DH10B (-80°C)
- (3) 25 v/v % glycerol stock in BL21 (-80°C)

**Plasmid Map:****Coding sequence:**

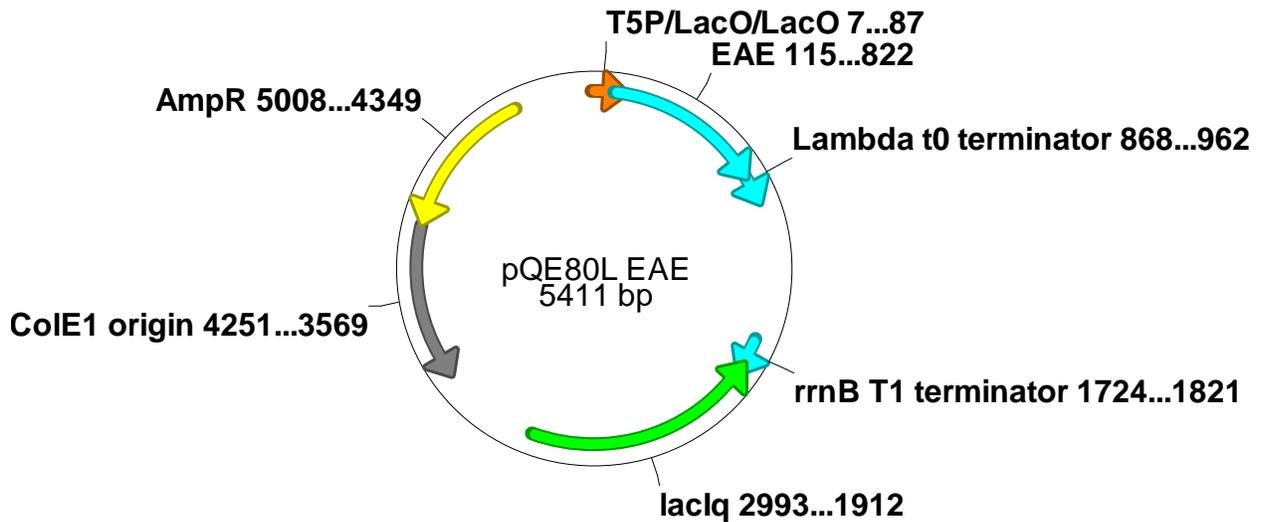
ATGAGATGCAGCAGCCATCATCATCATCACGTCGACGGCCACGGCGTGGGTGTT  
 CCGGGCGTCCGGTGTGCCGGGTGTGGGTGTGCCGGGCGAAGGTGTGCCGGGCGTCCG  
 TGTGCCGGGTGTTGGTGTTCGGGGCGTTGGTGTGCCGGGCGTTGGCGTGCCGGGGCGA  
 GGGTGTGCCGGGCGTTGGTGTTCGGGGCGTGGGTGTGCCGGGCGTGGGCGTGCCGG  
 GCGTCCGGTGTTCGGGGCGAGGGTGTGCCGGGCGTAGGTGTGCCGGGCGTTGGTGAG  
 CTCTATGCGGTTACCGGCCGTGGTGATTGTCCGGCCAGCTCTGCCCCGATCGCCACT  
 AGTGTGCCGGGCGTCCGGCGTGCCGGGCGTAGGTGTTCCGGGGCGAGGGTGTTCGGGG  
 CGTTGGTGTGCCGGGCGTCCGGCGTGCCGGGCGTGGGTGTTCCGGGGCGTAGGTGTGCC  
 GGGCGAGGGTGTGCCGGGCGTGGGCGTGCCGGGCGTAGGTGTTCCGGGGCGTAGGTG  
 TTCGGGGCGTAGGTGTTCCGGGTGAAGGCGTGCCGGGCGTTGGTGTGCCGGGTGTGG  
 GCGTGCCGGGCGGGCTGCTCGACGGTCCGCAAGGTATTTGGGGTCAGCTCGAGTGC  
 ATGTAA

**Translation:**

MRCSSHHHHHVDGHGVGVPGVGVPGVGVPGEVPGVGVPGVGVPGVGVPGVGVPGVGVPG  
 EGVPGVGVPGVGVPGVGVPGVGVPGEGVPGVGVPGVGVGELYAVTGRGDCPASSAPIATS  
 VPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPG  
 GVGVPGEVPGVGVPGVGVPGGLLDGPPQGIWGQLECM\*

**pQE-80L EAE**

<b><u>Submitted by:</u></b>	Larry Dooling
<b><u>Strain(s)/Plasmid:</u></b>	DH10B/pQE-80L EAE BL21/pQE-80L EAE
<b><u>Vector:</u></b>	pQE-80L
<b><u>Insert:</u></b>	EAE
<b><u>Antibiotic Resistance:</u></b>	Ampicillin
<b><u>Description:</u></b>	Hydrophilic elastin-like repeats [(VPGVG) <sub>2</sub> VPGE(VPGVG) <sub>2</sub> ] <sub>3</sub> flanking a A coiled coil/leucine zipper motif.
<b><u>Construction:</u></b>	The A domain was amplified by PCR from pQE-9 PC10A with <i>SacI</i> and <i>SpeI</i> overhangs, digested and subcloned into the similarly-digested pQE-80L EPE plasmid.
<b><u>Available Sources:</u></b>	(1) miniprep DNA (-20°C) (2) 25 v/v % glycerol stock in DH10B (-80°C) (3) 25 v/v % glycerol stock in BL21 (-80°C)

**Plasmid Map:****Coding sequence:**

```

ATGAGATGCAGCAGCCATCATCATCATCACGTCGACGGCCACGGCGTGGGTGTT
CCGGGCGTCCGGTGTGCCGGGTGTGGGTGTGCCGGGCGAAGGTGTGCCGGGCGTCCG
TGTGCCGGGTGTTGGTGTTCGGGGCGTTGGTGTGCCGGGCGTTGGCGTGCCGGGCGA
GGGTGTGCCGGGCGTTGGTGTTCGGGGCGTGGGTGTGCCGGGCGTGGGCGTGCCGG
GCGTCGGTGTTCGGGGCGAGGGTGTGCCGGGCGTAGGTGTGCCGGGCGTTGGTGAG
CTCATGCCGACTAGCGGTGACCTGGAAAACGAAGTGGCCCAGCTGGAAAGGGAAGT
TAGATCTCTGGAAGATGAAGCGGCTGAACTGGAACAAAAAGTCTCGAGACTGAAAA
ATGAAATCGAAGACCTGAAAGCCGAAATTGGTGACCATGTGGCGCCTCGAGACACT
AGTGTGCCGGGCGTCCGGCGTGCCGGGCGTAGGTGTTCCGGGCGAGGGTGTTCGGG
CGTTGGTGTGCCGGGCGTCCGGCGTGCCGGGCGTGGGTGTTCCGGGCGTAGGTGTGCC
GGGCGAGGGTGTGCCGGGCGTGGGCGTGCCGGGCGTAGGTGTTCCGGGCGTAGGTG
TTCGGGCGTAGGTGTTCCGGGTGAAGGCGTGCCGGGCGTTGGTGTGCCGGGTGTGG
GCGTGCCGGGCGGGCTGCTCGAGTGCATGTAA

```

**Translation:**

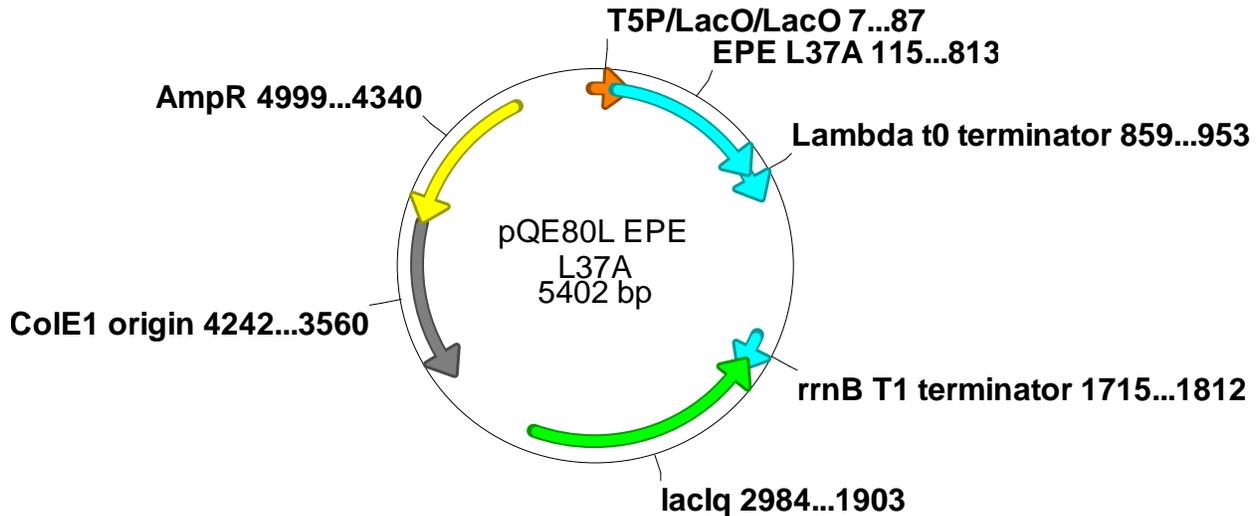
```

MRCSSHHHHHVDGHGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPG
EGVPGVGVPGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGELMPTSGDLENEVAQLEREV
RSLEDEAAELEQKVSRLKNEIEDLKAEIGDHVAPRDTSVPGVGVPGVGVPGEGVPGVGV
PGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPG
GLLECM*

```

**pQE-80L EPE L37A**

<b><u>Submitted by:</u></b>	Larry Dooling
<b><u>Strain(s)/Plasmid:</u></b>	DH10B1/pQE-80L EPE L37A BL21/pQE-80L EPE L37A
<b><u>Vector:</u></b>	pQE-80L
<b><u>Insert:</u></b>	EPE L37A
<b><u>Antibiotic Resistance:</u></b>	Ampicillin
<b><u>Description:</u></b>	Hydrophilic elastin-like repeats [(VPGVG) <sub>2</sub> VPGEG(VPGVG) <sub>2</sub> ] <sub>3</sub> flanking a P coiled coil motif containing the L37A mutation.
<b><u>Construction:</u></b>	The P domain was isolated from pUC19 P L37A by digestion with <i>SacI</i> and <i>SpeI</i> and subcloned into pQE-80L EPE digested with the same enzymes.
<b><u>Available Sources:</u></b>	(1) miniprep DNA (-20°C) (2) 25 v/v % glycerol stock in DH10B (-80°C) (3) 25 v/v % glycerol stock in BL21 (-80°C)

**Plasmid Map:****Coding sequence:**

```

ATGAGATGCAGCAGCCATCATCATCATCACGTCGACGGCCACGGCGTGGGTGTT
CCGGGCGTCGGTGTGCCGGGTGTGGGTGTGCCGGGCGAAGGTGTGCCGGGCGTCGG
TGTGCCGGGTGTTGGTGTTCGGGGCGTTGGTGTGCCGGGCGTTGGCGTGCCGGGCGA
GGGTGTGCCGGGCGTTGGTGTTCGGGGCGTGGGTGTGCCGGGCGTGGGCGTGCCGG
GCGTCGGTGTTCGGGGCGAGGGTGTGCCGGGCGTAGGTGTGCCGGGCGTTGGTGAG
CTCGGATCAGGACTTGGATCCGCGCCGCAAATGCTGCGTGAAGCGCAGGAAACCAA
TGCCGCGCTTCAGGATGTGCGGGAATTGCTTCGTCAACAGGTCAAGGAGATAACGTT
CTTGAAGAACACCGTCATGGAGTCGGATGCGTCCAAGCTTAATACTAGTGTGCCGGG
CGTCGGCGTGCCGGGCGTAGGTGTTCCGGGCGAGGGTGTTCGGGCGTTGGTGTGCC
GGGCGTCGGCGTGCCGGGCGTGGGTGTTCCGGGCGTAGGTGTGCCGGGCGAGGGTG
TGCCGGGCGTGGGCGTGCCGGGCGTAGGTGTTCCGGGCGTAGGTGTTCCGGGCGTA
GGTGTTCGGGTGAAGGCGTGCCGGGCGTTGGTGTGCCGGGTGTGGGCGTGCCGGG
CGGGCTGCTCGAGTGCATGTAA

```

**Translation:**

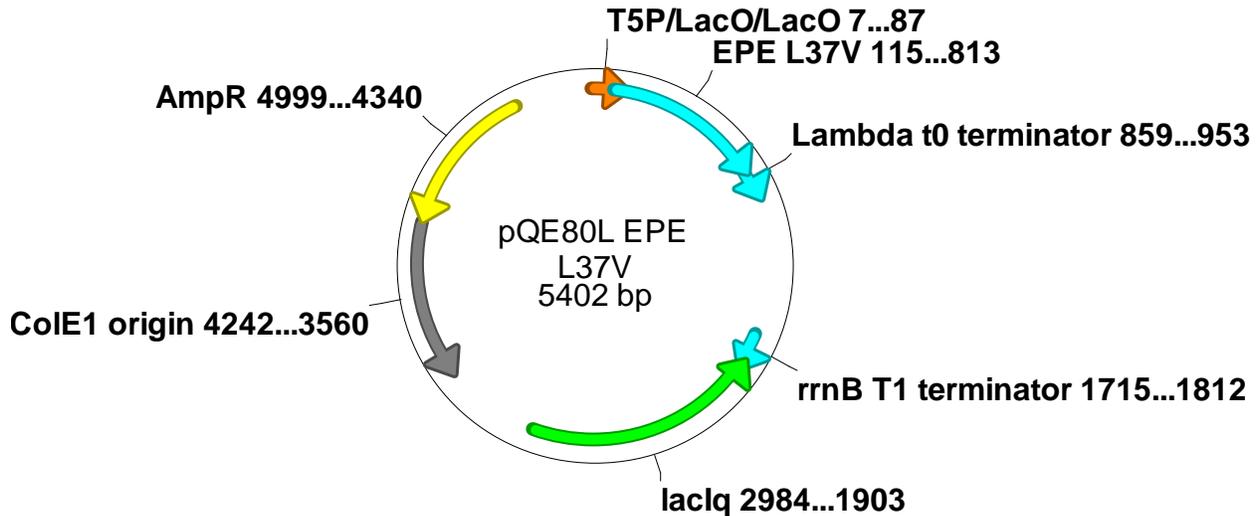
```

MRCSSHHHHHVDGHGVPGVPGVGPVPGEGVPGVPGVGPVPGVGPVPGVGPVPG
EGVPGVGPVPGVGPVPGVGPVPGEGVPGVGPVPGVGPVPGVGPVPGVGPVPGV
AALQDVRELLRQQVKEITFLKNTVMESDASKLNTSVPGVGPVGPVGPVGPVGPV
VGPVGPVGPVGPVGPVGPVGPVGPVGPVGPVGPVGPVGPVGPVGPVGPVGPVGP
LLECM*

```

**pQE-80L EPE L37V**

<b><u>Submitted by:</u></b>	Larry Dooling
<b><u>Strain(s)/Plasmid:</u></b>	DH10B/pQE-80L EPE L37V BL21/pQE-80L EPE L37V
<b><u>Vector:</u></b>	pQE-80L
<b><u>Insert:</u></b>	EPE L37V
<b><u>Antibiotic Resistance:</u></b>	Ampicillin
<b><u>Description:</u></b>	Hydrophilic elastin-like repeats [(VPGVG) <sub>2</sub> VPGEG(VPGVG) <sub>2</sub> ] <sub>3</sub> flanking a P coiled coil motif containing the L37V mutation.
<b><u>Construction:</u></b>	The P domain was isolated from pUC19 P L37V by digestion with <i>SacI</i> and <i>SpeI</i> and subcloned into pQE-80L EPE digested with the same enzymes.
<b><u>Available Sources:</u></b>	(1) miniprep DNA (-20°C) (2) 25 v/v % glycerol stock in DH10B (-80°C) (3) 25 v/v % glycerol stock in BL21 (-80°C)

**Plasmid Map:****Coding sequence:**

```

ATGAGATGCAGCAGCCATCATCATCATCACGTCGACGGCCACGGCGTGGGTGTT
CCGGGCGTCCGGTGTGCCGGGTGTGGGTGTGCCGGGCGAAGGTGTGCCGGGCGTCCG
TGTGCCGGGTGTTGGTGTTCGGGGCGTTGGTGTGCCGGGCGTTGGCGTGCCGGGCGA
GGGTGTGCCGGGCGTTGGTGTTCGGGGCGTGGGTGTGCCGGGCGTGGGCGTGCCGG
GCGTCCGGTGTTCGGGGCGAGGGTGTGCCGGGCGTAGGTGTGCCGGGCGTTGGTGAG
CTCGGATCAGGACTTGGATCCGCGCCGCAAATGCTGCGTGAAGTGCAGGAAACCAA
TGCCGCGCTTCAGGATGTGCCGGGAATTGCTTCGTCAACAGGTCAAGGAGATAACGTT
CTTGAAGAACACCGTCATGGAGTCGGATGCGTCCAAGCTTAATACTAGTGTGCCGGG
CGTCGGCGTGCCGGGCGTAGGTGTTCCGGGCGAGGGTGTTCGGGGCGTTGGTGTGCC
GGGCGTCGGCGTGCCGGGCGTGGGTGTTCCGGGCGTAGGTGTGCCGGGCGAGGGTG
TGCCGGGCGTGGGCGTGCCGGGCGTAGGTGTTCCGGGCGTAGGTGTTCCGGGCGTA
GGTGTTCGGGTGAAGGCGTGCCGGGCGTTGGTGTGCCGGGTGTGGGCGTGCCGGG
CGGGCTGCTCGAGTGCATGTAA

```

**Translation:**

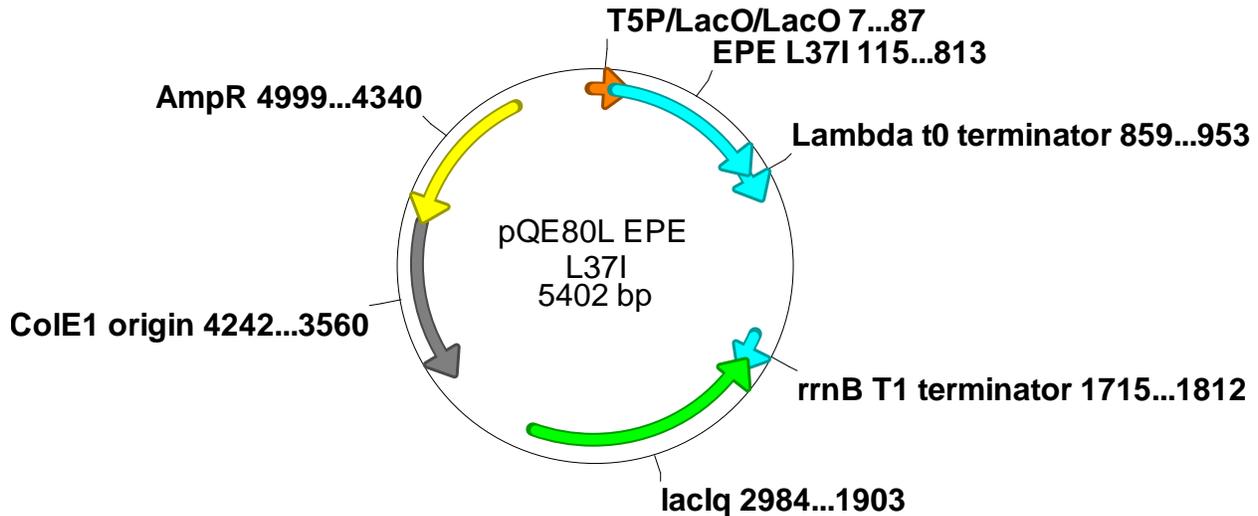
```

MRCSSHHHHHVDGHGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPG
EGVPGVGVPGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPGVGVPG
AALQDVRELLRQQVKEITFLKNTVMESDASKLNTSVPGVGVPGVGVPGEGVPGVGVPG
VGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGG
LLECM*

```

**pQE-80L EPE L37I**

<b><u>Submitted by:</u></b>	Larry Dooling
<b><u>Strain(s)/Plasmid:</u></b>	DH10B/pQE-80L EPE L37I BL21/pQE-80L EPE L37I
<b><u>Vector:</u></b>	pQE-80L
<b><u>Insert:</u></b>	EPE L37I
<b><u>Antibiotic Resistance:</u></b>	Ampicillin
<b><u>Description:</u></b>	Hydrophilic elastin-like repeats [(VPGVG) <sub>2</sub> VPGE(VPGVG) <sub>2</sub> ] <sub>3</sub> flanking a P coiled coil motif containing the L37I mutation.
<b><u>Construction:</u></b>	The P domain was isolated from pUC19 P L37I by digestion with <i>SacI</i> and <i>SpeI</i> and subcloned into pQE-80L EPE digested with the same enzymes.
<b><u>Available Sources:</u></b>	(1) miniprep DNA (-20°C) (2) 25 v/v % glycerol stock in DH10B (-80°C) (3) 25 v/v % glycerol stock in BL21 (-80°C)

**Plasmid Map:****Coding sequence:**

```

ATGAGATGCAGCAGCCATCATCATCATCACGTCGACGGCCACGGCGTGGGTGTT
CCGGGCGTCCGGTGTGCCGGGTGTGGGTGTGCCGGGCGAAGGTGTGCCGGGCGTCCG
TGTGCCGGGTGTTGGTGTTCGGGGCGTTGGTGTGCCGGGCGTTGGCGTGCCGGGCGA
GGGTGTGCCGGGCGTTGGTGTTCGGGGCGTGGGTGTGCCGGGCGTGGGCGTGCCGG
GCGTCCGGTGTTCGGGGCGAGGGTGTGCCGGGCGTAGGTGTGCCGGGCGTTGGTGAG
CTCGGATCAGGACTTGGATCCGCGCCGCAAATGCTGCGTGAAATTCAGGAAACCAA
TGCCGCGCTTCAGGATGTGCCGGGAATTGCTTCGTCAACAGGTCAAGGAGATAACGTT
CTTGAAGAACACCGTTCATGGAGTCGGATGCGTCCAAGCTTAATACTAGTGTGCCGGG
CGTCGGCGTGCCGGGCGTAGGTGTTCCGGGCGAGGGTGTTCGGGGCGTTGGTGTGCC
GGGCGTCGGCGTGCCGGGCGTGGGTGTTCCGGGCGTAGGTGTGCCGGGCGAGGGTG
TGCCGGGCGTGGGCGTGCCGGGCGTAGGTGTTCCGGGCGTAGGTGTTCCGGGCGTA
GGTGTTCGGGTGAAGGCGTGCCGGGCGTTGGTGTGCCGGGTGTGGGCGTGCCGGG
CGGGCTGCTCGAGTGCATGTAA

```

**Translation:**

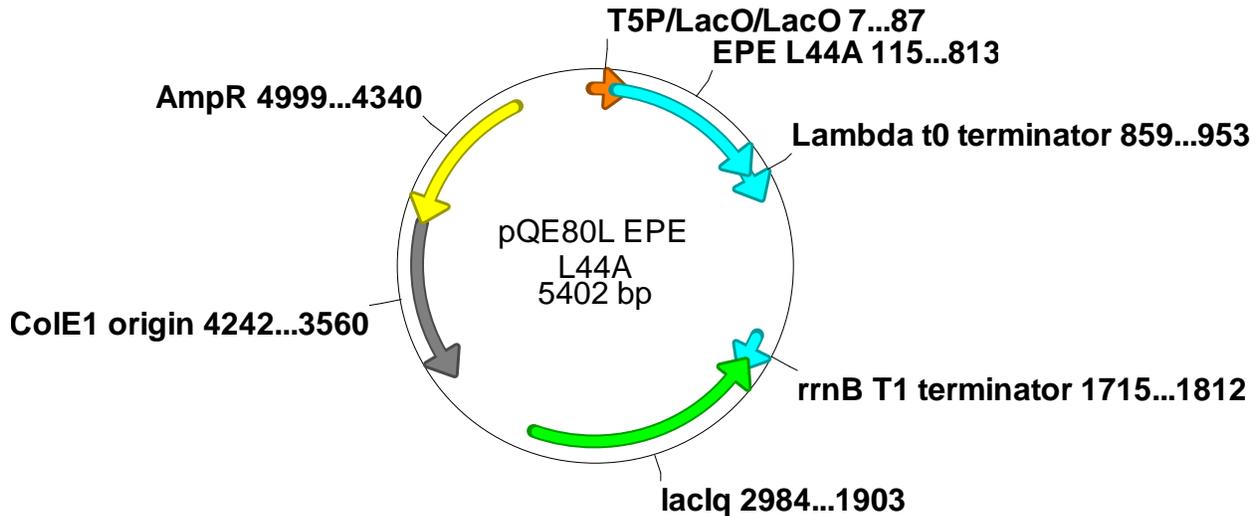
```

MRCSSHHHHHVDGHGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPG
EGVPGVGVPGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPGVGVPG
ALQDVRELLRQQVKEITFLKNTVMESDASKLNTSVPGVGVPGVGVPGEGVPGVGVPGV
GVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPGVGVPGVGVPGVGVPGV
ECM*

```

**pQE-80L EPE L44A**

<b><u>Submitted by:</u></b>	Larry Dooling
<b><u>Strain(s)/Plasmid:</u></b>	DH10B/pQE-80L EPE L44A BL21/pQE-80L EPE L44A
<b><u>Vector:</u></b>	pQE-80L
<b><u>Insert:</u></b>	EPE L44A
<b><u>Antibiotic Resistance:</u></b>	Ampicillin
<b><u>Description:</u></b>	Hydrophilic elastin-like repeats [(VPGVG) <sub>2</sub> VPGEG(VPGVG) <sub>2</sub> ] <sub>3</sub> flanking a P coiled coil motif containing the L44A mutation.
<b><u>Construction:</u></b>	The P domain was isolated from pUC19 P L44A by digestion with <i>SacI</i> and <i>SpeI</i> and subcloned into pQE-80L EPE digested with the same enzymes.
<b><u>Available Sources:</u></b>	(1) miniprep DNA (-20°C) (2) 25 v/v % glycerol stock in DH10B (-80°C) (3) 25 v/v % glycerol stock in BL21 (-80°C)

**Plasmid Map:****Coding sequence:**

```

ATGAGATGCAGCAGCCATCATCATCATCACGTCGACGGCCACGGCGTGGGTGTT
CCGGGCGTCCGGTGTGCCGGGTGTGGGTGTGCCGGGCGAAGGTGTGCCGGGCGTCCG
TGTGCCGGGTGTTGGTGTTCGGGGCGTTGGTGTGCCGGGCGTTGGCGTGCCGGGCGA
GGGTGTGCCGGGCGTTGGTGTTCGGGGCGTGGGTGTGCCGGGCGTGGGCGTGCCGG
GCGTCCGGTGTTCGGGGCGAGGGTGTGCCGGGCGTAGGTGTGCCGGGCGTTGGTGAG
CTCGGATCAGGACTTGGATCCGCGCCGCAAATGCTGCGTGAAGTGCAGGAAACCAA
TGCCGCGGCTCAGGATGTGCGGGAATTGCTTCGTCAACAGGTCAAGGAGATAACGT
TCTTGAAGAACACCGTCATGGAGTCGGATGCGTCCAAGCTTAATACTAGTGTGCCGG
GCGTCCGGCGTGCCGGGCGTAGGTGTTCCGGGGCGAGGGTGTTCGGGGCGTTGGTGTGC
CGGGCGTCCGGCGTGCCGGGCGTGGGTGTTCCGGGCGTAGGTGTGCCGGGCGAGGGT
GTGCCGGGCGTGGGCGTGCCGGGCGTAGGTGTTCCGGGCGTAGGTGTTCCGGGCGT
AGGTGTTCCGGGTGAAGGCGTGCCGGGCGTTGGTGTGCCGGGTGTGGGCGTGCCGG
GCGGGCTGCTCGAGTGCATGTAA

```

**Translation:**

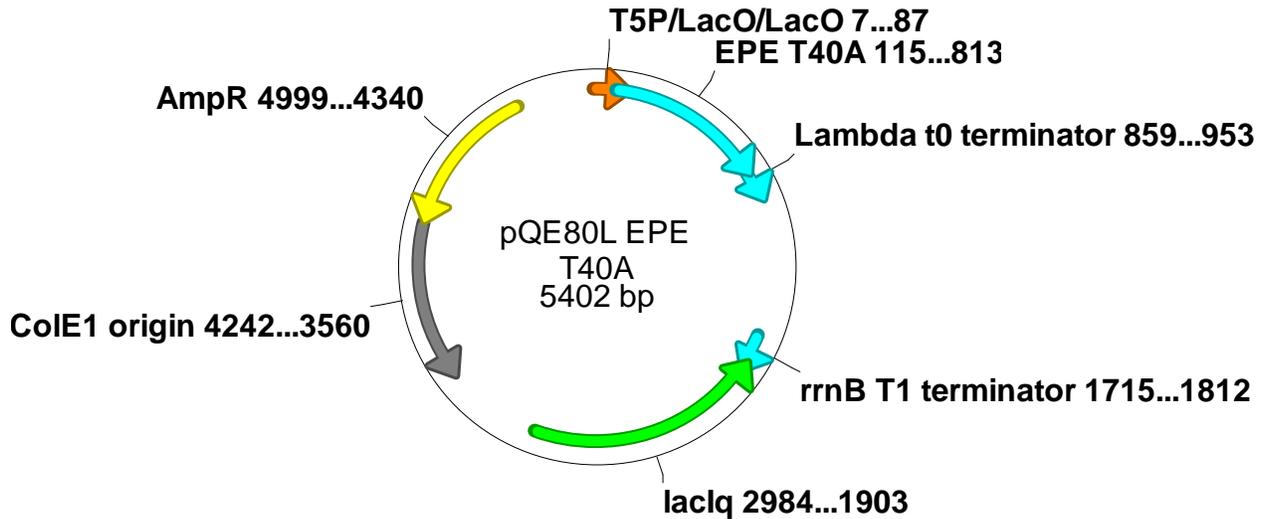
```

MRCSSHHHHHVDGHGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPG
EGVPGVGVPGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPGVGVPG
AAAQDVRELLRQQVKEITFLKNTVMESDASKLNTSVPGVGVPGVGVPGEGVPGVGVPG
VGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGG
LLECM*

```

**pQE-80L EPE T40A**

<b><u>Submitted by:</u></b>	Larry Dooling
<b><u>Strain(s)/Plasmid:</u></b>	DH10B/pQE-80L EPE T40A BL21/pQE-80L EPE T40A
<b><u>Vector:</u></b>	pQE-80L
<b><u>Insert:</u></b>	EPE T40A
<b><u>Antibiotic Resistance:</u></b>	Ampicillin
<b><u>Description:</u></b>	Hydrophilic elastin-like repeats [(VPGVG) <sub>2</sub> VPGEG(VPGVG) <sub>2</sub> ] <sub>3</sub> flanking a P coiled coil motif containing the T40A mutation.
<b><u>Construction:</u></b>	The P domain was isolated from pUC19 P T40A by digestion with <i>SacI</i> and <i>SpeI</i> and subcloned into pQE-80L EPE digested with the same enzymes.
<b><u>Available Sources:</u></b>	(1) miniprep DNA (-20°C) (2) 25 v/v % glycerol stock in DH10B (-80°C) (3) 25 v/v % glycerol stock in BL21 (-80°C)

**Plasmid Map:****Coding sequence:**

```

ATGAGATGCAGCAGCCATCATCATCATCACGTCGACGGCCACGGCGTGGGTGTT
CCGGGCGTCGGTGTGCCGGGTGTGGGTGTGCCGGGCGAAGGTGTGCCGGGCGTCCG
TGTGCCGGGTGTTGGTGTTCGGGGCGTTGGTGTGCCGGGCGTTGGCGTGCCGGGCGA
GGGTGTGCCGGGCGTTGGTGTTCGGGGCGTGGGTGTGCCGGGCGTGGGCGTGCCGG
GCGTCCGGTGTTCGGGGCGAGGGTGTGCCGGGCGTAGGTGTGCCGGGCGTTGGTGAG
CTCGGATCAGGACTTGGATCCGCGCCGCAAATGCTGCGTGAAGTGCAGGAAGCCAA
TGCCGCGCTTCAGGATGTGCCGGGAATTGCTTCGTCAACAGGTCAAGGAGATAACGTT
CTTGAAGAACACCGTCATGGAGTCGGATGCGTCCAAGCTTAATACTAGTGTGCCGGG
CGTCGGCGTGCCGGGCGTAGGTGTTCCGGGCGAGGGTGTTCGGGGCGTTGGTGTGCC
GGGCGTCGGCGTGCCGGGCGTGGGTGTTCCGGGCGTAGGTGTGCCGGGCGAGGGTG
TGCCGGGCGTGGGCGTGCCGGGCGTAGGTGTTCCGGGCGTAGGTGTTCCGGGCGTA
GGTGTTCGGGTGAAGGCGTGCCGGGCGTTGGTGTGCCGGGTGTGGGCGTGCCGGG
CGGGCTGCTCGAGTGCATGTAA

```

**Translation:**

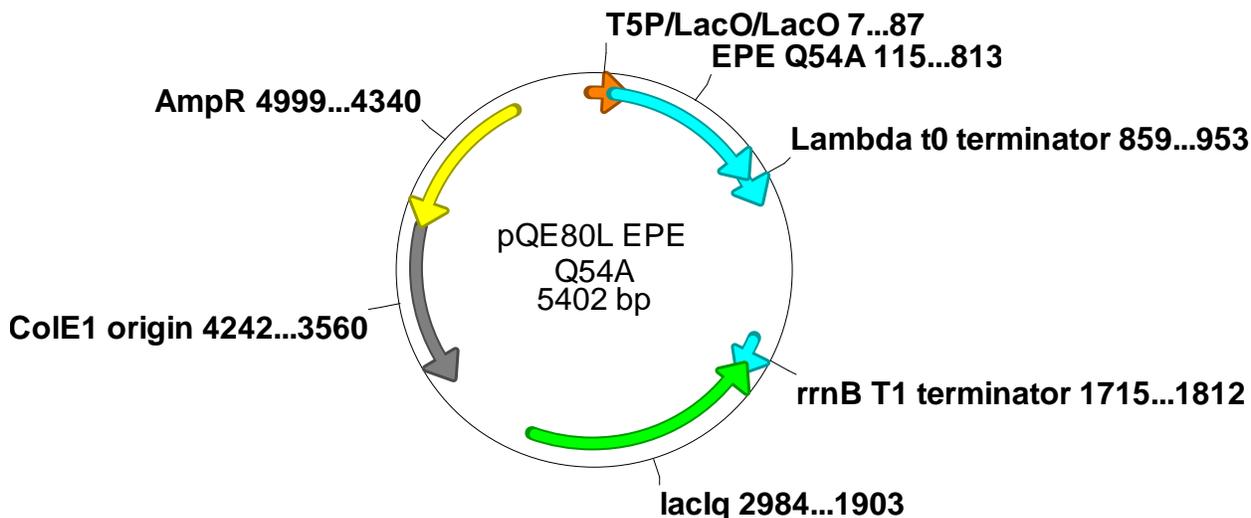
```

MRCSSHHHHHVDGHGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPG
EGVPGVGVPGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPGVGVPG
AALQDVRELLRQQVKEITFLKNTVMESDASKLNTSVPGVGVPGVGVPGEGVPGVGVPG
VGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGG
LLECM*

```

**pQE-80L EPE Q54A**

<b><u>Submitted by:</u></b>	Larry Dooling
<b><u>Strain(s)/Plasmid:</u></b>	DH10B/pQE-80L EPE Q54A BL21/pQE-80L EPE Q54A
<b><u>Vector:</u></b>	pQE-80L
<b><u>Insert:</u></b>	EPE Q54A
<b><u>Antibiotic Resistance:</u></b>	Ampicillin
<b><u>Description:</u></b>	Hydrophilic elastin-like repeats [(VPGVG) <sub>2</sub> VPGEG(VPGVG) <sub>2</sub> ] <sub>3</sub> flanking a P coiled coil motif containing the Q54A mutation.
<b><u>Construction:</u></b>	The P domain was isolated from pUC19 P Q54A by digestion with <i>SacI</i> and <i>SpeI</i> and subcloned into pQE-80L EPE digested with the same enzymes.
<b><u>Available Sources:</u></b>	(1) miniprep DNA (-20°C) (2) 25 v/v % glycerol stock in DH10B (-80°C) (3) 25 v/v % glycerol stock in BL21 (-80°C)

**Plasmid Map:****Coding sequence:**

```

ATGAGATGCAGCAGCCATCATCATCATCACGTCGACGGCCACGGCGTGGGTGTT
CCGGGCGTCCGGTGTGCCGGGTGTGGGTGTGCCGGGCGAAGGTGTGCCGGGCGTCCG
TGTGCCGGGTGTTGGTGTTCGGGGCGTTGGTGTGCCGGGCGTTGGCGTGCCGGGCGA
GGGTGTGCCGGGCGTTGGTGTTCGGGGCGTGGGTGTGCCGGGCGTGGGCGTGCCGG
GCGTCCGGTGTTCGGGGCGAGGGTGTGCCGGGCGTAGGTGTGCCGGGCGTTGGTGAG
CTCGGATCAGGACTTGGATCCGCGCCGCAAATGCTGCGTGAAGTGCAGGAAACCAA
TGCCGCGCTTCAGGATGTGCCGGGAATTGCTTCGTCAAGCGGTCAAGGAGATAACGTT
CTTGAAGAACACCGTCATGGAGTCGGATGCGTCCAAGCTTAATACTAGTGTGCCGGG
CGTCGGCGTGCCGGGCGTAGGTGTTCCGGGCGAGGGTGTTCGGGGCGTTGGTGTGCC
GGGCGTCGGCGTGCCGGGCGTGGGTGTTCCGGGCGTAGGTGTGCCGGGCGAGGGTG
TGCCGGGCGTGGGCGTGCCGGGCGTAGGTGTTCCGGGCGTAGGTGTTCCGGGCGTA
GGTGTTCGGGTGAAGGCGTGCCGGGCGTTGGTGTGCCGGGTGTGGGCGTGCCGGG
CGGGCTGCTCGAGTGCATGTAA

```

**Translation:**

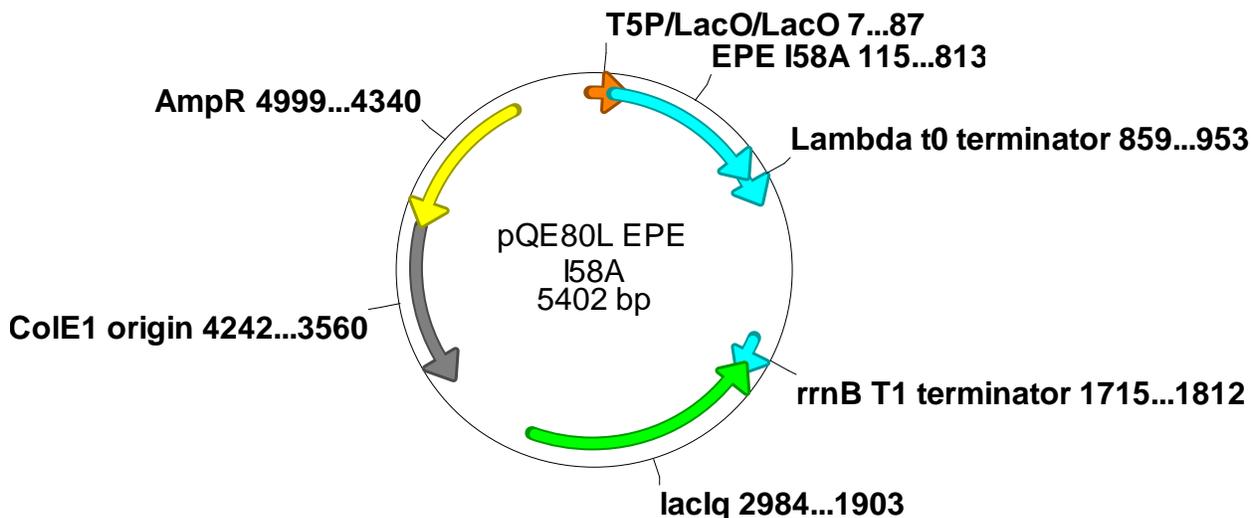
```

MRCSSHHHHHVDGHGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPG
EGVPGVGVPGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPGVGVPG
AALQDVRELLRQAVKEITFLKNTVMESDASKLNTSVPGVGVPGVGVPGEGVPGVGVPG
VGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGG
LLECM*

```

**pQE-80L EPE I58A**

<b><u>Submitted by:</u></b>	Larry Dooling
<b><u>Strain(s)/Plasmid:</u></b>	DH10B/pQE-80L EPE I58A BL21/pQE-80L EPE I58A
<b><u>Vector:</u></b>	pQE-80L
<b><u>Insert:</u></b>	EPE I58A
<b><u>Antibiotic Resistance:</u></b>	Ampicillin
<b><u>Description:</u></b>	Hydrophilic elastin-like repeats [(VPGVG) <sub>2</sub> VPGEG(VPGVG) <sub>2</sub> ] <sub>3</sub> flanking a P coiled coil motif containing the I58A mutation.
<b><u>Construction:</u></b>	The P domain was isolated from pUC19 P I58A by digestion with <i>SacI</i> and <i>SpeI</i> and subcloned into pQE-80L EPE digested with the same enzymes.
<b><u>Available Sources:</u></b>	(1) miniprep DNA (-20°C) (2) 25 v/v % glycerol stock in DH10B (-80°C) (3) 25 v/v % glycerol stock in BL21 (-80°C)

**Plasmid Map:****Coding sequence:**

```

ATGAGATGCAGCAGCCATCATCATCATCACGTCGACGGCCACGGCGTGGGTGTT
CCGGGCGTCCGGTGTGCCGGGTGTGGGTGTGCCGGGCGAAGGTGTGCCGGGCGTCCG
TGTGCCGGGTGTTGGTGTTCGGGGCGTTGGTGTGCCGGGCGTTGGCGTGCCGGGCGA
GGGTGTGCCGGGCGTTGGTGTTCGGGGCGTGGGTGTGCCGGGCGTGGGCGTGCCGG
GCGTCCGGTGTTCGGGGCGAGGGTGTGCCGGGCGTAGGTGTGCCGGGCGTTGGTGAG
CTCGGATCAGGACTTGGATCCCGCGCCGCAAATGCTGCGTGAAGTGCAGGAAACCAA
TGCCGCGCTTCAGGATGTGCCGGGAATTGCTTCGTCAACAGGTCAAGGAGGCAACGTT
CTTGAAGAACACCGTTCATGGAGTCGGATGCGTCCAAGCTTAATACTAGTGTGCCGGG
CGTCGGCGTGCCGGGCGTAGGTGTTCCGGGCGAGGGTGTTCGGGGCGTTGGTGTGCC
GGGCGTCGGCGTGCCGGGCGTGGGTGTTCCGGGCGTAGGTGTGCCGGGCGAGGGTG
TGCCGGGCGTGGGCGTGCCGGGCGTAGGTGTTCCGGGCGTAGGTGTTCCGGGCGTA
GGTGTTCGGGTGAAGGCGTGCCGGGCGTTGGTGTGCCGGGTGTGGGCGTGCCGGG
CGGGCTGCTCGAGTGCATGTAA

```

**Translation:**

```

MRCSSHHHHHVDGHGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPG
EGVPGVGVPGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPGVGVPG
AALQDVRELLRQQVKEATFLKNTVMESDASKLNTSVPGVGVPGVGVPGEGVPGVGV
GVGVPGVGVPGVGVPGEGVPGVGVPGVGVPGVGVPGVGVPGEGVPGVGVPGVGVPG
GLLECM*

```