CHAPTER 4

Enantio-, Diastereo- and Regioselective Iridium-Catalyzed Asymmetric Allylic Alkylation of Acyclic β -Ketoesters¹

4.1 INTRODUCTION

4.1.1 State of the art in the asymmetric construction of vicinal quaternary and tertiary carbon centers

As outlined in Chapter 3 (vide supra), the generation of enantioenriched allcarbon quaternary centers is complicated by the presence of vicinal tertiary stereocenters due to increased steric demands and the introduction of requisite diastereocontrol. Modern strategies for accessing these highly-congested stereochemical dyads have relied primarily on transition-metal catalysis, notably Pd-catalyzed enolate alkylation cascades,³⁰ Pd-catalyzed trimethylenemethane cycloadditions,^{73b,e} Cu-catalyzed asymmetric Claisen rearrangements,^{73f} and Mo-^{73c,d} and Ir-catalyzed⁸³ allylic alkylations. Common to the majority of these reports is the constraint that the nascent quaternary center be formed at a cyclic nucleophile. At the outset of our studies in this area, only two groups had reported success in employing linear nucleophiles to produce vicinal

¹ This work was performed in collaboration with Wen-Bo Lui, postdoctoral researcher in the Stoltz group. This work has been published. See: Liu, W. -B.; Reeves, C. M.; Stoltz, B. M. J. Am. Chem. Soc. **2013**, *135*, 17298.

quaternary/tertiary arrays. Namely, Trost's communication on the molybdenumcatalyzed allylic alkylation of β -cyanoesters^{73d} and Carreira's recent report on the allylic alkylation of aldehydes using stereodivergent dual catalysis.⁸³ To address these limitations, we initiated studies investigating the asymmetric allylic alkylation of linear β -ketoesters.

As described above, we have shown iridium-*N*-arylphosphoramidite catalysis⁸⁷ to be a powerful tool for accessing vicinal all-carbon quaternary and tertiary stereocenters. The success of our protocol for the regio-, diastereo- and enantioselective asymmetric allylic alkylation of cyclic β -ketoesters (Figure 4.1.1.1A)^{82, 88, 97} combined with the virtual absence of reports describing the application of this transformation to acyclic β ketoesters encouraged our further exploration of iridium catalysts in the domain of this important substrate class. In this chapter, we detail the development of the first highly regio-, diastereo- and enantioselective allylic alkylation of acyclic β -ketoesters to forge vicinal tertiary, quaternary centers (Figure 4.1.1.1B).

Figure 4.1.1.1. Representative Ir-catalyzed asymmetric allylic alkylation



4.2 DEVELOPMENT AND OPTIMIZATION OF AN IRIDIUM-CATALYZED ALLYLIC ALKYLATION OF LINEAR β -KETOESTERS

Our initial investigations in the domain of Ir-catalyzed allylic alkylation of linear β-ketoesters focused on identifying conditions that would afford both reaction efficiency and selectivity. We chose ethyl 2-methyl-3-oxo-3-phenylpropanoate (111a) and cinnamyl carbonate (98a) as standard coupling partners, and investigated several iridacycle complexes⁸⁴ at the outset of our studies. The result of these studies are shown in Table 4.2.1 (entries 1–6). We found that exposure of standard coupling components (111a) and (98a) to a combination of catalytic phosphoramidite ligand $L10 \cdot [Ir(cod)Cl]_2$ complex⁸⁶ and two equivalents of NaH in THF at ambient temperature afforded the desired product with good conversion, ee and regioselectivity, but low levels of diastereoselectivity (1:2) (entry 1). Use of either L11 or L16 under these conditions favored instead the reaction pathway yielding the undesired, linear allylic alkylation product (113a) in only modest conversion (entries 2 and 4). Ligands L13 and L17^{87,98} gave the desired branched product in good conversion but with diminished diastereoselectivity and enantioselectivity and protracted reaction times (entries 5 and 6). We were pleased to find that tetrahydroquinoline based ligand L12⁸⁷ rapidly furnished the desired α -quaternary β -ketoester (112a) in greater than 95% conversion, 95:5 regioselectivity, 13:1 dr and 99% enantiomeric excess (entry 3). Previous reports demonstrating the marked effect of metal cations over regio-99 and diastereoselectivity¹⁰⁰ in iridium-catalyzed allylic alkylations prompted further investigation of both bases and additives (entries 7–15). Contrary to our previous findings (vide infra, Chapter 3), a sluggish reaction was observed when LiBr was used in place of NaH (entry 7), presumably due to the decreased α -acidity of acyclic β -ketoesters relative to cyclic substrates. While amine and organic bases did not perform well in the chemistry, the use of alkoxide bases resulted in considerably reduced reaction times (entries 8–9). Ultimately, it was found that the use of LiO*t*-Bu as base was optimal, delivering β ketoester **112a** with an exceptional branched to linear ratio (93:7), >20:1 diastereoselectivity, and 98% enantioselectivity in only two hours (entry 8).

Table 4.2.1. Optimization of the Ir-catalyzed asymmetric allylic alkylation^a



^a Reactions performed with 0.1 mmol of **98a**, 0.2 mmol of **111a** at 0.1 M in THF at 20 °C. ^b Determined by ¹H NMR and UHPLC-MS analysis of the crude mixture. ^c Determined by chiral SFC analysis; parenthetical value is the ee of the alternate diastereomer. ^d Not determined.

4.3 EXPLORATION OF THE REACTION SCOPE AND SUBSTITUENT EFFECTS

4.3.1 Exploration of the iridium-catalyzed allylic alkylation of linear β -ketoesters with respect to allyl electrophile

With optimized conditions identified, the scope of the reaction with respect to the electrophile was next explored. A highly selective reaction was observed between β -

ketoester 111a and various cinnamyl carbonate-derived electrophiles (98) bearing electron-donating substitutents about the aryl group, R (Table 4.3.1.1, entries 2–4). 4-Me- (98i), 4-MeO- (98b), and 3-MeO-substitutions (98j) about the aryl ring gave the corresponding α -quaternary β -ketoesters (products **112b-112d**) in good to excellent yield, dr, ee and branched to linear ratio (Table 4.3.1.1). Electron deficient aryl substituents at the allyl group (entries 5-7) were also well tolerated, delivering the branched products **112e-112g**¹⁰¹ in good to excellent yield, outstanding ee and dr, and with only slightly diminished regioselectivies. Interestingly, (4-nitro)-aryl substitution at the allyl carbonate (entry 8, substrate 981) led to loss of regioselectivity in the reaction, giving equal amounts of products 112h (14:1 dr, 93% ee) and 113h (23% ee). We were pleased to discover, however, that heteroaryl-substituted allyl carbonates (substrates 98j and 98g) resulted in smooth reactions and delivered the alkylated products 112i and 112j with excellent yield, ee and regioselectivity and with good to excellent dr (entries 9–10). Finally, we found that sorbyl carbonate **98h** was also a suitable participant in the reaction, giving the corresponding β -ketoester product (112k) in good yield and dr and with excellent regio- and enantioselectivities (entry 11).

[Ir(cod)Cl]₂ *L12*, тво 111a CO₂Et LiOt-Bu THF, 25 °C CO₂Et R °OCO₂Me 112 113 98 dr of ee of yield entrv 98 product (112) 112:113° 112° 112 (%)d (%)^t 1 98a 112a: R = H 93:7 >20:1 98 97 2 98i 112b: R = 95:5 20:1 97 >99 Dh EtO₂C 3*e* 98b 99:1 >20:1 112c: R = MeO 85 >99 4 98j 90:10 17:1 99 >99 112d: R = MeO 5 98k 112e: R = CI 98 84:16 19:1 99 98c 6 112f: R = Br 98 86:14 14:1 >99 112g: R = 78:22 16:1 99 7 98d 86 981 8 112h: R = 50:50 14:1 93 9 98f 112i: X 97:3 8:1 95 99 10 98g 112j: X = O 93 95:5 13:1 >99 11 112 k 76 95:5 6:1 91 98h EtO₂C Ме

Table 4.3.1.1. Exploration of the reaction scope with respect to allyl electrophile^a

^a Reactions performed under the conditions of Table 1, entry 8. ^b Combined isolated yield of **112** and **113**. ^c Determined by ¹H NMR analysis of the crude mixture. ^d Determined by chiral SFC analysis of the major diastereomer. ^e Conditions of Table 1, entry 7. ^f 23% ee for the linear product.

4.3.2 Investigation of allyl electrophile substituent effects on reaction selectivity

During the course of this investigation, a trend relating regioselectivity and electrophile electron deficiency began to emerge. Specifically, the regioselectivity of the reaction diminished as the electron deficiency of the allyl substituent increased. In order to identify the linear free energy relationship governing the reaction, we performed a Hammett analysis relating the log of the ratio of branched to linear products, which is proportional to the relative rates of product formation, to the corresponding Hammett σ -values. The negative ρ value observed from this plot suggests that as the magnitude of electropositive charge generated at the putative allyl-Ir intermediate^{8b} increases, the reaction pathway yielding the branched allylation product becomes more favorable. This analysis can be extended to account, in part, for the poor regioselectivity observed in the case of *p*-nitro substituted allyl component **98**.

Figure 4.3.2.1. Hammett plot of the log of product ratios (112:113) from Table 2 versus Hammett σ -values



4.3.3 Exploration of the iridium-catalyzed allylic alkylation of linear β -ketoesters with respect to β -ketoester nucleophile

Having investigated reaction substrate scope with respect to the allyl electrophile, we next examined the diversity of nucleophilic coupling partners permitted in the chemistry (Table 4.3.3.1). β -Ketoesters (111) bearing either electron-donating or electron-withdrawing aryl substituents (R¹) at the ketone fared very well in the reaction, delivering products **112l** and **112m** in excellent yield, dr, ee and branched to linear ratio (entries 1 and 2). Gratifyingly, a wide variety of functional groups are readily permitted at the α -position (R²), including alkyl, benzyl, allyl, propargyl, heteroaryl and keto groups (substrates **111d–111i**, entries 3–8, respectively). The products of these reactions 112n–112s, respectively) were obtained with excellent (products ee and regioselectivities, and in good to excellent dr and yield. To the best of our knowledge, substrate **111g** represents the first example of a terminal propargyl-substituted nucleophile to undergo Ir-catalyzed allylic substitutions.¹⁰² Nitrile-containing substituents were tolerated in the reaction as well (substrate 111i), and α -quaternary β ketoester 112t was furnished in excellent yield, ee and regioselectivity, albeit with a diminished dr of 3:1. We were pleased to learn that use of certain of α -halogenated nucleophiles (substrates 111k and 111l) also resulted in an efficient and selective reaction as α -fluoro and α -chloro β -ketoesters **112u** and **112v** were obtained in excellent yields, dr, ee and regioselectivity. In addition to aryl ketones, cyclohexenyl β -ketoester **111m** was found to deliver the corresponding product **112w** in excellent yield, dr, ee and branched to linear ratio with no detectable products resulting from competitive bimolecular Michael addition. Although the use of alkyl β -ketoesters 111n and 1110 provided the desired products (112x and 112y, respectively) with excellent yields, ee's and regioselectivities, we were disappointed to find that the diastereoselectivities were diminished considerably. Lastly, we found that the use of a sterically-hindered ester moiety (substrate 111p) gave an efficient and highly enantioselective reaction but with a concurrent loss in regio- and diastereoselectivity (entry 15).

	Ph	$\begin{array}{c} 0 \\ R^2 \\ 111 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\$	od)Cl] ₂ TBD T-Bu 25 °C 11	R :0 ₂ R ³	() + R ¹	R ² CO ₂ F 113	R 13 3
entry	111	produc	t (112)	yield (%) ^b	112:113°	dr of <i>112°</i>	ee of 112 (%) ^d
1	111b	O Ph ↓ ↓	<i>112I</i> : R ³ = Et R ¹ = 4-MeO-C ₆ H ₄	93	93:7	17:1	99
2	111c	R ¹ Me ² CO ₂ R ³	<i>112m</i> : R ³ = Me R ¹ = 4-Br-C ₆ H ₄	92	92:8	17:1	>99
3	111d		<i>112n</i> : R ² = Et	94	90:10	>20:1	>99
4	111e		<i>1120</i> : R ² = Bn	99	90:10	13:1	>99
5	111f		<i>112p</i> : R ² = allyl	98	95:5	>20:1	>99
6 ^e	111g	O Ph ₹ _ 1	12q: R ² = propargyl	84	81:19	13:1	>99
7 ^f	111h	Ph 11. R ² CO ₂ Et 11.	2r: R ² = (CH ₂) ₂ COMe	98	93:7	20:1	99
8	111i	112	$s: R^2 = \bigcup_{N}$	88	95:5	7:1	>99
9	111j	1	n	99	95:5	3:1	>99 (>99)
10	111k		<i>112u</i> : R ² = F	92	96:4	13:1	95
11	1111		<i>112v</i> : R ² = Cl	96	96:4	>20:1	>99
12	111m	Me [°] CO ₂ Me	: 112w	85	90:10	12:1	99
13	111n	O Ph	<i>112x</i> : R ¹ = Cy R ² = Me	92	92:8	4:1	96
14	1110		<i>112y</i> : R ¹ = Me R ² = Et	90	93:7	1.5:1	90 (91) ^g
15	111p	Ph Me ^V CO ₂ t-Bu	112z	95	70:30	6:1	>99

Table 4.3.3.1. Exploration of the reaction scope with respect to β -ketoester nucleophile^{*a*}

^{*a*} Reactions performed under the conditions of Table 1, entry 8. ^{*b*} Combined isolated yield of **112** and **113**. ^{*c*} Determined by ¹H NMR analysis of the crude mixture. ^{*d*} Determined by chiral SFC analysis of the major diastereomer. ^{*e*} 4 mol % of [Ir(cod)Cl]₂ and 8 mol % of **L12** were used. ^{*f*} The reaction was run at 0.5 mmol scale. ^{*g*} ee for the minor diastereomer.

4.4 ELABORATION OF THE ALLYLIC ALKYLATION PRODUCTS

In order to exhibit the utility of our method for generating interesting and useful chiral building blocks, a number of selective transformations were carried out on products obtained in the course of our studies (Scheme 4.4.1). Aldol condensation of β -ketoester **112r** gave γ -quaternary cyclohexenone **114** in excellent yield. Ring-closing metathesis of diallyl β -ketoester **112p** cleanly furnished cyclohexene **115** in excellent yield. Finally, Pauson–Khand cyclization of progaryl-substituted β -ketoester **112q** smoothly delivered bicycle **116** in outstanding 99% yield.

Scheme 4.4.1. Rapid generation of molecular and stereochemical complexity employing allylic alkylation products



4.5 CONCLUDING REMARKS

In summary, the first enantioselective catalytic allylic alkylation of linear β ketoesters to generate vicinal quaternary and tertiary stereocenters in high yield, dr, ee and regioselectivity has been developed. The process hinges on the use of an Ir•*N*-arylphosphoramidite catalyst (**L12**). A variety of substitution patterns at the allyl electrophile and β -ketoester are well tolerated in the chemistry. A number of transformations were carried out on reaction products to demonstrate the value this method holds for the rapid generation of highly functionalized chiral building blocks. Studies utilizing this method toward the synthesis of complex biologically active natural products are underway in our laboratory and will serve to showcase the utility of the method in synthetic setting. Moreover, studies to extend the scope of functionality tolerated about the electrophile in the chemistry, in particular non-aromatic substituents, should greatly expand the potential applications of this method to total synthesis.

4.6 EXPERIMENTAL SECTION

4.6.1 Materials and Methods

Unless otherwise stated, reactions were performed in flame-dried glassware under an argon or nitrogen atmosphere using dry, deoxygenated solvents. Solvents were dried by passage through an activated alumina column under argon.⁶¹ Reaction progress was monitored by thin-layer chromatography (TLC) or Agilent 1290 UHPLC-MS. TLC was performed using E. Merck silica gel 60 F254 precoated glass plates (0.25 mm) and visualized by UV fluorescence quenching, *p*-anisaldehyde, or KMnO₄ staining. Silicycle Silia*Flash*® P60 Academic Silica gel (particle size 40–63 nm) was used for flash chromatography. ¹H NMR spectra were recorded on Varian Inova 500 MHz and 600 MHz spectrometers and are reported relative to residual CHCl₃ (δ 7.26 ppm) or C₆HD₅ (δ 7.16 ppm). ¹³C NMR spectra were recorded on a Varian Inova 500 MHz spectrometer (125 MHz) and are reported relative to CHCl₃ (δ 77.16 ppm) or C₆HD₅ (δ 128.06 ppm). ³¹P and ¹⁹F NMR spectra were recorded on a Varian Mercury 300 MHz (at 121 MHz and 282 MHz, respectively). ¹⁹F NMR spectra were reported relative to CFCl₃ (δ 0.0 ppm). Data for ¹H NMR

are reported as follows: chemical shift (δ ppm) (multiplicity, coupling constant (Hz), integration). Multiplicities are reported as follows: s = singlet, d = doublet, t = triplet, q =quartet, p = pentet, sept = septuplet, m = multiplet, br s = broad singlet, br d = broad doublet, app = apparent. Data for ${}^{13}C$ NMR are reported in terms of chemical shifts (δ ppm). IR spectra were obtained by use of a Perkin Elmer Spectrum BXII spectrometer or Nicolet 6700 FTIR spectrometer using thin films deposited on NaCl plates and reported in frequency of absorption (cm⁻¹). Optical rotations were measured with a Jasco P-2000 polarimeter operating on the sodium D-line (589 nm), using a 100 mm pathlength cell and are reported as: $[a]_D^T$ (concentration in g/100 mL, solvent). Analytical HPLC was performed with an Agilent 1100 Series HPLC utilizing a Chiralpak (AD-H or AS) or Chiralcel (OD-H, OJ-H, or OB-H) columns (4.6 mm x 25 cm) obtained from Daicel Chemical Industries, Ltd. Analytical SFC was performed with a Mettler SFC supercritical CO₂ analytical chromatography system utilizing Chiralpak (AD-H, AS-H or IC) or Chiralcel (OD-H, OJ-H, or OB-H) columns (4.6 mm x 25 cm) obtained from Daicel Chemical Industries, Ltd. High resolution mass spectra (HRMS) were obtained from Agilent 6200 Series TOF with an Agilent G1978A Multimode source in electrospray ionization (ESI+), atmospheric pressure chemical ionization (APCI+), or mixed ionization mode (MM: ESI-APCI+).

Reagents were purchased from Sigma-Aldrich, Acros Organics, Strem, or Alfa Aesar and used as received unless otherwise stated. Ligands L12, L13, L16 and L17⁸⁷ allyl carbonates 98,⁹⁵ and β -ketoesters 111^{103} were prepared by known methods.

4.6.2 Optimization of reaction parameters

 Table 4.6.2.1 Optimization of reaction parameters



^{*a*} Reactions performed with 0.1 mmol of **98a**, 0.2 mmol of **111a** in 1 mL of THF. ^{*b*} Determined by ¹H NMR or UHPLC-MS analysis of the crude reaction mixture. ^{*c*} Isolated yield. ^{*d*} (ee) of the alternative diastereomer.

General Procedure for Optimization Reaction (Table 4.6.2.1): All experiments were performed in a nitrogen-filled glove box.

Procedure A (for entries 1-9): $[Ir(cod)Cl]_2$ (1.4 mg, 0.002 mmol, 2 mol%), ligand L (0.004 mmol, 4 mol%), and TBD (1.4 mg, 0.01 mmol, 10 mol%) were added to a 2 dram scintillation vial (vial A) equipped with a magnetic stirring bar. The vial was then charged with THF (0.5 mL) and stirred at 25 °C for 10 min, generating an orange solution. To another 2 dram scintillation vial (vial B) was added base (0.2 mmol, 2 equiv), 0.5 mL of THF and β -ketoester **111a** (41.2 mg, 0.2 mmol, 2.0 equiv). After

stirring for 10 min at 25 °C, the pre-formed catalyst solution (vial A) was transferred to vial B, and cinnamyl carbonate (**98a**) (19.2 mg, 0.1 mmol, 1.0 equiv) was added. The vial was sealed and stirred at 25 °C until allylic carbonate **98a** was fully consumed, as indicated by UHPLC-MS analysis. Upon completion of the reaction the vial was removed from the glove-box and uncapped. Saturated NH₄Cl aqueous solution was added and the mixture was extracted with CH_2Cl_2 (10 mL x 3), the combined organic phase was washed with brine, dried over Mg₂SO₄, filtered and concentrated *in vacuo*. The regioselectivity (branched product to linear product, b:l) and diastereoselectivity (dr) were determined by ¹H NMR. The residue was purified by silica gel flash chromatography (gradient elution, $0\rightarrow 1\rightarrow 2\%$ EtOAc in hexanes) to afford the desired product.

Procedure B (for entries 10-15): [Ir(cod)Cl]₂ (1.4 mg, 0.002 mmol, 2 mol%), ligand L (0.004 mmol, 4 mol%), and TBD (1.4 mg, 0.01 mmol, 10 mol%) were added to a 2 dram scintillation vial equipped with a magnetic stirring bar. The vial was then charged with THF (0.5 mL) and stirred at 25 °C for 10 min, generating an orange solution. Cinnamyl carbonate (**98a**) (19.2 mg, 0.1 mmol, 1.0 equiv), β-ketoester **111a** (41.2 mg, 0.2 mmol, 2.0 equiv), base (0.2 mmol, 2 equiv) or additive (0.1 mmol, 1.0 equiv) and an additional 0.5 mL of THF were added. The vial was sealed and stirred at 25 °C until allylic carbonate **98a** was fully consumed, as indicated by UHPLC-MS analysis. Upon completion of the reaction the vial was removed from the glove-box and uncapped and THF evaporated under reduced pressure. Et₂O was added to the crude mixture and the resulting precipitate was filtered through a celite pad, rinsed with Et₂O and the filtrate was concentrated under reduced pressure. The regioselectivity (branched

product to linear product: b:l) and diastereoselectivity (dr) were determined by ¹H NMR. The residue was purified by silica gel flash chromatography (gradient elution, $0 \rightarrow 1 \rightarrow 2\%$ EtOAc in hexanes) to afford the desired product.

4.6.3. General procedure for the Ir-catalyzed asymmetric allylic alkylation of acyclic β -ketoesters

<u>Note</u> that the absolute configuration was determined only for compound **112f** via X-ray analysis of its derivative (vide infra). The absolute configuration for all other products **3** has been inferred by analogy. Isolated yields are reported in Tables 2 and 3 (see manuscript). For respective SFC conditions, please refer to Table 4.6.8.1. The relative configuration of product derivative **116** was determined by **2D** NMR studies, see appendix A6 for details. The relative configurations of all other products determined by anaolgy.



In a nitrogen-filled glove box, $[Ir(cod)Cl]_2$ (1.4 mg, 0.002 mmol, 2 mol%), ligand L12 (1.8 mg, 0.004 mmol, 4 mol%), and TBD (1.4 mg, 0.01 mmol, 10 mol%) were added to a 2 dram scintillation vial equipped with a magnetic stirring bar. The vial was then charged with THF (0.5 mL) and stirred at 25 °C for 10 min, generating an orange solution. To another 2 dram scintillation vial was added LiO*t*-Bu (16.0 mg, 0.2 mmol, 2

equiv) and 0.5 mL of THF, then β-ketoester **111a** (41.2 mg, 0.2 mmol, 2.0 equiv) was added. After stirring for 10 min, the above pre-formed catalyst solution was transferred to this vial, followed by cinnamyl carbonate (**98a**) (19.2 mg, 0.1 mmol, 1.0 equiv). The vial was sealed and stirred at 25 °C until allylic carbonate **98a** was fully consumed, as indicated by UHPLC-MS analysis. Upon completion of the reaction the vial was removed from the glove-box and uncapped and saturated NH₄Cl aqueous solution was added. The mixture was extracted with CH₂Cl₂ (10 mL x 3), the combined organic layers washed with brine, dried over Mg₂SO₄, filtered and concentrated under reduced pressure. The regioselectivity (branched product to linear product: b:1 = 93:7) and diastereoselectivity (dr >20:1) were determined by ¹H NMR of the crude reaction mixture. The residue was purified by silica gel flash chromatography (gradient elution, $0\rightarrow 1\rightarrow 2\%$ EtOAc in hexanes) to afford product (31.2 mg, 97% yield) as a colorless oil.

Ethyl-(2*R*,3*S*)-2-benzoyl-2-methyl-3-phenylpent-4-enoate (112a).

Ketoester **112a** was isolated by silica gel chromatography (gradient elution, $0 \rightarrow 1 \rightarrow 2\%$ EtOAc in hexanes) as a colorless oil. >99% ee, $[\alpha]_D^{25}$ +76.6 (*c* 0.77, CHCl₃); R_f = 0.4 (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.69–7.60 (m, 2H), 7.52–7.43 (m, 1H), 7.38–7.33 (m, 2H), 7.26–7.15 (m, 5H), 6.35 (ddd, *J* = 17.0, 10.3, 8.3 Hz, 1H), 5.13 (ddd, *J* = 10.2, 1.7, 0.9 Hz, 1H), 5.07 (ddd, *J* = 17.0, 1.8, 1.2 Hz, 1H), 4.39 (dd, *J* = 8.3, 1.1 Hz, 1H), 4.09 (qd, *J* = 7.2, 0.6 Hz, 2H), 1.53 (s, 3H), 1.04 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 198.0, 173.0, 139.5, 137.8, 137.2, 132.3, 130.1, 128.4, 128.20, 128.17, 127.1, 117.5, 61.8, 61.6, 54.8, 21.0, 13.8; IR (Neat Film, NaCl) 3062, 3028, 2981, 2939, 2902, 1731, 1686, 1682, 1597, 1582, 1493, 1452, 1446, 1377, 1311, 1243, 1218, 1186, 1096, 1018, 1001, 962, 920, 860, 758 cm⁻¹; HRMS (ESI+) *m/z* calc'd for C₂₁H₂₃O₃ [M+H]⁺: 323.1642, found 323.1647; SFC conditions: 5% IPA, 2.5 mL/min, Chiralpak IC column, $\lambda = 254$ nm, t_R (min): major = 4.12, minor = 6.14.

4.6.4. Spectroscopic data for Ir-catalyzed allylic alkylation products

Ethyl (2*R*,3*S*)-2-benzoyl-2-methyl-3-(*p*-tolyl)pent-4-enoate (112b).



Ketoester **112b** was isolated by silica gel chromatography (gradient elution, $0\rightarrow 1\rightarrow 2\%$ EtOAc in hexanes) as a colorless oil. 99% ee, $[\alpha]_D^{25}$ +80.4 (*c* 1.82, CHCl₃); $R_f = 0.4$ (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.69–7.65 (m, 2H), 7.47 (ddt, J = 8.6, 7.0, 1.3 Hz, 1H), 7.39–7.33 (m, 2H), 7.11–7.02 (m, 4H), 6.33 (ddd, J = 16.9, 10.2, 8.3 Hz, 1H), 5.11 (ddd, J = 10.3, 1.8, 1.0 Hz, 1H), 5.07 (ddd, J = 17.0, 1.9, 1.2 Hz, 1H), 4.38 (dd, J = 8.3, 1.2 Hz, 1H), 4.09 (qd, J = 7.1, 2.6 Hz, 2H), 2.29 (s, 3H), 1.54 (s, 3H), 1.05 (t, J =7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 198.1, 173.0, 137.9, 137.2, 136.6, 136.4, 132.3, 129.9, 128.9, 128.4, 128.2, 117.3, 61.8, 61.5, 54.3, 21.1, 20.9, 13.8; IR (Neat Film, NaCl) 3058, 3023, 2981, 2938, 1736, 1732, 1682, 1687, 1636, 1597, 1580, 1513, 1446, 1376, 1310, 1243, 1219, 1186, 1115, 1104, 1021, 1001, 963, 919, 820, 793 cm⁻¹; HRMS (MM: ESI-APCI+) m/z calc'd for C₂₂H₂₅O₃ [M+H]⁺: 337.1798, found 337.1805; SFC conditions: 5% IPA, 2.5 mL/min, Chiralpak IC column, $\lambda = 254$ nm, t_R (min): major = 6.71, minor = 9.45.

Ethyl (2*R*,3*S*)-2-benzoyl-3-(4-methoxyphenyl)-2-methylpent-4-enoate (112c).



Ketoester **112c** was isolated by silica gel chromatography (gradient elution, $2\rightarrow 5\%$ EtOAc in hexanes) as a colorless oil. >99% ee, $[\alpha]_D^{25}$ +81.5 (*c* 2.02, CHCl₃); $R_f = 0.2$ (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.70–7.63 (m, 2H), 7.52–7.43 (m, 1H), 7.38–7.32 (m, 2H), 7.15–7.08 (m, 2H), 6.85–6.74 (m, 2H), 6.32 (ddd, *J* = 17.0, 10.3, 8.2 Hz, 1H), 5.11 (ddd, *J* = 10.3, 1.8, 1.0 Hz, 1H), 5.05 (ddd, *J* = 17.0, 1.8, 1.2 Hz, 1H), 4.37 (d, *J* = 8.2 Hz, 1H), 4.09 (qd, *J* = 7.1, 0.9 Hz, 2H), 3.76 (s, 3H), 1.51 (s, 3H), 1.05 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 198.1, 173.0, 158.5, 138.0, 137.3, 132.3, 131.4, 131.1, 128.4, 128.2, 117.2, 113.5, 61.9, 61.5, 55.3, 53.9, 20.8, 13.9; IR (Neat Film, NaCl) 3067, 2981, 2937, 2904, 2835, 1731, 1686, 1682, 1610, 1597, 1581, 1511, 1446, 1376, 1302, 1245, 1218, 1181, 1114, 1101, 1034, 963, 922, 830 cm⁻¹; HRMS (ESI+) *m/z* calc'd for fragment C₁₀H₁₁O [M-C₁₁H₁₂O₃+H]⁺: 147.0804, found 147.0808; SFC conditions: 5% IPA, 2.5 mL/min, Chiralpak AD-H column, $\lambda = 254$ nm, t_R (min): minor = 11.17, major = 12.67.

Ethyl (2R,3S)-2-benzoyl-3-(-methoxyphenyl)-2-methylpent-4-enoate (112d).



Ketoester **112d** was isolated by silica gel chromatography (gradient elution, $0 \rightarrow 1 \rightarrow 2\%$ EtOAc in hexanes) as a colorless oil. >99% ee, $[\alpha]_D^{25}$ +85.1 (*c* 1.21, CHCl₃); $R_f = 0.3$ (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.70–7.63 (m, 2H), 7.52–7.43 (m, 1H), 7.42–7.32 (m, 2H), 7.23–7.11 (m, 1H), 6.82–6.71 (m, 3H), 6.32 (ddd, J = 16.9, 10.3,8.3 Hz, 1H), 5.13 (ddd, J = 10.3, 1.8, 0.9 Hz, 1H), 5.09 (ddd, J = 17.0, 1.8, 1.2 Hz, 1H), 4.37 (dt, J = 8.3, 1.1 Hz, 1H), 4.09 (qd, J = 7.2, 1.3 Hz, 2H), 3.74 (s, 3H), 1.53 (s, 3H), 1.05 (t, J = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 198.0, 172.9, 159.3, 141.0, 137.6, 137.3, 132.3, 129.1, 128.4, 128.2, 122.4, 117.6, 116.0, 112.4, 61.8, 61.6, 55.2, 54.8, 21.0, 13.8; IR (Neat Film, NaCl) 3078, 2982, 2940, 2835, 1731, 1683, 1598, 1583, 1488, 1454, 1377, 1315, 1245, 1217, 1186, 1162, 1096, 1049, 1019, 1001, 963, 922, 861, 781 cm⁻¹; HRMS (ESI+) *m/z* calc'd for C₂₂H₂₅O₄ [M+H]⁺: 353.1747, found 353.1761; SFC conditions: 2% MeOH, 2.5 mL/min, Chiralpak IC column, $\lambda = 254$ nm, t_R (min): major = 12.24, minor = 13.50.

Ethyl (2*R*,3*S*)-2-benzoyl-3-(3-chlorophenyl)-2-methylpent-4-enoate (112e).



Ketoester **112e** was isolated by silica gel chromatography (gradient elution, $0\rightarrow 1\rightarrow 2\%$ EtOAc in hexanes) as a colorless oil. 99% ee, $[\alpha]_D^{25}$ +99.1 (*c* 1.13, CHCl₃); $R_f = 0.3$ (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.70–7.63 (m, 2H), 7.48 (ddt, J = 7.8, 6.9, 1.2 Hz, 1H), 7.41–7.33 (m, 2H), 7.22–7.19 (m, 1H), 7.19–7.14 (m, 2H), 7.14–7.08 (m, 1H), 6.30 (ddd, J = 17.0, 10.2, 8.4 Hz, 1H), 5.15 (ddd, J = 10.3, 1.6, 0.9 Hz, 1H), 5.07 (dt, J = 17.0, 1.4 Hz, 1H), 4.34 (d, J = 8.4 Hz, 1H), 4.10 (q, J = 7.2 Hz, 2H), 1.52 (s, 3H), 1.06 (t, J = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 197.6, 172.6, 141.7, 137.0, 136.9, 133.9, 132.5, 130.3, 129.3, 128.5, 128.4, 128.3, 127.2, 118.1, 61.7, 61.7, 54.5, 20.9, 13.8; IR (Neat Film, NaCl) 3068, 2982, 2943, 1732, 1682, 1595, 1574, 1475, 1446, 1378, 1301, 1245, 1218, 1194, 1095, 1018, 1001, 963, 924, 784 cm⁻¹; HRMS (MM: ESI-APCI+) *m/z* calc'd for C₂₁H₂₂ClO₃ [M+H]⁺: 357.1252, found 357.1260; SFC conditions: 10% IPA, 4.0 mL/min, Chiralpak IC column, $\lambda = 254$ nm, t_R (min): major = 1.59, minor = 1.77.

Ethyl (2*R*,3*S*)-2-benzoyl-3-(4-bromophenyl)-2-methylpent-4-enoate (112f).



Ketoester **112f** was isolated by silica gel chromatography (gradient elution, $0 \rightarrow 1 \rightarrow 2\%$ EtOAc in hexanes) as a colorless oil. >99% ee, $[\alpha]_D^{25}$ +89.0 (*c* 1.42, CHCl₃); $R_f = 0.3$ (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.70–7.63 (m, 2H), 7.50–7.45 (m, 1H), 7.40–7.33 (m, 4H), 7.13–7.08 (m, 2H), 6.29 (ddd, J = 16.9, 10.2, 8.2 Hz, 1H), 5.16– 5.11 (m, 1H), 5.05 (dt, J = 17.0, 1.4 Hz, 1H), 4.36 (d, J = 8.2 Hz, 1H), 4.08 (q, J = 7.1 Hz, 2H), 1.51 (s, 3H), 1.05 (t, J = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 197.6, 172.7, 138.6, 137.2, 136.9, 132.5, 131.9, 131.2, 128.5, 128.3, 121.1, 117.9, 61.7, 61.6, 54.1, 20.8, 13.8; IR (Neat Film, NaCl) 3077, 2981, 2938, 1728, 1683, 1597, 1488, 1446, 1377, 1305, 1243, 1217, 1186, 1099, 1076, 1010, 963, 922, 822, 802, 716 cm⁻¹; HRMS (MM: ESI-APCI+) *m/z* calc'd for C₂₁H₂₂BrO₃ [M+H]⁺: 401.0747, found 401.0754; SFC conditions: 10% IPA, 4.0 mL/min, Chiralpak AD-H column, $\lambda = 210$ nm, t_R (min): minor = 2.95, major = 3.47.

Ethyl (2*R*,3*S*)-2-benzoyl-2-methyl-3-(4-(trifluoromethyl)phenyl)pent-4-enoate (112g).



Ketoester **112g** was isolated by silica gel chromatography (gradient elution, $0 \rightarrow 1 \rightarrow 2\%$ EtOAc in hexanes) as a colorless oil. >99% ee, [α]_D²⁵ +77.4 (*c* 0.94, CHCl₃); R_f = 0.3 (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.69–7.63 (m, 2H), 7.53–7.45 (m, 3H), 7.39–7.35 (m, 4H), 6.33 (ddd, *J* = 16.9, 10.2, 8.4 Hz, 1H), 5.16 (ddd, *J* = 10.2, 1.6, 0.9 Hz, 1H), 5.07 (dt, *J* = 17.0, 1.4 Hz, 1H), 4.44 (d, *J* = 8.4 Hz, 1H), 4.09 (q, *J* = 7.1 Hz, 2H), 1.53 (s, 3H), 1.04 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 197.5, 172.6, 143.9, 136.9, 136.8, 132.6, 130.6, 129.2 (q, *J* = 32.4 Hz), 128.5, 128.3, 125.0 (q, *J* = 3.7 Hz), 124.3 (d, *J* = 271.8 Hz), 118.3, 61.8, 61.6, 54.6, 20.8, 13.8; IR (Neat Film, NaCl) 3070, 2984, 2941, 1732, 1687, 1682, 1617, 1597, 1581, 1446, 1413, 1379, 1327, 1245, 1220, 1166, 1123, 1069, 1018, 964, 924, 846 cm⁻¹; HRMS (ESI+) *m/z* calc'd for C₂₂H₂₂F₃O₃ [M+H]⁺: 391.1516, found 391.1517; SFC conditions: 2% IPA, 2.5 mL/min, Chiralpak AD-H column, $\lambda = 254$ nm, t_R (min): minor = 5.20, major = 6.68.

Ethyl (2*R*,3*S*)-2-benzoyl-2-methyl-3-(4-nitrophenyl)pent-4-enoate (112h).



Ketoester **112h** was isolated by silica gel chromatography (5% EtOAc in hexanes) as a colorless oil. 93% ee, $[α]_D^{25}$ +96.8 (*c* 0.64, CHCl₃); R_f = 0.3 (10% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 8.13–8.09 (m, 2H), 7.70–7.63 (m, 2H), 7.52–7.48 (m, 1H), 7.46–7.43 (m, 2H), 7.40–7.35 (m, 2H), 6.31 (ddd, *J* = 16.9, 10.2, 8.5 Hz, 1H), 5.23–5.16 (m, 1H), 5.08 (dt, *J* = 17.0, 1.3 Hz, 1H), 4.50 (d, *J* = 8.5 Hz, 1H), 4.10 (qd, *J* = 7.1, 2.4 Hz, 2H), 1.54 (s, 3H), 1.05 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 197.0, 172.3, 147.4, 146.8, 136.3, 136.1, 132.6, 131.0, 128.4, 128.2, 123.1, 118.8, 61.8, 61.4, 54.3, 20.5, 13.7; IR (Neat Film, NaCl) 3080, 2982, 2942, 1731, 1686, 1682, 1597, 1523, 1519, 1446, 1379, 1346, 1245, 1219, 1111, 1001, 1015, 964, 926, 853 cm⁻¹; HRMS (MM: ESI-APCI+) *m/z* calc'd for C₂₁H₂₂O₅N [M+H]⁺: 368.1492, found 368.1508; SFC conditions: 5% MeOH, 3.0 mL/min, Chiralpak AD-H column, λ = 254 nm, t_R (min): minor = 4.47, major = 5.71.

Ethyl (E)-2-benzoyl-2-methyl-5-(4-nitrophenyl)pent-4-enoate (113h).



Ketoester **113h** was isolated by silica gel chromatography (5% EtOAc in hexanes) as a yellow solid. 23% ee, $[\alpha]_D^{25}$ +4.7 (*c* 0.96, CHCl₃); $R_f = 0.3$ (10% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 8.15 (dt, *J* = 9.0, 2.5 Hz, 2H), 7.86 (dt, *J* = 8.5, 2.0 Hz, 2H), 7.59–7.49 (m, 1H), 7.47–7.40 (m, 4H), 6.45 (dt, *J* = 15.9, 1.2 Hz, 1H), 6.32 (dt, *J* = 15.7, 7.5 Hz, 1H), 4.12 (qd, *J* = 7.2, 1.1 Hz, 2H), 3.00 (ddd, *J* = 14.2, 7.3, 1.3 Hz, 1H), 2.90 (ddd, *J* = 14.2, 7.7, 1.2 Hz, 1H), 1.58 (s, 3H), 1.05 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 197.0, 173.6, 146.9, 143.6, 135.4, 133.1, 132.1, 130.0, 128.7, 126.8, 124.1, 61.7, 57.3, 40.6, 21.6, 14.0; IR (Neat Film, NaCl) 2981, 2936, 1732, 1686, 1682, 1596, 1519, 1515, 1446, 1342, 1298, 1267, 1239, 1184, 1108, 973, 857 cm⁻¹; HRMS (MM: ESI-APCI+) *m/z* calc'd for C₂₁H₂₂O₅N [M+H]⁺: 368.1492, found 368.1499; SFC conditions: 10% MeOH, 3.0 mL/min, Chiralpak AD-H column, λ = 254 nm, t_R (min): major = 7.13, minor = 8.06.

Ethyl (2*R*,3*R*)-2-benzoyl-2-methyl-3-(thiophen-2-yl)pent-4-enoate (112i).



Ketoester **112i** was isolated by silica gel chromatography (gradient elution, $0\rightarrow 1\rightarrow 2\%$ EtOAc in hexanes) as a colorless oil. 95% ee, $[\alpha]_D^{25}$ +53.0 (*c* 1.63, CHCl₃); $R_f = 0.4$ (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.74–7.68 (m, 2H), 7.51–7.47 (m, 1H), 7.42–7.35 (m, 2H), 7.16 (dd, J = 5.1, 1.2 Hz, 1H), 6.89 (dd, J = 5.1, 3.5 Hz, 1H), 6.84 (ddd, J = 3.5, 1.2, 0.7 Hz, 1H), 6.23 (ddd, J = 16.4, 10.6, 9.0 Hz, 1H), 5.23–5.16 (m, 2H), 4.77 (d, J = 9.0 Hz, 1H), 4.11 (q, J = 7.1 Hz, 2H), 1.52 (s, 3H), 1.09 (t, J = 7.2 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 197.5, 172.3, 142.1, 136.8, 136.6, 132.5, 128.5, 128.4, 126.9, 126.2, 124.9, 118.2, 62.2, 61.9, 49.8, 19.4, 13.9; IR (Neat Film, NaCl) 3069, 2981, 2937, 1732, 1687, 1682, 1597, 1580, 1446, 1383, 1298, 1246, 1221, 1107, 1018, 1001, 968, 924, 851, 795, 747 cm⁻¹; HRMS (ESI+) *m/z* calc'd for C₁₉H₂₁O₃S [M+H]⁺: 329.1206, found 329.1214; SFC conditions: 5% IPA, 2.5 mL/min, Chiralpak IC column, $\lambda = 254$ nm, t_R (min): major = 7.92, minor = 11.24.

Ethyl (2R,3R)-2-benzoyl-3-(furan-2-yl)-2-methylpent-4-enoate (112j).



Ketoester **112j** was isolated by silica gel chromatography (gradient elution, $0 \rightarrow 1 \rightarrow 2\%$ EtOAc in hexanes) as a colorless oil. >99% ee, $[\alpha]_D^{25}$ +67.3 (*c* 1.34, CHCl₃); $R_f = 0.4$ (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.73–7.67 (m, 2H), 7.52–7.45 (m, 1H), 7.40–7.35 (m, 2H), 7.27 (dd, J = 1.8, 0.9 Hz, 1H), 6.25 (dd, J = 3.3, 1.8 Hz, 1H), 6.17 (ddd, J = 17.0, 10.2, 8.5 Hz, 1H), 6.11 (dt, J = 3.3, 0.7 Hz, 1H), 5.19 (ddd, J = 10.2, 1.7, 0.8 Hz, 1H), 5.14 (ddd, J = 17.0, 1.7, 1.1 Hz, 1H), 4.62 (dd, J = 8.5, 0.9 Hz, 1H), 4.12 (q, J = 7.1 Hz, 2H), 1.52 (s, 3H), 1.10 (t, J = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 196.9, 172.1, 152.9, 141.5, 136.6, 134.4, 132.2, 128.3, 128.2, 118.3, 110.1, 108.7, 61.6, 61.4, 47.9, 19.3, 13.8; IR (Neat Film, NaCl) 3081, 2983, 2941, 2904, 1732, 1686, 1597, 1581, 1501, 1446, 1378, 1301, 1246, 1222, 1149, 1096, 1013, 968, 926, 885, 860, 797, 736 cm⁻¹; HRMS (MM: ESI-APCI+) *m/z* calc'd for C₁₉H₂₁O₄ [M+H]⁺: 313.1431, found 313.1438; SFC conditions: 5% IPA, 2.5 mL/min, Chiralpak IC column, $\lambda = 254$ nm, t_R (min): major = 5.09, minor = 9.14.

Ethyl (2R,3S,E)-2-benzoyl-2-methyl-3-vinylhex-4-enoate (112k).

Ketoester **112k** was isolated by silica gel chromatography (gradient elution, $0\rightarrow 1\rightarrow 2\%$ EtOAc in hexanes) as a colorless oil. 91% ee, $[\alpha]_D^{25}$ +43.5 (*c* 1.86, CHCl₃); $R_f = 0.4$ (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.80–7.71 (m, 2H), 7.52–7.46 (m, 1H), 7.42–7.36 (m, 2H), 5.98 (ddd, J = 16.9, 10.5, 7.5 Hz, 1H), 5.47–5.27 (m, 2H), 5.15–5.00 (m, 2H), 4.09 (qd, J = 7.1, 1.5 Hz, 2H), 3.73–3.69 (m, 1H), 1.65–1.58 (m, 3H), 1.48 (s, 3H), 1.07 (t, J = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 197.4, 173.0, 137.2, 136.8, 132.4, 129.0, 128.5, 128.4, 128.3, 117.1, 61.4, 60.5, 51.6, 19.2, 18.2, 14.0; IR (Neat Film, NaCl) 3077, 2981, 2939, 2913, 1732, 1687, 1682, 1597, 1580, 1446, 1377, 1299, 1245, 1221, 1100, 1018, 1001, 970, 921, 859 cm⁻¹; HRMS (ESI+) *m/z* calc'd for C₁₈H₂₃O₃ [M+H]⁺: 287.1642, found 287.1654; SFC conditions: 2% IPA, 2.5 mL/min, Chiralpak AD-H column, $\lambda = 210$ nm, t_R (min): minor = 4.40, major = 5.52.

Ethyl (2*R*,3*S*)-2-(4-methoxybenzoyl)-2-methyl-3-phenylpent-4-enoate (112l).

Ketoester **1121** was isolated by silica gel chromatography (4% EtOAc in hexanes) as a colorless oil. 99% ee, $[\alpha]_D^{25}$ +53.9 (*c* 1.51, CHCl₃); $R_f = 0.3$ (10% EtOAc in hexanes);

¹H NMR (500 MHz, CDCl₃) δ7.76–7.69 (m, 2H), 7.25–7.17 (m, 5H), 6.87–6.82 (m, 2H), 6.36 (ddd, J = 17.0, 10.3, 8.1 Hz, 1H), 5.12 (ddd, J = 10.3, 1.8, 1.0 Hz, 1H), 5.05 (ddd, J = 17.0, 1.8, 1.2 Hz, 1H), 4.39 (dt, J = 8.1, 1.2 Hz, 1H), 4.10 (q, J = 7.1 Hz, 2H), 3.84 (s, 3H), 1.53 (s, 3H), 1.07 (t, J = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 195.8, 173.4, 162.9, 139.6, 138.0, 130.8, 130.2, 129.5, 128.1, 127.0, 117.3, 113.6, 61.5, 61.3, 55.5, 54.7, 21.0, 13.9; IR (Neat Film, NaCl) 3083, 2981, 2940, 2842, 1727, 1677, 1600, 1576, 1513, 1454, 1417, 1376, 1310, 1247, 1221, 1176, 1118, 1031, 964, 842 cm⁻¹; HRMS (ESI+) m/z calc'd for C₂₂H₂₅O₄ [M+H]⁺: 353.1747, found 353.1752; SFC conditions: 10% IPA, 4.0 mL/min, Chiralpak IC column, $\lambda = 254$ nm, t_R (min): major = 2.67, minor = 3.51.

Methyl (2*R*,3*S*)-2-(4-bromobenzoyl)-2-methyl-3-phenylpent-4-enoate (112m).

Ketoester **112m** was isolated by silica gel chromatography (gradient elution, $1\rightarrow 2$ Et₂O in hexanes) as a colorless oil. >99% ee, $[\alpha]_D^{25}$ +65.9 (*c* 0.50, CHCl₃); $R_f = 0.3$ (10% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.51–7.48 (m, 4H), 7.29–7.09 (m, 5H), 6.31 (ddd, J = 16.9, 10.3, 8.4 Hz, 1H), 5.21–4.96 (m, 2H), 4.37 (d, J = 8.5 Hz, 1H), 3.62 (s, 3H), 1.51 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 196.6, 173.3, 139.1, 137.3, 135.7, 131.7, 130.0, 129.7, 128.2, 127.5, 127.1, 117.7, 61.8, 54.7, 52.5, 20.8; IR (Neat Film, NaCl) 3355, 3028, 2997, 2948, 1735, 1685, 1584, 1484, 1452, 1395, 1245, 1221, 1117, 1073, 966, 922, 841, 756 cm⁻¹; HRMS (ESI+) *m/z* calc'd for C₂₀H₂₀BrO₃ [M+H]⁺:

387.0590, found 387.0612; SFC conditions: 5% IPA, 2.5 mL/min, Chiralpak AD-H column, $\lambda = 254$ nm, t_R (min): major = 5.35, minor = 5.88.

Ethyl (2*R*,3*S*)-2-benzoyl-2-ethyl-3-phenylpent-4-enoate (112n).



Ketoester **112n** was isolated by silica gel chromatography (gradient elution, $0\rightarrow 2$ Et₂O in hexanes) as a colorless oil. >99% ee, $[\alpha]_D^{25}$ +77.4 (*c* 0.25, CHCl₃); $R_f = 0.4$ (10% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.61–7.51 (m, 2H), 7.48–7.38 (m, 1H), 7.35–7.11 (m, 7H), 6.33 (ddd, *J* = 17.0, 10.3, 7.8 Hz, 1H), 5.07 (ddd, *J* = 10.3, 1.8, 1.1 Hz, 1H), 4.96 (dt, *J* = 17.1, 1.5 Hz, 1H), 4.38 (dd, *J* = 7.9, 1.4 Hz, 1H), 4.06 (qd, *J* = 7.2, 3.3 Hz, 2H), 2.15 (dq, *J* = 15.0, 7.5 Hz, 1H), 1.89 (dq, *J* = 14.9, 7.5 Hz, 1H), 0.99 (t, *J* = 7.1 Hz, 3H), 0.78 (t, *J* = 7.5 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 198.1, 172.9, 139.5, 138.4, 137.9, 132.0, 129.8, 128.2, 128.1, 127.9, 127.0, 116.6, 65.2, 61.0, 52.2, 27.6, 13.6, 8.5; IR (Neat Film, NaCl) 3061, 3028, 2980, 1729, 1679, 1597, 1446, 1386, 1303, 1228, 1208, 1097, 1028, 993, 917, 759 cm⁻¹; HRMS (ESI+) *m*/z calc'd for C₂₂H₂₅O₃ [M+H]⁺: 337.1798, found 337.1813; SFC conditions: 5% MeOH, 2.5 mL/min, Chiralpak AD-H column, $\lambda = 254$ nm, t_R (min): major = 2.48, minor = 2.20.

Ethyl (2*R*,3*S*)-2-benzoyl-2-benzyl-3-phenylpent-4-enoate (112o).



Ketoester **1120** was isolated by silica gel chromatography (gradient elution, $1\rightarrow 2$ EtOAc in hexanes) as a white solid. >99% ee, $[\alpha]_D^{25}$ +37.1 (*c* 1.29, CHCl₃); R_f = 0.4 (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.33–7.29 (m, 1H), 7.29–7.21 (m, 4H), 7.17– 7.10 (m, 8H), 7.04–7.00 (m, 2H), 6.37 (ddd, *J* = 17.0, 10.3, 8.1 Hz, 1H), 5.13 (ddd, *J* = 10.2, 1.8, 1.0 Hz, 1H), 5.02 (dt, *J* = 17.0, 1.5 Hz, 1H), 4.42 (d, *J* = 7.9 Hz, 1H), 3.99 (dq, *J* = 10.7, 7.1 Hz, 1H), 3.80 (dq, *J* = 10.7, 7.1 Hz, 1H), 3.28 (AB, *J* = 15.5, 13.5 Hz, 2H), 0.82 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 200.5, 172.0, 139.7, 139.3, 138.3, 136.8, 131.4, 131.2, 130.0, 128.4, 128.1, 127.7, 127.6, 127.4, 126.9, 117.6, 67.5, 61.0, 56.8, 42.1, 13.4; IR (Neat Film, NaCl) 3063, 3029, 2982, 2929, 1729, 1673, 1600, 1582, 1496, 1448, 1367, 1299, 1241, 1209, 1082, 1066, 1025, 925, 757 cm⁻¹; HRMS (ESI+) *m/z* calc'd for C₂₇H₂₇O₃ [M+H]⁺: 399.1955, found 399.1956; SFC conditions: 4% IPA, 4.0 mL/min, Chiralpak AD-H column, $\lambda = 254$ nm, t_R (min): major = 5.06, minor = 8.94.

Ethyl (2*R*,3*S*)-2-allyl-2-benzoyl-3-phenylpent-4-enoate (112p).



Ketoester **112p** was isolated by silica gel chromatography (2% EtOAc in hexanes) as a colorless oil. >99% ee, $[\alpha]_D^{25}$ +80.7 (*c* 0.31, CHCl₃); $R_f = 0.4$ (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.54–7.49 (m, 2H), 7.43 (ddt, *J* = 7.7, 7.0, 1.2 Hz, 1H),

7.33–7.27 (m, 2H), 7.25–7.19 (m, 3H), 7.18–7.13 (m, 2H), 6.36 (ddd, J = 17.0, 10.3, 8.0 Hz, 1H), 5.71 (dddd, J = 17.0, 10.2, 7.8, 7.0 Hz, 1H), 5.10 (ddd, J = 10.3, 1.8, 1.1 Hz, 1H), 5.03 (ddt, J = 10.2, 2.1, 1.0 Hz, 1H), 4.99 (ddd, J = 17.0, 1.8, 1.2 Hz, 1H), 4.92 (dq, J = 17.1, 1.7 Hz, 1H), 4.35 (dt, J = 8.0, 1.2 Hz, 1H), 4.14–3.96 (m, 2H), 2.86 (ddt, J = 14.6, 7.0, 1.3 Hz, 1H), 2.63 (ddt, J = 14.6, 7.7, 1.2 Hz, 1H), 0.99 (t, J = 7.2 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 198.0, 172.4, 139.4, 138.3, 138.2, 132.5, 132.1, 130.1, 128.3, 128.14, 128.12, 127.2, 119.5, 117.2, 65.2, 61.2, 53.4, 39.4, 13.7; IR (Neat Film, NaCl) 3079, 3029, 2981, 2933, 1728, 1679, 1638, 1597, 1581, 1493, 1446, 1367, 1257, 1216, 1181, 1144, 1045, 1023, 1001, 921, 758 cm⁻¹; HRMS (MM: ESI-APCI+) *m/z* calc'd for C₂₃H₂₅O₃ [M+H]⁺: 349.1808, found 349.1808; SFC conditions: 3% IPA, 4.0 mL/min, Chiralpak IC column, $\lambda = 254$ nm, t_R (min): major = 7.16, minor = 8.60.

Ethyl (2*R*,3*S*)-2-benzoyl-3-phenyl-2-(prop-2-yn-1-yl)pent-4-enoate (112q).



Ketoester **112q** was isolated by silica gel chromatography (2% EtOAc in hexanes) as a colorless oil. >99% ee, $[\alpha]_D^{25}$ +57.7 (*c* 0.94, CHCl₃); $R_f = 0.3$ (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.62–7.55 (m, 2H), 7.46 (ddt, *J* = 8.6, 7.0, 1.2 Hz, 1H), 7.35–7.29 (m, 2H), 7.25–7.23 (m, 5H), 6.37 (ddd, *J* = 17.0, 10.3, 7.8 Hz, 1H), 5.15 (ddd, *J* = 10.3, 1.8, 1.2 Hz, 1H), 5.09 (dt, *J* = 17.0, 1.5 Hz, 1H), 4.58 (dt, *J* = 7.8, 1.3 Hz, 1H), 4.19–4.05 (m, 2H), 2.97 (dd, *J* = 17.4, 2.7 Hz, 1H), 2.69 (dd, *J* = 17.4, 2.7 Hz, 1H), 2.09

(t, J = 2.7 Hz, 1H), 1.05 (t, J = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 196.0, 171.4, 138.9, 137.6, 137.3, 132.5, 130.1, 128.4, 128.32, 128.27, 127.4, 117.6, 79.3, 72.9, 64.0, 61.7, 52.5, 25.7, 13.8; IR (Neat Film, NaCl) 3288, 3060, 3030, 2981, 1728, 1682, 1597, 1580, 1446, 1255, 1213, 1183, 1046, 1001, 925 cm⁻¹; HRMS (ESI+) *m/z* calc'd for C₂₃H₂₃O₃ [M+H]⁺: 347.1642, found 347.1647; SFC conditions: 5% IPA, 2.5 mL/min, Chiralcel OD-H column, $\lambda = 254$ nm, t_R (min): minor = 5.67, major = 6.44.

Ethyl (E)-2-benzoyl-5-phenyl-2-(prop-2-yn-1-yl)pent-4-enoate (113q).



Ketoester **113q** was isolated by silica gel chromatography (2% EtOAc in hexanes) as a colorless oil. 34% ee, $[\alpha]_D^{25}$ -23.4 (*c* 0.42, CHCl₃); $R_f = 0.3$ (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.88–7.85 (m, 2H), 7.59–7.54 (m, 1H), 7.48–7.42 (m, 2H), 7.30–7.24 (m, 4H), 7.21 (ddd, *J* = 8.7, 4.9, 3.8 Hz, 1H), 6.43 (dt, *J* = 15.6, 1.3 Hz, 1H), 5.90 (dt, *J* = 15.5, 7.7 Hz, 1H), 4.20 (qq, *J* = 7.1, 3.6 Hz, 2H), 3.19 (ddd, *J* = 14.5, 7.9, 1.3 Hz, 1H), 3.11 (ddd, *J* = 14.5, 7.5, 1.4 Hz, 1H), 2.98 (d, *J* = 2.7 Hz, 2H), 2.07 (t, *J* = 2.7 Hz, 1H), 1.13 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 195.2, 171.7, 137.1, 135.7, 134.9, 133.2, 128.8, 128.60, 128.57, 127.6, 126.4, 122.9, 79.0, 72.4, 62.1, 60.6, 36.5, 23.9, 14.1; IR (Neat Film, NaCl) 3287, 3059, 3026, 2979, 2933, 1729, 1679, 1596, 1580, 1446, 1367, 1285, 1270, 1239, 1192, 1180, 1094, 1064, 1023, 966, 937 cm⁻¹; HRMS (ESI+) *m/z* calc'd for C₂₃H₂₃O₃ [M+H]⁺: 347.1642, found 347.1651; SFC

conditions: 5% IPA, 2.5 mL/min, Chiralpak IC column, $\lambda = 254$ nm, t_R (min): minor = 9.81, major = 10.67.

Ethyl (*R*)-2-benzoyl-5-oxo-2-((*S*)-1-phenylallyl)hexanoate (112r).



Ketoester **112r** was isolated by silica gel chromatography (gradient elution, $5\rightarrow10\%$ EtOAc in hexanes) as a colorless oil. 99% ee, $[\alpha]_D^{25}$ +66.8 (*c* 1.20, CHCl₃); $R_f = 0.4$ (25% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.59–7.55 (m, 2H), 7.48–7.43 (m, 1H), 7.37–7.30 (m, 2H), 7.26–7.14 (m, 5H), 6.34 (ddd, J = 17.0, 10.3, 8.1 Hz, 1H), 5.09 (ddd, J = 10.3, 1.7, 1.0 Hz, 1H), 4.97 (ddd, J = 17.0, 1.7, 1.2 Hz, 1H), 4.36–4.32 (m, 1H), 4.13–3.99 (m, 2H), 2.46–2.38 (m, 1H), 2.34–2.29 (m, 2H), 2.25–2.17 (m, 1H), 1.99 (s, 3H), 1.00 (t, J = 7.2 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 206.9, 198.0, 172.4, 139.1, 137.9, 137.8, 132.4, 129.7, 128.4, 128.3, 128.0, 127.3, 117.2, 64.2, 61.3, 53.2, 38.3, 29.9, 28.0, 13.7; IR (Neat Film, NaCl) 3063, 3030, 2981, 2905, 1733, 1717, 1681, 1637, 1596, 1582, 1495, 1446, 1419, 1367, 1243, 1214, 1137, 1093, 1029, 1002, 925, 860, 761 cm⁻¹; HRMS (ESI+) m/z calc'd for C₂₄H₂₇O₃ [M+H]⁺: 379.1904, found 379.1911; SFC conditions: 5% IPA, 2.5 mL/min, Chiralcel OD-H column, $\lambda = 254$ nm, t_R (min): minor = 8.00, major = 10.08.

Ethyl (2R,3S)-2-((1H-indol-3-yl)methyl)-2-benzoyl-3-phenylpent-4-enoate (112s).



Ketoester **112s** was isolated by silica gel chromatography (gradient elution, $5\rightarrow10\%$ EtOAc in hexanes) as a colorless oil. >99% ee, $[\alpha]_D^{25}$ -7.7 (*c* 1.60, CHCl₃); $R_f = 0.4$ (25% EtOAc in hexanes); ¹H NMR (300 MHz, C₆D₆) δ 7.70–7.65 (m, 1H), 7.36 (dd, J = 7.2, 1.8 Hz, 2H), 7.24–7.16 (m, 2H), 7.08–6.93 (m, 5H), 6.90–6.63 (m, 6H), 6.56 (br s, 1H), 5.17–5.12 (m, 1H), 5.10 (t, J = 1.0 Hz, 1H), 4.82 (d, J = 7.8 Hz, 1H), 3.82–3.63 (m, 3H), 3.62–3.45 (m, 1H), 0.40 (td, J = 6.7, 1.3 Hz, 3H); ¹³C NMR (126 MHz, C₆D₆) δ 199.7, 172.2, 139.7, 139.6, 138.9, 135.5, 130.8, 130.2, 128.4, 128.1, 127.9, 127.8, 127.6, 126.9, 124.3, 121.6, 119.7, 119.2, 116.9, 110.5, 66.6, 60.4, 56.0, 31.9, 12.9; IR (Neat Film, NaCl) 3411, 3060, 1724, 1673, 1456, 1241, 1216, 1096, 1012, 923, 747 cm⁻¹; HRMS (ESI+) *m/z* calc'd for C₂₉H₂₈NO₃ [M+H]⁺: 438.2064, found 438.2070; SFC conditions: 10% MeOH, 2.5 mL/min, Chiralpak IC column, $\lambda = 254$ nm, t_R (min): minor = 8.26, major = 9.30.

Ethyl (2*R*,3*S*)-2-benzoyl-2-(2-cyanoethyl)-3-phenylpent-4-enoate (112t) and ethyl (2*S*,3*S*)-2-benzoyl-2-(2-cyanoethyl)-3-phenylpent-4-enoate (112t').

Products **112t** and **112t'** were isolated by silica gel chromatography (5% EtOAc in hexanes) as a mixture of diastereomers (3:1), which were separated by preparative HPLC (20% EtOAc in hexanes).

The major diastereomer **112t** was isolated as a white solid, >99% ee, $[\alpha]_D^{25}$ +59.1 (*c* 1.81, CHCl₃); $R_f = 0.2$ (10% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.52–7.44 (m, 3H), 7.36–7.30 (m, 2H), 7.26–7.20 (m, 3H), 7.15–7.11 (m, 2H), 6.33 (ddd, J = 17.0, 10.2, 8.2 Hz, 1H), 5.14 (ddd, J = 10.2, 1.6, 1.0 Hz, 1H), 5.01 (dt, J = 17.0, 1.3 Hz, 1H), 4.32 (dt, J = 8.0, 1.0 Hz, 1H), 4.15–4.06 (m, 2H), 2.50–2.36 (m, 2H), 2.28 (td, J = 8.0, 0.8 Hz, 2H), 1.01 (t, J = 7.2 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 197.2, 171.8, 138.3, 137.5, 137.1, 132.8, 129.7, 128.7, 128.5, 128.1, 127.7, 119.1, 118.1, 64.0, 61.9, 53.6, 30.6, 13.7, 13.1; IR (Neat Film, NaCl) 3061, 3027, 2981, 2248, 1729, 1675, 1596, 1580, 1446, 1243, 1214, 1185, 1085, 1017, 971, 924, 757 cm⁻¹; HRMS (MM: ESI-APCI+) *m/z* calc'd for C₂₃H₂₄NO₃ [M+H]⁺: 362.1751, found 362.1766; SFC conditions: 5% IPA, 2.5 mL/min, Chiralpak AD-H column, $\lambda = 254$ nm, t_R (min): major = 9.77, minor = 11.61.



The minor diastereomer was isolated as a white solid, >99% ee, $[\alpha]_{D}^{25}$ +22.3 (*c* 0.11, CHCl₃); $R_f = 0.2$ (10% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.69–7.63 (m, 2H), 7.55–7.47 (m, 1H), 7.41–7.32 (m, 2H), 7.31–7.22 (m, 4H), 7.22–7.16 (m, 1H), 6.29 (dt, *J* = 16.8, 10.2 Hz, 1H), 5.21–5.11 (m, 2H), 4.35 (d, *J* = 10.3 Hz, 1H), 4.03 (dq, *J* = 10.8, 7.2 Hz, 1H), 3.84 (dq, *J* = 10.8, 7.2 Hz, 1H), 2.74–2.63 (m, 1H), 2.46–2.28 (m, 2H), 2.14–2.02 (m, 1H), 0.81 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 196.0, 171.3, 139.7, 136.7, 136.0, 133.0, 129.7, 128.7, 128.5, 128.3, 127.4, 119.2, 118.9, 63.8, 61.7, 53.0, 30.7, 13.3, 12.9; IR (Neat Film, NaCl) 3062, 3029, 2982, 2248, 1728, 1678, 1596, 1580, 1494, 1446, 1367, 1275, 1242, 1215, 1093, 1018, 972, 928, 757 cm⁻¹; HRMS (MM: ESI-APCI+) *m/z* calc'd for C₂₃H₂₄NO₃ [M+H]⁺: 362.1751, found 362.1757; SFC conditions: 5% IPA, 2.5 mL/min, Chiralpak AD-H column, $\lambda = 254$ nm, t_R (min): major = 10.65, minor = 12.02.

Ethyl (25,3S)-2-benzoyl-2-fluoro-3-phenylpent-4-enoate (112u).



Ketoester **112u** was isolated by silica gel chromatography (gradient elution, $1\rightarrow 2\%$ EtOAc in hexanes) as a colorless oil. 95% ee, $[\alpha]_D^{25}$ +82.0 (*c* 1.19, CHCl₃); $R_f = 0.4$ (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.77 (ddt, J = 7.6, 2.0, 1.2 Hz, 2H), 7.49 (ddt, J = 7.7, 7.0, 1.3 Hz, 1H), 7.38–7.30 (m, 4H), 7.31–7.14 (m, 3H), 6.20 (ddd, J =17.0, 10.2, 9.4 Hz, 1H), 5.27 (ddt, J = 17.0, 1.5, 0.8 Hz, 1H), 5.23 (ddd, J = 10.1, 1.5, 0.6 Hz, 1H), 4.65 (dd, J = 33.3 (J_{H-F}), 9.4 Hz, 1H), 4.31 (ddq, J = 39.5, 10.8, 7.2 Hz, 2H), 1.30 (t, J = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 192.6 (d, J = 27.1 Hz), 166.3 (d, J = 26.4 Hz), 137.4, 134.9 (d, J = 3.5 Hz), 134.5 (d, J = 4.0 Hz), 133.6, 129.9 (d, J = 2.6Hz), 129.5 (d, J = 6.7 Hz), 128.5, 128.4, 127.4, 119.1, 103.1 (d, J = 208.4 Hz), 63.1, 54.4 (d, J = 18.3 Hz), 14.2; ¹⁹F NMR (282 MHz, CDCl₃) δ -169.39 (d, J = 33.2 Hz); IR (Neat Film, NaCl) 3062, 3030, 2982, 2934, 1749, 1694, 1597, 1454, 1447, 1367, 1230, 1187, 1129, 1038, 929, 856, 743 cm⁻¹; HRMS (ESI+) m/z calc'd for C₂₀H₂₀FO₃ [M+H]⁺: 327.1391, found 327.1401; SFC conditions: 5% IPA, 4.0 mL/min, Chiralpak IC column, $\lambda = 254$ nm, t_R (min): major = 2.02, minor = 3.18.

Ethyl (2*S*,3*S*)-2-benzoyl-2-chloro-3-phenylpent-4-enoate (112v).



Ketoester **112v** was isolated by silica gel chromatography (gradient elution, $2\rightarrow 3\%$ EtOAc in hexanes) as a colorless oil. >99% ee, $[\alpha]_D^{25}$ +93.8 (*c* 1.43, CHCl₃); $R_f = 0.4$ (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.84–7.78 (m, 2H), 7.54–7.47 (m, 1H), 7.47–7.40 (m, 2H), 7.40–7.33 (m, 2H), 7.33–7.18 (m, 3H), 6.36 (ddd, *J* = 16.9, 10.2, 8.7 Hz, 1H), 5.20 (ddd, *J* = 10.2, 1.5, 0.8 Hz, 1H), 5.15 (ddd, *J* = 16.9, 1.6, 1.0 Hz, 1H), 4.66 (d, *J* = 8.6 Hz, 1H), 4.18 (q, *J* = 7.1 Hz, 2H), 1.12 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 189.8, 167.3, 138.0, 135.9, 134.8, 133.1, 130.7, 129.3, 128.3, 128.0, 127.5, 118.7, 77.3, 63.4, 55.8, 13.8; IR (Neat Film, NaCl) 3062, 3029, 2982, 1754, 1696,

1597, 1581, 1446, 1367, 1299, 1239, 1207, 1186, 1094, 1075, 1025, 1007, 928, 749 cm⁻¹; HRMS (ESI+) *m/z* calc'd for C₂₀H₂₀ClO₃ [M+H]⁺: 343.1095, found 343.1099; SFC conditions: 10% MeOH, 2.5 mL/min, Chiralpak IC column, $\lambda = 254$ nm, t_R (min): major = 2.52, minor = 2.77.

Ethyl (2*R*,3*S*)-2-(cyclohex-1-ene-1-carbonyl)-2-methyl-3-phenylpent-4-enoate (112w).



Ketoester **112w** was isolated by silica gel chromatography (1% EtOAc in hexanes) as a colorless oil. 99% ee, $[\alpha]_D^{25}$ +80.8 (*c* 1.00, CHCl₃,); R_f = 0.3 (10% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.33–7.13 (m, 5H), 6.47 (td, *J* = 3.9, 2.0 Hz, 1H), 6.25 (ddd, *J* = 16.8, 10.3, 8.0 Hz, 1H), 5.09 (dt, *J* = 10.2, 1.4 Hz, 1H), 5.00 (dt, *J* = 16.9, 1.4 Hz, 1H), 4.29 (dd, *J* = 8.0, 1.4 Hz, 1H), 3.68 (d, *J* = 0.8 Hz, 3H), 2.24–2.03 (m, 3H), 1.97–1.81 (m, 1H), 1.55 (tq, *J* = 7.0, 4.0, 2.3 Hz, 4H), 1.43 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 198.1, 173.8, 139.4, 138.3, 137.9, 137.2, 130.0, 128.0, 126.8, 117.1, 61.1, 54.5, 52.2, 25.9, 24.3, 22.0, 21.3, 20.7; IR (Neat Film, NaCl) 2938, 1732, 1671, 1636, 1452, 1433, 1234, 1113, 984, 917, 703 cm⁻¹; HRMS (ESI+) *m/z* calc'd for C₂₀H₂₅O₃ [M+H]⁺: 313.1798, found 313.1785; SFC conditions: 5% MeOH, 4.0 mL/min, Chiralpak IC column, $\lambda = 254$ nm, t_R (min): minor = 5.83, major = 6.55.

Methyl (2*R*,3*S*)-2-(cyclohexanecarbonyl)-2-methyl-3-phenylpent-4-enoate (112x).



Ketoester **112x** was isolated by silica gel chromatography (5% EtOAc in hexanes) as a mixture of diastereomers (4:1 dr). *For the major isomer*: 96% ee, $[\alpha]_D^{25}$ +57.7 (*c* 1.54, CHCl₃, measured with 4:1 dr mixture); R_f = 0.3 (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.29–7.18 (m, 5H), 6.26 (ddd, *J* = 17.0, 10.2, 8.5 Hz, 1H), 5.08 (ddd, *J* = 10.2, 1.8, 0.9 Hz, 1H), 5.00 (ddd, *J* = 17.0, 1.8, 1.1 Hz, 1H), 4.18 (dt, *J* = 8.5, 1.0 Hz, 1H), 3.71 (s, 3H), 2.50 (tt, *J* = 11.4, 3.2 Hz, 1H), 1.79–1.56 (m, 5H), 1.53–1.46 (m, 1H), 1.33 (s, 3H), 1.32–1.11 (m, 4H); ¹³C NMR (126 MHz, CDCl₃) δ 210.3, 172.4, 139.6, 137.8, 129.9, 128.3, 127.0, 117.4, 64.7, 54.1, 52.2, 48.4, 30.3, 29.6, 25.8, 25.73, 25.71, 18.7; IR (Neat Film, NaCl) 2933, 2854, 1729, 1708, 1635, 1495, 1453, 1432, 1380, 1315, 1228, 1144, 1101, 989, 919 cm⁻¹; HRMS (ESI+) *m/z* calc'd for C₂₀H₂₆O₃ [M+H]⁺: 315.1955, found 315.1955; SFC conditions: 100% CO₂, 4.0 mL/min, Chiralcel OJ-H column, $\lambda = 210$ nm, t_R (min): minor = 7.28, major = 7.96.

Methyl (2*R*,3*S*)-2-acetyl-2-ethyl-3-phenylpent-4-enoate (112y) and methyl (2*S*,3*S*)-2acetyl-2-ethyl-3-phenylpent-4-enoate (112y').

Ketoesters **112y** and **112y'** were isolated by silica gel chromatography (gradient elution, $0 \rightarrow 4\%$ EtOAc in hexanes) as mixture of two diastereomers (1.5:1 dr). The diastereomers were separated by preparative HPLC (gradient elution, $60 \rightarrow 70\%$ MeCN in H₂O).



The major diastereomer **112y** was isolated as a colorless oil. 90% ee, $[\alpha]_D^{25}$ +72.7 (*c* 0.60, CHCl₃); $R_f = 0.4$ (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.32–7.25 (m, 2H), 7.25–7.18 (m, 1H), 7.17–7.07 (m, 2H), 6.28 (ddd, *J* = 17.0, 10.2, 8.4 Hz, 1H), 5.08 (ddd, *J* = 10.2, 1.7, 0.9 Hz, 1H), 4.97 (ddd, *J* = 17.0, 1.7, 1.1 Hz, 1H), 4.03 (d, *J* = 8.5 Hz, 1H), 3.76 (s, 3H), 2.06 (s, 3H), 1.86–1.68 (m, 2H), 0.77 (t, *J* = 7.5 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 205.8, 172.4, 139.5, 137.7, 129.3, 128.4, 127.3, 117.3, 68.2, 53.5, 51.9, 29.8, 27.4, 9.1; IR (Neat Film, NaCl) 3083, 3029, 2978, 2950, 2883, 1709, 1601, 1494, 1434, 1385, 1353, 1301, 1220, 1116, 1029, 993, 919, 755 cm⁻¹; HRMS (MM: ESI-APCI+) *m*/z calc'd for C₁₆H₂₁O₃ [M+H]⁺: 261.1485, found 261.1492; SFC conditions: 0.5% IPA, 2.5 mL/min, Chiralcel OJ-H column, λ = 210 nm, t_R (min): minor = 4.79, major = 7.02.



The minor diastereomer **112y'** was isolated as a colorless oil. 91% ee, $[\alpha]_D^{25}$ +25.3 (*c* 0.26, CHCl₃); $R_f = 0.4$ (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.26–7.23 (m, 2H), 7.21–7.17 (m, 1H), 7.17–7.13 (m, 2H), 6.31 (ddd, J = 16.9, 10.2, 9.3 Hz, 1H), 5.15 (ddd, J = 10.2, 1.6, 0.7 Hz, 1H), 5.10 (ddd, J = 17.0, 1.7, 1.0 Hz, 1H), 4.11 (d, J = 9.4 Hz, 1H), 3.63 (s, 3H), 2.12 (s, 3H), 2.09–1.98 (m, 1H), 1.80 (dq, J = 14.8, 7.5 Hz, 1H), 0.77 (t, J = 7.5 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 205.3, 172.3, 140.1, 137.3,

129.4, 128.2, 127.0, 117.6, 68.0, 52.5, 51.9, 30.1, 26.9, 8.8; IR (Neat Film, NaCl) 3063, 3030, 2962, 2925, 2850, 1710, 1600, 1446, 1354, 1286, 1260, 1223, 1182, 1118, 1095, 1023, 921, 864, 801 cm⁻¹; HRMS (MM: ESI-APCI+) *m/z* calc'd for C₁₆H₂₁O₃ [M+H]⁺: 261.1485, found 261.1494; SFC conditions: 0.5% IPA, 2.5 mL/min, Chiralcel OJ-H column, $\lambda = 210$ nm, t_R (min): minor = 6.29, major = 7.02.

tert-Butyl (2*R*,3*S*)-2-benzoyl-2-methyl-3-phenylpent-4-enoate (112z).



Ketoester **112z** was isolated by silica gel chromatography (gradient elution, $0\rightarrow 2\rightarrow 5\%$ EtOAc in hexanes) as a colorless oil. >99% ee, $[\alpha]_D^{25}$ +67.4 (*c* 1.54, CHCl₃); $R_f = 0.4$ (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.74–7.70 (m, 2H), 7.49–7.46 (m, 1H), 7.39–7.34 (m, 2H), 7.25–7.16 (m, 5H), 6.34 (ddd, *J* = 17.0, 10.3, 7.9 Hz, 1H), 5.12 (ddd, *J* = 10.3, 1.8, 1.1 Hz, 1H), 5.03 (dt, *J* = 17.0, 1.6 Hz, 1H), 4.42 (dt, *J* = 7.9, 1.2 Hz, 1H), 1.50 (s, 3H), 1.29 (s, 9H); ¹³C NMR (126 MHz, CDCl₃) δ 198.1, 171.8, 139.7, 138.1, 137.3, 132.2, 130.4, 128.5, 128.3, 128.1, 126.9, 117.1, 82.6, 62.1, 54.6, 27.8, 21.0; IR (Neat Film, NaCl) 3061, 3027, 2978, 2934, 1728, 1716, 1687, 1682, 1598, 1454, 1446, 1251, 1155, 1115, 961, 918, 844 cm⁻¹; HRMS (ESI+) *m/z* calc'd for C₂₃H₂₇O₃ [M+H]⁺: 351.1955, found 351.1955; SFC conditions: 5% IPA, 2.5 mL/min, Chiralpak IC column, $\lambda = 254$ nm, t_R (min): major = 4.14, minor = 5.80.

tert-Butyl (E)-2-benzoyl-2-methyl-5-phenylpent-4-enoate (113z).

Ph
$$Me CO_2 t$$
-Bu 113z

Ketoester **113z** was isolated by silica gel chromatography (gradient elution, $0\rightarrow 2\rightarrow 5\%$ EtOAc in hexanes) as a colorless oil. 38% ee, $[\alpha]_D^{25}$ +3.9 (*c* 0.74, CHCl₃); $R_f = 0.4$ (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.92–7.89 (m, 2H), 7.56–7.51 (m, 1H), 7.46–7.41 (m, 2H), 7.33–7.27 (m, 4H), 7.23–7.18 (m, 1H), 6.39 (dt, *J* = 15.8, 1.3 Hz, 1H), 6.16–6.06 (m, 1H), 2.93 (ddd, *J* = 14.2, 7.5, 1.4 Hz, 1H), 2.84 (ddd, *J* = 14.2, 7.7, 1.3 Hz, 1H), 1.55 (s, 3H), 1.28 (s, 9H); ¹³C NMR (126 MHz, CDCl₃) δ 197.8, 172.8, 137.3, 135.9, 133.9, 132.7, 128.8, 128.6, 128.5, 127.4, 126.3, 124.7, 82.2, 58.0, 40.7, 27.8, 21.5; IR (Neat Film, NaCl) 3058, 3026, 2977, 2933, 1728, 1686, 1682, 1597, 1579, 1447, 1368, 1249, 1212, 1152, 1111, 970, 845 cm⁻¹; HRMS (ESI+) *m/z* calc'd for C₂₃H₂₇O₃ [M+H]⁺: 351.1955, found 351.1949; SFC conditions: 5% IPA, 2.5 mL/min, Chiralpak IC column, $\lambda = 254$ nm, t_R (min); major = 9.88, minor = 8.48.

4.6.5 Procedures for derivatization of allylic alkylation products and spectroscopic data of derivatives

Synthesis of cyclohexenone 114:



To a solution of ketoester 112r (60.4 mg, 0.6 mmol) in t-BuOMe (2 mL) was added pyrrolidine (13.7 mg, 0.19 mmol) and AcOH (11.6 mg, 0.19 mmol). The mixture was stirred for 12 h at 25 °C then heated to reflux for 4 h. The solvent was removed under reduced pressure and the residue was subjected to column chromatography on silica gel (gradient elution, $10 \rightarrow 25\%$ EtOAc in hexanes) to give cyclohexenone 114 (55.1 mg, 95% yield) as a colorless oil. $[\alpha]_{D}^{25}$ +151.4 (c 1.19, CHCl₃); R_f = 0.5 (25% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.34-7.28 (m, 1H), 7.25-7.19 (m, 2H), 7.13-7.04 (m, 3H), 7.04–6.97 (m, 2H), 6.69–6.63 (m, 2H), 6.33 (ddd, J = 16.9, 10.2, 7.8 Hz, 1H), 6.07 (s, 1H), 5.10 (ddd, J = 10.2, 1.6, 1.0 Hz, 1H), 4.87 (dt, J = 17.0, 1.5 Hz, 1H), 4.31 (q, J = 7.1 Hz, 2H), 4.26 (dt, J = 7.5, 1.5 Hz 1H), 2.91–2.77 (m, 1H), 2.62–2.53 (m, 3H), 1.32 (t, J = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 198.5, 173.2, 161.6, 139.5, 139.2, 138.0, 131.9, 129.8, 128.7, 128.2, 128.1, 128.0, 126.7, 118.1, 62.0, 54.0, 53.8, 34.7, 30.9, 14.3; IR (Neat Film, NaCl) 3058, 3030, 2981, 1725, 1673, 1602, 1492, 1444, 1326, 1240, 1214, 1170, 1016, 921, 882 cm⁻¹; HRMS (ESI+) m/z calc'd for C₂₄H₂₅O₃ [M+H]⁺: 361.1798, found 361.1798.

Synthesis of bicyclic enone 116:



A dried flask was charged with a solution of envne 112q (8.6 mg, 0.025 mmol) and Co₂(CO)₈ (11.8 mg, 0.034 mmol) in CH₂Cl₂ (2 mL) and the mixture was stirred at room temperature for 12 h under an atmosphere of argon. After full consumption of **112g** was observed by TLC analysis, Me₃NO•2H₂O (7.6 mg, 0.068 mmol) was added. The mixture was stirred for 20 min and an additional portion of Me₃NO•2H₂O (30.1 mg, 0.27 mmol) was added. Stirring was continued until complete consumption of the cobalt-alkyne complex was observed by TLC analysis (about 4 h). The mixture was filtered through a celite pad, washed with CH₂Cl₂, the solvent removed under reduced pressure, and the residue subjected to column chromatography on silica gel (25% EtOAc in hexanes) to give the bicyclic enone 116 (9.3 mg, 99% yield) as a colorless oil. The relative stereochemistry of **116** was assigned by 2D-NOESY. $[\alpha]_D^{25}$ -178.8 (c 0.79, CHCl₃); $R_f =$ 0.3 (25% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.33–7.26 (m, 1H), 7.26– 7.18 (m, 4H), 7.13–7.02 (m, 5H), 6.03 (td, J = 2.2, 1.1 Hz, 1H), 4.11–3.91 (m, 3H), 3.82 (d, J = 12.3 Hz, 1H), 3.73-3.63 (m, 1H), 3.01 (dq, J = 18.2, 1.1 Hz, 1H), 2.54 (ddd, J = 1.1 Hz, 1H)18.0, 6.4, 0.8 Hz, 1H), 2.14 (ddd, J = 18.0, 2.5, 1.7 Hz, 1H), 0.86 (t, J = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 209.3, 197.1, 183.8, 173.3, 137.1, 136.7, 132.4, 129.5, 128.5, 128.3, 127.9, 127.8, 126.1, 68.6, 62.0, 55.8, 50.4, 41.7, 39.2, 13.6; IR (Neat Film, NaCl) 3034, 2979, 2927, 1733, 1714, 1668, 1636, 1600, 1583, 1449, 1409, 1255, 1211, 1183,

1071, 1043, 927, 914, 819 cm⁻¹; HRMS (MM: ESI-APCI+) m/z calc'd for C₂₄H₂₃O₄ [M+H]⁺: 375.1596, found 375.1592.

Synthesis of compound 115:



To a flame-dried Schlenk flask was added a solution of **112p** (20.8 mg, 0.06 mmol in 1.5 mL of CH₂Cl₂) and Hoveyda-Grubbs II catalyst (3.7 mg, 10 mol%). The reaction mixture was stirred for 3 h at 40 °C, filtered through a short silica pad and purified by silica gel chromatography (gradient elution, $1\rightarrow5\%$ EtOAc in hexanes) to give ketoester **115** (18.3 mg, 96% yield) as a white solid. $[\alpha]_D^{25}$ -613.7 (*c* 0.94, CHCl₃); $R_f = 0.4$ (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.61–7.55 (m, 2H), 7.38 (ddt, J = 8.6, 7.1, 1.2 Hz, 1H), 7.28–7.21 (m, 2H), 6.97–6.93 (m, 3H), 6.89–6.83 (m, 2H), 5.90 (ddt, J= 6.1, 2.6, 1.8 Hz, 1H), 5.75 (dtd, J = 5.5, 2.6, 1.5 Hz, 1H), 4.92 (q, J = 2.3 Hz, 1H), 4.11 (q, J = 7.1 Hz, 2H), 4.03 (dtd, J = 18.0, 2.6, 1.9 Hz, 1H), 2.73 (ddd, J = 18.0, 2.6, 1.6 Hz, 1H), 1.00 (t, J = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 194.2, 174.4, 138.4, 136.6, 133.8, 132.5, 129.5, 128.5, 128.1, 128.0, 127.4, 127.0, 68.3, 61.9, 58.0, 41.5, 13.8; IR (Neat Film, NaCl) 3060, 3028, 2959, 2932, 2871, 1736, 1732, 1686, 1682, 1598, 1582, 1492, 1447, 1365, 1258, 1243, 1220, 1157, 1087, 1048, 1004, 966, 922, 876, 761 cm⁻¹; HRMS (MM: ESI-APCI+) *m/z* calc'd for C₂₁H₂₁O₃ [M+H]⁺: 321.1485, found 321.1489.

4.6.6 Determination of the absolute confirmation of compound 112f



To a flame-dried flask was added **112f** (98% ee, 62.1 mg, 0.16 mmol), CH₂Cl₂ (5 mL) and this solution was cooled to -78 °C. DIBAL-H (0.62 mL, 1.0 M solution in hexane) was added dropwise by syringe. The mixture was stirred for 2 h at -78 °C, then allowed to warm to 25 °C and stirred for an additional 12 h. The reaction was then cooled to 0 °C, and another 0.62 mL of DIBAL-H solution was added, followed by stirring at 25 °C for 3 h. The reaction mixture was then guenched with saturated aqueous Rochelle's salt (20 mL) and stirred for another 3 h. The aqueous layer was partitioned with a total of 100 mL of CH₂Cl₂ and the combined organic phases washed with brine, dried over MgSO₄, filtered and concentrated under reduced pressure. The crude product was purified by silica gel chromatography (gradient elution, $5 \rightarrow 20\%$ EtOAc in hexanes) to give 40.2 mg (72% yield) of **117** as white solid mixture (1:1 dr). The diastereomers were separated by preparative HPLC (gradient elution, $60 \rightarrow 90\%$ MeCN in H₂O). For isomer a: white solid, $[\alpha]_{D}^{25}$ +58.9 (c 0.47, CHCl₃); R_f = 0.4 (25% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.46–7.39 (m, 4H), 7.36–7.31 (m, 2H), 7.31–7.27 (m, 1H), 7.27–7.23 (m, 2H), 6.79 (dt, J = 16.9, 10.1 Hz, 1H), 5.20–5.10 (m, 2H), 5.07 (s, 1H), 3.67 (d, J = 10.1 Hz, 1H), 3.34 (d, J = 11.2 Hz, 1H), 3.11 (dd, J = 11.1, 0.7 Hz, 1H), 1.56 (br s, 2H), 0.72 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 141.5, 141.0, 140.4, 131.6, 131.4, 128.04, 128.79,

127.8, 120.5, 116.5, 76.0, 66.6, 55.8, 46.2, 15.3; IR (Neat Film, NaCl) 3423, 3070, 2969, 2923, 1486, 1452, 1403, 1342, 1074, 1012, 916, 827 cm⁻¹; HRMS (ESI+) *m/z* calc'd for fragment $C_{19}H_{18}Br$ [M-H₄O₂+H]⁺: 325.0586, found 325.0585. *For isomer b*: white solid, $[\alpha]_{D}^{25}$ +53.4 (*c* 0.43, CHCl₃); $R_f = 0.4$ (24% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.50–7.45 (m, 2H), 7.35–7.27 (m, 5H), 7.25–7.20 (m, 2H), 6.30 (dt, *J* = 16.8, 10.0 Hz, 1H), 5.35 (ddd, *J* = 16.8, 1.9, 0.8 Hz, 1H), 5.24 (ddd, *J* = 10.0, 1.9, 0.5 Hz, 1H), 4.39 (d, *J* = 10.0 Hz, 1H), 4.26 (s, 1H), 3.64 (d, *J* = 11.6 Hz, 1H), 3.51 (dd, *J* = 11.6, 1.8 Hz, 1H), 1.58 (br s, 2H), 0.48 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 141.0, 140.2, 137.2, 131.7, 131.3, 128.02, 127.97, 127.8, 120.6, 118.3, 80.0, 65.6, 49.7, 44.5, 15.5; IR (Neat Film, NaCl) 3372, 2973, 2938, 2888, 1637, 1486, 1473, 1454, 1402, 1348, 1266, 1203, 1101, 1077, 1024, 1010, 921, 894, 831, 782 cm⁻¹; HRMS (ESI+) *m/z* calc'd for fragment $C_{19}H_{18}Br$ [M-H₄O₂+H]⁺: 325.0586, found 325.0588.

4.6.7 Determination of enantiomeric excess

entry	compound	SFC analytic conditions	ee (%)
1	Ph Ph Me [°] CO ₂ Et 112a	Chiralpak IC, λ = 254 nm 5% IPA/CO ₂ , 2.5 mL/min t _R (min): major 4.12, minor 6.14	98
2	Ph Me Me Co_2Et 112b	Chiralpak IC, λ = 254 nm 5% IPA/CO ₂ , 2.5 mL/min t _R (min): major 6.71, minor 9.45	>99
3	Ph Me [°] CO ₂ Et 112c	Chiralcel OJ-H, λ = 254 nm 4% IPA/CO ₂ , 2.5 mL/min t _R (min): minor 5.64, major 6.84	>99
4	Ph Me ⁱ CO ₂ Et 112d	Chiralpak IC, λ = 254 nm 2% MeOH/CO ₂ , 2.5 mL/min t _R (min): major 12.24, minor 13.50	>99
5	Ph Cl Me ^{CO_2Et} 112e	Chiralpak IC, λ = 254 nm 10% IPA/CO ₂ , 4.0 mL/min t _R (min): major 1.59, minor 1.77	99
6	Ph Me ^S CO ₂ Et 112f	Chiralpak AD-H, λ = 254 nm 10% IPA/CO ₂ , 4.0 mL/min t _R (min): minor 2.95, major 3.17	>99
7	Ph CF_3 Ph CO_2Et 112g	Chiralpak AD-H, λ = 254 nm 2% IPA/CO ₂ , 2.5 mL/min t _R (min): minor 5.20, major 6.68	>99

Table 4.6.7.1. Determination of enantiomeric excess

entry	compound	SFC analytic conditions	ee (%)
8	Ph Me [°] CO ₂ Et 112h	Chiralpak AD-H, λ = 254 nm 5% MeOH/CO₂, 3.0 mL/min t _R (min): minor 4.47, major 5.71	93
9	$Ph \underbrace{Me}^{O}_{CO_2Et} \underbrace{NO_2}_{113h}$	Chiralpak AD-H, λ = 254 nm 10% MeOH/CO ₂ , 3.0 mL/min t _R (min): major 7.13, minor 8.06	23
10	Ph Me [°] CO ₂ Et 112i	Chiralpak IC, λ = 254 nm 5% IPA/CO ₂ , 2.5 mL/min t _R (min): major 7.92, mino 11.24	95
11	Ph Me [*] CO ₂ Et 112j	Chiralpak IC, λ = 254 nm 5% IPA/CO ₂ , 2.5 mL/min t _R (min): major 5.09, minor 9.14	>99
12	Ph Me Me [°] CO ₂ Et 112k	Chiralpak AD-H, λ = 210 nm 2% IPA/CO $_2$, 2.5 mL/min t _R (min): minor 4.40, major 5.52	91
13	MeO Ph Me ^C CO ₂ Et	Chiralpak IC, λ = 254 nm 10% IPA/CO ₂ , 4.0 mL/min t _R (min): major 2.67, minor 3.51	99
14	$H2I$ Ph $He^{2}CO_{2}Me$ $112m$	Chiralpak AD-H, λ = 254 nm 5% IPA/CO2, 2.5 mL/min $t_{\rm R}$ (min): major 5.35, minor = 5.88	>99
15	$Ph \xrightarrow{\text{O} Ph}_{\text{Et}} CO_2 Et$	Chiralpak AD-H, λ = 254 nm, 5% MeOH/CO $_2$, 2.5 mL/min, $t_{\rm R}$ (min): major = 2.48, minor = 2.20	>99

entry	compound	SFC analytic conditions	ee (%)	
16	$Ph \xrightarrow{Ph}_{Bn^{*} CO_{2}Et}$ 1120	Chiralpak AD-H, λ = 254 nm 4% MeOH/CO ₂ , 4.0 mL/min t _R (min): major 5.06, minor 8.94	>99	
17	Ph Ph CO ₂ Et	Chiralpak IC, λ = 254 nm 3% IPA/CO ₂ , 4.0 mL/min t _R (min): major 7.16, minor 8.60	>99	
18	Ph Ph CO ₂ Et	Chiralcel OD-H, λ = 254 nm 5% IPA/CO ₂ , 2.5 mL/min t _R (min): minor 5.67, major 6.44	>99	
19	Ph CO ₂ Et	Chiralpak IC, λ = 254 nm 5% IPA/CO ₂ , 2.5 mL/min t _R (min): major 10.67, minor 9.81	34	
20	$Ph \xrightarrow{O \qquad Ph}_{CO_2Et}$ $O = 112r$	Chiralcel OD-H, λ = 254 nm 5% IPA/CO ₂ , 2.5 mL/min t _R (min): minor 10.8, major 8.0	99	
21	Ph Ph CO ₂ Et NH 112s	Chiralpak IC, λ = 254 nm 10% MeOH/CO₂, 2.5 mL/min t _R (min): major 8.26, minor 9.30	>99	
22	Ph Ph CO ₂ Et NC 112t	Chiralpak AD-H, λ = 254 nm 5% IPA/CO ₂ , 2.5 mL/min t _R (min): major 9.77, minor 11.60	>99	
23	Ph Ph NC	Chiralpak AD-H, λ = 254 nm 5% IPA/CO ₂ , 2.5 mL/min t _R (min): major 10.65, minor 12.00	>99	

entry	compound	SFC analytic conditions	ee (%
24	$Ph \xrightarrow{O Ph}_{F CO_2Et}$	Chiralpak IC, λ = 254 nm 5% IPA/CO ₂ , 4.0 mL/min t _R (min): major 2.02, minor 3.18	95
25	$Ph \xrightarrow{O Ph}_{Cl} Co_2Et$ $112v$	Chiralpak IC, λ = 254 nm 10% IPA/CO ₂ , 2.5 mL/min t _R (min): major 2.52, minor 2.76	>99
26	O Ph Me ^v CO ₂ Me	Chiralpak IC, λ = 254 nm 5% MeOH/CO ₂ , 4 mL/min t _R (min): major 6.56, minor 5.83	99
27	O Ph Me [°] CO ₂ Me 112x	Chiralcel OJ-H, λ = 210 nm 100% CO ₂ , 4.0 mL/min t _R (min): minor 7.28, major 7.96	96
28	$Me \xrightarrow{\text{CO}_2Me} Determines} Determines Det$	Chiralcel OJ-H, λ = 210 nm 0.5% IPA/CO ₂ , 2.5 mL/min t _R (min): minor 4.79, major 5.33	90
29	$Me \xrightarrow{\text{Ph}}_{\text{Et}} CO_2Me$	Chiralcel OJ-H, λ = 210 nm 0.5% IPA/CO ₂ , 2.5 mL/min t _R (min): minor 6.29, major 7.02	91
30	Ph Ph Me ⁱ CO ₂ t-Bu 112z	Chiralpak IC, λ = 254 nm 5% IPA/CO ₂ , 2.5 mL/min t _R (min): major 4.14, minor 5.80	>99
31		Chiralpak IC, $\lambda = 254$ nm 5% IPA/CO ₂ , 2.5 mL/min	38

4.7 REFERENCES AND NOTES

- (97) For the first example see reference 75; for reviews see references 76.
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