

Chapter 1

Introduction

Introduction

Melatonin and adenosine are two small molecules that have been reported to have an important effect on sleep wake behavior. This study examines how these two molecules individually contribute to the regulation of sleep-wake behavior and proposes a model by which one regulates the other to bring about the sleep/wake states. In order to study their role on sleep, a scientist's definition of sleep is first required. Sleep can be defined as a rapidly reversible state of behavioral quiescence which must have the following features to be called sleep: (1) consolidated periods of immobility exhibiting circadian rhythmicity (2) a specific posture, specific to individual species (3) a decrease in sensory responsiveness/ or an increased arousal threshold (4) a homeostatic regulatory mechanism or a mechanism by which deprivation of the behavior results in an accompanying rebound once the deprivation stimulus has been removed (Campbell and Tobler, 1984; Hendricks et al., 2000).

Using these criteria, sleep has been observed in a variety of the members of the animal kingdom including humans, zebrafish and fruit flies (Campbell and Tobler, 1984; Hendricks et al., 2000). Sleep deprivation assays have shown that prolonged deprivation of sleep results in deterioration of performance in memory related tasks. The absence of sleep can result in deterioration in performance and eventually prolonged deprivation can result in death (Everson et al., 1989; Li et al., 2009; Seugnet et al., 2008; Shaw et al., 2002). These results have lead scientists to theorize that during sleep episodes maintenance processes must be occurring in the brain, which are crucial to brain performance. One theory suggests that the purpose of sleep is to replenish depleted brain energy stores such as glycogen (Benington and Craig Heller, 1995). This theory places

low the low energy molecule adenosine in a central role, as a readout of the brain energy levels, and a regulator of sleep wake circuits (Benington et al., 1995). Another theory suggests that the purpose of sleep may be to trim the majority of the connections formed in the brain over the course of the day in order to be able to absorb new information the next day, and to also strengthen specific important connections, which has been supported by evidence from *drosophila* (Tononi and Cirelli, 2003) . In either case, regulating the occurrence of sleep then can be considered to be very important for animals.

What is the neural circuitry that regulates sleep? Studies show that the hypothalamic circuitry plays a major role in the control of sleep-wake behavior. There are two neural systems in the brain that control sleep-wake behavior. The first is the ascending arousal system situated in the brainstem/hypothalamus region. It includes a group of nuclei such as the locus coeruleus (Noradrenergic), Dorsal Raphae nucleus (Serotonergic), laterodorsal tegmental nucleus (Cholinergic), Pedunculopontine nucleus (cholinergic) and Tuberomamillary nucleus (Histaminergic). The ascending arousal system also includes the orexinergic and melanin concentrating hormone neurons in the lateral hypothalamus, the cholinergic neurons in the basal forebrain and the dopaminergic neurons in the ventral periaqueductal gray region. The second system is the sleep active neurons in the ventero-lateral pre optic area (VLPO). The projections from these neurons inhibit the wake active areas, according to the flip-flop model of sleep regulation (Saper et al., 2001, 2005). At any one time, either the ascending arousal system is active or the sleep system is active since they inhibit each others, which results in sleep state and the wake state being mutually exclusive. The molecules that bring about this flip are unclear

however Adenosine has been proposed to be a candidate. (Basheer et al., 2000; Radulovacki et al., 1984; Saper et al., 2005).

What then controls the timing of the two states? According to the two-process model proposed by (Borbély, 1982), sleep is controlled by two processes, process C and process S. Process C is the circadian process, whose levels vary under the control of the circadian clock. It is regulated by external factors such as light. Process S is the homeostatic process, which responds to internal cues of sleep need. Its levels rise with wakefulness and decrease with sleep. While the molecular workings of process C have been well characterized, its output molecule was unknown. In contrast, for process S, there have been several candidates for the output molecule, including Adenosine, AMP, NOS (Brown et al., 2012), however where in the brain they are produced and how their release is regulated is quite unclear. Melatonin however is a good candidate for the process C output molecule. Its levels are under the control of the circadian clock in humans, rodents as well as in zebrafish (Klein, 2007). Also exogenous melatonin induces sleep in vertebrates (Cajochen et al., 1997, 2003; Zhdanova et al., 1996).

We decided to use zebrafish to study the role played by melatonin and adenosine in sleep wake behavior. Why use zebrafish? Zebrafish is a comparatively new model system developed in the 1970s. Zebrafish can be housed compactly; moreover they produce many hundreds of embryos at a time. They are amenable to genetic manipulations and they are transparent, making them ideal for imaging. In addition to all these characteristics, they also exhibit the hallmarks of sleep behavior by the age of 4dpf (Prober et al., 2006; Zhdanova, 2006). Zebrafish are vertebrates, but they possess a simpler nervous system than mammals. They possess the components of the sleep wake

circuitry in higher vertebrates (Chiu and Prober, 2013), but they have lesser numbers of neurons, for example only 10-12 orexin neurons as compared to the over 100 neurons in rats (Liu et al., 2015; Prober et al., 2006). They also exhibit similar responses to agonists and antagonists as higher vertebrates (Rihel et al., 2010). The zebrafish blood brain barrier is formed during 3-10 days post fertilization. There is therefore, good brain penetrance of sleep-wake affecting drugs added directly to the fish-water, during this period (Fleming et al., 2013). The small size of zebrafish larvae allows for high-throughput behavioral assays to be performed, which allows for statistically solid behavioral experiments (Kokel and Peterson, 2008; Rihel et al., 2010). Moreover they have a fully functional circadian system at 5dpf (Hirayama et al., 2005; Kazimi and Cahill, 1999). Also zebrafish are amenable to the ZFN, TALEN and CRISPR technologies, making it possible to generate desired gene mutants (Chen et al., 2013; Hwang et al., 2013; Sander et al., 2011a, 2011b). All these advantages result in the zebrafish model system being well poised to answer many questions relating to sleep-wake regulation.

Bibliography

- Alam, M.N., Szymusiak, R., Gong, H., King, J., and McGinty, D. (1999). Adenosinergic modulation of rat basal forebrain neurons during sleep and waking: neuronal recording with microdialysis. *J. Physiol.* *521*, 679–690.
- Alam, M.N., Kumar, S., Rai, S., Methippara, M., Szymusiak, R., and McGinty, D. (2009). Role of adenosine A1 receptor in the perifornical–lateral hypothalamic area in sleep–wake regulation in rats. *Brain Res.* *1304*, 96–104.
- Barbhaiya, H., McClain, R., Ijzerman, A., and Rivkees, S.A. (1996). Site-directed mutagenesis of the human A1 adenosine receptor: influences of acidic and hydroxy residues in the first four transmembrane domains on ligand binding. *Mol. Pharmacol.* *50*, 1635–1642.
- Basheer, R., Porkka-Heiskanen, T., Strecker, R.E., Thakkar, M.M., and McCarley, R.W. (2000). Adenosine as a biological signal mediating sleepiness following prolonged wakefulness. *Biol. Signals Recept.* *9*, 319–327.
- Benington, J.H., and Craig Heller, H. (1995). Restoration of brain energy metabolism as the function of sleep. *Prog. Neurobiol.* *45*, 347–360.
- Benington, J.H., Kodali, S.K., and Heller, H.C. (1995). Stimulation of A1 adenosine receptors mimics the electroencephalographic effects of sleep deprivation. *Brain Res.* *692*, 79–85.
- Blanco-Centurion, C., Xu, M., Murillo-Rodriguez, E., Gerashchenko, D., Shiromani, A.M., Salin-Pascual, R.J., Hof, P.R., and Shiromani, P.J. (2006). Adenosine and Sleep Homeostasis in the Basal Forebrain. *J. Neurosci.* *26*, 8092–8100.
- Boehmler, W., Petko, J., Woll, M., Frey, C., Thisse, B., Thisse, C., Canfield, V.A., and Levenson, R. (2009). Identification of zebrafish A2 adenosine receptors and expression in developing embryos. *Gene Expr. Patterns* *9*, 144–151.
- Borbély, A.A. (1982). A two process model of sleep regulation. *Hum. Neurobiol.* *1*, 195–204.
- Brown, R.E., Basheer, R., McKenna, J.T., Strecker, R.E., and McCarley, R.W. (2012). Control of Sleep and Wakefulness. *Physiol. Rev.* *92*, 1087–1187.
- Cajochen, C., Kräuchi, K., and Wirz-Justice, A. (1997). The acute soporific action of daytime melatonin administration: effects on the EEG during wakefulness and subjective alertness. *J. Biol. Rhythms* *12*, 636–643.
- Cajochen, C., Kräuchi, K., and Wirz-Justice, A. (2003). Role of Melatonin in the Regulation of Human Circadian Rhythms and Sleep. *J. Neuroendocrinol.* *15*, 432–437.

Van Calker, D., Müller, M., and Hamprecht, B. (1979). Adenosine regulates via two different types of receptors, the accumulation of cyclic AMP in cultured brain cells. *J. Neurochem.* *33*, 999–1005.

Campbell, S.S., and Tobler, I. (1984). Animal sleep: A review of sleep duration across phylogeny. *Neurosci. Biobehav. Rev.* *8*, 269–300.

Cermak, T., Doyle, E.L., Christian, M., Wang, L., Zhang, Y., Schmidt, C., Baller, J.A., Somia, N.V., Bogdanove, A.J., and Voytas, D.F. (2011). Efficient design and assembly of custom TALEN and other TAL effector-based constructs for DNA targeting. *Nucleic Acids Res.* *39*, e82–e82.

Chen, S., Oikonomou, G., Chiu, C.N., Niles, B.J., Liu, J., Lee, D.A., Antoshechkin, I., and Prober, D.A. (2013). A large-scale in vivo analysis reveals that TALENs are significantly more mutagenic than ZFNs generated using context-dependent assembly. *Nucleic Acids Res.* *41*, 2769–2778.

Chiu, C.N., and Prober, D.A. (2013). Regulation of zebrafish sleep and arousal states: current and prospective approaches. *Front. Neural Circuits* *7*.

Choi, H.M.T., Beck, V.A., and Pierce, N.A. (2014). Next-generation in situ hybridization chain reaction: higher gain, lower cost, greater durability. *ACS Nano* *8*, 4284–4294.

Dixon, A.K., Gubitza, A.K., Sirinathsinghji, D.J., Richardson, P.J., and Freeman, T.C. (1996). Tissue distribution of adenosine receptor mRNAs in the rat. *Br. J. Pharmacol.* *118*, 1461–1468.

El Yacoubi, M., Ledent, C., Parmentier, M., Costentin, J., and Vaugeois, J.-M. (2000). SCH 58261 and ZM 241385 differentially prevent the motor effects of CGS 21680 in mice: evidence for a functional “atypical” adenosine A_{2A} receptor. *Eur. J. Pharmacol.* *401*, 63–77.

Everson, C.A., Bergmann, B.M., and Rechtschaffen, A. (1989). Sleep deprivation in the rat: III. Total sleep deprivation. *Sleep* *12*, 13–21.

Fang, J., Payne, L., and Krueger, J.M. (1995). Pituitary adenylate cyclase activating polypeptide enhances rapid eye movement sleep in rats. *Brain Res.* *686*, 23–28.

Fleming, A., Diekmann, H., and Goldsmith, P. (2013). Functional characterisation of the maturation of the blood-brain barrier in larval zebrafish. *PloS One* *8*, e77548.

Force, A., Lynch, M., Pickett, F.B., Amores, A., Yan, Y., and Postlethwait, J. (1999). Preservation of Duplicate Genes by Complementary, Degenerative Mutations. *Genetics* *151*, 1531–1545.

Fredholm, B.B., IJzerman, A.P., Jacobson, K.A., Klotz, K.-N., and Linden, J. (2001). International Union of Pharmacology. XXV. Nomenclature and Classification of Adenosine Receptors. *Pharmacol. Rev.* *53*, 527–552.

- Freissmuth, M., Schütz, W., and Linder, M.E. (1991). Interactions of the bovine brain A1-adenosine receptor with recombinant G protein alpha-subunits. Selectivity for rGi alpha-3. *J. Biol. Chem.* *266*, 17778–17783.
- Gallopin, T., Luppi, P.-H., Cauli, B., Urade, Y., Rossier, J., Hayaishi, O., Lambolez, B., and Fort, P. (2005). The endogenous somnogen adenosine excites a subset of sleep-promoting neurons via A2A receptors in the ventrolateral preoptic nucleus. *Neuroscience* *134*, 1377–1390.
- Halldner, L., Lopes, L.V., Daré, E., Lindström, K., Johansson, B., Ledent, C., Cunha, R.A., and Fredholm, B.B. (2004). Binding of adenosine receptor ligands to brain of adenosine receptor knock-out mice: evidence that CGS 21680 binds to A1 receptors in hippocampus. *Naunyn. Schmiedebergs Arch. Pharmacol.* *370*, 270–278.
- Hendricks, J.C., Finn, S.M., Panckeri, K.A., Chavkin, J., Williams, J.A., Sehgal, A., and Pack, A.I. (2000). Rest in *Drosophila* Is a Sleep-like State. *Neuron* *25*, 129–138.
- Higashijima, S.-I., Mandel, G., and Fetcho, J.R. (2004). Distribution of prospective glutamatergic, glycinergic, and GABAergic neurons in embryonic and larval zebrafish. *J. Comp. Neurol.* *480*, 1–18.
- Hirayama, J., Kaneko, M., Cardone, L., Cahill, G., and Sassone-Corsi, P. (2005). Analysis of Circadian Rhythms in Zebrafish. In *Methods in Enzymology*, M.W. Young, ed. (Academic Press), pp. 186–204.
- Huang, Z.-L., Qu, W.-M., Eguchi, N., Chen, J.-F., Schwarzschild, M.A., Fredholm, B.B., Urade, Y., and Hayaishi, O. (2005). Adenosine A2A, but not A1, receptors mediate the arousal effect of caffeine. *Nat. Neurosci.* *8*, 858–859.
- Hwang, W.Y., Fu, Y., Reyon, D., Maeder, M.L., Tsai, S.Q., Sander, J.D., Peterson, R.T., Yeh, J.-R.J., and Joung, J.K. (2013). Efficient genome editing in zebrafish using a CRISPR-Cas system. *Nat. Biotechnol.* *31*, 227–229.
- Jacobson, K.A., and Gao, Z.-G. (2006). Adenosine receptors as therapeutic targets. *Nat. Rev. Drug Discov.* *5*, 247–264.
- Kazimi, N., and Cahill, G.M. (1999). Development of a circadian melatonin rhythm in embryonic zebrafish. *Dev. Brain Res.* *117*, 47–52.
- Klein, D.C. (2007). Arylalkylamine N-Acetyltransferase: “the Timezyme.” *J. Biol. Chem.* *282*, 4233–4237.
- Kokel, D., and Peterson, R.T. (2008). Chemobehavioural phenomics and behaviour-based psychiatric drug discovery in the zebrafish. *Brief. Funct. Genomic. Proteomic.* *7*, 483–490.
- Kumar, S., Rai, S., Hsieh, K.-C., McGinty, D., Alam, M.N., and Szymusiak, R. (2013). Adenosine A2A receptors regulate the activity of sleep regulatory GABAergic neurons in

the preoptic hypothalamus. *Am. J. Physiol. - Regul. Integr. Comp. Physiol.* *305*, R31–R41.

Li, X., Yu, F., and Guo, A. (2009). Sleep Deprivation Specifically Impairs Short-term Olfactory Memory in *Drosophila*. *Sleep* *32*, 1417–1424.

Liu, J., Merkle, F.T., Gandhi, A.V., Gagnon, J.A., Woods, I.G., Chiu, C.N., Shimogori, T., Schier, A.F., and Prober, D.A. (2015). Evolutionarily conserved regulation of hypocretin neuron specification by *Lhx9*. *Dev. Camb. Engl.* *142*, 1113–1124.

Liu, W., Guo, F., Lu, B., and Guo, A. (2008). amnesiac regulates sleep onset and maintenance in *Drosophila melanogaster*. *Biochem. Biophys. Res. Commun.* *372*, 798–803.

Lohse, M.J., Klotz, K.N., Lindenborn-Fotinos, J., Reddington, M., Schwabe, U., and Olsson, R.A. (1987). 8-Cyclopentyl-1,3-dipropylxanthine (DPCPX)--a selective high affinity antagonist radioligand for A1 adenosine receptors. *Naunyn. Schmiedebergs Arch. Pharmacol.* *336*, 204–210.

Martin, J.V., Berman, K.F., Skolnick, P., and Mendelson, W.B. (1989). Behavioral and electroencephalographic effects of the adenosine1 agonist, L-PIA. *Pharmacol. Biochem. Behav.* *34*, 507–510.

Mertens, I., Husson, S.J., Janssen, T., Lindemans, M., and Schoofs, L. (2007). PACAP and PDF signaling in the regulation of mammalian and insect circadian rhythms. *Peptides* *28*, 1775–1783.

Methippara, M.M., Kumar, S., Alam, M.N., Szymusiak, R., and McGinty, D. (2005). Effects on sleep of microdialysis of adenosine A1 and A2a receptor analogs into the lateral preoptic area of rats. *Am. J. Physiol. - Regul. Integr. Comp. Physiol.* *289*, R1715–R1723.

Morgan, J.I., and Curran, T. (1991). Stimulus-Transcription Coupling in the Nervous System: Involvement of the Inducible Proto-Oncogenes *fos* and *jun*. *Annu. Rev. Neurosci.* *14*, 421–451.

Müller, C.E., and Jacobson, K.A. (2011). Recent developments in adenosine receptor ligands and their potential as novel drugs. *Biochim. Biophys. Acta* *1808*, 1290–1308.

Oishi, Y., Huang, Z.-L., Fredholm, B.B., Urade, Y., and Hayaishi, O. (2008). Adenosine in the tuberomammillary nucleus inhibits the histaminergic system via A1 receptors and promotes non-rapid eye movement sleep. *Proc. Natl. Acad. Sci.* *105*, 19992–19997.

Olah, M.E., and Stiles, G.L. (1992). Adenosine Receptors. *Annu. Rev. Physiol.* *54*, 211–225.

Olah, M.E., and Stiles, G.L. (2000). The role of receptor structure in determining adenosine receptor activity. *Pharmacol. Ther.* *85*, 55–75.

Palmer, T.M., and Stiles, G.L. (1997). Structure-function analysis of inhibitory adenosine receptor regulation. *Neuropharmacology* 36, 1141–1147.

Porkka-Heiskanen, T., Strecker, R.E., Thakkar, M., Bjørkum, A.A., Greene, R.W., and McCarley, R.W. (1997). Adenosine: A Mediator of the Sleep-Inducing Effects of Prolonged Wakefulness. *Science* 276, 1265–1268.

Porkka-Heiskanen, T., Strecker, R.E., and McCarley, R.W. (2000). Brain site-specificity of extracellular adenosine concentration changes during sleep deprivation and spontaneous sleep: an in vivo microdialysis study. *Neuroscience* 99, 507–517.

Postlethwait, J.H., Yan, Y.-L., Gates, M.A., Horne, S., Amores, A., Brownlie, A., Donovan, A., Egan, E.S., Force, A., Gong, Z., et al. (1998). Vertebrate genome evolution and the zebrafish gene map. *Nat. Genet.* 18, 345–349.

Prober, D.A., Rihel, J., Onah, A.A., Sung, R.-J., and Schier, A.F. (2006). Hypocretin/Orexin Overexpression Induces An Insomnia-Like Phenotype in Zebrafish. *J. Neurosci.* 26, 13400–13410.

Radulovacki, M., Virus, R.M., Djuricic-Nedelson, M., and Green, R.D. (1984). Adenosine analogs and sleep in rats. *J. Pharmacol. Exp. Ther.* 228, 268–274.

Reppert, S.M., Weaver, D.R., Stehle, J.H., and Rivkees, S.A. (1991). Molecular Cloning and Characterization of a Rat A1-Adenosine Receptor that is Widely Expressed in Brain and Spinal Cord. *Mol. Endocrinol.* 5, 1037–1048.

Reyon, D., Tsai, S.Q., Khayter, C., Foden, J.A., Sander, J.D., and Joung, J.K. (2012). FLASH assembly of TALENs for high-throughput genome editing. *Nat. Biotechnol.* 30, 460–465.

Rihel, J., Prober, D.A., Arvanites, A., Lam, K., Zimmerman, S., Jang, S., Haggarty, S.J., Kokel, D., Rubin, L.L., Peterson, R.T., et al. (2010). Zebrafish Behavioral Profiling Links Drugs to Biological Targets and Rest/Wake Regulation. *Science* 327, 348–351.

Rivkees, S.A. (1995). The ontogeny of cardiac and neural A1 adenosine receptor expression in rats. *Dev. Brain Res.* 89, 202–213.

Sander, J.D., Maeder, M.L., Reyon, D., Voytas, D.F., Joung, J.K., and Dobbs, D. (2010). ZiFiT (Zinc Finger Targeter): an updated zinc finger engineering tool. *Nucleic Acids Res.* 38, W462–W468.

Sander, J.D., Cade, L., Khayter, C., Reyon, D., Peterson, R.T., Joung, J.K., and Yeh, J.-R.J. (2011a). Targeted gene disruption in somatic zebrafish cells using engineered TALENs. *Nat. Biotechnol.* 29, 697–698.

Sander, J.D., Yeh, J.-R.J., Peterson, R.T., and Joung, J.K. (2011b). Chapter 3 - Engineering Zinc Finger Nucleases for Targeted Mutagenesis of Zebrafish. In *Methods in Cell Biology*, M.W. and L.I.Z. H. William Detrich, ed. (Academic Press), pp. 51–58.

- Saper, C.B., Chou, T.C., and Scammell, T.E. (2001). The sleep switch: hypothalamic control of sleep and wakefulness. *Trends Neurosci.* *24*, 726–731.
- Saper, C.B., Scammell, T.E., and Lu, J. (2005). Hypothalamic regulation of sleep and circadian rhythms. *Nature* *437*, 1257–1263.
- Satoh, S., Matsumura, H., Suzuki, F., and Hayaishi, O. (1996). Promotion of sleep mediated by the A2a-adenosine receptor and possible involvement of this receptor in the sleep induced by prostaglandin D2 in rats. *Proc. Natl. Acad. Sci. U. S. A.* *93*, 5980–5984.
- Satoh, S., Matsumura, H., and Hayaishi, O. (1998). Involvement of adenosine A2A receptor in sleep promotion. *Eur. J. Pharmacol.* *351*, 155–162.
- Satoh, S., Matsumura, H., Kanbayashi, T., Yoshida, Y., Urakami, T., Nakajima, T., Kimura, N., Nishino, S., and Yoneda, H. (2006). Expression pattern of FOS in orexin neurons during sleep induced by an adenosine A2A receptor agonist. *Behav. Brain Res.* *170*, 277–286.
- Scammell, T.E., Gerashchenko, D.Y., Mochizuki, T., McCarthy, M.T., Estabrooke, I.V., Sears, C.A., Saper, C.B., Urade, Y., and Hayaishi, O. (2001). An adenosine A2a agonist increases sleep and induces Fos in ventrolateral preoptic neurons. *Neuroscience* *107*, 653–663.
- Seugnet, L., Suzuki, Y., Vine, L., Gottschalk, L., and Shaw, P.J. (2008). D1 receptor activation in the mushroom bodies rescues sleep-loss-induced learning impairments in *Drosophila*. *Curr. Biol. CB* *18*, 1110–1117.
- Shaw, P.J., Cirelli, C., Greenspan, R.J., and Tononi, G. (2000). Correlates of Sleep and Waking in *Drosophila melanogaster*. *Science* *287*, 1834–1837.
- Shaw, P.J., Tononi, G., Greenspan, R.J., and Robinson, D.F. (2002). Stress response genes protect against lethal effects of sleep deprivation in *Drosophila*. *Nature* *417*, 287–291.
- Stenberg, D., Litonius, E., Halldner, L., Johansson, B., Fredholm, B.B., and Porkka-Heiskanen, T. (2003). Sleep and its homeostatic regulation in mice lacking the adenosine A1 receptor. *J. Sleep Res.* *12*, 283–290.
- Thakkar, M.M., Winston, S., and McCarley, R.W. (2003). A1 Receptor and Adenosinergic Homeostatic Regulation of Sleep-Wakefulness: Effects of Antisense to the A1 Receptor in the Cholinergic Basal Forebrain. *J. Neurosci.* *23*, 4278–4287.
- Thakkar, M.M., Engemann, S.C., Sharma, R., and Sahota, P. (2010). Role of Wake-Promoting Basal Forebrain and Adenosinergic Mechanisms in Sleep-Promoting Effects of Ethanol. *Alcohol. Clin. Exp. Res.* *34*, 997–1005.
- Thisse, C., and Thisse, B. (2008). High-resolution in situ hybridization to whole-mount zebrafish embryos. *Nat. Protoc.* *3*, 59–69.

- Ticho, S.R., and Radulovacki, M. (1991). Role of adenosine in sleep and temperature regulation in the preoptic area of rats. *Pharmacol. Biochem. Behav.* *40*, 33–40.
- Tononi, G., and Cirelli, C. (2003). Sleep and synaptic homeostasis: a hypothesis. *Brain Res. Bull.* *62*, 143–150.
- Ukena, D., Schudt, C., and Sybrecht, G.W. (1993). Adenosine receptor-blocking xanthines as inhibitors of phosphodiesterase isozymes. *Biochem. Pharmacol.* *45*, 847–851.
- Urade, Y., Eguchi, N., Qu, W.-M., Sakata, M., Huang, Z.-L., Chen, J.-F., Schwarzschild, M.A., Fink, J.S., and Hayaishi, O. (2003). Sleep regulation in adenosine A2A receptor-deficient mice. *Neurology* *61*, S94–S96.
- Urnov, F.D., Rebar, E.J., Holmes, M.C., Zhang, H.S., and Gregory, P.D. (2010). Genome editing with engineered zinc finger nucleases. *Nat. Rev. Genet.* *11*, 636–646.
- Vaudry, D., Falluel-Morel, A., Bourgault, S., Basille, M., Burel, D., Wurtz, O., Fournier, A., Chow, B.K.C., Hashimoto, H., Galas, L., et al. (2009). Pituitary adenylate cyclase-activating polypeptide and its receptors: 20 years after the discovery. *Pharmacol. Rev.* *61*, 283–357.
- Weber, R.G., Jones, C.R., Lohse, M.J., and Palacios, J.M. (1990). Autoradiographic visualization of A1 adenosine receptors in rat brain with [3H]8-cyclopentyl-1,3-dipropylxanthine. *J. Neurochem.* *54*, 1344–1353.
- Wen, L., Wei, W., Gu, W., Huang, P., Ren, X., Zhang, Z., Zhu, Z., Lin, S., and Zhang, B. (2008). Visualization of monoaminergic neurons and neurotoxicity of MPTP in live transgenic zebrafish. *Dev. Biol.* *314*, 84–92.
- Woods, I.G., Kelly, P.D., Chu, F., Ngo-Hazelett, P., Yan, Y.-L., Huang, H., Postlethwait, J.H., and Talbot, W.S. (2000). A Comparative Map of the Zebrafish Genome. *Genome Res.* *10*, 1903–1914.
- Zhdanova, I.V. (2006). Sleep in Zebrafish. *Zebrafish* *3*, 215–226.
- Zhdanova, I.V., Wurtman, R.J., Morabito, C., Piotrovskaya, V.R., and Lynch, H.J. (1996). Effects of low oral doses of melatonin, given 2-4 hours before habitual bedtime, on sleep in normal young humans. *Sleep* *19*, 423–431.
- Zhdanova, I.V., Wang, S.Y., Leclair, O.U., and Danilova, N.P. (2001). Melatonin promotes sleep-like state in zebrafish. *Brain Res.* *903*, 263–268.