# Understanding the Chemical Basis of Neuronal Development and Communication:

I. The Role of Fucose  $\alpha(1-2)$  Galactose Carbohydrates in Neuronal Growth

II. Structure-Function Analysis of Chondroitin Sulfate in the Brain

Thesis by

Cristal Ivette Gama

In Partial Fulfillment of the Requirements

for the Degree of

Doctor of Philosophy

California Institute of Technology

Pasadena, California

2009

(Defended 17 July 2008)

© 2009

Cristal Ivette Gama

All Rights Reserved

...for my family...

#### Acknowledgments

Without the help and support of many people, both scientific and personal, the work in this thesis would not have been possible. I would like to thank my advisor, Linda Hsieh-Wilson, for her advice and guidance. I would also like to thank the members of my committee, Judy Campbell, Mary Kennedy, and Paul Patterson. Without any of them, this thesis would not have come to completion. I would also like to thank Dennis Dougherty, even though he is not technically on my committee, he was like a second mentor and really helped our lab in the beginning, letting us join his group meetings and giving us much advice.

My undergraduate professors deserve special thanks as well since it is because of them that I came to graduate school. Dr. Robert Vellanoweth at Cal State LA was such a great mentor, a young professor with such an amazing mind, brilliant really. He was such an inspiration to me, being of Mexican decent, having gone to Cal State LA himself as an undergrad and then back as a professor leading his own lab and directing his own unique research. It was a wonderful learning experience and he showed me how to design my own experiments and think independently about the projects I worked on. Of course, I am indebted to Dr. Carlos Gutierrez at Cal State LA. I met Dr. Gutierrez as a senior in high school trying to decide which college to go to. He was the first academic professional I ever met, doing "lab research," which is what I wanted to do, even though at that time I wasn't even sure what exactly that meant. Dr. Gutierrez is such a wonderful human being, caring, easy going and great, I think it was because of him that I thought I could stick it out in the science field and make it through to get my Ph.D. There are many more wonderful people at Cal State LA that made me getting to Caltech possible, it's hard to mention everyone. However, I cannot go without special thanks to Vicki Kubo-Anderson. Without Vicki, nothing would be possible. Without Vicki, the biochemistry department would crumble. Without Vicki, we students would be in such a terrible state we wouldn't function either. She was like a mom, a mentor, a counselor, but most importantly, a friend.

The members of the Hsieh-Wilson lab, both past and present, have been a great group to work with. All the original members were unforgettable each in their own way, Raymond Doss, Sarah Tully, Nelly Khidekel, Katherine Poulin-Kerstien, Sherry Tsai, Nathan Lamarre-Vincent and Lori Lee. We helped each other get through some rough and tough times, learning the basics and going from there. Lori Lee was my first partner on the fucose project and, more than that, we became good friends. Sarah Tully did all the synthesis on the chondroitin sulfate project and without her this work couldn't have happened. It was really tough, her getting the synthesis to work and me trying to get those neurons to grow and behave. There were some very long nights and crazy "losing my mind" moments, it was great! Isn't that what science is really about? After the first group of students then came Heather Murrey. What can I say about Heather, other than she is the craziest, most intelligent scientist I think I have ever met. Through the years we have become wonderful friends and I will never forget all the times we've shared, both in and out of lab. Dr. Marian Bryan joined the lab and changed my life. Not only did we share time in the lab, she became my own personal doctor, counselor, therapist and everything in between. Katie Saliba is an amazing chemist and an even better being. She is, through and through, one of the most wonderful, caring people in this world. I would also like to thank everyone else that I have met and worked with. Each and

everyone of you has helped me get to this point: Maria Chiriac, Dr. Stacey Kalovidouris, Dr. Eric Shipp, Dr. Manish Rawat, Tammy Campbell, Dr. Helen Cheng, Dr. Ross Mabon, Monica Luo, Bruce Tai, Rob Moncure, Peter Clark, Claude Rogers, Dr. Mike Chang, Dr. Song-Gil Lee, Dr. Seok-Ho Yu, Dr. Jiang Xia, Jessica Rexach, Jessica Dweck, Long Phan, Gloria Sheng, Joshua Brown, and Joelle Radford. Of course I have to give special mention to some of the newest members in the lab, Young In Oh, Chithra Krishnamurthy, and Arif Wibowo. Although it hasn't been that long, I consider you close friends and I am thankful for all your support.

There are many more people outside of lab who have supported me and helped me survive. First and foremost, I would like to thank my parents, Bonifacia and Guadalupe. It is because of them that I am who I am today and it is for them that I have moved forward and continued my education to this point. My sister Marisol and brother Eric have always provided undying support, ever since my first day in elementary to my last day in graduate school. I know they have thought I was crazy for being in school this long, but they have always been there for me in every way. To all of my nephews and nieces, those here and those to come, this is for you. My dear friend Xiomara Padilla has been wonderful ever since we met. We have grown up together, cried through Quant, made it through graduation and now share our lives over Thai food, Shabu Shabu, chili cheese fries and, of course, Roscoe's. I have to thank Callie Bryan again. She has been the most wonderful friend, helping me get through these last years, pushing me forward, and buying me coffee, or anything else. I also have to thank one of my best friends, Wilbert Preyer. He has supported and believed in me every moment. Thank you all soo much from the bottom of my heart, we did it!

#### Abstract

Although carbohydrates are known to participate in many important processes including inflammation, cancer metastasis and pathogenic infection, their functional roles are only beginning to be understood on a molecular level. The challenge is that carbohydrates and glycoproteins are inherently difficult to study. Unlike DNA and proteins, carbohydrate structures are not template-encoded, and the modifications are challenging to detect *in vivo* and manipulate for structure-function analyses. As such, new tools are needed to complement the traditional biochemical and genetic approaches in order to advance our understanding of carbohydrates and their physiological roles. Here, we seek to understand the roles of carbohydrates in regulating the structure and function of proteins in the brain. Our major focus will be on two carbohydrate modifications that play important roles in neuronal communication, development and memory storage: fucosylation (Part I) and chondroitin sulfate glycosaminoglycan modifications (Part II).

In Part I, we describe our progress in elucidating the molecular mechanisms by which fucosyl saccharides regulate neuronal communication in the brain. Information flow in the brain is regulated by synapses, which are specialized sites of contact between neurons. Synaptic connections involve numerous molecular recognition events among proteins, carbohydrates, and small molecules. One of the molecules enriched at the synapse is the sugar L-fucose. Previous studies have suggested that fucose $\alpha$ (1-2)galactose (Fuc $\alpha$ (1-2)Gal) saccharides play essential roles in learning and memory. For instance, preventing formation of Fuc $\alpha$ (1-2)Gal linkages has been shown to cause reversible amnesia in animals. Despite these intriguing observations, proteins that express the Fuc $\alpha(1-2)$ Gal epitope (glycoproteins) or proteins that bind this epitope (lectins) have not been identified. Through the use of several chemical probes, we have established that Fuc $\alpha(1-2)$ Gal associated proteins participate in a novel carbohydrate-mediated pathway for regulating neuronal growth. Additionally, we have found that Fuc $\alpha(1-2)$ Gal glycoproteins are prevalent in the developing brain and that synapsin Ia/Ib are the major Fuc $\alpha(1-2)$ Gal glycoproteins in the adult brain. In our attempts to identify Fuc $\alpha(1-2)$ Gal lectins, we have established that multivalent polymers enhance our ability to capture and characterize such proteins.

In Part II, we describe our efforts toward understanding the role of chondroitin sulfate glycosaminoglycans in neuronal development. Chondroitin sulfate (CS) glycosaminoglycans are linear, sulfated polysaccharides implicated in cell division, neuronal growth, and spinal cord injury. The structural complexity and heterogeneity of CS has hampered efforts to understand its precise biological roles. Although they exist as a heterogeneous mix in nature, it is thought that CS activity is dictated by a sulfation code where distinct sulfation sequences are spatially and temporally regulated and direct the biological activity of CS glycosaminoglycans. We have developed a chemical approach to evaluate the structure-activity relationship of CS as it effects neuronal growth. We generated the first synthetic library of well-defined CS oligosaccharides containing various sulfation sequences and have demonstrated that the CS-E sulfation sequence is a stimulatory motif that promotes the growth of several neuron types. Moreover, we determined that CS-E mediated stimulation of neurite outgrowth was facilitated by activation of midkine/PTP5 and BDNF/TrkB signaling pathways.

#### **Table of Contents**

| Acknowledgm            | ents  | iv  |
|------------------------|---|-----|
| Abstract               |   | vii |
| Table of Conte         | nts   | ix  |
| List of Figures        |   | X   |
| List of Abbrev         | iations   | xiv |
| Part I :<br>Chapter 1  | Fucosylation in the Brain   | 1   |
| Chapter 2              | Fucose α(1-2) Galactose Carbohydrates Regulate Neuronal Growth  | 16  |
| Chapter 3              | Fucose $\alpha(1-2)$ Galactose-Containing Glycoproteins are Prevalent in the Brain and Regulate Neuronal Morphology | 42  |
| Chapter 4              | Discovery of Fucose $\alpha(1-2)$ Galactose-Specific Lectins in the Developing Brain                                | 61  |
| Part II :<br>Chapter 5 | The Biological Activity of Chondroitin Sulfate<br>Glycosaminoglycans  | 98  |
| Chapter 6              | Investigations into the Sulfation Code of Chondroitin Sulfate<br>Glycosaminoglycans                                 | 118 |
| Chapter 7              | Elucidating the Mechanism of CS-E Mediated Neuronal Outgrowth   | 134 |

## List of Figures

| Chapter 1  |   | Page |
|------------|---|------|
| Figure 1.1 | The monosaccharide L-fucose   | 3    |
| Figure 1.2 | In mammalian cells, the biosynthesis of GDP-fucose occurs through two distinct pathways             | 4    |
| Figure 1.3 | Common fucose-galactose linkages found on the terminal ends of carbohydrate chains                  | 5    |
| Figure 1.4 | Incorporation of 2-deoxy-D-Galactose (2-dGal) inhibits formation of Fuc $\alpha$ (1-2)Gal linkages. | 5    |
| Figure 1.5 | The trisaccharide Fuc $\alpha$ (1-2)Gal $\beta$ (1-4)GlcNAc recognized by antibody A46-B/B10        | 8    |
| Figure 1.6 | Proposed mechanisms by which the Fuc $\alpha$ (1-2)Gal epitope acts in neurons                      | 11   |

| Figure 2.1  | Fuc $\alpha$ (1-2)Gal-biotin probe 1 was designed to mimic<br>endogenous glycoproteins                               | 18 |
|-------------|--|----|
| Figure 2.2  | Fuc $\alpha$ (1-2)Gal probe 1 binds to the cell surface of hippocampal neurons                                       | 20 |
| Figure 2.3  | Fuc $\alpha(1-2)$ Gal probe 1 binds to hippocampal neurons   | 20 |
| Figure 2.4  | Lipid extraction does not alter labeling with $Fuc\alpha(1-2)Gal$ probe 1  | 21 |
| Figure 2.5  | Costaining of neurons with $Fuc\alpha(1-2)Gal$ probe 1 and an anti-<br>tau antibody.                                 | 22 |
| Figure 2.6  | Costaining of neurons with $Fuca(1-2)Gal$ probe 1 and a MAP2 antibody  | 22 |
| Figure 2.7  | Costaining of neurons with $Fuca(1-2)Gal$ probe 1 and an anti-<br>synapsin antibody                                  | 23 |
| Figure 2.8  | Costaining of neurons with UEA-I lectin and an anti-tau antibody   | 24 |
| Figure 2.9  | Lipid extraction using MeOH/CHCl <sub>3</sub> did not diminish UEA-I lectin labeling                                 | 24 |
| Figure 2.10 | Fuc $\alpha$ (1-2)Gal promotes neuronal growth   | 26 |
| Figure 2.11 | Only the Fuc $\alpha$ (1-2)Gal-selective lectins UEA-I and LTL stimulate neuronal growth                             | 27 |
| Figure 2.12 | Treatment with 2-dGal diminishes the expression of the Fuc $\alpha(1-2)$ Gal epitope on glycoproteins                | 28 |
| Figure 2.13 | Hippocampal neurons treated with varying concentrations of 2-<br>dGal exhibit increasing defects in neuronal growth. | 29 |

| Figure 2.14 | Treatment of hippocampal neurons with 2-dGal (15 mM), but | 30 |  |
|-------------|---|----|--|
|             | not 3-dGal (15 mM), for 2 days inhibits neuronal growth   |    |  |

## Chapter 3

| Figure 3.1 | Trisaccharide Fuc $\alpha$ (1-2)Gal $\beta$ (1-4)GlcNAc recognized by antibody A46-B/B10   | 44 |
|------------|--|----|
| Figure 3.2 | Costaining of neurons with $Fuc\alpha(1-2)$ Gal antibody A46-B/B10<br>and an anti-tubulin antibody   | 45 |
| Figure 3.3 | Lipid extraction of cellular membranes does not abolish<br>staining with antibody A46-B/B10  | 45 |
| Figure 3.4 | Costaining of neurons cultured for 14 DIV with $Fuc\alpha(1-2)Gal$<br>antibody A46-B/B10 and either an anti-synapsin antibody or an<br>anti-spinophilin antibody | 46 |
| Figure 3.5 | Fuca $(1-2)$ Gal is expressed on several glycoproteins in the hippocampus and is developmentally regulated   | 47 |
| Figure 3.6 | Synapsins Ia and Ib are Fuc $\alpha$ (1-2)Gal glycoproteins  | 49 |
| Figure 3.7 | Expression of synapsin I was reduced by treatment with 2-dGal but not 6-dGal   | 50 |
| Figure 3.8 | The morphology of hippocampal neurons is modulated by 2-<br>dGal in a concentration-dependent manner   | 51 |
| Figure 3.9 | Synapsin-deficient neurons display reduced neurite retraction relative to wild-type neurons upon treatment with 2-dGal   | 52 |

| Figure 4.1 | Monovalent capture probe 2 and control molecule 3  | 64 |
|------------|--|----|
| Figure 4.2 | Capture probe <b>2</b> specifically labels the fucose-binding lectins AAA and UEA-I  | 66 |
| Figure 4.3 | Capture probe 2 labels distinct proteins in dissociated neurons  | 67 |
| Figure 4.4 | Probe <b>2</b> specifically labels $Fuc\alpha(1-2)Gal$ lectins in dissociated neurons  | 68 |
| Figure 4.5 | Fuc $\alpha$ (1-2)Gal lectins were specifically captured by probe <b>2</b> and isolated on a streptavidin column                         | 69 |
| Figure 4.6 | Coomassie stain analysis of total protein captured on streptavidin column after probe <b>2</b> labeling                                  | 70 |
| Figure 4.7 | Subcellular fractionation of protein lysates labeled with probe $2$ and control molecule $3$   | 71 |
| Figure 4.8 | Scheme depicting the strategy by which probe 2 is used to label<br>and identify $Fuc\alpha(1-2)Gal$ lectins from neuronal protein lysate | 72 |
| Figure 4.9 | Distinct proteins from rat pup lysate were captured and isolated   | 73 |

|             | by probe <b>2</b>   |    |
|-------------|---|----|
| Figure 4.10 | Protein lysate labeled with probe 2 did not confirm                     | 74 |
|             | identification of potential Fuc $\alpha$ (1-2)Gal lectins               |    |
| Figure 4.11 | Multivalent polymer 4 displays multiple $Fuc\alpha(1-2)Gal$ epitopes    | 75 |
|             | (red) and multiple biotin moieties (green)                              |    |
| Figure 4.12 | Labeling of proteins from dissociated embryonic neurons using           | 76 |
|             | the multivalent or monovalent probe                                     |    |
| Figure 4.13 | Strategy for identification of Fuc $\alpha$ (1-2)Gal lectins from       | 76 |
|             | neuronal protein lysate using multivalent polymer 4                     |    |
| Figure 4.14 | Capture of purified UEA-I lectin using the multivalent Fuc $\alpha$ (1- | 77 |
|             | 2)Gal polymer 4   |    |
| Figure 4.15 | Capture of UEA-I and Fuc $\alpha$ (1-2)Gal lectins from rat pup lysate  | 78 |
| Figure 4.16 | Capture of Fuc $\alpha$ (1-2)Gal lectins from rat pup lysate using the  | 79 |
| 8           | Fuc $\alpha$ (1-2)Gal affinity column                                   |    |
| Figure 4.17 | Design of multivalent capture polymer 5                                 | 80 |
| 0           |   |    |

## Chapter 5

| Figure 5.1 | Proposed mechanism of GAG function at the cell surface                    | 99  |
|------------|---|-----|
| Figure 5.2 | Structures of representative classes of GAGs                              | 100 |
| Figure 5.3 | Biosynthesis of HS/heparin and CS GAGs takes place in the Golgi apparatus | 102 |
| Figure 5.4 | Structures of the most common CS motifs found <i>in vivo</i>              | 103 |
| Figure 5.5 | Heterogeneous GAG polysaccharides exhibit areas of low and high sulfation | 104 |
| Figure 5.6 | Crystallographic models of the FGF-FGFR-HS complex                        | 107 |

| Figure 6.1 | Structures of the initial synthetic CS oligosaccharides                            | 120 |
|------------|--|-----|
| Figure 6.2 | CS-E tetrasaccharide <b>8</b> stimulates the outgrowth of hippocampal neurons      | 121 |
| Figure 6.3 | Structures of synthetic CS oligosaccharides containing distinct sulfation patterns | 123 |
| Figure 6.4 | The sulfation pattern directs the neuritogenic activity of CS                      | 124 |
| Figure 6.5 | The CS-E sulfation motif stimulates the outgrowth of DRG (A)                       | 126 |

# and dopaminergic neurons (B)

| Figure 7.1  | A substratum of CS-E polysaccharide stimulates the outgrowth of hippocampal neurons   | 136 |
|-------------|---|-----|
| Figure 7.2  | CS-E polysaccharide added in solution inhibits the outgrowth of hippocampal neurons   | 137 |
| Figure 7.3  | Schematic representation of (A) natural CS and (B) CS glycopolymers   | 138 |
| Figure 7.4  | CS-E glycopolymers inhibit the outgrowth of hippocampal neurons   | 139 |
| Figure 7.5  | CS-E tetrasaccharide glycopolymer inhibits hippocampal neurite outgrowth  | 140 |
| Figure 7.6  | Comparison of the inhibitory potencies of CS glycoplymer <b>15</b> and the natural polysaccharide at various concentrations             | 141 |
| Figure 7.7  | Each sulfation pattern exhibits a distinct structural conformation  | 143 |
| Figure 7.8  | A specific sulfation pattern promotes the interaction of CS with neuronal growth factors  | 144 |
| Figure 7.9  | The CS-E sulfation motif stimulates neuronal growth through activation of the midkine-PTPζ and BDNF-TrkB signaling pathways             | 146 |
| Figure 7.10 | Function-blocking antibodies against BDNF and TrkB, but not class-matched control antibodies, disrupt the neuritogenic activity of CS-E | 147 |

#### List of Abbreviations

| 1D                 | one-dimensional   |
|--------------------|---|
| 2D                 | two-dimensional   |
| 2-dGal             | 2-deoxy-D-galactose   |
| 2-fucosyllactose   | L-fucose $\alpha(1-2)$ galactose $\beta(1-4)$ glucose       |
| 3-dGal             | 3-deoxy-D-galactose   |
| 6-dGal             | 6-deoxy-D-Galactose   |
| AAA                | Anguilla anguilla agglutinin                                |
| Ab                 | antibody  |
| Ac                 | acetyl, acetate   |
| aq                 | aqeous  |
| BCA                | bicinchoninic acid  |
| BDNF               | brain derived neurotrophic factor                           |
| BSA                | bovine serum albumin  |
| °C                 | degree Celsius  |
| CaCl <sub>2</sub>  | calcium chloride  |
| cAMP               | cyclic adenosine monophosphate                              |
| CH <sub>3</sub> N  | acetonitrile  |
| CHCl <sub>3</sub>  | chloroform  |
| CMF-HBSS           | Calcium and Magnesium Free Hank's Balanced Salt Solution    |
| CNS                | central nervous system                                      |
| $CO_2$             | carbon dioxide  |
| CRD                | carbohydrate recognition domain                             |
| CS                 | chondroitin sulfate   |
| ddH <sub>2</sub> O | double distilled water                                      |
| D-Gal              | D-galactose   |
| diazirine          | trifluoromethylphenyldiazirine                              |
| DIV                | days in vitro   |
| DMEM               | Dulbecco's Minimal Eagle's medium                           |
| DMSO               | dimethylsulfoxide   |
| DNA                | deoxyribonucleic acid                                       |
| DP                 | degree of polymerization                                    |
| DRG                | dorsal root ganglion  |
| DTT                | dithiothreitol  |
| E18                | embryonic day 18  |
| ECL                | enhance chemiluminescence                                   |
| EDTA               | ethylenediaminetetraacetic acid                             |
| endo N             | endoneuraminidase   |
| EXT1 and 2         | exostoses enzymes 1 and 2                                   |
| FGF                | fibroblast growth factor                                    |
| FGFR               | fibroblast growth factor receptor, tyrosine kinase receptor |
| Fuc                | L-fucose  |
| Fuca(1-2)Gal       | fucose $\alpha(1-2)$ galactose                              |
| FX                 | epimerase-reductase enzyme                                  |

| g                 | gram, gravitational force                                  |
|-------------------|--|
| GAG               | glycosaminoglycan  |
| Gal               | galactose  |
| GalNAc            | <i>N</i> -acetylgalactosamine                              |
| GDP-fucose        | guanosine diphosphatyl-fucose                              |
| GlcA              | D-glucuronic acid  |
| GlcN              | D-glucosamine  |
| GlcNAc            | <i>N</i> -acetylglucosamine                                |
| Gluc              | glucose  |
| GluR1             | glutamate receptor 1                                       |
| GMD               | GDP-mannose-4,6-dehydratase                                |
| GPC               | gas phase chromatography                                   |
| GTP               | guanosine triphosphate                                     |
| h                 | hour   |
| HIO <sub>4</sub>  | periodate  |
| HRP               | horse-radish peroxidase                                    |
| HS                | heparan sulfate  |
| IC <sub>50</sub>  | inhibition concentration at 50%                            |
| IdoA              | L-iduronic acid  |
| IgG               | immunoglobulin   |
| IP                | immunoprecipitated   |
| $K^+$             | potassium ion  |
| Kassoc            | association constant                                       |
| KCl               | potassium chloride   |
| kDa               | kilodalton   |
| L                 | liter  |
| LTL               | Lotus tetragonolobus lectin                                |
| LTP               | long-term potentiation                                     |
| M                 | molar  |
| MALDI-TOF         | matrix-assisted laser desorption/ionization time-of-flight |
| MAP2              | microtubule associated protein 2                           |
| MAPK              | mitogen-associated protein kinase                          |
| MEM               | Minimal Eagle's Medium                                     |
| МеОН              | methanol   |
| μg                | microgram  |
| MgCl <sub>2</sub> | magnesium chloride   |
| min               | minutes  |
| m                 | milli or meter   |
| μ                 | micro  |
| mol               | mole   |
| MS                | mass spectrometry  |
| n                 | nano   |
| Ν                 | normal   |
| Na <sup>+</sup>   | sodium ion   |
| NaCl              | sodium chloride  |
| NaOH              | sodium hydroxide   |
|                   |  |

| NCAM                             | neural cell adhesion molecule             |
|----------------------------------|---|
| NDST                             | N-deacetylase-N-sulfotransferase          |
| NH <sub>4</sub> HCO <sub>3</sub> | ammonium bicarbonate                      |
| NP-40                            | nonidet P-40 detergent                    |
| NSF                              | <i>N</i> -ethylmaleimide sensitive factor |
| OEt                              | <i>O</i> -ethyl                           |
| P2                               | insoluble fraction 2                      |
| PAA                              | polyacrylamide polymer lacking saccharide |
| PAGE                             | polyacrylamide gel electrophoresis        |
| PAO                              | phenyl arsine oxide                       |
| PAPS                             | 3'phosphoadenosine 5'phosphosulfate       |
| PBS                              | phosphate buffered saline                 |
| PI3-K                            | phosphatidylinositol 3-kinase             |
| PSA                              | polysialic acid                           |
| PSD-95                           | post synaptic density protein 95          |
| ΡΤΡζ                             | protein tyrosine phosphatase zeta         |
| PVDF                             | polyvinylidene difluoride                 |
| Qeq                              | charge equilibrium                        |
| RNA                              | ribonucleic acid                          |
| rpm                              | revolutions per minute                    |
| rt                               | room temperature                          |
| S2                               | soluble fraction 2                        |
| SDS                              | sodium dodecyl sulfate                    |
| SEM                              | standard error of the mean                |
| Syn KO                           | synapsin knockout                         |
| TBST                             | tris buffered saline with Tween-20        |
| TNFα                             | tumor necrosis factor alpha               |
| Tris-Cl                          | tris chloride                             |
| TrkA                             | tyrosine kinase A receptor                |
| TrkB                             | tyrosine kinase B receptor                |
| U                                | unit                                      |
| UEA I                            | Ulex europeaus agglutinin I               |
| UV                               | ultraviolet                               |
| vol                              | volume                                    |
| W/V                              | weight per volume                         |
| WGA                              | wheat germ agglutinin                     |
| WT                               | wild type                                 |
|                                  |   |