The development of RNA-based control systems to regulate signaling and dictate cell fate in a model MAPK pathway

Thesis by Kate Elizabeth Galloway

In Partial Fulfillment of the Requirements for the Degree of Doctor of Philosophy



California Institute of Technology Pasadena, California 2012 (Defended May 18, 2012)

© 2012 Kate Elizabeth Galloway All Rights Reserved

Acknowledgments

This thesis has been seven years in the making. It represents not only my efforts, but also the efforts of many people who have provided valuable academic, professional, and technical assistance and guidance, not to mention personal support. While this list is not exhaustive, I would like to particularly thank the people below for their much appreciated help.

I am so very grateful to my advisor, Prof. Christina Smolke, who has been a great mentor and inspiration to me. Since I arrived in Christina's lab in early 2006, I have been impressed by her diligence in research and commitment to bringing out the best in her students. Christina's tireless help with editing this manuscript has given me a much deeper appreciation of the nuances of research and the power of language. In my experience, it is rare to find someone who is so excellent in her own work and simultaneously so patient with the development of others. When I think about Christina, I recall a conversation we had in late 2008, only six weeks before our lab's scheduled move to Stanford. I walked into Christina's office to inform her that I couldn't join the lab in moving to Stanford as I had just received the surprising news that I was pregnant, and I needed to stay in Pasadena to keep my family together. Christina never expressed any sentiment other than her concern and unwavering commitment to help me figure out a way to finish my Ph.D. at Caltech. She helped me arrange to remain at Caltech and organized grants and other funding so that I could finish my research here. Additionally, she has spent extra hours on the side coordinating my benefits and pay and meeting both via Skype and in person to continue guiding my work. Throughout all of these challenges, Christina has continued to support my development as a scientist and has

given me great flexibility to be around for my family. I am so grateful to have had Christina as my advisor and for her compassion and guidance throughout the past six years.

I would also like to thank my committee: Prof. Michael Elowitz, Prof. Richard Murray, and Prof. Dave Tirrell. I am particularly thankful to Michael for allowing me to use his lab's Quanta flow cytometer, which was critical for characterizing my system. When the Quanta needed to be moved, Michael willingly released it to its new home in the Murray lab. I am so thankful to Richard for coordinating and financing the move of the Quanta as well as providing space in his lab for it. Thanks to Dave for his willingness to help when I needed equipment in his lab and for serving as the chair of my thesis committee.

The Smolke lab has been a wonderful group of friends and colleagues. I am particularly grateful to Dr. Joe Liang for many encouraging discussions via Google Chat, his efforts as the Stanford-to-Caltech supply train, and his excellent work in developing more stringent switches. My work would not be possible without the new methods and switches that he developed. I am also thankful to Dr. Josh Michener for our biocontrol discussions and for his efforts to keep me awake in seminars. I will remember Leo d'Espaux for modeling work-life balance, practicing the beach entry for the Hermosa Beach triathlon, and sharing his stories of growing up in Cuba, including the one about stroking Fidel Castro's beard as a child. To Dr. Yvonne Chen: I am thankful for your friendship and unparalleled model of diligence and efficiency. I am excited to read about the work that will be coming out of your lab starting in 2013.

The Smolke lab move to Stanford in 2009 gave me the privilege of being adopted by a number of labs over the last several years. I am so thankful to Dr. Anand Asthagiri and the Asthagiri lab for providing me a lab home for two years. I am especially thankful to Dr. Steve Chapman for teaching me his Western blot protocol and sharing his stocks with me. I have been more recently adopted by Dr. Frances Arnold's lab and the German contingent of postdocs. Thanks to Dr. Sabine Bastian for her thoughtfulness in helping me maintain a bench through all the remodeling chaos on the second floor and overcrowding upstairs. I will also remember the reliable Ernie's lunch bunch, Dr. Kersten Rabe, Dr. Ryan Lauchli, Dr. Sebastian Schoof, Dr. Martin Engqvist, and Chris Farwell for our varied discussions ranging from new cloning methods to global politics. The last few years have found me increasingly borrowing time, space, and equipment across the campus. Thanks to the Tirrell, Elowitz, and Murray labs for being welcoming and friendly when I came to borrow things.

To the biocontrol reading group, I am grateful not only for our meetings which broadened my appreciation of synthetic biology, systems biology and modeling, but also for the wonderful people who have contributed so much to me both professionally and personally. I am especially grateful to Dr. Elisa Franco for her help in providing a sounding board for ideas about network architecture that changed my approach to Chapter 3. I am also thankful for her generosity in helping me to develop a model of my system. Thanks also to Ophelia Venturelli for the interesting discussions on all things yeast and particularly the galactose pathway. I look forward to hearing the final word on the systems biology of the galactose network response.

v

There is a lot of work that goes into administering the grad program at Caltech. I want to say thank you to the people who work behind the scenes on these critical tasks. I am particularly grateful to Kathy Bubash, Laura Lutz-King, and Martha Hepworth. In addition to their work supporting students, staff, and faculty, these three ladies made my transition back to work easier following the births of my children. For IT support, I am thankful to Suresh Gupta for the many times he rescued my computer and even saved the computer for the aforementioned flow cytometer.

I am very thankful for the timely emergence of the unofficial Caltech gradstudent-mom's club, whose members include Dr. Katie Brenner, Dr. Melissa Pope, and Dr. Kristin Gleitzman. I am especially glad that I witnessed Katie's pioneering efforts as a grad-student-mom. Her example and encouragement gave me the inspiration and the hope that I needed to complete this journey and without which I am not sure that I would have had the courage to attempt it. Also, I am grateful for Melissa and her friendship as we journeyed together figuring out which doctor to see, how to manage infant feedings, and how to finally get some work done in lab. Sometimes it felt like a wild ride. I am glad I had someone else who was there to share in the unique joys and trials of being a mom in grad school.

I could not have come so far without the help of my family. Each one has helped in their own special way by bringing food, babysitting, or simply emailing me words of encouragement. My parents, Steve and Jane Gropp, have always supported my education, whether it was homeschooling me or driving me 30 miles to high school. They have also been an inspiration to me to work diligently and creatively. I am so grateful for their love and encouragement. My dad tirelessly helped me build my science poster every year and taught me how to use an X-Acto knife and rubber cement. He instilled his love of design within me. My mom dared to dream big dreams for me and believed I could achieve them long before I could. I hope I have made them proud. I am also grateful for the support of my siblings Anna Johansen and Michael Gropp. Thanks for letting me wear the lab coat and goggles when we were kids and for lots of fun with our "secret chemistry labs", which I am sure horrified Mom. Also, I am so glad for my mother-in-law, Rosemary Galloway, who has helped me finish my thesis by coming and taking care of our family, especially when the children were very little. Additionally, I am thankful to SaraJean Wright, who began as Allie's nanny over two years ago and has since become like family. Thank you for taking excellent care of the babies so I could go to lab and get work done.

Nine years ago, I realized I had met the man I was going to marry. Gordie and I have been on this grad school adventure for all but six weeks of our marriage. What will it be like without grad school? I am sure we will find something to do. I am so grateful for his unwavering support. He has believed in me even when I doubted myself. He has sacrificed his time (and probably some sanity) editing my fellowship applications and research write-ups and patiently listened to whole conversations about thermodynamics, network architectures, and other technical boredom to be by my side at Caltech events. He has coached me through some of the most difficult, scary, and joyous moments that I can remember. He is a wonderful father to our two delightful children, Allie (almost 3 years) and Aaron (almost 3 months). When I entered Caltech, I did not expect to be a mother while in grad school, but I would not wish life to be any other way. Allie and, more recently, Aaron have given me such joy and an ability to enjoy the simple things

through their eyes. Allie recently told me, "Mommy, if you have a dream in your heart, you should get it out." Over these last seven years, I have realized many of the dreams in my heart, including their births, and now a thesis.

Finally, and most importantly, I am thankful to the one who puts dreams in our hearts to inspire us to strive for these goals: Jesus Christ, creator of the universe and my personal savior. I have been so delighted to look into nature and see the mind of God displayed in the intricately imbedded designs. Moreover, it is miraculous to me that we search out and understand the inner workings of nature and apply that understanding to improve other's well-being and life here. I am thankful for this beautiful world and for my brief time to work and delight in it, knowing that it is but a shadow of things to come.

Abstract

Cells integrate extracellular information via native signaling pathways to spatially and temporally coordinate complex tasks such as development and the immune response. Cellular programming holds the potential of harnessing the sophisticated and complex biological processes of living cells for diverse applications. In the last decade, cellular reprogramming has emerged as a viable therapeutic strategy. In large, reprogramming strategies have relied on statically programmed levels of gene expression to alter cellular behaviors. To construct more sophisticated programs requires dynamic control of expression and strategies for the facile construction of complex control architectures. Additionally, the application of synthetic programs to the control of native regulatory pathways requires the development of tools for interfacing with these pathways, as well as the construction of stringent controllers. Further, control systems composed of modular and tunable elements will facilitate the expansion of synthetic circuitry to a wide array of natural networks with varying system properties.

Here we describe the development of RNA-based control systems to regulate signaling and dictate cell fate in a model mitogen-activated protein kinase (MAPK) pathway. We construct networks of RNA-based control systems that interface with the *Saccharomyces cerevisiae* mating pathway to dictate entry into one of three programmed alternative fates dependent on environmental stimuli. We present a readily translatable method for identifying control points within natural networks that enable the construction of a modular interface between synthetic circuitry and native networks. In building these networks, we demonstrate the rational tuning of circuit performance via the exchange of well-defined parts to compose networks capable of actuating changes in cellular behavior

in response to environmental cues. Further, we construct network architectures which facilitate reduced interference from simultaneously integrated opposing programs and identified sensitive parameters for engineering robust circuit performance. Finally, we present the development of a novel RNA-based control element for the regulation of both synthetic and endogenous transcripts. This work provides a model for engineering systems that regulate signaling and direct cell fate which may be applied to additional decision-making pathways to advance tissue engineering strategies, treat diseases, and study the behavior of natural regulatory networks.

Acknowledgments	iii
Abstract	ix
Table of Contents	xi
List of Tables	xiii
List of Figures	xiv
Chapter 1: Introduction	I-1
Synthetic gene regulatory networks to control biological systems	I-2
Building a modular interface from non-coding RNAs	I-4
Selecting for RNA-based sensors	I-7
Natural RNA switches as gene expression control systems.	I-8
Synthetic RNA switches that act through ribozyme-based cleavage	
mechanism	I-9
MAPK cascades as universal signaling modules in eukarvotes	I-13
Combining synthetic biology and systems biology to build gene regulatory	
networks that control cellular fate	I-16
Thesis organization	I 10 I_10
Deferences	I-17
control systems to control decision-making in the yeast mating pathway	II-1
Abstract	II-2
Introduction	II-4
Results	II-6
	II-31 II-37
Iviateriais and methods.	11-57 11-42
References	II-43 II ЛЛ
Supplementary figures	II-44 II_/0
Supplementary tables	II-47 II_59
Supplementary tables	11-37
Chapter 3: Constructing synthetic gene networks to control decision-making	
in the yeast mating pathway	III-1
Abstract	III-2
Introduction	III-4
Results	III-9
Discussion	III-34
Materials and methods	III-4(

Table of Contents

Acknowledgments References Supplementary figures	III-43 III-44 III-48
Supplementary tables	111-52
Chapter 4: Development of trans-ribozymes as actuators controlling gene	
expression.	IV-1
Abstract	IV-2
Introduction	IV-4
Results	IV-9
Discussion	IV-22
Materials and Methods	IV-30
Acknowledgments	IV-34
References	IV-35
Supplementary figures	IV-40
Supplementary tables	IV-48
Chapter 5: Conclusions	V-1
Immediate challenges	V-3
Future directions.	V-7
References	V-11

List of Tables

- Supplementary Table 2.1 GFP plasmids
- Supplementary Table 2.2 Primer sequences
- Supplementary Table 2.3 Galactose-titration plasmids
- Supplementary Table 2.4 Primers for mutagenesis
- Supplementary Table 2.5 Switch sequence information
- Supplementary Table 2.6 Msg5 plasmids
- Supplementary Table 2.7 Ste4 plasmids
- Supplementary Table 2.8 Plasmids for strain construction
- Supplementary Table 2.9 Yeast strains
- Supplementary Table 2.10 Primers for integration
- Supplementary Table 3.1 pCS2094-based dual-expression cassette plasmids
- Supplementary Table 3.2 pCS1128-based dual-expression cassette plasmids
- Supplementary Table 3.3 pCS1128-based single-module booster plasmids
- Supplementary Table 4.1 Plasmids
- Supplementary Table 4.2 Ribozyme sequences
- Supplementary Table 4.3 Yeast strains
- Supplementary Table 4.4 Cloning primers
- Supplementary Table 4.5 *In vitro* assay primers

List of Figures

- Figure 1.1 Building synthetic circuitry that interfaces with the environment and native regulatory networks to control cellular behavior
- Figure 1.2 Components for interfacing with the environment and native regulatory networks
- Figure 1.3 RNA-based switches regulate gene expression in response to smallmolecule concentration.
- Figure 1.4 Converting cis-acting actuators to trans-acting requires engineering an intramolecular reaction into an intermolecular reaction.
- Figure 1.5 MAPK cascades as universal signaling modules in eukaryotes
- Figure 1.6 Yeast mating pathway and phenotypic response
- Figure 1.7 Induced network topology shapes the dynamics in a natural regulatory pathway dictating cellular fate
- Figure 1.8. Potential application of molecular network diverters to tissue engineering via small-molecule regulated patterning of cell fate.
- Figure 2.1 Regulator expression modulates pathway activity and over a narrow range of expression transitions to an alternative fate
- Figure 2.2 Identifying titratable regulators of pathway activity in the yeast mating pathway
- Figure 2.3 Overexpression of Msg5 and Ste4 modulates pathway activity and routes cells to an alternative fate
- Figure 2.4 Identifying titratable regulators in the yeast osmolarity pathway
- Figure 2.5 Reshaping the native molecular network with molecular network diverters
- Figure 2.6 Implementation of a molecular network diverter from various genetic parts
- Figure 2.7 Composition of molecular network diverter from well-defined parts
- Figure 2.8 Optimal configuration of the molecular network diverter requires tuning via the selection of components with the requisite metrics.

- Figure 2.9 Tetracycline-inducible positive network diverters of different architectures conditionally route cells to the promiscuous phenotype.
- Figure 2.10 Tracing the pathway response curve of Ste4 expression to the promiscuous fate
- Figure 2.11 Theophylline-inducible molecular network diverters constructed with constitutive Msg5 expression conditionally route cells to the chaste phenotype.
- Figure 2.12 Theophylline-inducible molecular network diverters constructed with feedback expression of Msg5 route cells to the chaste phenotype.
- Figure 2.13 Tracing the pathway response curve of Msg5 to the chaste fate
- Figure 3.1 Composition of an expression module from well-defined parts
- Figure 3.2 Molecular network diverters are composed from single- and doubleexpression modules that allow fate-routing dependent on small-molecule input.
- Figure 3.3 Integration of positive and negative single-module diverters fails to achieve dual-fate routing.
- Figure 3.4 Addition of booster module to positive feedback diverter enhances fate switching in the presence of the resistance diverter.
- Figure 3.5 Pathway activation ratio is enhanced for positive feedback diverters in network configurations including a low-strength resistance module.
- Figure 3.6 Structuring networks to amplify switching by layering positive feedback with a resistance module and a booster module
- Figure 3.7 Networks configured with an amplifying diverter and various attenuating diverters show that pathway attenuation is a weak function of the strength of the negative feedback module.
- Figure 3.8 A dual-module positive diverter shows that pathway activation is sensitive to the activity of the resistance module.
- Figure 3.9 A dual-module negative diverter shows that pathway attenuation is a strong function of the activity of the resistance module.
- Figure 3.10 A dual-module negative diverter shows that pathway attenuation and activation are sensitive to the strength of the positive feedback module.

- Figure 3.11 Benchmarking dual diverters against various positive MAAAs indicates strong performance by Diverter A.
- Figure 3.12 Benchmarking dual diverters against various negative MAAAs indicates strong performance from Diverter B and D.
- Figure 3.13 Higher small-molecule inputs improve dual-fate routing.
- Figure 3.14. Metabolic modulation restores wild-type halo in the absence of either trigger and enhances promiscuous fate routing from dual-diverters.
- Figure 3.15. Routing genetically identical cells to divergent fates in response to smallmolecule triggers supported by metabolic cues
- Figure 3.16 Reducing expression from the positive feedback and resistance modules in Diverter B may optimize the dual diverter network.
- Figure 4.1 The hammerhead ribozyme
- Figure 4.2 The anatomy of a cis-acting hammerhead ribozyme and trans-acting hammerhead ribozyme with target transcript
- Figure 4.3 Various thRz designs
- Figure 4.4 Initial ribozyme designs with pCS933
- Figure 4.5 Initial ribozyme designs with extended targeting arms show improved *in vitro* efficiency but fail to knockdown expression *in vivo*.
- Figure 4.6 Redesigned expression system
- Figure 4.7 Improved ribozymes with chRz-processing expression cassette show only modest *in vivo* knockdown despite significant improvement *in vitro*.
- Figure 4.8 Controlling target expression via the galactose-inducible promoter demonstrates that knockdown increases at higher expression levels of target transcript.
- Supplementary Figure 2.1 Engineered strain linearly increases the mean level of expression from the galactose-inducible promoter in response to increasing galactose.
- Supplementary Figure 2.2 Promoter characterization
- Supplementary Figure 2.3 Range of switch expression strengths

Supplementary Figure 2.4	Constitutive expression with low-strength promoter shows high pFUS1-GFP levels, yet switches fail to cross the phenotypic transitory range.
Supplementary Figure 2.5	Theophylline-inducible positive network diverters mirror response of tetracycline-responsive diverters.
Supplementary Figure 2.6	Selection of mating-resistant cells occurs over time at super- threshold levels of positive feedback.
Supplementary Figure 2.7	Constitutive expression of Msg5 routes cells to chaste for entire range of promoter strengths.
Supplementary Figure 2.8	Histograms show population distribution differences for constitutive and feedback network diverters.
Supplementary Figure 2.9	Neglecting the low pFUS1-GFP population, both feedback architectures show similar mean levels of pathway activity are required for diverting pathway response to the chaste phenotype.
Supplementary Figure 2.10	Tracing the pathway response curve of constitutive Msg5 expression including the low GFP population shows similar transitory range for fate divergence
Supplementary Figure 2.11	Original plasmids for construction of molecular network diverter and reporters
Supplementary Figure 2.12	Original plasmids for construction of yeast strains
Supplementary Figure 3.1	Range of switch expression strengths
Supplementary Figure 3.2	Single-module diverters fail to achieve dual-fate routing.
Supplementary Figure 3.3	Reducing the strength of the feedback module from S3tc to S2tc yields weak promiscuous routing while modestly improving chaste routing.
Supplementary Figure 3.4	Reducing the strength of the booster module from S4tc to S3tc yields weak promiscuous routing and does not significantly improve chaste routing.
Supplementary Figure 3.5	Loci characterization
Supplementary Figure 3.6	Plasmid maps

xviii

Supplementary Figure 4.1	Comparison of minimal ribozyme folded with truncated and extended target sequence in RNAstructure
Supplementary Figure 4.2	Control ribozyme sequences from pCS933 folded with extended 137 nucleotide yEGFP target sequence
Supplementary Figure 4.3	Canonical ribozyme sequences from pCS933 folded with extended 137 nucleotide yEGFP target sequence
Supplementary Figure 4.4	Initial ribozyme designs with chRZ processing expression cassette (pCS975)
Supplementary Figure 4.5	Control ribozyme sequences from pCS975 folded with extended 137 nucleotide yEGFP target sequence
Supplementary Figure 4.6	Canonical ribozyme sequences from pCS975 folded with extended 137 nucleotide yEGFP target sequence
Supplementary Figure 4.7	Canonical ribozyme sequences from pCS975 folded with extended 137 nucleotide yEGFP target sequence
Supplementary Figure 4.8	Promoter characterization
Supplementary Figure 4.9	Ribozyme designs with extended arms in pCS933
Supplementary Figure 4.10	Plasmid maps for ribozyme expression vectors
Supplementary Figure 4.11	Plasmid maps for construction of CSY341 expression vectors