I. STUDIES ON MIXED ANHYDRIDE SYSTEMS

II. STUDIES OF THE ALPHA CHYMOTRYPSIN CATALYZED HYDROLYSIS OF SPECIFIC SUBSTRATES

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ABSTRACT

The synthesis and purification of p-nitrobenzoic-acetic mixed anhydride, p-methoxybenzoic-acetic mixed anhydride and benzoic-acetic mixed anhydride is described. The thermodynamic constants for the disproportionation reaction have been determined. A mechanism for this reaction is discussed. Measurements of the interaction of acetic and benzoic acids with the acetic-benzoic anhydride system have been made.

The cleavage of the mixed anhydrides with nucleophilic agents is described.

The synthesis of the following compounds is described:

N-trimethylacetyl-L-tyrosinhydrazide, N-dichloroacetyl-L
tyrosinhydrazide, N-formyl-L-tyrosinhydrazide, N-trichloroacetyl-L-tyrosinamide and N-acetyl-L-cysteic acid carboxylamide. The kinetic constants for the chymotrypsin hydrolysis of these substances are reported.

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PART I. STUDIES OF MIXED ANHYDRIDE SYSTEMS

INTRODUCTION

Mixed anhydrides I, formed by the loss of water from two different carboxylic acids, have been known or postu-

lated for a long time. Until the advent of high vacuum techniques the liquid members of this class of compounds could not be prepared in a pure state. Any attempt at fractionation of these compounds at temperatures above one hundred degrees centigrade resulted in the production of a distillate, whose boiling range was continuous from the boiling point of one symmetrical anhydride to the boiling point of the other symmetrical anhydride. Therefore, it was not certain whether this material represented a new compound or a mixture of equal portions of the two symmetrical anhydrides. This point was partially clarified by Behal, (1) who prepared several crystalline solid mixed anhydrides and showed them to have discrete properties distinct from either of the symmetrical anhydrides. Other evidence for the existence of these substances was obtained by Kilpatrick, (2) who showed that acetic-proponic mixed anhydride hydrolyzed at a rate which was intermediate between the rate of hydrolysis of acetic and proponic anhydrides. The reaction accurately followed first order kinetics throughout its course, while an equimolar mixture of acetic and proponic anhydrides gave kinetics which was not homogeneously first order.

Except for their relationship to the Perkin synthesis (3) and certain acylation reactions, (4) interest in these substances has lagged until the last few years. Recent developments in the field of peptide synthesis have greatly increased the importance of these compounds and have stimulated efforts to understand their properties.

Wieland (5) and Vaughn (6) have developed new peptide syntheses based upon the conversion of an acylated amino acid or peptide into a mixed anhydride. The mixed anhydride is then allowed to react with the free amino group of another amino acid to form a peptide bond. It was soon found very convenient to form the acylated amino acid mixed anhydride with an alkyl chlorocarbonate.

$$R'-CH-C'-OH$$
 $R'-CH-C'-O-C-OR'''$
 $R'-CH-C'-O-C-OR''$
 $R'-C'-O-C-OR''$
 $R'-C'-O-C-OR'$
 $R'-C'-O-C-OR'$
 $R'-C'-O-C-OR'$
 $R'-C'-O-C-OR'$
 $R'-C'-O-C-OR'$
 $R'-C'-O-C-OR'$
 $R'-C'-O-C-OR'$
 $R'-C'-O-C-OR'$
 $R'-C'-O-C-OR'$
 $R'-C'-O-C-OR'$

The rate and the magnitude of equilibrium constant of the disproportionation reaction and the products obtained by heterolytic cleavage with nucleophilic agents are the properties of a mixed anhydride which one is most interested in being able to predict.

Mixed anhydrides all show a tendency to disporportionate into the two symmetrical anhydrides

It is possible to show that this reaction is reversible and that an equilibrium state is reached. However, quantitative measurements of this equilibrium mixture were not possible until the advent of infrared spectophotometric techniques, since it was not possible to devise a chemical test which would quantitatively distinguish between the mixed anhydride and an equal molar mixture of the two symmetrical anhydrides.

Brown and Trotter⁽⁷⁾ were the first to carry out quantitative measurements on a mixed anhydride system. They showed that if a mixture of acetic and butyric anhydrides is allowed to stand and its infrared spectrum examined as a function of time, the spectrum changes from an additive mixture of acetic and butyric anhydrides to that of a mixture of the two symmetrical anhydrides and a third substance. During this change a prominent new peak appears in the 8-9 micron region [the region of the C-O-C stretching frequency of anhydrides]. The appearance of this peak is accompanied by a corresponding decrease in the peaks associated with

either symmetrical anhydride. Therefore, the new peak is assigned the C-O-C stretching frequency of acetic-butyric mixed anhydride. It was shown that after several hours the mixed anhydride peak attains constant dimensions and that it is invariant with changes in temperature. From this fact it was determined that the change in enthalpy for the disproportionation reaction was zero and that only entropy considerations should affect the over-all free energy change. Thus the disproportionation reaction seems to be similar to certain processes of ester interchange, of metal alkyl interchange and of the distribution of halogens between glycol dihalides. These reactions were first recognized by Calingaert (8) and called redistribution reactions. they are characterized by a statistical distribution of organic radicals about a central moiety.

If the mixed anhydride disproportionation reaction is a redistribution reaction, then the equilibrium constant should be four. This hypothesis was tested and found to be consistent with the data obtained for the acetic-butyric anhydride system.

Thus Brown and Trotter were able to measure the equilibrium constant for the acetic-butyric anhydride system without ever having isolated or purified the mixed anhydride.

In like manner, they were able to measure the rate of disproportionation of the acetic-butyric mixed anhydride and found the second order rate constant to be

-4 = 4.125 Liters moles⁻¹ min.⁻¹ at 398 K

One of the most important reactions of mixed anhydrides is their cleavage by nucleophilic agents. This reaction constitutes the basis for many acylation reactions.

By the work of Gold (9) it has been shown that for most mixed anhydrides this reaction can be considered as an $\mathrm{S}_{\mathrm{N}}\mathrm{2}$ attack by the nucleophilic reagent upon the carbonyl carbonatom which carries the greatest positive charge and which is the least sterically hindered for such an attack. Often it is found that the factors which increase the positive charge of the carbonyl carbon also increase its steric shielding. Thus in the series: acetic-monochloroacetic, acetic-dichloroacetic, acetic-trichloroacetic mixed anhydride, it is the monochloroacetic mixed anhydride which has the best combination of characteristics to give the highest yield of \(\omega\) haloacylation. Often the effective steric size of substituents on the \(\mathbf{q}\) carbon will vary with the reaction If there is a delicate balance between the susceptibility of the two carbonyls to nucleophilic attack, then striking variations in product composition can be obtained by changing the reaction medium. Thus, in benzene, chloroacetic-acetic mixed anhydride gives six times as much chloroacetylation as acetylation, but in aqueous solvents two times as much acetylation as chloroacetylation is obtained.

In the absence of strongly protolytic solvents it is believed that acylium ions do not play an important part in the reactions of carboxylic anhydrides. However, Tedder et al. (10) have shown that trifluoroacetic mixed anhydrides seem to constitute an exception to this rule. In this case some reactions seem to proceed by a preliminary ionization into trifluoroacetate and acylium ions.

STUDIES ON SYSTEMS OF PARA SUBSTITUTED BENZOIC-ACETIC MIXED ANHYDRIDES

Aim and Scope of the Studies

The purpose of this work is to study the properties of three para substituted benzoic-acetic mixed anhydride systems; the benzoic-acetic, the p-nitrobenzoic-acetic, and the p-methoxybenzoic-acetic mixed anhydride systems. The investigation of the disproportionation reaction and the heterolytic cleavage with nucleophilic reagents was undertaken to learn whether the behavior of these substances was compatible with the ideas that were advanced in the introduction. In addition these studies were extended to include the interaction of acetic and benzoic acids with the benzoic-acetic mixed anhydride system.

It is hoped that the results of this work will be of some help in the design of future synthetic procedures in the field of amino acid chemistry.

The Disproportionation Reaction

Introduction

The work of Brown and Trotter⁽⁷⁾ suggested that the nature of the disproportionation reaction might be relatively independent of the structure of the mixed anhydride and regardless of the character of the organic rest attached to the carbonyl carbon, the equilibrium constant would be

four and the temperature independent.

To test this hypothesis, benzoic-acetic, p-nitrobenzoic-acetic, and p-methoxybenzoic-acetic mixed anhydrides were prepared in a pure state. These compounds differ from one another in regard to the positive character of the phenyl carbonyl carbon atom and should demonstrate any effect which arises from this cause. It is fortunate that these mixed anhydrides are of sufficient stability to allow their purification. This allows one to remove any doubt as to the interpretation of the change in the infrared spectrum that occurs when the two symmetrical anhydrides are heated together. The presence of the pure mixed anhydride allows one to approach the equilibrium point from two directions and to establish its value with greater certainty.

Results and Discussion

By use of the infrared spectrophotometric technique described in the experimental section, the concentration of the three species comprising an equilibrium mixture can be measured and an equilibrium constant, K, determined. The results are shown in Tables 1, 2, and 3.

$$K = \frac{\text{(mixed anhydride)}^2}{\text{(Acetic anhyd) (Phenyl anhyd)}}$$

<u>Table 1</u>

Benzoic-acetic Anhydride System

Equilibrium constant derived from mixtures with initial compositions of:

Temperature	Mixed Anhydride	Symmetrical Anhydride	Acid and an Anhyd
70°C	1.07	1.04	
75 [°] C	-~	~~~	1.08 1.25
100°C	1.85	1.98	3.12 2.00
150°C	4.17	4.31	4.94 3.74

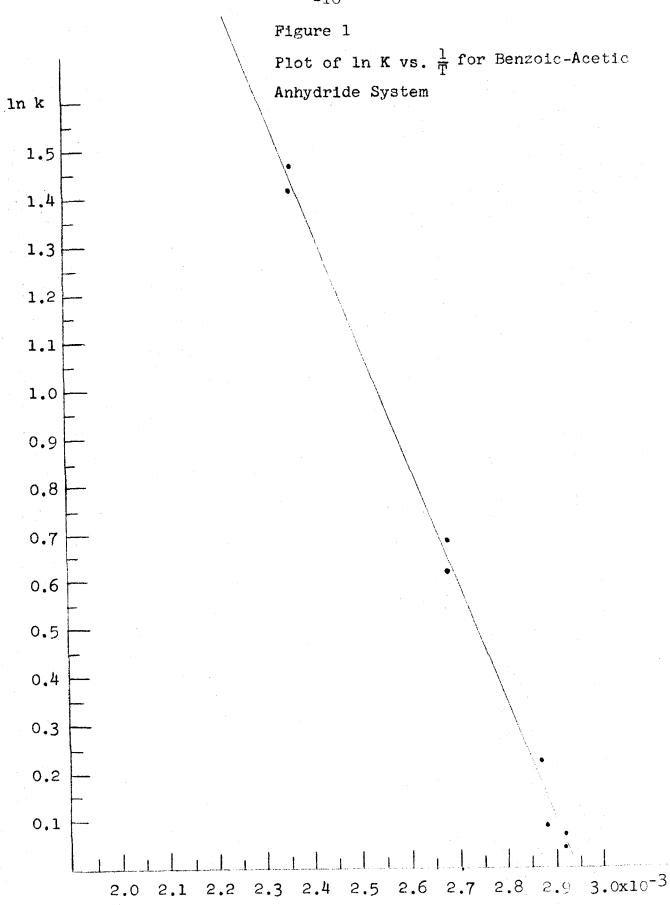
A plot of the ln K vs. $\frac{1}{T}$ is shown in Figure 1. From this plot, the change in enthalpy for the disproportionation reaction was determined as \triangle H = 5.0 Kcal.

<u>Table 2</u> <u>p-Methoxybenzoic-acetic Anhydride System</u>

Equilibrium constant derived from mixtures with initial compositions of:

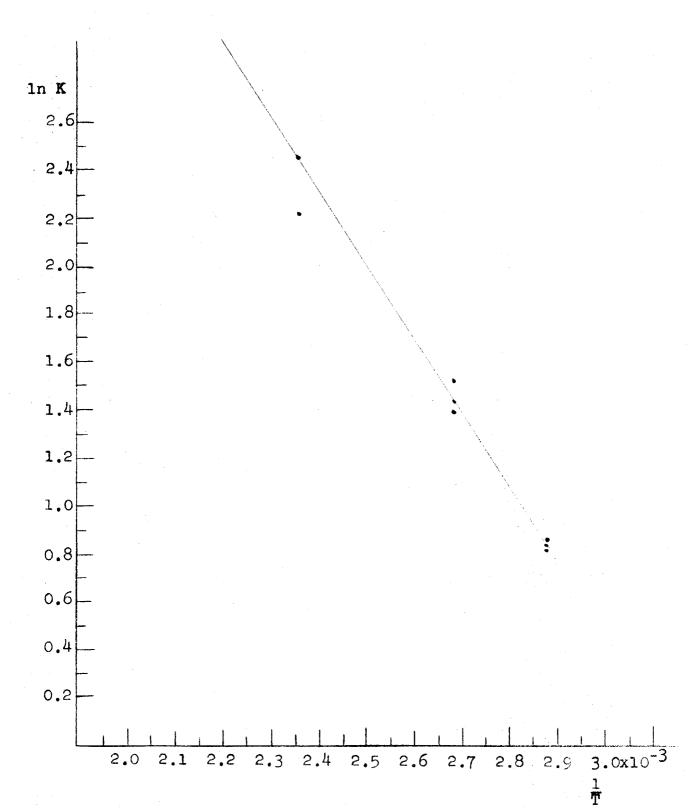
Temperature	Mixed Anhydride	Symmetrical Anhydrides
75 ⁰ 0	2.31	2.27
100 ⁰ C	4.25	4.22 4.69
105 ⁰ C	9.13	11.93

A plot of ln K vs. $\frac{1}{T}$ is shown in Figure 2. From this plot, the change in enthalpy for the disproportionation reaction was determined as \triangle H = 5.8 Kcal.



l T

Figure 2 Plot of ln K vs. $\frac{1}{T}$ for <u>p-Methoxybenzoic-Acetic</u> Anhydride System



 $\frac{\texttt{Table 3}}{\texttt{p-Nitrobenzoic-acetic Anhydride System}}$

Equilibrium constant derived from mixtures with initial compositions of:

Temperature	Mixed Anhydride	Symmetrical Anhydrides
75°C	1.27	1.20
100°C	1.29	1.32 1.38
150°C	1.70	1.78

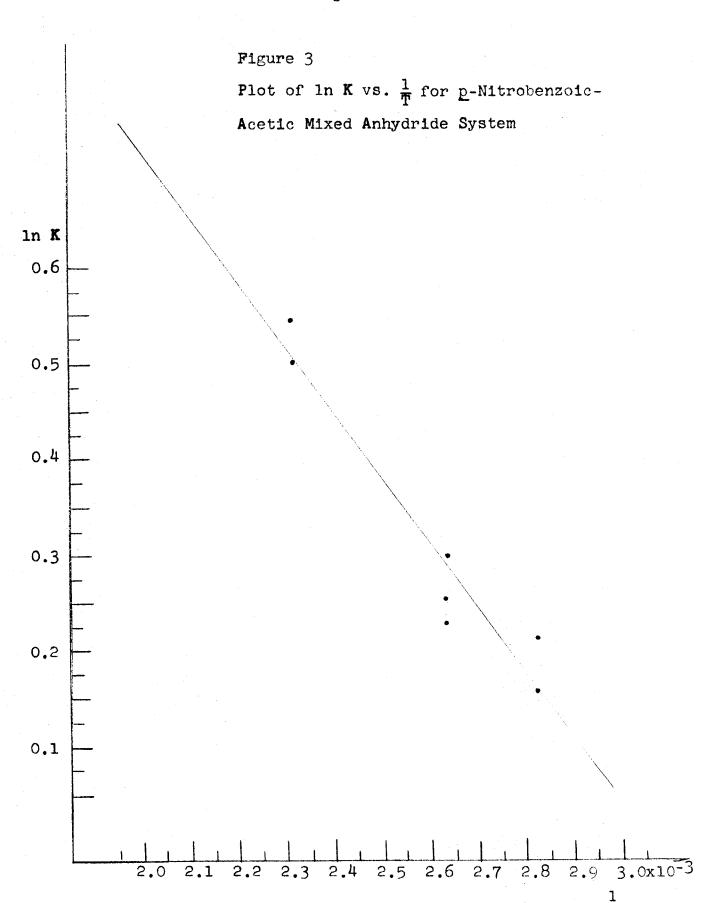
A plot of ln K vs. $\frac{1}{T}$ is shown in Figure 3. From this plot the change in enthalpy for the disproportionation reaction was determined as Δ H = 1.04 Kcal.

The above results show that the equilibrium constants for the disproportionation reaction of each of the anhydride systems have a decided temperature dependency and hence are in marked contrast to the aliphatic mixed anhydrides.

It appears at the present time that there is no simple explanation either of the differences of the substituted benzoic-acetic mixed anhydrides between one another or of the differences between the substituted benzoic-acetic mixed anhydrides and the aliphatic mixed anhydrides.

Hammett (11) has set forth the following relation which describes the change in free energy which takes place during a reaction:

$$\Delta F = -RT ln K = \Delta E_p + \Delta E_z - RT ln \frac{\sum f_p}{\sum f_R}$$



The term Δ E_p is the difference in potential energy between the reactants and the products. ΔE_2 is the difference in zero point energies of the various reaction species present and $f_{\rm D}$ and $f_{\rm r}$ are the partition functions of the products and reactants. The term $\Delta E_{\mathbf{p}}$ is the energy change which would occur if the reaction could take place so that none of the reactive species would possess the energy of motion that is associated with the various vibrations and rotations that the molecules are able to undergo. The logarithmic term and the quantity $\Delta \mathbf{E}_{z}$ are concerned then, with the kinds of rotational and vibrational motion that the molecules can execute and with the various energy-level spacings of the reactants and products. The term $\Delta \mathbf{F}_{\mathbf{p}}$ is the quantity which is influenced by changes in resonance energy, electron displacements, dipole interactions, etc. Unless the kinetic terms of the Hammett equation can be calculated or shown to be insignificant, it is not possible to discuss the effect of changes in structure upon reaction rate, equilibrium constant or heat of reaction entirely in terms of changes in bond strength, resonance energy, dipole interactions, etc.

If the products and reactants of a reaction are so similar that the energy level spacings are nearly the same then,

$$ln \frac{\sum f_p}{\sum f_R} \approx 0$$
 $\Delta E_p \approx 0$

and $\Delta F = \Delta E_{\rho}$

Since ΔV and ΔP are small;

 $\Delta F = \Delta H$ $\Delta S = 0$

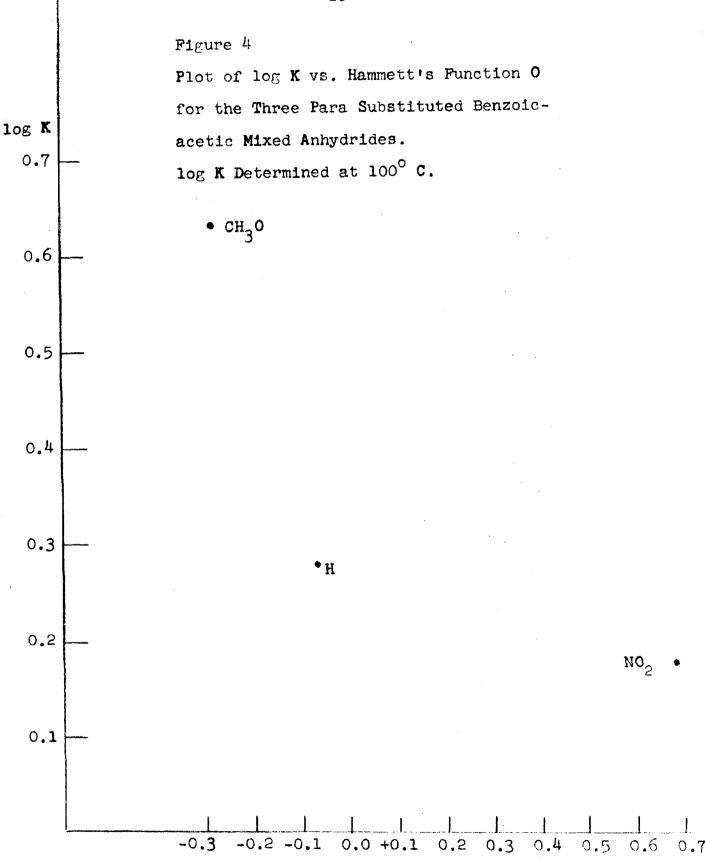
Reactions between compounds which satisfy these conditions can be described by the use of the Hammett functions ρ and σ . Meta and para substituted benzene derivatives, in which the reaction takes place at a reactive center adjacent to the benzene ring, constitute the classical examples in which this type of analysis is possible. Aliphatic compounds in general do not display these properties.

Table 4 shows the thermodynamic constants of the disproportionation reaction for the three anhydride systems. It is evident that ΔS is far from being zero and ΔF is not even of the same sign as ΔH . Figure 4 shows the logarithm of the equilibrium constant for the disproportionation reaction plotted against Hammett's constant for the para nitro, para methoxy and the unsubstituted benzoic-acetic mixed anhydride. It is seen that the deviations from a linear relationship are rather large.

Table 4

Thermodynamic Constants for the Disproportionation Reaction of the Para Substituted Benzoic-acetic Mixed Anhydrides at 100° C.

	▲ F(cal)	▲ H(cal)	AScal/OC
<pre>p Nitro p Methoxy p Hydrogen</pre>	-212	1,252	3.4
	-1.088	5,800	18.0
	-480	5,000	14.7



It was hoped that because the number and types of bonds in the mixed anhydride as compared to the symmetrical anhydrides are so nearly identical, this system could be analyzed in terms of changes in potential energy alone. The failure for this partly lies in the importance of the acetyl carbonyl group, which is removed from the phenyl group by a carbon and oxygen atom, as a reactive center in the disproportionation reaction.

Although it is probably not realistic to describe the differences in the thermodynamic characteristics of the mixed anhydrides in terms of changes in potential energy alone, it should be pointed out that a certain amount of correlation can be obtained by discussing these substances in terms of classical resonance structures.

It is noted that in each case the sign of the ΔH term is positive and hence the mixed anhydride is less stable than the symmetrical anhydrides. Proceeding from the para methoxy compound to the para nitro compound the differences in ΔH become smaller.

Weiland (12) has calculated the resonance energy for acetic anhydride as 41 Kcal/mole and a value of 29 Kcal/mole has been estimated by Franklin. (13)

Wieland suggests that this resonance energy is due to the following interactions:

It is to be expected that substituents in the para position of the symmetrical benzoic anhydrides might well affect the importance of structures such as III.

Structure III involves the creation of a charge on the central oxygen atom, and this charge is adjacent to a carbonyl carbon which already carries a partial positive charge. This situation tends to decrease the stability of III. If the para substituent X is a methoxyl group, then the partial positive charge on the carbonyl carbon is diminished and the resonance form III is of increased stability. A para nitro group exerts the opposite effect, and with an increased charge on the carbonyl carbons the contribution of III to the structure of p-nitrobenzoic anhydride is smaller. At the same time the effect of the para nitro group is to diminish the stability of only one of the corresponding structures (IV - A) in the mixed anhydride.

The effect of the substitution of a para nitro group is to decrease the energy difference between the symmetrical and the mixed anhydride by decreasing the resonance energy of the symmetrical anhydride to a greater extent than it decreases the resonance energy of the corresponding mixed anhydride and thus in a rough way to decrease the value of ΔH .

In the mixed anhydride the phenyl group may interact with the resonating system of the carbonyl groups so as to destroy the equivalence of the two structures corresponding to III. This would generally tend to decrease the stability of the mixed anhydrides relative to the symmetrical anhydrides.

Kinetics of the Disproportionation Reaction

Using the infrared spectrophotometric techniques described in the experimental section, it was possible to follow the disproportionation of a mixed anhydride into the symmetrical anhydrides and to examine the kinetic consequences of this reaction.

The p-nitrobenzoic-acetic mixed anhydride disproportionated at a rate that was considerably faster than either the p-methoxybenzoic-acetic or the benzoic-acetic mixed anhydrides. Because of its rather low solubility in chloroform, this was not a convenient case to study.

With \underline{p} -methoxybenzoic-acetic and benzoic-acetic mixed anhydrides the disproportionation reaction displayed kinetics

which could be unambiguously determined as a second order with respect to the mixed anhydride. See Figure 5A and B. Only the initial 20% of the reaction was followed, as at this point the back reaction became important and the apparent rate constant continuously decreased to zero.

Second Order Rate Constants for the Disproportionation of p-Methoxybenzoic-acetic Mixed Anhydride

A. Reaction followed by the disappearance of the mixed anhydride at $100^{\circ}\mathrm{C}$

$$k_2 = 1.29 \times 10^{-3} \text{ moles}^{-1} \text{ liters min.}^{-1}$$

B. Reaction followed by the appearance of \underline{p} -methoxybenzoic anhydride at 100°C

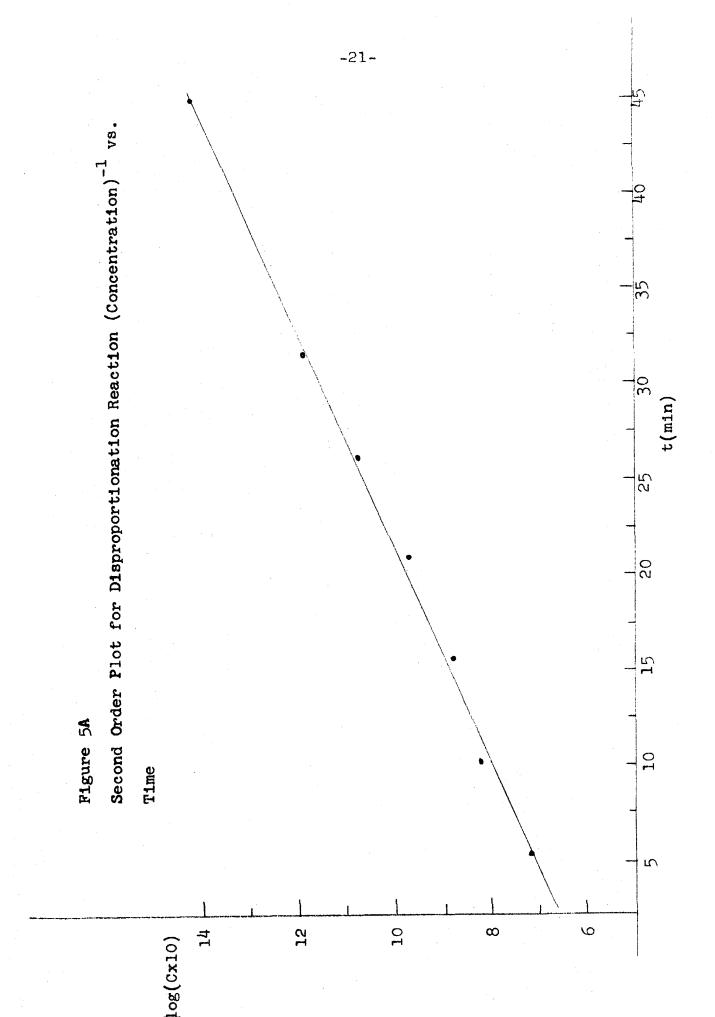
$$k_2 = 1.34 \times 10^{-3} \text{ moles}^{-1} \text{ liters min.}^{-1}$$

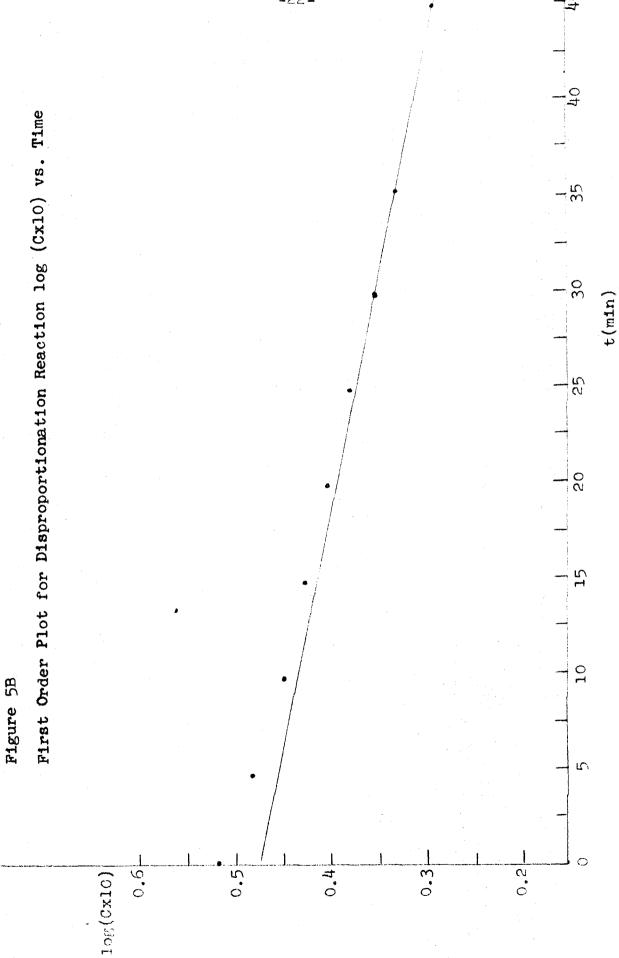
Second Order Rate Constants for the Disproportionation of Benzoic-acetic Mixed Anhydride

Reaction followed by the disappearance of the mixed anhydride at 100° C

$$k_2 = 2.27 \times 10^{-3} \text{ moles}^{-1} \text{ liters min.}^{-1}$$
 2.82×10^{-3} " " "
 2.43×10^{-3} " " "
 2.16×10^{-3} " " "

From the additional data presented in the experimental section, it is seen that the reaction is homogeneous, i.e., powdered glass does not affect the rate constant. The addition of sodium acetate increases the rate somewhat (p. 137).





This is easily explained in terms of the work of Evans, (14) who showed that acetate ion labeled with C¹⁴ very rapidly exchanges with acetic anhydride. Acetic or benzoic acid also increases the rate somewhat. This is understandable in view of the work in the next section.

The evidence set forth effectively excludes from consideration of the mechanism for the disproportionation reaction any scheme which involves as a rate determining step the production of ions as suggested below.

$$CH_3-C = 0$$

$$CH_$$

Rather it is believed that the second order reaction kinetics represent a manifestation of a true bimolecular mechanism. It is suggested that the approach of the two reacting molecules is facilitated by the dipole-dipole interaction of the carbonyl groups and that the "reactive intermediate" has the structure shown below.

The breakdown of the active intermediate is favored by a concerted shift of carbon-oxygen bonds throughout the pseudo six-membered ring.

The initial orientation of the reacting molecules is important and determines whether the combination could lead to the same or different species. Because of steric considerations, most reactions which are capable of resulting in a disproportionation probably proceed through an orientation of the activated complex in which the initial contact is formed by the carbonyl functions which have the methyl groups attached (see p. 29).

After the initial interaction of the carbonyl functions to which the methyl groups are attached, one can imagine that the facility with which the activated complex breaks down to give the disproportionated products is governed by the ease with which the carbon-oxygen bond two-three is formed and the carbon-oxygen bond three-four is broken. It is suggested that the mechanism of the disproportionation reaction is such that the rate determining step is the nucleophilic attack of the oxygen atom number two upon the carbonyl carbon atom number three. It is expected that the greater the susceptibility of this carbonyl atom to nucleophilic attack the faster will the rate of disproportionation proceed. The susceptibility of the carbonyl carbon atom to this type of attack will be profoundly affected by the nature of the para substituent X. If this substituent is a nitro group, the nucleophilic attack would be expected to proceed more readily than if the substituent is a methoxyl group.

The experimental results confirm the prediction, formulated on the basis of this hypothesis, that the rate of disproportionation of the p-nitrobenzoic-acetic mixed anhy-

dride is greater than the rate of disproportionation of <u>p</u>-methoxybenzoic-acetic mixed anhydride, and that the rate of disproportionation of the benzoic-acetic mixed anhydride has a value intermediate between these two.

Brown and Trotter⁽⁷⁾ found that the rate of disproportionation of butyric-acetic mixed anhydride was one thousand times greater than the rate of disproportionation of benzoic-acetic mixed anhydride.

The Interaction of Acetic and Benzoic Acids with the Benzoic-Acetic Anhydride System.

By extending the infrared spectrophotometric techniques used to measure the anhydride systems to a five component mixture, it is possible to quantitatively measure the interaction of acetic or benzoic acids with the benzoic-acetic anhydride system. The para-methoxybenzoic or para-nitrobenzoic-acetic anhydride systems were not suitable for this study, as para-nitrobenzoic and para-methoxybenzoic acids are not soluble in chloroform.

In addition to the equilibrium involving only the anhydrides, the system can be described by defining two other equilibrium constants corresponding to reactions I and II.

$$T = \frac{1}{CH_{3}C-OH} + \frac{1}{CH$$

Table 5

Temperature	K (anhyd)	K (acid-I)	K (acid-II)	Composition of Initial Solution
75°C	1.25	1.58	1.97	acetic anhyd, ben- zoic anhyd, acetic acid
75°C	1.08	3.36	3.61	benzoic-acetic mixed anhyd, ben- zoic acid
100°C	2.00	1.85	2.04	benzoic-acetic mixed anhyd, ben- zoic acid
100°C	3.12	2.23	6.95	benzoic anhyd acetic acid
150°C	3.74	1.27	4.75	benzoic-acetic mixed anhyd, acetic acid
150 ⁰ C	4.94	1.29	6.33	acetic anhyd, benzoic acid

Unfortunately the accuracy of the analytical technique does not permit, with this complicated mixture, a very accurate determination of the equilibrium constants associated

with the interaction of the acids with the anhydrides. It is thus not possible to describe the details of the mechanism either in terms of a redistribution reaction or in terms of the disproportionation reaction of the para substituted benzoic-acetic mixed anhydrides. If the acid interaction followed the principles of a redistribution reaction, then the values of the first acid equilibrium constant should be one half and those for the second acid equilibrium constant should be two.

A source of ambiguity is the uncertainty over which molecular species of the acid is able to react with the anhydrides. Recent work (15) has indicated that in the concentration range in which these measurements were made acetic acid is almost entirely converted to the dimeric form. It is possible to write a mechanism for the reaction of the monomeric acid with an anhydride which is formally quite similar to that proposed for the disproportionation reaction.

From this point of view it does not appear likely that the dimeric form of the acid would react as readily as the monomeric species. If the monomeric form of the acid is the only one which reacts, then the true acid equilibrium

constants will be proportional to the square roots of the observed (or dimeric) acid concentrations. If the values for K (acid-I) and K (acid-II) are corrected by using the square roots of the observed acid concentration, no improvement in the consistency of the results is observed. To settle this point a more sensitive analytical procedure will have to be employed.

The Nucleophilic Cleavage of the Benzoic-acetic, p-Methoxy-benzoic-acetic, and p-Nitrobenzoic-acetic Mixed Anhydrides.

In this section attention is directed towards the reaction of a mixed anhydride with nucleophilic agents in general and amines (p-toluidine) in particular. In the experimental section an analytical scheme is presented which enables one to determine the product composition of the reaction between a mixed anhydride and p-toluidine. From this procedure a quantity called the acetylation ratio is determined. It is defined as the amount of N-acetyl-p-toluidine found in the gross product divided by the amount of N-(p-substituted benzoyl)-p-toluidine found in the gross product.

Table 6
Acetylation Ratio

Reaction Medium	Benzoic-acetic Mixed Anhyd	<pre>p-Methoxybenzoic- acetic Mixed Anhyd</pre>	
Benzene	43.9	102.4	2.7
50 w/w Acetone- Benzene	37.7	140.7	4.7
Acetone	29.6	120.5	3.5
50 w/w Acetone- Water	42.5	93.8	3.3

It is evident that in this case the acetylation ratio is relatively independent of the polarity of the reaction solvent. This result can be expected when there are no substituents which could shield the carbonyl groups and whose steric size could be altered through changes in the amount of their solvation. Thus one would not expect the results that Gold (9) obtained with a chloroacetic-acetic mixed anhydride whose acetylation ratio was highly dependent upon the polarity of the reaction medium.

All three of the para substituted benzoic-acetic mixed anhydrides yield a preponderant of N-acetyl-p-toluidine, demonstrating the steric effectiveness of the phenyl group in hindering the attack of p-toluidine at the carbonyl function which is attached to the phenyl group.

The acetylation ratio decreases as one passes from p-methoxybenzoic-acetic mixed anhydride through benzoic-

acetic mixed anhydride to the p-nitrobenzoic-acetic mixed anhydride. The marked difference between the p-nitro and p-methoxy compounds is probably due to the combination of mesomeric and inductive effects, which for the p-nitro group will increase the positive charge at the carbonyl carbon atom adjacent to the phenyl ring and for a p-methoxy group will decrease the charge. It is expected that as the charge on one of the carbonyl carbon atoms is increased by substitution in some part of the molecule, the nucleophilic attack will be increasingly favored at that position, and hence the acetylation ratio will change accordingly.

In like manner it would be expected that the rates of hydrolysis of a mixed anhydride should be affected in a similar fashion. The more the positive charge of the carbonyl carbon atom adjacent to the phenyl ring is decreased by the action of substituents in the phenyl ring the slower the rate of hydrolysis. As the positive charge on the carbonyl group is increased, it is expected that the rate of hydrolysis will increase, since the nucleophilic attack would not tend to discriminate against one of the reactive centers. This prediction has been verified, as is seen in Table 7, where apparent first order hydrolysis constants for the mixed anhydrides are given. Berliner (16) also obtained similar results with the symmetrical para substituted anhydrides.

Table 6

Hydrolysis Constants for Para Substituted Benzoic-acetic Mixed Anhydrides in 50% w/w Tetrahydrofuran-water

Benzoic-acetic $\frac{p}{acetic}$ Mixed $\frac{p}{acetic}$ Mixed $\frac{p}{acetic}$ benzoic-acetic $\frac{p}{acetic}$ Mixed $\frac{p}{acetic}$ Mixed $\frac{p}{acetic}$ Mixed Anhyd Mixed Anhyd Mixed Anhyd $\frac{p}{acetic}$ Order $\frac{p}{acetic}$ Anhydride $\frac{p}{acetic}$ Mixed Anhyd $\frac{p}{acetic}$ Mixed Anhyd $\frac{p}{acetic}$ Anhydride $\frac{p}{acetic}$ Mixed Anhyd $\frac{p}{acetic}$ Order $\frac{p}{acetic}$ Anhydride $\frac{p}{acetic}$ Mixed Anhyd $\frac{p}{acetic}$ Anhydride $\frac{p}{acetic}$ Mixed Anhyd $\frac{p}{acetic}$ Anhydride $\frac{p}{acetic}$

EXPERIMENTAL SECTION.

PREPARATION OF STARTING MATERIALS

Symmetrical Anhydrides

A brief survey of available methods for the preparation of symmetrical anhydrides was made. At that time it was judged that the procedure suggested by Wheland (5) was the most convenient and gave high yields consistent with purity.

This method is based upon the reaction of a carboxylic acid with phosgene in the presence of a tertiary amine and in an inert solvent to form an unstable mixed anhydride of carbonic acid. The mixed anhydride then undergoes rapid disproportionation forming the symmetrical carboxylic anhydride and carbon dioxide.

It was found necessary to carry out the reaction at -10°C where yields of 80% were obtained. At 0°C only 10% of the desired product was formed.

Procedure

In the following manner benzoic anhydride, symmetrical \underline{p} -methoxybenzoic, and symmetrical \underline{p} -nitrobenzoic anhydrides were synthesized:

One tenth mole of the desired acid was dissolved in 250 ml. of dry tetrahydrofuran. The solution was placed in a 500 ml. three necked round bottomed flask equipped with a sealed mechanical stirrer and protected from the atmosphere with a calcium chloride drying tube. Twenty-four one hundredths mole of dry triethylamine was added and the mixture cooled to -10° C in an ice salt bath.

A solution of 0.11 mole of phosgene in 50 ml. tetrahydro-furan was prepared at room temperature and then cooled to -10° C. The cooled phosgene solution was slowly added by means of a dropping funnel to the stirred solution of acid and triethylamine. The process of addition requires one half hour.

The solid triethylamine hydrochloride was separated by rapid filtration and the solvent removed by distillation under reduced pressure in the presence of nitrogen. The results are shown in Table 7.

In order to detect small quantities of the parent acid

^{*}Tetrahydrofuran is preferred as a reaction solvent, since it dissolves larger quantities of the aromatic acids than do the other low polling inert solvents. It may be conveniently dried by distillation over sodium wire but cannot be stored in the presence of sodium without further treatment with lithium aluminum hydride to avoid slow deterioration.

in a sample of anhydride, recourse can be made to spectrophotometric analysis in the infrared spectra. The carbonyl
stretching frequencies in the neighborhood of 5.7-5.9 microns
are used, since they are very intense bands. The absorption
bands of the anhydride are shifted 0.13 microns toward
shorter wave length relative to the corresponding bands of
the acid. Using this method it is possible to detect five
parts in 1000 of acid in anhydride. The final criterion of
purity of a sample of anhydride was the complete absence of
any absorption bands of the acid in the spectrum of the
anhydride.

Table 7
Preparation of Symmetrical Anhydrides

Anhydride	M.P. after First Recrystallization	Final <u>Yield</u>	Final M.P.	Charac	ength of teristic tion Anhyd.
Benzoic	43.0-44.5	71%	44.5-45.0	5.89	5.78
<u>p</u> -Methoxy- benzoic	87-92	53%	95.5-96.2	5.87	5.74
p-Nitro- benzoic	188-190	48%	191.5-192.0	5.90	5.79

Recrystallization Solvent Benzoic Anhydride 2 Vol. Petroleum Ether . 1 Vol. Benzene p-Methoxybenzoic Carbon Tetrachloride p-Nitrobenzoic Purified Acetone

The acetic anhydride used in the study was purified by distillation over sodium wire and fractionated through 150 cm. of helices in a Todd column. The middle third portion was retained. The material was stored over sodium wire. Each preparation was used for two weeks only, as acetic anhydride seems to attack sodium wire slowly at room temperature.

Mixed Anhydrides (Para Substituted Benzoic-Acetic Mixed Anhydrides)

An old method of obtaining mixed anhydrides involves the reaction between the sodium salt of one acid and the acid chloride of another. This method suffers from the fact that the sodium chloride forms a finely divided precipitate and a colloidal suspension usually results. Separation of this reaction mixture usually involves lengthy filtration or contracted centrifugations, which result in lower yields. The method of choice is therefore the reaction of ketene and the desired aryl carboxylic acid. This results in the case of benzoic and p-methoxybenzoic acids in rapid quantitative conversions to the para substituted benzoic-acetic anhydride.

p-Nitrobenzoic acid, however, failed to react in a straight forward manner with ketene, and it was necessary to fall back upon the reaction between sodium p-nitrobenzoate and acetyl chloride.

Preparation of Mixed Anhydrides by Reaction of Aryl Carboxylic Acids and Ketene

One hundredth mole of the desired acid wasdissolved in 200 ml. of dry tetrahydrofuran contained in a 500 ml. Erlenmeyer flask. The flask was then placed on a magnetic stirrer and mildly agitated, while a slow steady stream of ketene was passed into the solution. The reaction was followed by dipping a stirring rod into the solution and allowing the adhering solution to evaporate. As the reaction proceeded smaller quantities of solid remained on the stirring rod after the evaporation of the solvent, and at the end of the reaction only a heavy oil was produced. Near the end of the reaction the rate of addition of ketene was decreased and the solution closely observed. As soon as a slight brown color had appeared, the reaction was stopped. This brown color is due to a slight excess of ketene in the solution, which is quickly converted into dark colored condensation products.

The ketene was produced by the pyrolysis of acetone as described by $\operatorname{Hurd.}^{(17)}$ The time for addition of ketene was 45 minutes.

The solvent was then removed from the reaction mixture by distillation under reduced pressure and the residue purified by fractionation or recrystallization.

Yield after (B.P. 88-89°at Fractionation 80% 0.2 mm. (M.P. 1.5°C 7.5 = 1.5152

<u>p</u>-Methoxybenzoic-acetic Mixed Anhydride Yield after
Recrystallization 79% (M.P.)
from Benzene (43-44°C)

Synthesis of p-Nitrobenzoic-acetic Mixed Anhydride

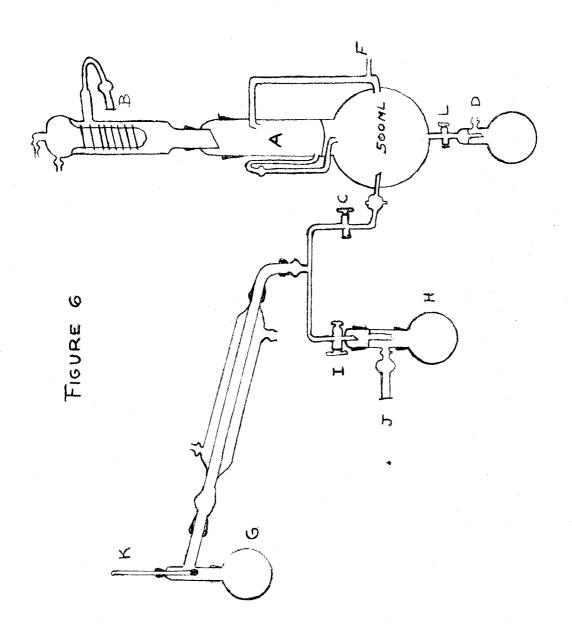
When p-nitrobenzoic acid was treated in the above manner with ketene, only starting material could be obtained. Therefore, it was necessary to utilize the reaction between sodium p-nitrobenzoate and acetyl chloride to obtain the desired product.

The mixed anhydride is a relatively unstable substance. It reacts quickly with any moisture in its environment and upon heating is rapidly disproportionated to the symmetrical anhydrides. Therefore, in order to prepare pure samples of this anhydride, it is necessary to carry out the reaction at room temperature and in the complete absence of moisture. Sodium p-nitrobenzoic and acetyl chloride react vigorously with the evolution of heat, and hence it is necessary to introduce the acid chloride diluted with an inert solvent.

To bring about these conditions the apparatus shown in Figure 6 was designed. The reaction unit is fundamentally a Soxhlet extractor modified in such a manner that liquid is transferred from the lower chamber to the upper by means of a vapor lift using nitrogen gas.

Procedure

A 75 x 150 mm. Soxhlet extraction thimble is soaked over night in reagent grade acetyl chloride, washed with anhydrous ether and dried in the oven at 120° for 15 min. Without being cooled, it is filled with dry sodium p-nitro-



benzoate and placed in the upper chamber of the modified Soxhlet extractor shown at A.*

In the flask G was placed 500 ml. of reagent grade chloroform, which was then slowly distilled; the distillate being collected in receiver H until 100 ml. had been col-The stop cock at C was opened and the one at I closed, and 120 ml. of chloroform was collected in the lower chamber of the reaction vessel. Stop cock C was then closed. Reagent grade acetyl chloride in the amount of 150 ml. was placed in flask G and distilled into receiver H until no hydrogen chloride gas was given off through the drying tube J and the reading of the thermometer K indicated that pure acetyl chloride was distilling. (About 20 ml. of the material collected.) Stop cock C was then opened and stop cock I closed. A stream of nitrogen was then admitted through F in such a manner that a slow steady stream of liquid contained in the lower chamber of the reaction vessel was circulated over the sodium p-nitrobenzoate contained in the extraction thimble in the upper part of the reaction In the course of 15 minutes, 30 ml. of acetyl chloride was transferred into the reaction chamber. Stop cock C was then closed and the material circulated for an

Before the reaction, the apparatus was thoroughly dried by placing a rubber stopper in the drying tube at B, the stop cock at C closed, and the reaction vessel evacuated through the port at D. Under reduced pressure the apparatus was flamed and then returned to atmospheric pressure by admission of dry nitrogen through port F.

additional five minutes. Stop cock L was then opened, and the liquid was transferred to the flask beneath the reaction vessel.

Seventy-five milliliters of dry chloroform was again distilled into the reaction vessel and circulated over the sodium p-nitrobenzoate for 15 minutes. This solution was then added to the main reaction mixture through stop cock L. The solvent was then removed by distillation under vacuum applied through port D.

The product was extracted twice with cold carbon tetrachloride, leaving behind a small amount of insoluble residue. An over-all yield of about 30% based upon sodium p-nitrobenzoate was obtained.

The purified material melted sharply at 86° C. Shortly after melting, a solid phase separated out of the clear melt. This material was identified as <u>p</u>-nitrobenzoic anhydride and presumably arose through rapid disproportionation of the mixed anhydride.

Each of the mixed anhydrides was purified until its spectrum did not change upon three consecutive purification operations. It was noted that distinct and prominent bands of the symmetrical anhydrides and of the acids were always missing from the spectra of the mixed anhydrides.

Choice and Purification of Solvent Materials

Of the various solvent materials available for use in infrared work, chloroform was found to be the most suitable.

For the <u>p</u>-nitro and <u>p</u>-methoxy systems it was a much better solvent than carbon tetrachloride, yet it did not have a strong absorption in the spectral region under investigation.

Pure chloroform has the rather distinct disadvantage of undergoing a slow photochemical conversion with the oxygen of the air to form phosgene.

The chloroform was purified by taking reagent grade material and washing it several times with 6-8 N sodium hydroxide, then several times with 6 N hydrochloric acid and finally with several portions of water. The solvent was then dried over calcium chloride and fractionated through a 300 cm. packed column under an atmosphere of nitrogen. The purified chloroform was stored in 25 ml. dark colored glass stoppered bottles at 5° C.

Only freshly prepared material was used for equilibrium studies, and once a sample was opened to the air it was discarded. With the above precautions the conversion of chloroform to phospene was not a troublesome factor.

Demonstration of the Interconversion of the Symmetric and Mixed Anhydrides

When a sample of a mixed anhydride is dissolved in an inert solvent and allowed to stand under anhydrous conditions, its infrared spectrum slowly changes and assumes a new configuration. The greatest change occurs in the region of the C-O-C stretching frequency between eight and nine microns and is characterized by the decrease in size of some of the

absorption peaks and the appearance of new peaks rather close to those which decreased. This change in configuration of the infrared absorption spectrum for benzoic-acetic mixed anhydride is illustrated in Figure 7. It is possible to identify all of these new peaks with those in the spectrum of one or the other of the symmetrical anhydrides.

In a converse manner it is observed that when the two symmetrical anhydrides are mixed in equal proportions, the initial spectrum of the mixture changes in a manner analogous to the way that the spectrum of the mixed anhydride changed. The new peaks that arise can be identified with those of the mixed anhydride. If given sufficient time, the final spectrum of the mixed anhydride and the spectrum of a mixture of equal portions of the symmetrical anhydrides become identical.

Therefore, it is judged that what is being observed is the equilibration of the three anhydride species among one another.

-45-

The pertinent spectra for the three anhydride systems studied are shown in Figures 8 to 19.

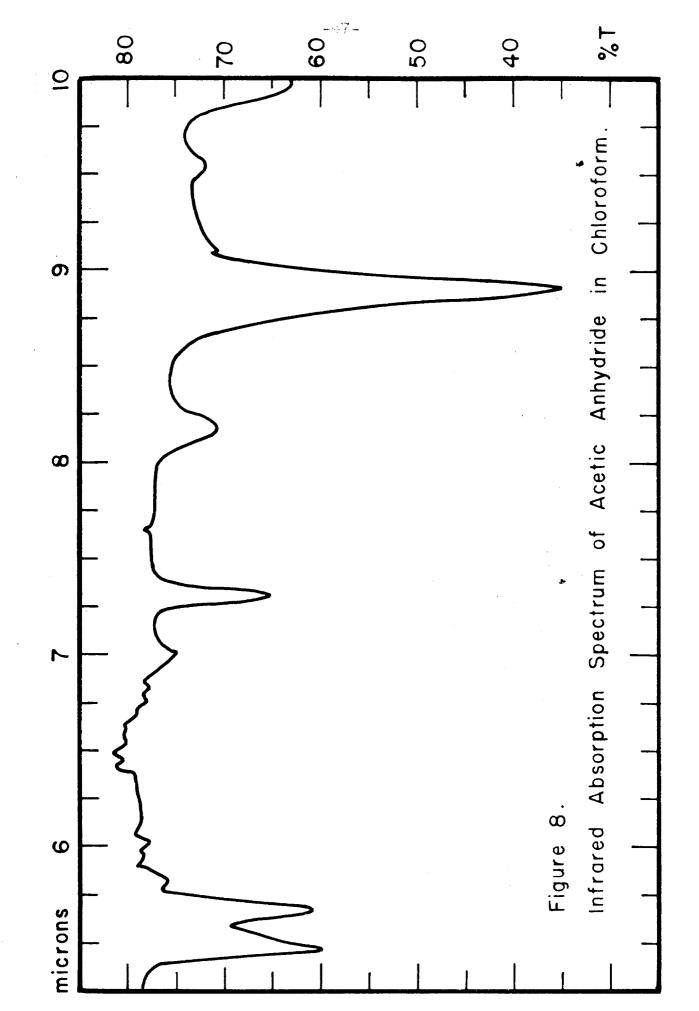
In exactly the same way it can be demonstrated that an acid and an anhydride can equilibrate to form a mixture of three anhydrides and two acids. See Figure 20.

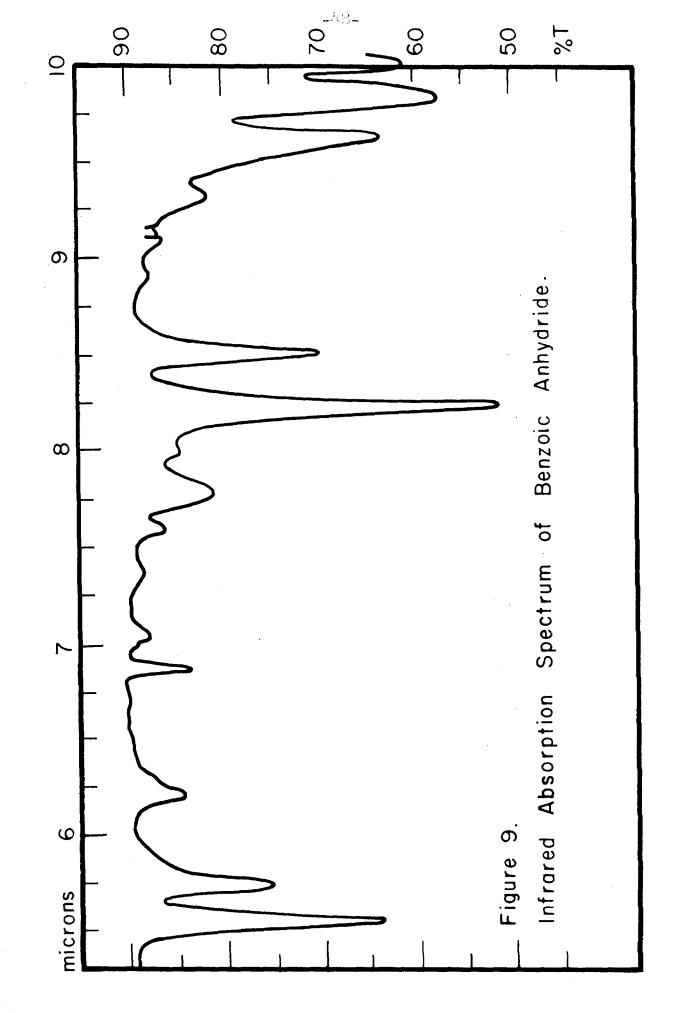
Quantitative Measurements Made with the Perkin-Elmer Model 21 Infrared Spectrophotometer

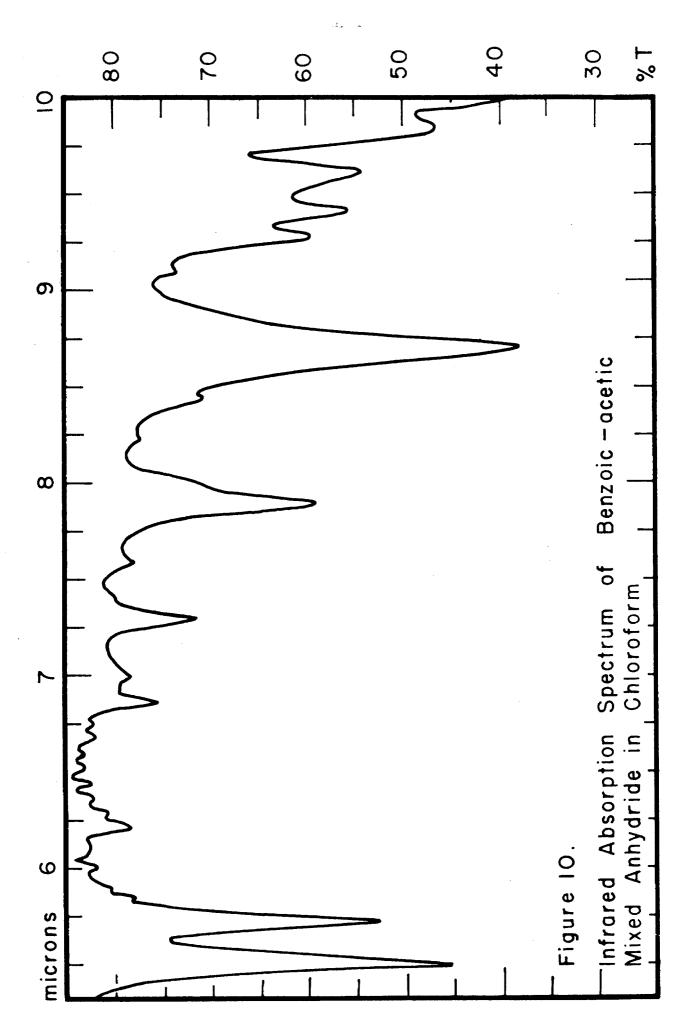
The Perkin-Elmer instrument is designed primarily for qualitative use and is most convenient to use in this capacity. When quantitative measurements are sought it is necessary to devise a special operating procedure, which is often quite tedious in execution.

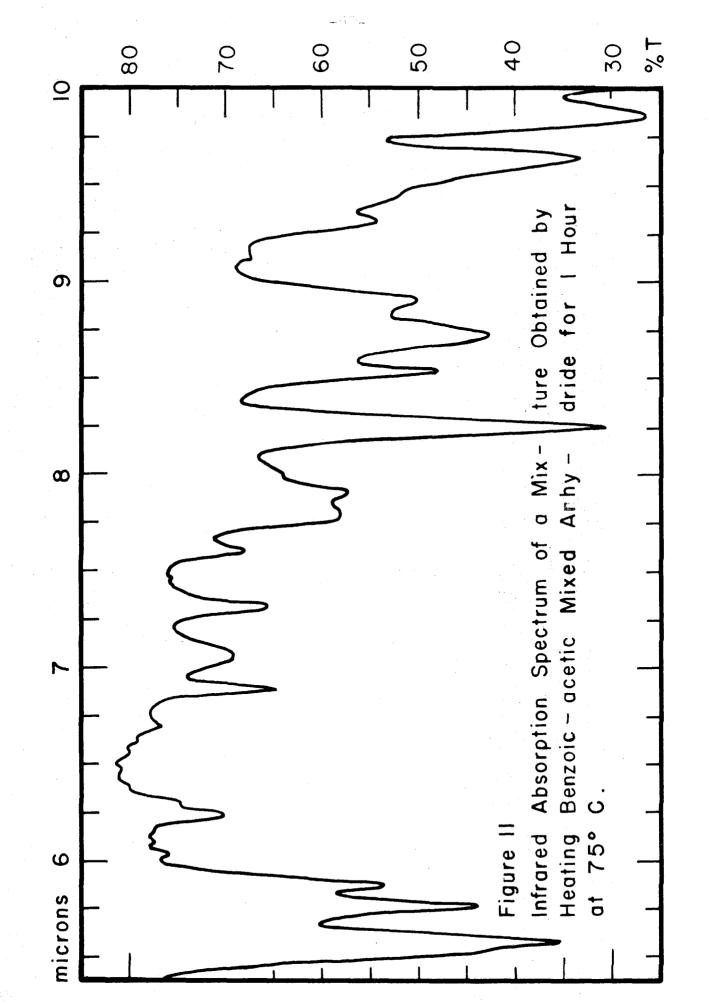
When operating in the eight and one half to nine micron region, the resolution control was set so that a slit width of 60-70 microns was obtained.

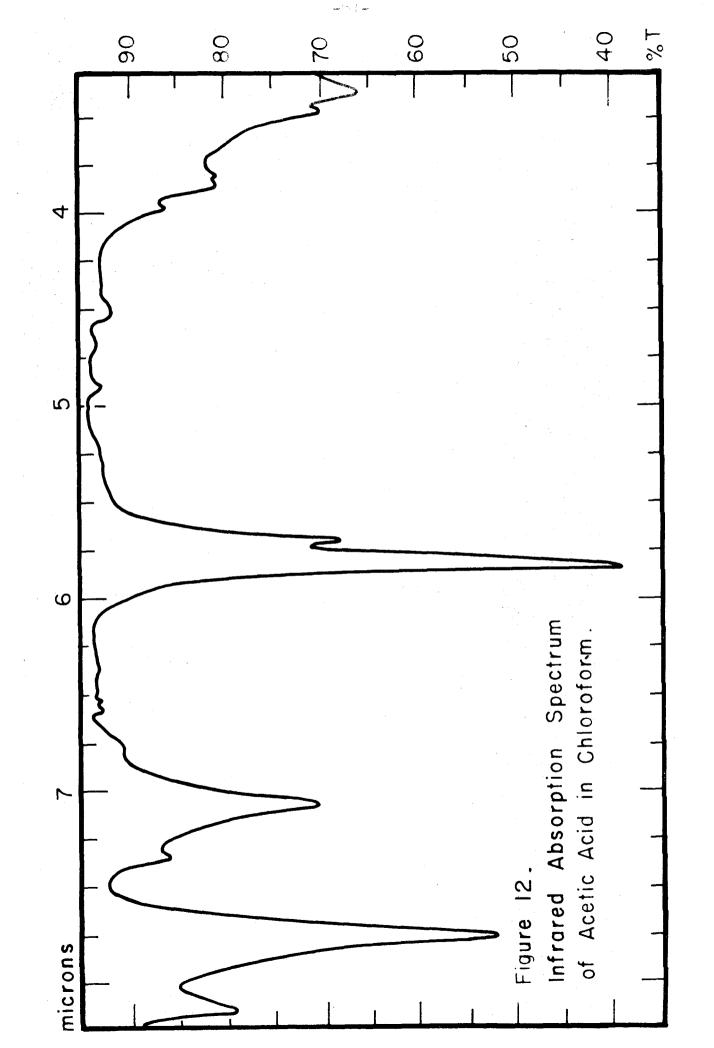
Reliance on the wave length scale was never used in setting the machine at the position of an absorption peak which was under consideration. Rather the outline of the peak was traced out each time, the machine reversed and allowed to retrace its first outline of the peak until the maximum was again reached. At this point of maximum absorption the wave length drivewas stopped and the machine readied for the reading of the optical density. It was found that the position of a maximum might vary about 0.02 microns from day to day.

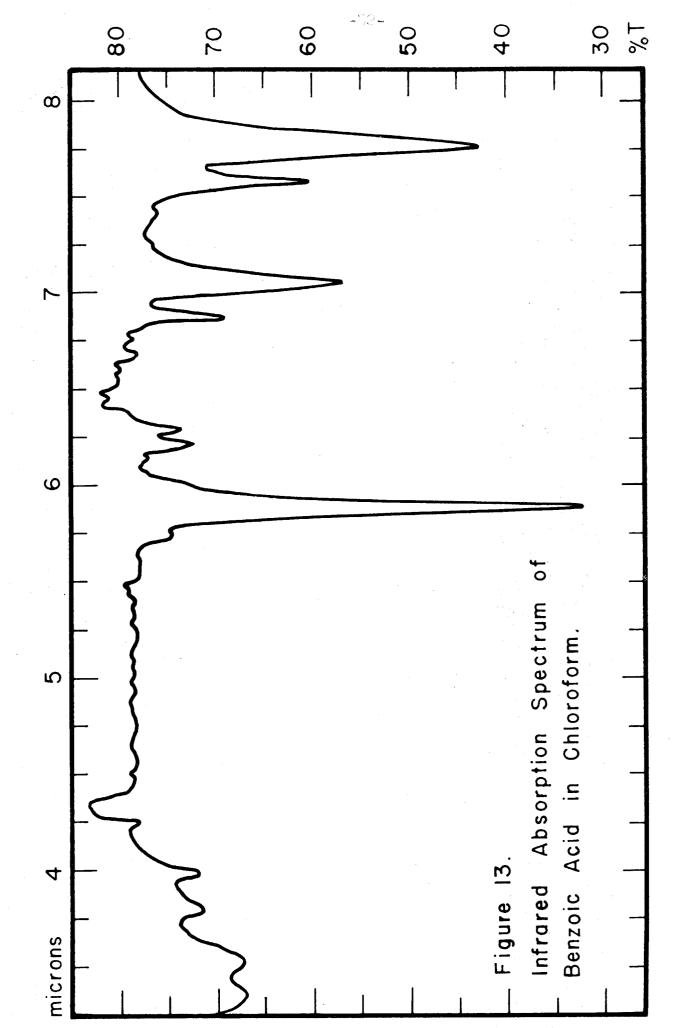


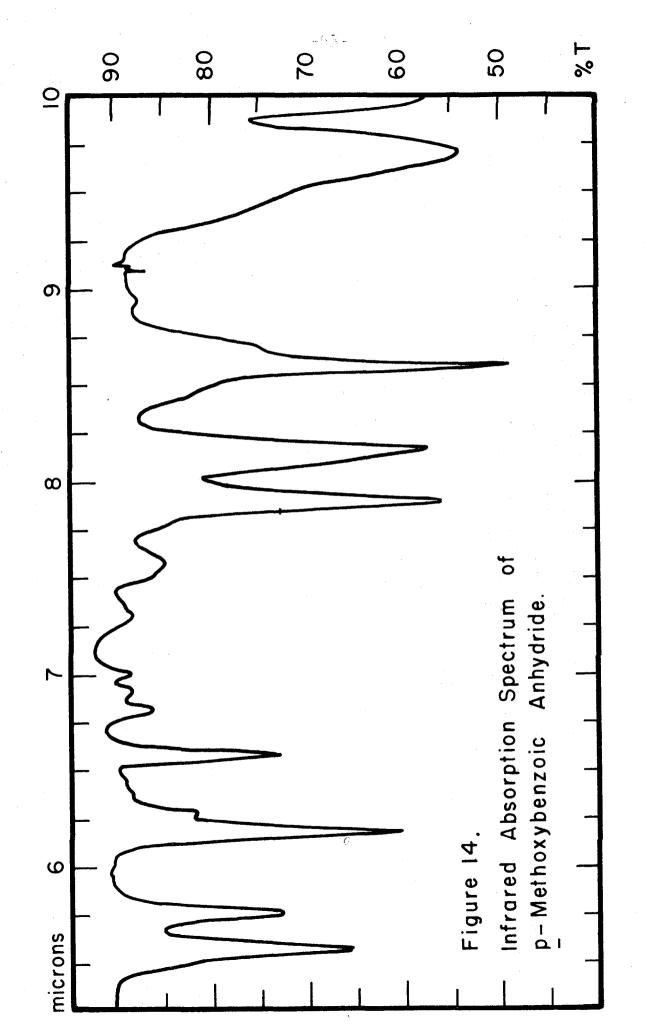


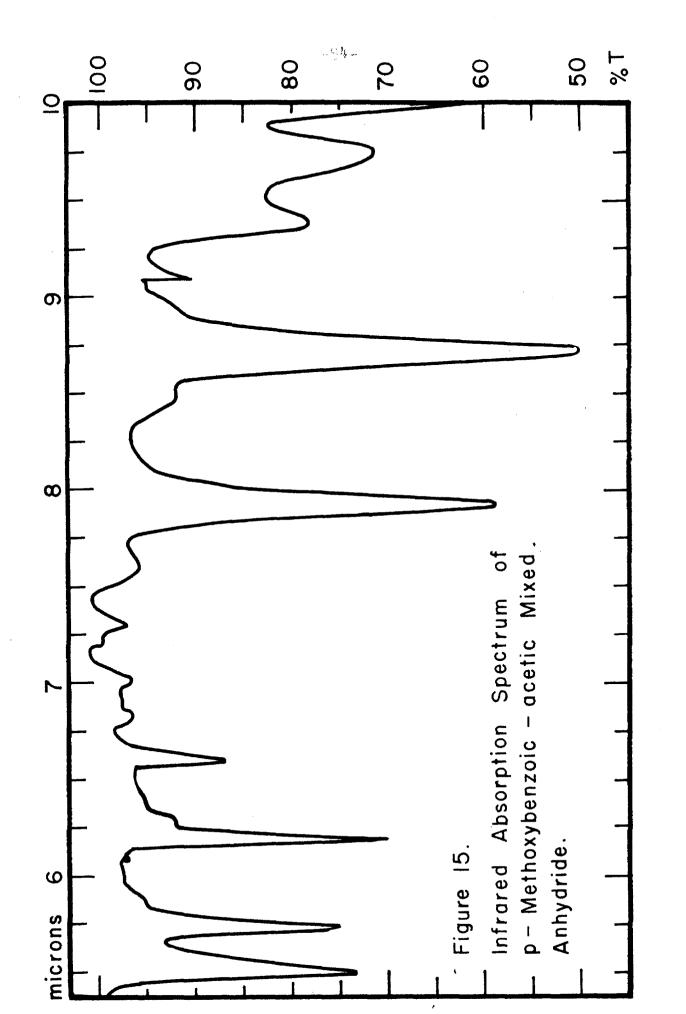


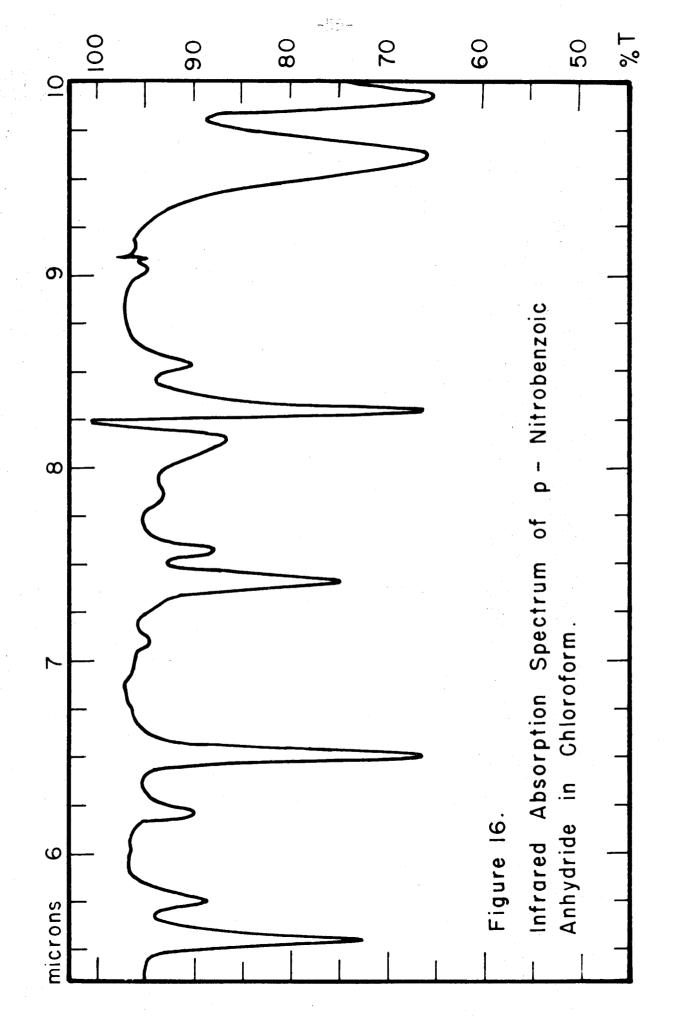


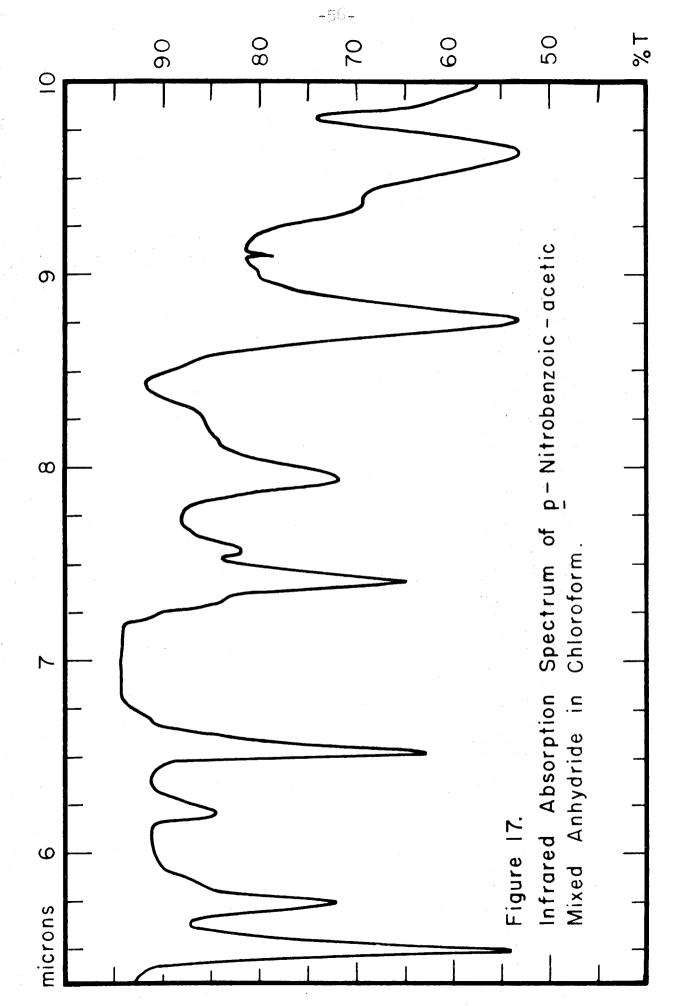


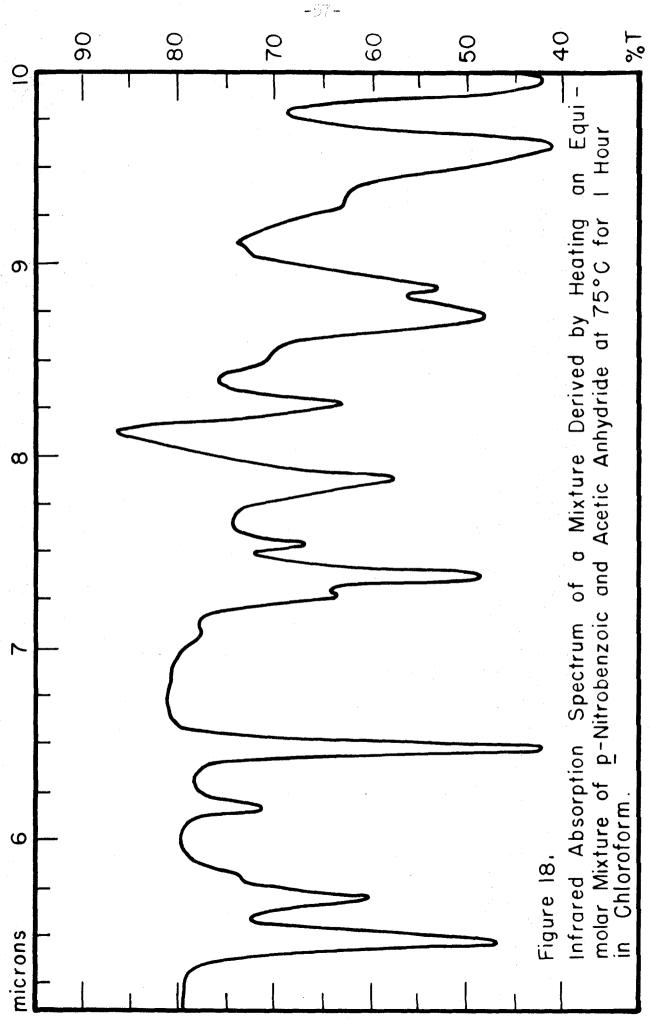


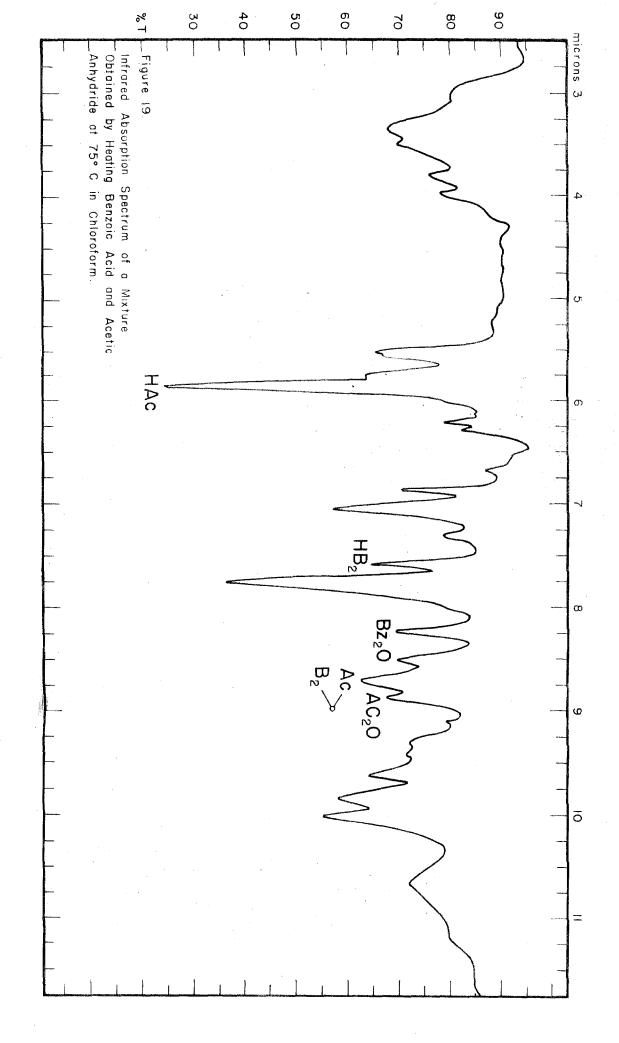


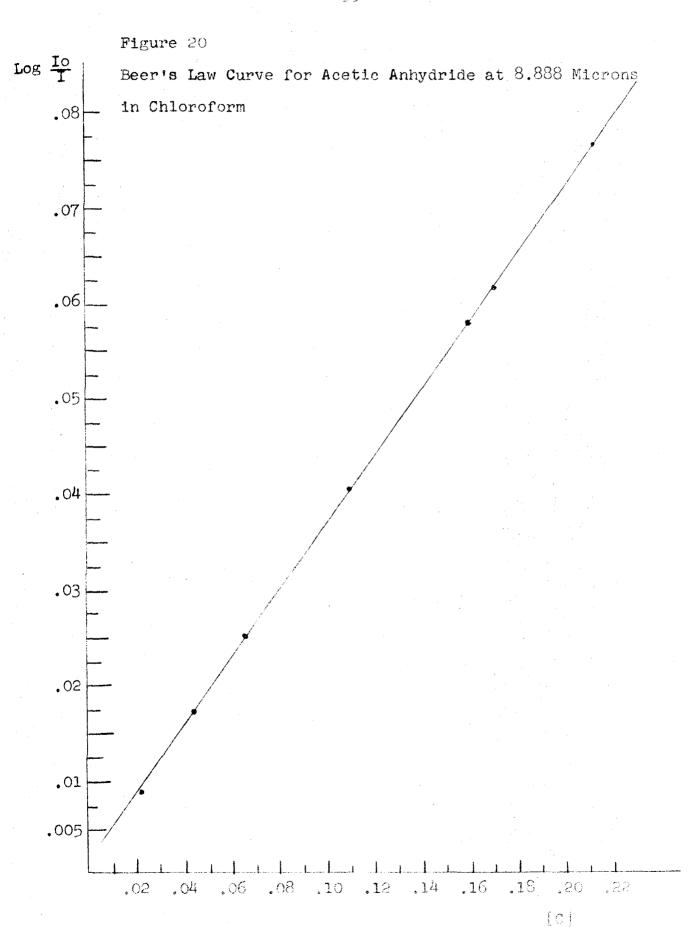












After the machine was adjusted so that it was reading the absorption of the desired peak, the "Gain," "Zero point," and "Balance" controls were adjusted with both reference and sample cells in place and filled with solvent (chloroform).

The "sample" light path was closed and the pen carriage was allowed to travel to the low end of the percent transmission scale. The "Gain" was then adjusted until a perceptible oscillation about an equilibrium point was just obtained. The "Gain" setting producing this effect was between five and six. The pen carriage was then moved on its drive cable until the equilibrium point of the small oscillations coincided with the zero point of the percent transmission scale. These adjustments were checked at the conclusion of each series of measurements.

After setting the "Gair" and the "Zero point" controls, the sample light path was opened and the pen allowed to travel to the high end of the percent transmission scale. The "Balance" control was then adjusted until a reading of 90-98% transmission was obtained.

If the pen carriage is displaced momentarily from its position at the high end of the percent transmission scale by slowly blocking the sample light path, and then by reopening the light path allowing the carriage to return, it is observed that the pointer does not always return to the same position at the high end of the percent transmission scale

that it formerly occupied. If a number of observations of this kind are made, it is observed that five or six consecutive readings are relatively constant (varying ± 0.1 of a transmission unit) and then that a period of instability is observed when the consecutive readings vary about 0.5 to 1.0 transmission units. After this brief period of instability, a period of quiescence is again reached, which may or may not be identical with the reading of the former period.

It was planned that the initial blank reading, the sample reading, and the final blank reading would all take place during one of these quiescent periods. To try to achieve this the following procedure was used in obtaining the value of the percent transmission of an unknown sample:

The sample cell was first filled with chloroform and several readings of %T were observed. When three consecutive values were obtained which did not differ more than ± 0.1 transmission units, the average value was noted and recorded as the first blank value. The sample cell was removed and quickly filled with the sample. The cell was then placed in its holder and the light paths reopened. The pen carriage was allowed to come to rest and then slightly displaced and again allowed to come to rest. Following this, the sample reading was obtained. The sample cell then was removed and quickly flushed and filled with solvent and the second blank reading was obtained. If the

second blank reading was greater than 0.4 transmission units from the first, the observations were discarded and the operations repeated.

Three sets of these observations were obtained for each quantitative measurement taken, and an average value of these observations was computed.

Another variable which must be controlled is the position of the cells in the cell holder. The cell holder is slightly larger than the flange dimensions of the absorption cells. It is necessary that the cells always occupy the same position relative to the light beam for each observation. This was brought about by making certain that the cell flanges always were flush against the side of the cell holder which is nearest the operator.

The Determination of the Various Beer's Law Curves

To determine the absorption characteristics of the various anhydride species, a given quantity of the anhydride was dissolved in chloroform. By dilution of this stock solution and by the use of the procedure which was outlined in the previous section, the optical density of the anhydride at various concentrations was obtained. In general the range of concentrations over which measurements were determined was chosen so as to cover the limits between which most of the future measurements would be made.

A typical plot of optical density vs. concentration is shown in figure 20.

It is to be noted that while a linear relationship is obtained, the curve does not pass through the origin. Therefore, a strict proportional relationship between optical density and concentration does not describe the situation which is presented by several of the anhydride species. It is therefore necessary to use the value of the intercept on the concentration axis to obtain a relationship which is useful over the concentration range of interest.

It does not seem possible at the present time to ascribe a cause for this non-proportionality. It is interesting to note that when another series of measurements were made using cells with a much wider absorption path (1.0 mm.), a linear relationship was obtained which passed through the origin. These cells permitted the use of much more dilute solutions (0.004 M). This suggests that perhaps the relationship is not strictly linear, but due to the insensitiveness of the analytical technique, this second order effect is not observed.

All the measurements in this study were made with a single set of absorption cells. The absorption path of the sample cell was 108.5 microns, and that of the blank was 105.3 microns. The data for all the Beer's law curves pertain to the use of these cells and do not strictly apply to any other set of cells.

The data obtained from these measurements and a summary of the various Beer's law constants can be found on pages to 111.

Measurements of the Equilibrium Constants

The absorption measurements were made as described previously. It was found that at 70°C approximately three days were necessary to insure that the various species present had reached a state of equilibrium. However, it was adopted as standard practice to allow four days for the equilibrium state to be established.

Measurements at high temperatures were made by sealing the samples under nitrogen in glass ampules made of 10 mm. pyrex tubing. The ampules were then placed in a small steel bomb (a war surplus bail out oxygen cylinder). The bomb was then partially filled with chloroform and inserted in a cotton seed oil bath, heated by a 200 watt immersion heater and controlled by a mercury thermoregulator to ± 0.5°C. After the period of heating, the sample tube was quickly cooled to -10°C and the values of the infrared absorption at the various wave lengths determined as soon as possible. The data from these measurements are presented on pages 112 to 128.

Determination of the Rate of Disproportionation

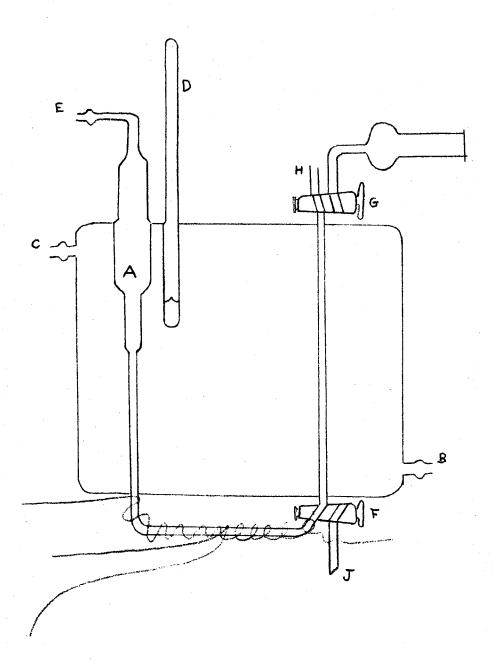
The rate of disproportionation was determined by heating a sample of the mixed anhydride at 100°C and withdrawing samples of the hot material at various times for measurement of the mixed anhydride content. Since the mixed anhydrides react rather rapidly with moisture, it is necessary to carry out the heating and sampling under anhydrous

conditions. To bring this about the special apparatus shown in Figure 21 was constructed.

The chamber where the samples of mixed anhydride were heated while undergoing disproportionation is shown at A. The temperature is controlled by the flow of steam through B-C. and recorded by thermometer D. A sample is withdrawn for measurement by applying a slight positive pressure of nitrogen at E. Stop cocks F and G are adjusted such that the anhydride can flow out of exit port H. The mixed anhydride is allowed just beyond the stop cock G into H and then F is turned 90°. G is turned 180°. When the desired time has arrived to take a sample, stop cock F is turned another 90° and the mixed anhydride allowed to flow directly into a volumetric flask containing chloroform. The exit tip J is placed under the surface of the chloroform, so that the transfer of the mixed anhydride is made without it ever coming in contact with the air. After the transfer is made, the sample is quickly adjusted to volume and then quenched to -10°C.

The anhydride sample, when passing from the reaction zone to the measuring tube between stop cocks F and G, must pass outside of the steam heated zone. To avoid errors introduced by this passage, the external section is wrapped with a nichrome heating element and a thermocouple attached to the glass wall at the center of the external passage. The whole array is wrapped with asbestos tape. The temperature in the external segment is controlled by varying the passage of current through the heating element in response to the

FIGURE 21



thermocouple reading.

The volume of the tube between stop cocks F and G was found to be $0.1154~\mathrm{cm}^3$.

The samples were kept in the -10°C bath until immediately before they were placed in the sample cell of the infrared spectrophotometer.

The data obtained by this procedure are shown on pages 129 to 138.

Determination of the Nature of the Reaction of the Mixed Anhydride with Nucleophilic Agents

In order to determine the composition of the product mixture obtained from the reaction of a mixed anhydride with p-toluidine, recourse was again made to the infrared spectrophotometric analytical technique outlined in a previous section. With p-methoxybenzoic and p-nitrobenzoicacetic mixed anhydrides, the absorption bands associated with the characteristic vibrations of the methoxyl and nitro group served as a convenient means for measuring the amount of N-(p-methoxybenzoyl)-p-toluidine and of N-(p-nitrobenzoyl)p-toluidine. N-Benzoyl-p-toluidine was determined by the use of its carbonyl stretching absorption. The frequency of this absorption corresponded to the same absorption of the acetyl compound, but its intensity was greater. The acetyl compound was determined by the use of a relatively minor peak in its infrared spectrum. The assignment of this peak to a specific interaction does not seem possible at this

time.

The generalized procedure for determining the acetylation ratio is as follows:

A 0.05 formal solution of the mixed anhydride in the organic solvent was prepared, and an equal volume of an 0.11 molar solution of p-toluidine in the same or different solvent was added. The homogeneous mixture was allowed to stand for 24 hours and then was evaporated to dryness. The solids were then treated with 10% excess of the amount of 1 N sodium carbonate necessary to convert all the free acid present to its sodium salt. The mixture evaporated to dryness at room temperature. The mixture of the sodium salts, p-toluidine, and the toluidides was then extracted with two 50 ml. portions of hot benzene. The benzene solutions were evaporated to dryness and the dry solids treated with a sufficient quantity of 6 N hydrochloric acid to neutralize all of the p-toluidine present. The heterogeneous mixture was again evaporated to dryness at room temperature. The mixture of p-toluidine hydrochloride and the toluidides was again extracted with two portions of warm benzene. The benzene was again removed by evaporation. Finally the remaining solids were taken up in chloroform, and from this solution the value of the acetylation ratio was obtained.

This procedure removes any chance for the alteration of the acetylation ratio, which might occur through several partitions between an aqueous and organic solvent. The results of this procedure are shown on pages 149 to 154.

The Determination of the Rate of Hydrolysis of the Mixed Anhydrides

The procedure for this determination has been well worked out. (2,18) It consisted, as modified for the present situation, of dissolving the mixed anhydride in a tetrahydrofuran-water mixture and pipetting samples of this solution at given times into an arresting solution. The arresting solution was composed of aniline and tetrahydrofuran. Its composition was adjusted so that a homogeneous phase was maintained upon the addition of the anhydride solution. The concentration of aniline in the arresting solution was at least twice that of the anhydride. The aniline reacted very quickly with the anhydride present in the aqueous solvent. Titration of the arresting solution with standard hydrochloric acid easily gave the concentration of the anhydride in the aliquot portion.

The values of the apparent first order rate constants listed on page 155 have been corrected for the amount of tetrohydrofuran used to obtain homogeneity in the initial solution.

The Estimation of the Probable Error in the Determination of an Equilibrium Constant

The probable error in the slope and intercept of a line drawn through a series of points, which deviate from the line in a random fashion, can be estimated by the following series of formulae.

For y = a + bxProbable errors in a and b are (19)

$$P_{o} = \pi_{e} \sqrt{\frac{\Sigma x_{i}^{2}}{D}}$$
; $P_{b} = \pi_{e} \sqrt{\frac{m}{D}}$
where
$$\pi_{e} = 0.6745 \sqrt{\frac{\Sigma d_{i}^{2}}{(m-2)}} \quad D = m \Sigma x_{i}^{2} - (\Sigma x_{i})^{2}$$

Using the above formulation the probable errors in the slopes and intercepts of the Beer's law functions were computed for the benzoic-acetic anhydride system and are compiled in the table:

Probable Error in the Beer's Law Functions for the Benzoic-Acetic Anhydride System

Substance	Wavelen 8.224 microns Slope Intercept	gth 8.888 microns Slope Intercept	8.690 microns Slope Intercept
Acetic	0.360 0.007	3.49 0.024	0.662 0.009
Anhydride	±0.002 ±0.004	±0.05 ±0.002	±0.017 ±0.005
Benzoic	3.598 0.000	0.151 0.000	0.0912 0.000
Anhydride	±0.051 ±0.006	±0.001±0.002	±0.0004±0.0007
Mixed	0.290 0.004	0.663 0.004	2.292 0.005
Anhydride	±0.006 ±0.002	±0.004±0.003	±0.05 ±0.007

For the final computation of the concentration of an anhydride species the following third order determinant must be solved:

[anhyd_I]
$$a_1$$
 + [anhyd_I] b_1 + [anhyd_II] c_1 = N1
[anhyd_I] a_2 + [anhyd_II] b_2 + [anhyd_III] c_2 = N2
[anhyd_I] a_3 + [anhyd_II] b_3 + [anhyd_III] c_3 = N3

Where N1, N2, and N3 are the corrected optical densities

The probable error in a function (f) derived from a series of quantities (S $_{
m i}$) whose error is Q $_{
m i}$ is given by $^{(20)}$

P.E. in
$$f = \sqrt{\sum \left(\frac{\partial f}{\partial s_i}\right)^2 Q_i^2}$$

Therefore, for the anhydride species Anhyd I the probable error would be given by:

$$P.E.in andyd_{1} = \frac{\begin{vmatrix} N_{1} & b_{1} & c_{1} \\ N_{2} & b_{2} & c_{2} \\ N_{3} & b_{3} & c_{3} \end{vmatrix}}{\begin{vmatrix} \alpha_{1} & b_{1} & c_{1} \\ N_{3} & b_{3} & c_{3} \end{vmatrix}} + \frac{\begin{vmatrix} N_{1} & b_{1} & c_{1} \\ N_{2} & b_{2} & c_{2} \\ N_{3} & b_{3} & c_{3} \end{vmatrix}}{\begin{vmatrix} \alpha_{1} & b_{1} & c_{1} \\ \alpha_{2} & b_{2} & c_{2} \\ \alpha_{3} & b_{3} & c_{3} \end{vmatrix}} + \frac{\begin{vmatrix} N_{1} & b_{1} & c_{1} \\ N_{3} & b_{3} & c_{3} \end{vmatrix}}{\begin{vmatrix} \alpha_{1} & b_{1} & c_{1} \\ \alpha_{2} & b_{2} & c_{2} \\ \alpha_{3} & b_{3} & c_{3} \end{vmatrix}}$$

Where P. F. in
$$\begin{vmatrix} N_1 & b_1 & c_1 \\ N_2 & b_2 & c_2 \\ N_3 & b_3 & c_3 \end{vmatrix} = \sqrt{\frac{\left[(N_1)(b_2)(dc_3)\right]^2 + \left[(N_1)(c_3)(db_3)\right]^2 + \left[(C_3)(b_2)(dN_1)\right]^2 - - - + \left[(N_1)(b_3)(dc_3)\right]^2 + \left[(N_1)(c_2)(db_3)\right]^2 + \left[(C_3)(b_3)(dN_1)\right]^2}$$

When the above calculations were carried out for the data on the top half of page 120, the following results were obtained:

(Acetic Anhydride) = 0.0826 ± 0.0015

(Benzoic Anhydride) = 0.0340 ± 0.0015

(Benzoic-Acetic Mixed

Anhydride) = 0.0466 ± 0.0041

Equilibrium Constant = 4.311 + 0.144

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Summary of Data for the Benzoic-acetic Anhydride And Benzoic-acetic Acid Systems

Principal Wave length

Benzoic-acetic mixed anhydride ---- 8.690 microns

Acetic anhydride ----- 8.888 microns

Benzoic anhydride ----- 8.224 microns

Acetic acid ------ 5.823 microns

Benzoic acid ----- 7.570 microns

Beer's Law Curves

- A. Benzoic-acetic mixed anhydride
 - 1. At mixed anhydride wave length (8.690 microns) $Log \frac{Io}{T} = 2.292(M-B_z) + 0.0050$
 - 2. At acetic anhydride wave length (8.888 microns) $Log \frac{Io}{T} = 0.6625(M-Bz) + 0.0100$
 - 3. At benzoic anhydride wave length (8.224 microns) $Log \frac{Io}{T} = 0.2901 (M-Bz) + 0.0040$
 - 4. At acetic acid wave length (5.823 microns) $Log \frac{Io}{I} = 0.2818(M-Bz) + 0.0000$
 - 5. At benzoic acid wave length (7.570 microns) $Log \frac{Io}{I} = 0.2550(M-Bz) + 0.0000$
- B. Benzoic anhydride
 - 1. At mixed anhydride wave length (8.690 microns) $Log \frac{Io}{I} = 0.0912(Bz_20) + 0.000$
 - 2. At acetic anhydride wave length (8.888 microns) $Log \frac{Io}{I} = 0.1507(Bz_20) + 0.000$

- 3. At benzoic anhydride wave length (8.224 microns) $Log \frac{Io}{T} = 3.598(Bz_2O) + 0.000$
- 4. At acetic acid wave length (5.823 microns) $Log \frac{Io}{I} = 0.3090(Bz_20) + 0.000$
- 5. At benzoic acid wave length (7.570 microns) $Log \frac{Io}{T} = 0.2260(Bz_20) + 0.000$
- C. Acetic Anhydride
 - 1. At mixed anhydride wave length (8.690 microns) $\log \frac{Io}{T} = 0.6623(Ac_{0}0) + 0.0089$
 - 2. At benzoic anhydride wave length (8.224 microns) $Log \frac{Io}{T} = 0.3602(Ac_20) + 0.0075$
 - 3. At acetic anhydride wave length (8.888 microns) $Log \frac{Io}{I} = 3.491(Ac_2O) + 0.024$
 - 4. At acetic acid wave length (5.823 microns) $Log \frac{Io}{T} = 0.0575(Ac_2O) + 0.000$
 - 5. At benzoic acid wave length (7.570 microns) $Log \frac{Io}{I} = 0.0468(Ac_20) + 0.000$

D. Acetic Acid

- 1. At mixed anhydride wave length (8.690 microns) $Log \frac{Io}{I} = 0.0191(HAc) + 0.0050$
- 2. At acetic anhydride wave length (8.888 microns) $Log \frac{Io}{I} = 0.0339(HAc) + 0.0030$
- 3. At benzoic anhydride wave length (8.224 microns) $\log \frac{Io}{T} = 0.0384(HAc) + 0.000$
- 4. At acetic acid wave length (5.823 microns) $Log \frac{Io}{T} = 2.140(HAc) + 0.000$
- 5. At benzoic acid wave length (7.570 microns) $Log \frac{To}{T} = 0.0778(HAc) + 0.0065$

E. Benzoic Acid

- 1. At mixed anhydride wave length (8.690 microns) $Log \frac{Io}{I} = 0.0530 (HBz) + 0.000$
- 2. At acetic anhydride wave length (8.888 microns) $Log \frac{Io}{I} = 0.2458 (HBz) + 0.000$
- 3. At benzoic anhydride wave length (8.224 microns) $Log \frac{Io}{I} = 0.0404 (HBz) + 0.000$
- 4. At acetic acid wave length (5.823 microns) $Log \frac{Io}{T} = 1.710(HBz) + 0.0200$
- 5. At benzoic acid wave length (7.570 microns) $Log \frac{Io}{I} = 0.7790(HBz) + 0.000$

Simultaneous Equations Derived from the Beer's Law Data

A. No Acid Present

- 1. At 8.224 microns $3.598(Bz_20) + 0.3602(Ac_20) + 0.2901(M-Bz) =$ $Log \frac{Io}{T} 0.0115$
- 2. At 8.888 microns $0.1507(Bz_20) + 3.491(Ac_20) + 0.6625(M-Bz) =$ $\log \frac{Io}{I} 0.034$
- 3. At 8.690 microns $0.0912(Bz_20) + 0.6025(Ac_20) + 2.292(M-Bz) =$ $Log \frac{Io}{T} 0.014$

B. Acids Present

- 1. At 8.690 microns $2.292(M-Bz) + 0.0912(Bz_20) + 0.6025(Ac_20) + 0.0191(HAc) + 0.0530(HBz) = Log \frac{Io}{I} 0.019$
- 2. At 8.888 microns $0.6625(M-Bz) + 0.1507(Bz₂0) + 3.491(Ac₂0) + 0.0339(HAc) + 0.2458(HBz) = Log \frac{Io}{T} 0.0370$
- 3. At 8.224 microns $0.2901(M-Bz) + 3.598(Bz₂0) + 0.3602(Ac₂0) + \\ 0.0384(HAc) + 0.0404(HBz) = Log \frac{Io}{T} 0.0115$
- 4. At 5.823 microns $0.2818(M-Bz) + 0.3090(Bz_20) + 0.3602(Ac_20) + 2.140(HAc) + 1.710(HBz) = Log \frac{Io}{T} 0.0200$
- .5. At 7.570 microns $0.2550(M-Bz) + 0.2260(Bz₂0) + 0.0468(Ac₂0) + 0.0778(HAc) + 0.7790(HBz) = Log \frac{Io}{I} 0.0065$

Beer's Law Curve For Acetic Anhydride At 8.888 Microns

Concentration moles per liter	Blank	Sample	Blank	<u>%T</u>	Ave. Io	Log ave. Io
0.2122	92.6 92.6 92.4	15.9	92.4	17.367 17.189 17.226	5.7934	0.762932
0.1700	97.0 97.0 96.9	23.6	97.1	24.329 24.317 24.329	4.1109	0.613939
0.1591	90.6 90.9 91.7		91.3	26.350 26.344 26.310	3.8021	0.580021
0.1061	93.6 93.5 93.9	37.4 37.5 37.4	94.0	39.935 40.000 39.808	2.5095	0.399585
c.0636	91.4 91.0 91.2		91.1	55.969 56.123 56.030	1.7844	0.251456
0.0424	91.5 91.2 91.2	61.6	91.3	67.468 67.506 67.616	1.4808	0.170202
0.0212	91.6 91.1 91.2	74.1	91.0	81.331 81.383 81.229	1.2298	o.089838
	т.					

 $Log \frac{Io}{I} = 3.491(Ac_20) + 0.024$

Beer's Law Curve For Acetic Anhydride At 8.224 Microns In Chloroform

Concentrat moles per		Blank	Sample	Blank	<u>%T</u>	Ave. Io	Log ave. Io
0.6166		92.7 93.3 93.0		93.0 93.0 93.0	59.235 59.259 59.247	1.6850	0.22660
0.4144		93.0 93.1 93.2	65.0	93.0 93.4 93.1	69.784 70.270 69.672	1.4304	0.155460
0.3083		92.7 92.6 92.2	70.8 70.8 70.2		76.457 76.416 76.138	1.3099	0.117237
0.2122		93.0 93.1 93.3		93.2 93.4 93.4	82.706 83.002 82.806	1.2066	0.081566
0.1061		90.7 91.0 91.0	82.3	91.0 91.1 91.9	90.588 90.389 90.642	1.1044	0.043126
	Log	$S = \frac{Io}{T} =$	0.3602	(Ac ₂ 0)	+ 0.00	75	

Beer's Law Curve For Acetic Anhydride At 8.690 Microns In Chloroform

Concentrati		Blank	Sample	Blank	<u>%T</u>	Ave. Io	Log ave. To
0.4244		91.8 91.7 91.7	48.7 48.6 48.5	92.0 91.8 91.6	52.992 52.970 52.918	1.8882	0.276046
0.3183		91.3 90.8 91.3		90.7 91.2 91.0	60.439 59.890 60.230		0.220400
0.2122		92.0 92.0 91.3		92.3 91.4 91.1	71.296 70.883 71.052		0.148260
0.1061		93.9 93.0 92.4		93.5 93.0 92.4	83.564 83.870 83.874	1.1937	0.076892
0.06375		90.4 90.2 91.0		90.7 90.1 91.0	89.122 89.073 89.120	1.1222	0.050068
	Lo	g <u>Io</u> =	0.6623	(Ac ₂ 0)	+ 0.00	89	

Beer's Law Curve For Acetic Anhydride At 5.823 Microns In Chloroform

Concentration moles per liter	Blank	Sample	Blank	<u>%</u> T	Ave. To	Log ave. Io
1.6958	90.2 90.3 90.0	71.6 71.5 71.3		79.247 79.356 79.310	1.2609	0.100685
1.2718	90.5 90.3 90.4	76.2	90.3 90.3 90.5	84.623 84.385 84.466	1.1183	0.073065
1.0599	91.3 91.4 91.8			86.904 86.856 86.880	1.1510	v.061080
0.8479	90.2 90.0 90.1	80.5 80.2 80.3	90.5 90.2 90.0	89.196 89.101 89.172	1.1216	0.049638
Lo	g <u>Io</u> =	0.0575	(Ac ₂ 0)	+ 0.00) C	

Beer's Law Curve For Acetic Anhydride At 7.570 Microns in Chloroform

Concentration moles per liter	Blank	Sample	Blank	%T	Ave.Io	Log ave. Io
1.6958	93.5 93.6 93.3			83.422 83.484 83.556	1.1982	0.078532
1.2718	94.8 94.7 93.7	82.7 82.8 81.9	94.6 94.5 94.0	87.382 87.480 87.266	1.1447	0.058796
1.0599	95.9 94.3 95.0	85.5 94.3 84.7	95.7 94.6 94.8	89.248 89.253 89.251	1,1208	0.049508
0.8479	95.0 95.0 95.2	86.6 86.8 86.9		91.157 91.224 91.281	1.0949	0.039371
Lo	g <u>io</u> =	0.0468	(Ac ₂ O)	+ 0.000	00	

Beer's Law Curve For Benzoic Anhydride
At 8.224 Microns In Chloroform

Concentration moles per liter	Blank	Sample	Blank	%T	Ave. Io	Log ave. Io
0.1210	97.5 97.4 96.9	35.6 35.6 35.4	97.5 97.6 96.9	36.512 36.512 36.532	2.7385	0.43751
0.1061	93.8 94.1 9 3. 5	38.8 39.0 39.0	93.8 94.1 93.7	41.364 41.445 41.655	2.4124	0.382448
0.0975	97.5 97.4 98.0	43.3 43.3 43.5	97.5 97.6 97.8	44.410 44.410 44.433	2.2513	0.352437
0.08488	93.2 93.0 92.9	46.2	93.1 93.1 93.0	49.919 49.650 49.704	2.0097	0.303034
0.06366	92.4 92.0 92.6	54.5	92.8 92.5 92.4	59.179 59.078 59.027	1.6921	0.228530
0.04240	92.8 92.7 93.0	64.9	92.7 93.2 93.0	70.080 69.822 69.784		0.155510
0.00408	92.0 92.3 92.0	89.5		96.585 96.913 96.663	1.0339	0.014478
Lc	og <u>Io</u> =	3.598(Bz ₂ 0)	+ 0.000		

Beer's Law Curve For Benzoic Anhydride At 8.888 Microns In Chloroform

Concentrat moles per		Blank	Sample	Blank	<u>%T</u>	Ave. To	Log ave. Io
0.3183		95.5 95.7 94.5	85.4 85.3 84.4	95.7 95.6 94.8	89.330 89.179 89.170	1.1207	0.049493
0.19098		92.5 91.8 92.0		92.0 91.7 91.9	93.333 93.732 93.529	1.0691	0.029020
0.15612		92.5 92.7 92.6	87.5 87.5 87.5	92.5 92.5 92.4	94.594 94.492 94.594	1.0576	0.024322
0.10610		93.5 94.0 94.0		94.0 94.0 94.0	96.223 96.170 96.276	1.0392	0.016702
0.0795		92.2 92.1 92.2		92.3 92.2 92.2	97.018 97.124 97.180	1.0297	0.011864
	Lo	g <u>Io</u> =	0.1507	(Bz ₂ 0)	÷ 0.000	0	

Beer's Law Curve For Benzoic Anhydride At 8.690 Microns In Chloroform

Concentration moles per liter	· Blank	Sample	Blank	<u>%T</u>	Ave. Io	Log ave. Io
0.1061	92.5	90.7 90.5 90.6	92.7	97.842 97.732	-	0.009536
0.09549	92.1	90.4 90.4 90.3	92.0		1.0197	0.008471
0.08488	92.6	91.0 91.0 91.1	92.5	98.219 98.325 98.327	1.0173	0.007449
Lo	og <u>Io</u> =	0.0912	(Bz) +	0.000		

Beer's Law Curve For Benzoic Anhydride At 5.823 Microns In Chloroform

Concentration moles per liter	Blank	Sample	Blank	%T	Ave. <u>Io</u>	Log ave. Io
0.7250	96.0 95.8 95.9	57.4 57.3 57.3	96.4 96.0 96.2	59.667 59.749 59.656	1.6752	0.224062
0.4350	95.5	70.0 69.5 69.4	95.5	73.107 72.774 73.516	1.3673	0.135866
0.3625	94.6 94.9 94.7		95.1	77.085 76.906 77.085	1.2982	0.113340
0.2900	96.5 95.5 95.4	77.9 78.0 77.9	96.5 95.7 96.0	80.725 81.589 81.400	1.2309	0.090225
0.18125	95.5	84.0 84.0 84.1	95.4	87.958 88.004 88.062	1.1362	0.055446
Log	$\frac{Io}{I} =$	0.3090(Bz ₂ 0)	+ 0.000		

Beer's Law Curve For Benzoic Anhydride At 7.570 Microns In Chloroform

Concentration moles per liter	Blank	Sample	Blank	<u>ZT</u>	Ave. Io	Log Ave. To
0.7250	95.2 95.1 95.0		95.1	67.962 67.928 68.105	1.4707	0.167523
0.4350	94.4 94.1 94.4	75.1 75.1 75.0	94.3	79.639 79.723 79.533	1.2557	0.098885
0.3625	93.7 93.9 94.0	77.4 77.5 77.4	93.7 94.0 93.7	82.604 82.490 82.472	1.2117	0.083392
0.2900	93.8 93.9 94.2			85.729 85.790 85.259	1.1683	0.067551
0.18125	93.5 93.4 93.7	85.2 85.2 85.2	93.7 93.7 93.5	91.025 91.074 91.025	1.0984	0.040760
0.1450	93.7 93.6 94.1	86.9 87.0 86.9		92.643 92.750 92.643	1.0674	0.032380

 $Log \frac{Io}{I} = 0.2261(Bz_20) + 0.000$

Beer's Law Curve For Benzoic-acetic Mixed Anhydride at 8.690 Microns In Chloroform

Concentration moles per liter	Blank	Sample	Blank	<u> </u>	Ave. <u>Io</u>	Log ave. Io		
0.29095	98.6 98.0 98.0	21.3 21.2 21.2	98.7 98.0 98.1	21.591 21.632 21.621	4.6264	0.665246		
0.2645	91.3 91.3 91.2	21.8	91.4 91.3 91.1	23.864 23.877 23.916	4.1867	0.621870		
0.23276	98.4 98.2 98.0	25.5	98.3 98.1 98.1	26.029 25.980 26.007	3.8454	0.584944		
0.1980	91.5 90.8 90.1	31.5	90.6 90.5 90.4	34.696 34.749 34.570	2.8841	0.460015		
0.13225	92.3 92.0 91.8	45.3 45.1 45.0	92.4 92.2 91.9	49.052 48.968 48.992	2.0391	0.309441		
0.0990	91.2 91.1 91.2		91.6 91.4 91.3	58.469 58.520 58.520	1.7093	0.232818		
0.0661	91.1 91.3 91.4		91.4 91.2 91.3	67.945 67.945 67.870	1.4723	0.168003		
0.02695	91.5	77.0 76.8 77.0	91.0	84.164	1.1817	0.072549		
$Log \frac{Io}{I} = 2.292(M) + 0.005$								

Beer's Law Curve For Benzoic-acetic Mixed Anhydride At 8.224 Microns In Chloroform

Concentration moles per liter	Blank	Sample	Blank	<u>%T</u>	Ave. Io	Log ave. Io
0.5745	91.5 91.4 91.5	60.9	91.5 91.5 91.5	66.557 66.592 66.666	1.5013	0.176467
0.4220	91.2 91.5 91.5	67.7	91.5 91.5 91.4	74.110 73.989 74.029	1.3505	0.130495
0.3591	91.5 94.1 91.4	71.4	91.5 93.9 91.5	75.956 75.957 75.888	1.3164	0.119392
0.2680	92.2 91.5 92.0	75.6 75.5 75.5	92.0 92.0 92.0	82.084 82.288 82.065	1.2163	0.085038
0.1325	93.5 92.6 92.7	83.8	93.1 92.7 92.6	90.568 90.447 90.447	1.1063	0.043877
Lo	$\frac{Io}{I} =$	0.2901	(M) + (0.0040		

Beer's Law Curve For Benzoic-acetic Mixed Anhydride At 8.888 Microns In Chloroform

Concentration moles per lit		Sample	Blank	<u>%T</u>	Ave. To	Log ave. To
0.5745	93.5 93.5 93.6	43.5 43.6 43.6	93.6	46.524 46.606 46.556	2.1476	0.331950
0.4220	94.2 94.0 94.5	54.0	94.3 94.2 94.2	57.355 57.285 57.339	1.7444	0.24165
0.2680	93.6 93.0 93.0	64.2 64.3 63.7	93.9 93.2 93.0	68.480 69.065 68.494	1.4553	0.16295
0.1325	93.3 93.9 93.5		93.6 94.0 93.7	82.504 82.810 82.478	1.2136	0.084076
0.06625	93.5 93.8 93.9	84.6 84.5 84.5		90.336 89.941 89.941	1.1102	0.045398
	$Log \frac{Io}{T} =$	0.6625	(M) + (0.0100		

Beer's Law Curve For Benzoic-acetic Mixed Anhydride At 5.823 Microns In Chloroform

Concer	ntrat	tion						
			Blank	Sample	Blank	%T	Ave .Io	Log ave. $\frac{Io}{T}$
0.7835	5		98.5 98.6 98.5	59.3 59.2 59.3	98.5 98.6 98.6	60,203 60,040 60,172	1.6637	0.221072
0.6612	25		97.0 96.9 97.2		97.0 97.2 97.1	65.567 65.636 65.568	1.5246	0.183154
0.5290)		95.3 95.4 95.5	68.5 68.5 68.4		71.765 71.765 71.585	1.3946	0.144446
0.3967	'5		98.0 97.9 97.8	74.5	98.3 98.0 97.7	75.904 76.059 76.317	1.3084	0.116742
0.2645)		95.5 95.4 95.2	81.5	95.4 95.2 95.2	85.385 85.519 84.978	1.1724	0.069078
		Log	; Io =	0.2818(M) + 0	0.000		

Beer's Law Curve For Benzoic-acetic Mixed Anhydride At 7.570 Microns In Chloroform

Concentration moles per liter	<u>Blank</u>	Sample	<u>Blank</u>	%T	Ave Io	Log ave Io
0.66125	97.1 97.0 97.3	65.0 65.0 65.1	97.2 97.3 97.2	66.906 66.906 66.940	1.4944	0.174466
0.5290	96.5 97.0 96.5	70.0 70.0 70.0	96.7 97.0 96.7	72.463 72.164 72.463	1.3819	0.140378
0.2645	97.0 97.2 97.4	82.1 82.1 82.0	97.3 97.1 97.3	84.508 84.508 84.232	1.1846	0.073572
0.19838	97.1 97.2 97.4	86.6 86.6 86.7	97.2 97.2 97.5	89.140 89.094 88.968	1.1227	0.050263
0.13225	97.6 96.3 96.0	90.7 90.1 89.9	96.4 96.2 95.8	93.989 93.610 93.743	1.0701	0.029421
Log	$\frac{Io}{T} =$	0.2550((M) + (0.000		

Beer's Law Curve For Acetic Acid At 8.690 Microns In Chloroform

Concentration moles per lite	er Blank	Sample	<u> Blank</u>	<u> ST</u>	Ave. <u>Io</u>	Log ave. Io
1.424	98.2	85.4 85.4 85.5	97.9	87.098	1.1491	0.060358
1.187	97.5 97.3 97.7	87.6 87.5 88.0	97.3 97.2 97.9	89.938 89.974 89.979	1.1115	0.045905
0.7120	96.7	89.7 89.9 89.1	96.9	92.569 92.871 92.764	1.0783	0.03274
I	$\log \frac{Io}{I} =$	0.0191	(HAc) -	+ 0.005		

Beer's Law Curve For Acetic Acid At 8.888 Microns In Chloroform

Concentration moles per liter	Blank	<u>Sample</u>	<u> 3lank</u>	<u> ZT</u>	Ave. Io	Log ave. Io
1.424	98.5 98.4 98.0	78.4	98.2	79.634 79.755 80.061	1.2528	0.097880
1.187	98.6 98.3 98.0	82.9	98.3 98.0 98.2	84.408 84.462 84.403	1.1844	0.073498
0.7120	95.5 95.5 95.5	85.2		89.250 89.167 89.167	1.1213	0.049724
0.3560	95.3 95.5 95.6	89.4		93.501 93.514 93.913	1.0678	0.028488
Log	; <u>Io</u> =	0.03390	(HAc) -	- 0.003		

Beer's Law Curve For Acetic Acid At 8.224 Microns In Chloroform

Concentration moles per lite	er Blank	Sample	Blank	<u>%T</u>	Ave. Io	Log ave. Io
1.424	94.0	73.1	94.2	77.224 77.683 77.825	1.2890	0.11025
0.1187	93.9	82.8	93.7	88.342 88.272 88.263	1.1326	0.054078
0.3560	93.7	88.2	93.8	94.186 94.080 94.192	1.0621	0.026162
I	$\log \frac{Io}{I} =$	c.o384	(HAc)	+ 0.000		

Beer's Law Curve For Acetic Acid At 5.823 Microns In Chloroform

Concentration moles per liter	Blank	Sample	Blank	<u>%T</u>	Ave. To	Log ave. Io			
0.3560	94.5 94.0 94.1	19.4 19.3 19.3	94.5 94.0 94.2	20.529 20.531 20.499	4.8732	0.687818			
0.2848	95.4 95.0 94.8	23.7 23.6 23.5	95.2 95.1 94.7	24.868 24.829 24.802	4.0268	0.604958			
0.2541	92.0 91.8 91.7	25.9 25.9 25.9	91.5 91.9 91.6	28.383 28.198 28.259	3.5360	0.54851			
0.2136	93.4 93.0 93.2	31.6 31.5 31.6	93.0 93.2 93.1	33.905 33.834 33.923	2.9509	c.469955			
0.1825	96.2 96.5 96.0	38.3 38.4 38.2	96.5 96.8 96.1	39.750 39.730 39.770	2.5157	0.406590			
0.1762	93.0 92.8 93.1	37.5 37.5 37.6	92.6 92.5 93.2	40.409 40.474 40.365	2.4742	0.393436			
0.1460	95.2 95.7 94.8	45.1 45.2 44.9	95.6 95.5 95.0	47.274 47.280 47.312	2.1159	0.325499			
0.1250	93.2 93.7 93.5	48.8 49.0 48.9	93.1 93.5 93.3	52.388 52.322 52.355	1.9100	0.281030			
0.0701	93.5 93.3 92.9	63.0 62.7 62.7	93.5 92.9 92.9	67.379 67.346 67.491	1,4835	0.171290			
0.0350	92.5 92.2 92.0	75.9 75.5 75.4	92.5 92.0 92.3	82.054 81.976 81.823	1.2202	0.086432			
$Log \frac{Io}{I} = 2.140(HAc) + 0.000$									

(Concentration Limit 0.24 molar)

Beer's Law Curve For Acetic Acid At 7.570 Microns In Chloroform

Concentration moles per liter	Blank	Sample	Blank	%T	Ave Io	Log ave. Io
1.5150	95.0 95.6 95.8	70.0 70.1 69.9		73.529 73.364 73.002	1.3642	0.134874
1.1120	93.0 96.2 96.5		96.5	79.506 79.709 79.274	1.2579	0.109646
0.727	94.4 93.4 93.4	80.2	94.5 93.4 93.6	85.442 85.867 85.454	1.1644	0.066102
0.539	95.9 96.0 95.8	85.0 85.1 85.0	95.9 96.3 95.7	88.633 88.507 88.772	1.1281	0.052348
0.359	96.5 96.2 96.0	88.7 88.5 88.4	96.8 96.1 95.9	91.774 92.043 92.131	1.0871	0.036270
	т.	_	,	_		

 $Log \frac{Io}{I} = 0.0778(HAc) + 0.0065$

Beer's Law Curve For Benzoic Acid At 8.690 Microns In Chloroform

Concentration moles per lite	r Blank	Sample	<u>Blank</u>	<u> </u>	Ave. Io	Log ave. Io
0.7366	95.2 95.0 95.3	86.9	95.1	91.368 91.425 91.023	1.0956	0.039650
0.58928	96.8 97.1 97.2	90.7	97.3	93.347 93.321 93.319	1.0715	0.030190
0.3183	97.5 97.8 97.0	93.9		96.205 96.061 96.448	1.0391	0.01661
I	$\log \frac{Io}{I} =$	0.0530	(HBz) -	+ 0.000		

Beer's Law Curve For Benzoic Acid At 8.888 Microns In Chloroform

Concentration moles per liter	Blank	Sample	Blank	%T	Ave. Io	Log ave. Io
0.7366	97.2 97.0 96.9	64.9	96.9 97.1 96.8	66.769 66.872 66.804	1.4966	0.175004
0.58928	96.2 96.5 96.0		96.2	76.207 76.236 72.254	1.3844	0.141268
0.43196	96.2 96.5 96.0	75.1 75.5 75.2		77.944 78.400 78.211	1.2791	1.06904
0.3183	96.0 96.5 96.3	81.I	96.0 96.3 96.8	84.687 84.128 84.308	1.1852	0.073792
Lo	$g \frac{Io}{I} =$	0.2458	(HBz)	+ 0.000		

Beer's Law Curve For Benzoic Anhydride At 8.224 Microns In Chloroform

Concentration moles per liter	Blank	Sample	Blank	<u>%T</u>	Ave. To	Log ave. Io
0.7366	97.5	90.6 90.7 90.6	97.2	93.161 93.168 93.018	1.0737	0.03088
0.58928	97.0	92.0	97.5	94.633 94.601 94.644	1.0567	0.023947
0.3183	96.0	93.3	96.3	97.023 97.035 97.032	1.0306	0.013032
Lo	$g \frac{Io}{I} =$	0.0404	(HBz)	+ 0.000		

Beer's Law Curve For Benzoic Acid At 5.823 Microns In Chloroform

Concentratio moles per li		Sample	Blank	<u>%T</u>	Ave. Io	Log ave Io
0.3183	96.1 96.3 96.7	42.2	96.1 96.5 96.6	44.016 43.798 43.559	2.2837	0 . 358643
0.1912	94.5 95.4 94.8	57.2 57.3 57.3	94.2 95.2 94.9	60.625 60.125 60.411	1.6559	c.219034
0.1592	92.8 93.1 93.3	60.6 60.5 60.5	92.7 93.3 93.3	65.336 64.914 64.844	1.5377	0.186766
0.1274	93.5 93.7 93.5	66.0 66.0 66.0	93.5 93.5 93.4	70.588 70.512 70.626	1.4169	0.15308
0.1061	92.7 92.7 92.7	68.5	92.5 93.1 92.7	73.974 73.735 73.894	1.3537	0.131524
0.0795	92.1 92.2 91.9	73.0	92.2 92.3 92.0	79.110 79.132 79.499	1.2602	0.10034
0.0637	92.0 92.2 92.3	75.8		81.956 82.168 81.709	1.2203	0.08468
	$Log \frac{Io}{I} =$	1.710(HBz) +	0.020		

Beer's Law Curve For Benzoic Acid At 7.570 Microns In Chloroform

Concentration moles per liter	<u>Blank</u>	<u>Sample</u>	Blank	<u> </u>	Ave To	Log ave. To
0.3183	96.1 95.3 95.4	53.5 53.6 53.5	96.2 95.3 95.5	55.642 56.243 56.050	1.7885	0.25249
0.25464	95.3 93.4 93.6	60.1 59.0 59.1	95.2 93.6 93.8	63.097 63.101 63.073	1.5850	0.20003
0.1912	92.8 93.0 92.9	65.5 65.3	93.0 93.2 92.5	70.505 70.354 70.442	1.4197	0.152197
0.15915	92.4 91.6 91.4			74.186 74.426 74.548	1.3443	0.128464
0.1273	94.3 94.7 94.3	74.0 74.1 73.8	94.5 94.3 94.0	78.389 78.412 78.385	1.2755	0.105680
0.0795	94.6 94.8 94.6	81.6 81.7 81.6		86.166 86.136 86.257	1.1602	0.064534
0.0637	95.1 94.5 95.0			88.538 88.783 88.678	1.1278	0.051462
Lo	g <u>Io</u> =	0.7790	(HBz) -	+ 0.000		

Summary Of Data For The Para-Methoxybenzoic-Acetic Anhydride System

Principal Wave length

- A. p-Methoxybenzoic-acetic mixed anhydride -- 8.711 microns
- B. Acetic anhydride ----- 8.888 microns
- C. p-Methoxybenzoic anhydride ----- 8.1770 microns

Beer's Law Curves

- A. p-Methoxybenzoic-acetic mixed anhydride
 - 1. At mixed anhydride wave length (8.711 microns) $Log \frac{Io}{T} = 4.220(M-pMeOBz) + 0.015$
 - 2. At acetic anhydride wave length (8.888 microns) $Log \frac{Io}{T} = 0.9541 (M-pMeOBz) + 0.0042$
 - 3. At p-methoxybenzoic anhydride wave length (8.1770 microns) $Log \frac{Io}{I} = 0.1732 (M-p-MeOBz) + 0.00225$
- B. p-Methoxybenzoic anhydride
 - 1. At mixed anhydride wave length (8.711 microns) $Log \frac{Io}{I} = 0.3945 \text{ (p-MeOBz)} + 0.000$
 - 2. At acetic anhydride wave length (8.888 microns) $\log \frac{Io}{I} = 0.1885 (p-MeOBz) + 0.000$
 - 3. At p-methoxybenzoic anhydride wave length (8.1770 microns) Log $\frac{\text{Io}}{\text{T}}$ = 4.350(p-MeOBz) + 0.0135

C. Acetic anhydride

- 1. At mixed anhydride wave length (8.711 microns) $Log \frac{Io}{T} = 0.857(Ac) + 0.003$
- 2. At acetic anhydride wave length (8.888 microns) $\log \frac{Io}{I} = 3.491(Ac) + 0.024$
- 3. <u>p</u>-Methoxybenzoic anhydride wave length (8.1770 microns) $Log \frac{Io}{I} = 0.5045(Ac) + 0.002$

Simultaneous Equations Derived From The Above Data

At 8.711 microns

$$4.220(M-p-MeOBz) + 0.3945(pMeOBz) + 0.857(Ac) =$$
 $Log \frac{Io}{I} - 0.018$

At 8.888 microns

$$0.954(M-p-MeOBz) + 0.1885(p-MeOBz) + 3.491(Ac) =$$
 $\log \frac{Io}{T} - 0.028$

At 8.1770 microns

$$0.1732(M-p-MeOBz) + 4.35(p-MeOBz) + 0.5045(Ac) =$$
 $Log \frac{Io}{I} - 0.018$

Beer's Law Curve For p-Methoxybenzoic Anhydride At 8.7110 Microns In Chloroform

Concentrate moles per		Blank	Sample	Blank	<u>%T</u>	$\frac{\text{Ave}, \overline{\text{Io}}}{\overline{\text{I}}}$	Log ave. Io
0.0941		97.4 97.6 97.8		97.7 97.8 98.0	91.132 90.992 90.909	1.0987	0.040880
0.07528		97.2 97.3 97.4	90.1	97.3 97.2 97.0	92.647 92.647 92.592	1.0795	0.03220
0.05646		97.1 97.2 97.1	91.9		94.541 94.450 94.487	1.0582	0.024572
0.04705		98.5 98.5 98.5	93.9	98.5 98.5 98.4	95.329 95.329 95.378	1.0488	0.020696
	Lo	$g \frac{Io}{I} =$	0.3945	(p-Meo	Bz) + 0.	000	

Beer's Law Curve For p-Methoxybenzoic Anhydride At 8.888 Microns In Chloroform

Concentration moles per liter	<u>Blank</u>	Sample	Blank	<u>%T</u>	Ave. Io Log ave. Io
0.1414	97.6	91.8 91.8 91.7	97.6	94.057	1.0608 0.025638
0.10605	97.0	92.7	97.2	95.482 95.468 95.459	1.0474 0.020014
0.0707	97.5	94.5 94.6 94.5	97.6	96.972 96.975 96.923	1.0317 0.013554
Lo	g <u>I</u> e =	0.1888	(p-Meo	3z) + 0.	000

Beer's Law Curve For p-Methoxybenzoic Anhydride At 8.177 Microns In Chloroform

Concentration moles per liter	<u>Blank</u>	Sample	Blank	<u>%T</u>	Ave. Io	Log ave. Io
0.0707	97.4 96.9 96.7	46.6 46.3 46.2	97.2 96.7 96.5	47.893 47.830 47.826	2.0913	0,320210
0.0565	94.6 94.6 94.7	52.5 52.5 52.6	94.7 94.9 95.0	55.467 55.408 55.455	1.8043	0.256312
0.0424	95.6 95.6 95.8	60.6 60.6 60.5	95.7 95.7 95.9	63.355 63.355 63.119	1.5803	0.198768
0.03535	95.7 95.4 95.0	65.0 64.8 64.9	95.5 95.5 95.5	67.991 67.888 68.136	1.4704	0.167436
0.0283	95.5 95.0 95.1	69.8 69.7 69.5	95.2 95.2 94.7	73.203 73.291 73.234	1.3653	0.135196
o.C1415	94.8 94.3 94.5	79.6	95.0 94.3 94.7	84.404 84.411 84.460	1.1844	0.073498
0.007075	95.7 95.4 95.8	86.7	95.5 95.7 96.0	90.794 90.737 90.719	1.1019	0.042141

 $Log \frac{Io}{I} = 4.350(p-MeoBz) + 0.0135$

Beer's Law Curve For p-Methoxybenzoic-Acetic Mixed Anhydride at 8.7110 Microns In Chloroform

Concentration moles per liter E	<u>Blank</u>	Sample	Blank	<u>%T</u>	Ave. Io	Log ave. Io
	97.7 98.0 97.5	24.5 24.6 24.5	97.7 97.6 97.9	25.076 25.153 25.076	3.9836	0.600166
0.11140	98.3 98.5 98.3	32.5 32.4 32.3	98.5 98.4 98.2	33.028 32.910 32.875	3.0329	0.481856
0.09320	98.0 97.9 97.9	38.4 38.1 38.3	98.0 97.8 97.8	89.183 38.937 39.141	2.5583	0.407951
0.05592	97.7 97.8 98.1	53.8 54.9 54.9	97.7 97.8 98.3	56.090 56.134 55.906	1.7843	0.251465
0.04464	95.7 96.0 95.8	58.9 59.0 58.9	95.9 95.8 95.6	61.482 61.522 61.546	1.6255	0.210985
0.02796	98.4 97.6 98.3	72.6 72.3 72.6	98.1 98.0 98.2	73.893 73.926 73.893	1.3531	0.131335
0.011160	98.0 98.1 97.9	85.7 85.9 86.0	98.2 97.9 98.2	87.461 87.653 87.710	1.1414	0.057434
	ŤO			,		

 $Log \frac{Io}{I} = 4.220(M-p-MeoBz) + 0.015$

Beer's Law Curve For p-Methoxybenzoic-Acetic Mixed Anhydride At 8.888 Microns In Chloroform

Concentration moles per lite	er Blank	Sample	Blank	%T	$\frac{\text{Ave.}}{\text{I}}$	Log ave. Io
0.12140	97.9 97.5 97.6		97.6 97.6 97.6	75.498 75.448 75.512	1.3265	0.122705
0.09712	98.1 97.5 97.7		97.8 97.7 97.4	79.530 79.610 79.548	1.2600	0.10037
0.07284	97.9 97.8 97.7	82.6 82.6 82.7	97.7 97.9 97.7	84.458 83.414 84.646	1.1861	0.074117
0.04856	98.1 98.1 98.0		98.1 98.2 98.3	88.685 88.741 88.639	1.1275	0.052115
0.03642	98.3 98.0 98.0	89.7 89.4 89.4	98.2 97.9 98.0	91.297 91.271 91.224	1.0957	0.039697
	$Log \frac{Io}{I} =$	0.9541	M-q-M)	eoBz) +	0.0042	

Beer's Law Curve For p-Methoxybenzoic-Acetic Mixed Anhydride At 8.177 Microns In Chloroform

Concentration moles per liter	Blank	Sample	Blank	<u>%T</u>	Ave To	Log ave. Io
0.01398	98.0 98.1 97.8		97.9 98.1 98.0	88.922 88.888 88.968	1.1245	0.050960
0.01118	97.8 97.5 97.4		97.6 97.5 97.2	90.992 90.974 90.955	1.0992	0.041078
0.00839	97.0 97.5 97.0	90.1 90.5 90.1	96.8 97.2 97.3	92.982 92.963 92.743	1.0764	0.031974
0.00699	96.5 96.5 96.4	90.9 90.9 91.0	96.5 96.5 96.3	94.196 94.196 94.447	1.0606	0.025556
Lo	$g \frac{I_0}{I} =$	0.1732	M-p-M)	eoBz) +	0.00225	

Beer's Law Curve For Acetic Anhydride
At 8.7110 Microns In Chloroform

Concentration moles per liter	<u>Blank</u>	Sample	Blank	<u> </u>	Ave.Io	Log ave. Io
0.4242	97.3 97.4 97.2	40.3 40.1 40.2	97.4 97.4 97.1	41.294 41.170 41.379	2.4224	0.384242
0.3610	97.4 97.3 97.3	46.7 46.8 46.6	97.3 97.2 97.0	47.971 48.123 47.967	2.0838	0.318858
c.2760	97.4 97.3 97.3	55.5 55.4	97.3 97.3 97.5	57.010 56.834 56.878	1.7572	0.244820
0.2122	96.8 96.6 96.5	63.3 63.1 63.0	96.8 96.5 96.3	65.392 65.354 65.352	1.5306	0.184864
0.1488	96.7 96.5 96.4	71.5 71.5 71.4	96.5 96.5 96.1	74.016 74.093 74.181	1.3495	0.130138
0.10561	96.4 96.3 96.2	77.6 77.5 77.4	96.4 96.4 96.0	80.597 80.435 80.541	1.2418	0.094050
0.06375	96.5 96.6 96.6		96.6 96.6 96.8	87.519 87.577 87.487	1.1424	0.057822
0.04244	96.4 96.4 96.5	88.0	96.3 96.3 96.6	91.333 91.333 91.040	1.0960	0.03981
Lo	$g = \frac{Io}{T} =$	0.8570	(Ac) +	0.003		

Beer's Law Curve For Acetic Anhydride
At 8.177 Microns In Chloroform

Concentration moles per liter	· <u>Blank</u>	Sample	Blank	<u>%T</u>	Ave. Io	Log ave. Io
0.4244	98.4 98.3 98.5	59.3 59.2 59.3	98.4 98.4 98.4	60.264 60.193 60.233	1.6571	0.219346
0.2548	98.3 98.2 98.0	72.1		73.384 73.421 73.506	1.3617	0.133764
0.2122	98.0 98.0 97.6	75.8 75.8 75.7	98.0 97.6 97.4	77.346 77.505 77.641	1.2903	0.110692
0.1488	98.4 98.4 98.5	82.0 82.0 82.1	98.4 98.4 98.4	83.333 83.333 83.392	1.1997	0.079072
0.1061	97.7 98.3 98.3	85.8 86.2 86.1	97.4 98.2 98.4	87.954 87.735 87.545	1.1396	0.056748
0.06375	98.4 98.5 98.5		98.4 98.5 98.7	92.276 92.385 92.190	1.0771	0.03226

 $Log \frac{Io}{I} = 0.5045(Ac) + 0.002$

Summary Of Data For The Para-Nitrobenzoic-acetic Anhydride System

Principal Wave Length

- A. p-Nitrobenzoic-acetic mixed anhydride --- 7.897 microns
- B. Acetic anhydride ----- 8.888 microns
- C. <u>p</u>-Nitrobenzoic anhydride ----- 6.486 microns

Beer's Law Curves

- A. p-Nitrobenzoic-acetic mixed anhydride
 - 1. At mixed anhydride wave length (7.897 microns) $Log \frac{Io}{I} = 3.341(M-p-Nitro Bz) + 0.000$
 - 2. At acetic anhydride wave length (8.888 microns) $Log \frac{Io}{I} = 1.601 (M-p-Nitro Bz) + 0.0036$
 - 3. At p-Nitrobenzoic anhydride wave length (6.486 microns) $Log \frac{Io}{T} = 5.30(M-p-Nitro Bz) + 0.004$
- B. p-Nitrobenzoic anhydride
 - 1. At mixed anhydride wave length (7.897 microns) $Log \frac{Io}{T} = 0.559 \text{ (p-Nitro Bz)} + 0.0025$
 - 2. At acetic anhydride wave length (8.888 microns) $\text{Log } \frac{\text{Io}}{\text{I}} = 0.181 \text{ (p-Nitro Bz)} + 0.000$
 - 3. At p-nitrobenzoic anhydride wave length (6.486 microns) $\text{Log } \frac{\text{Io}}{\text{I}} = 11.09 (\text{p-Nitro Bz}) + 0.000$

C. Acetic Anhydride

- 1. At mixed anhydride wave length (7.897 microns) $Log \frac{Io}{T} = 0.2535(Ac) + 0.000$
- 2. At acetic anhydride wave length (8.888 microns) $Log \frac{Io}{I} = 3.491(Ac) + 0.024$
- 3. At p-nitrobenzoic anhydride wave length (6.486 microns) $Log \frac{Io}{T} = 0.587(Ac) + 0.002$

Simultaneous Equations Derived From The Above Data

At 7.897 microns

3.341(M-p-Nitro Bz) + 0.559(p-Nitro Bz) + 0.2535(Ac) =
$$log \frac{Io}{I} - 0.0025$$

At 8.888 microns

1.601(M-p-Nitro Bz) + 0.181(p-Nitro Bz) + 3.491(Ac) =
$$Log \frac{Io}{T}$$
 - 0.028

At 6.486 microns

$$5.30(M-p-Nitro Bz) + 11.09(p-Nitro Bz) + 0.587(Ac) =$$
 $Log \frac{Io}{T} - 0.006$

Beer's Law Curve For <u>p</u>-Nitrobenzoic Anhydride At 7.897 Microns In Chloroform

Concentration moles per liter	Blank	Sample	Blank	<u>%T</u>	Ave. Io	Log ave. Io
0.0381	98.9 98.7 98.6	93.7 93.6 93.7	98.5 98.6 98.8	94.934 94.880 94.934	1.0535	0.022635
0.03422	98.1 98.4 97.8		97.8 98.3 97.6	95.354 95.068 95.496	1.0492	0.020846
0.02285	98.9 98.7 98.8		98.9 98.5 98.9	96.663 96.855 96.713	1.0336	0.014352
0.01123	98.9 98.8 98.9	97.0 97.0 97.0	98.9 98.9 98.9	98.078 98.128 98.078	1.0194	0.008346
Lo	$\frac{Io}{I} =$	0.5590	(p-Nit	ro Bz) +	0.0025	

Beer's Law Curve For p-Nitrobenzoic Anhydride At 8.888 Microns In Chloroform

Concentration moles per liter	Blank	Sample	Blank	<u>%T</u>	$\frac{\text{Ave.}}{I}$	Log ave. Io
0.0960	98.6 98.9 98.9	94.3 94.4 94.3	98.8 98.7 98.8	95.542 95.546 95.493	1.0468	0.019866
0.0480	98.6 98.5 98.4	96.5 96.4 96.3	98.7 98.6 98.4	97.820 97.818 97.865	1.0221	0.009493
0.0384	98.4 98.6 98.4	96.7	98.3	98.172 98.222 98.322	1.0168	0.007234

 $Log \frac{Io}{T} = 0.1810(p-Nitro Bz) + 0.000$

Beer's Law Curve For <u>p</u>-Nitrobenzoic Anhydride At 6.486 Microns In Chloroform

Concentration moles per liter	Blank	Sample	Blank	<u>%T</u>	$\frac{\text{Ave.}}{I}$	Log ave. I
0.03060	98.2 98.5 98.1	45.1 45.2 45.1	98.1 98.4 98.3	45.950 45.911 45.926	2.1915	0.340740
0.02295	98.5 98.9 98.3		98.7 98.6 98.1	55.679 55.724 55.702	1.7952	0.254110
0.01530	97.7 97.7 97.5	66.2 66.2 66.1	97.6 97.5 97.8	67.793 67.827 67.690	1.4755	0.168940
0.00922	97.5 97.4 97.3			79.055 79.096 79.155	1.2641	0.101784
0.00765	97.5 97.7 97.8	80.4 80.4 80.5	97.5 97.6 97.9	82.461 82.334 82.268	1.2142	c.084288
0.00614	97.2 97.1 97.3		97.1 97.2 97.5	85.743 85.743 85.626	1.1668	0.066996
0.00307	97.1 97.2 97.3	89.9		92.584 92.537 92.449	1.0808	0.033748
T	Io	77 00/-	NT 2 4		0.0000	

 $Log \frac{Io}{I} = 11.09(p-Nitro Bz) + 0.0000$

Beer's Law Curve For <u>p</u>-Nitrobenzoic-acetic Mixed Anhydride At 7.897 Microns In Chloroform

Concentration moles per liter	<u>Blank</u>	Sample	Blank	<u>%T</u>	Ave Io	Log ave. Io
.0.0783	90.8 91.0 91.2		91.1 91.2 90.9	54.755 54.665 54.426	1.8309	0.262466
0.06264	91.2 91.3 91.3		91.4 91.0 91.5	61.664 61.656 61.597	1.6223	0.212531
0.0426	89.9 90.1 90.1		90.0 89.9 90.0	72.595 72.444 72.044	1.3819	0.140168
0.3915	92.3 92.1 92.5	68.7	92.1 92.5 92.6	74.511 74.431 74.446	1.3429	c.128047
0.02555	90.4 90.2 90.4		90.2 90.4 90.3	82.059 82.392 82.014	1.2172	0.085362
0.01704	90.4 90.3 90.1	78.8	90.3 90.1 89.9	87.216 87.361 87.444	1.1449	0.058763
0.00852	98.0 98.1 97.9		97.9 97.9 98.3	93.415 93.319 93.374	1.0710	0.02979
Lo	$g \frac{Io}{I} =$	3.341(rtN-q-N	tro Bz)	+ 0.000	

Beer's Law Curve For <u>p</u>-Nitrobenzoic-acetic Mixed Anhydride At 8.888 Microns In Chloroform

Concentration moles per lite	er Blank	Sample	Blank	%T	Ave <u>Io</u>	Log ave. To
0.0475	97.8 97.6 97.6		97.7 97.5 97.3	83.069 83.136 83.222	1.2027	0.080159
0.0380	97.4 97.0 97.1	83.8	97.0 97.0 97.0	86.213 86.391 86.347	1.1585	0.063895
0.0285		86.8 86.5 86.6	97.1 97.0 97.0	89.392 89.083 89.232	1.1206	0.049454
0.0190	97.1 97.1 97.0		97.1 97.0 97.2	92.481 92.426 92.481	1.0815	0.034030
0.0095	97.1 97.1 97.0			95.674 95.826 95.777	1.0442	0.018784
I	$\log \frac{Io}{I} =$	1.601(1	M-p-Ni	tro Bz)	+ 0.0036	

Beer's Law Curve For <u>p</u>-Nitrobenzoic-acetic Anhydride At 6.486 Microns In Chloroform

Concentrat moles per		<u>Blank</u>	Sample	Blank	<u>%T</u>	Ave. Io	Log ave. To
0.0480	·	97.1 97.3 97.5	52.6 52.7 52.8	97.1 97.3 97.5	54.170 54.162 54.153	1.8475	c.266585
0.0384		96.5 96.4 96.7		96.6 96.8 96.8	60.693 60.662 60.671	1.6481	0.216986
0.0288		98.2 98.2 98.0		98.2 98.1 97.9	68.635 68.670 68.708	1.4562	0.16220
0.0192		98.2 98.1 98.0	75.9 75.9 75.8	98.1 98.1 98.2	77.330 77.370 77.268	1.2932	0.111666
0.0096		98.1 98.0 98.2		98.0 98.0 98.1	87.608 87.653 87.519	1.1416	0.057134
	Log	s <u>Io</u> =	5.300(1	N-p-Ni	tro Bz)	+ 0.004	

Beer's Law Curve For Acetic Anhydride
At 7.897 Microns In Chloroform

Concentrate moles per		Blank	Sample	Blank	<u>%T</u>	Ave. Io	Log ave. Io
0.5244		98.8 98.6 98.7			73.924 73.897 73.822	1.3535	0.13146
0.3486		98.2 98.8 98.1		98.5 98.8 98.2	81.443 81.477 81.609	1.2268	0.088500
0.2622		98.8 98.8 98.6	84.9	98.9 98.5 98.7	85.887 86.061 85.960	1.1632	c.065654
0.1748		98.5 98.4 98.7	88.6	98.5 98.7 98.6	90.050 89.903 89.913	1.1116	0.045956
0.1049		98.8 98.7 98.7		98.8 98.7 98.8	93.927 93.920 93.974	1.0645	0.027145
	Log	$rac{Io}{I} =$	0.2535	(Ac) +	0.0000		

Beer's Law Curve For Acetic Anhydride At 6.486 Microns In Chloroform

Concentration moles per liter	Blank	Sample	Blank	%T	Ave. Io	Log ave. Io
0.5244				92.762 92.817 92.762	1.0778	0.032540
0.4370	98.1 98.1 98.1	92.0 92.0 92.1	98.1 98.0 98.3	93.781 93.829 93.788	1.0661	0.027800
0.3486	97.9 97.9 97.9	93.0	97.9 97.9 97.9	94.994 94.994 94.994	1.0526	0.022225
0.31464	97.8 97.8 97.6	93.3		95.549 95.496 95.545	1.0467	0.019824
Log	$g \frac{Io}{I} =$	0.5870	(Ac) +	0.002		

Equilibrium Mixture Derived From Acetic Anhydride And Benzoic Acid In Chloroform At 150°C

Wave length (microns)		Sample	Blank	%T	Average <u>Io</u>	Log ave. Io		
8.690	96.8 97.0 97.3	45.4 45.5 45.6	96.9 97.2 97.2	46.876 46.858 46.889	*	0.32905		
8.888	96.0 96.2 96.5	44.8 44.9 45.0		46.642 46.673 46.656	2.1433	0.33108		
8.224	96.5 96.7 96.9	58.5 58.6 58.7	96.7 96.9 97.1	60.559 60.537 60.515	1.6518	0.217958		
5.823	96.0 96.3 96.2	63.0 63.1 63.1	96.2 96.2 96.4	65.556 65.558 65.524	1 . 5256	0.183438		
7.570	95.9 96.0 95.9		95.9	87.870 87.858 87.845	1.1386	0.056368		
Concentratio	on benz	oic-ace	etic mi	ixed ani	nydride C	.1213350		
Concentration	on bėnz	oic anh	nydride		·	.0433680		
Concentratio	on acet	ic anhy	dride		C	.0686636		
Concentratio	on benz	oic aci	Id		C	.0121030		
Concentratio	on acet	ic acid	l		C	.0437800		
Equilibrium Constant (anhydride) 4.94								
	Equili	brium C	onstar	it (acid	l-I) 1	.29		
	Equili	brium C	onstar	it (acid	I-II) 6	•33		

Equilibrium Mixture Derived From Acetic Anhydride, Benzoic Anhydride And Acetic Acid In Chloroform At 75°C

Wave length (microns)	Blank	Sample	Blank	<u>%T</u>	Average Io	Log ave. Io			
8.690	97.6 97.5 97.8		97.8 97.6 97.7	68.5772 68.5802 68.5423	2 1.45843	0.163881			
8.888	97.8 97.8 97.6	63.8 63.7 63.6	97.9 97.6 97.5	65.2018 65.1995 65.1973	1.53375	0.185755			
8.224		49.7		50.9501 50.9482 50.9462	2 1.96277	0.292874			
5.823	97.8 98.1 98.0	57.1 57.2 57.1	98.0 98.2 97.9	58.3248 58.2781 58.2950	1.71528	0.234338			
7.570	97.9 97.8 97.7	84.7 84.6 84.6	97.9 97.7 97.8	86.5168 86.5473 86.5473	3 1.15557	0.062601			
Concentration	on benz	oic-ace	etic mi	Lxed anh	nydride O	.056873			
Concentration	on benz	oic an	nydride	e	O	.0721210			
Concentratio	on acet	ic anhy	/dride			.038472			
Concentratio	on benz	oic act	id		0	.0418791			
Concentration	Concentration acetic acid 0.0520808								
	Equilibrium Constant (anhydride) 1.25								
	Equili	.brium (Constar	nt (acid	d-I) 1	.58			
	Equili	.brium (Constar	nt (acid	d-II) 1	•97			

Equilibrium Mixture Derived From Benzoic-acetic Mixed Anhydride And Benzoic Acid In Chloroform At 75°C

Wave length (microns)	Blank	Sample	Blank	<u> </u>	Average Io	Log ave. Io		
8.690	95.7 95.8 95.9	56.5 56.6 56.6	95.8 95.9 96.0	59.0078 59.0509 58.9890	5 1.69436	0.229013		
8.888	96.1 96.5 96.4	46.4 46.5 46.5	96.3 96.5 96.4	48.207 48.186 48.236	2.07711	0.317462		
8.224	96.7 96.5 96.6	50.1 50.1 50.1		51.8633 51.8903 51.9170	2 1.92962	0.285470		
5.823	96.3 96.1 96.2	48.7 48.6 48.7		50.5474 50.5986 50.5713	5 1.97671	0.295834		
7.570	96.4 96.5 96.6	83.1 83.2 83.2	96.5 96.6 96.7	86.1586 86.1729 86.0838	9 1.16092	0.064794		
Concentration	on benz	oic-ac∈	etic mi	ixed anh	nydride O	.0752055		
Concentratio								
Concentratio	on acet	ic anhy	rdride		O	.0708808		
Concentration benzoic acid 0.0250481								
Concentration acetic acid 0.0857286								
Equilibrium Constant (anhydride) - 1.08								

Equilibrium Constant (anhydride) - 1.08

Equilibrium Constant (acid-I) ---- 3.36

Equilibrium Constant (acid-II) --- 3.61

Equilibrium Mixture Derived From Benzoic-acetic Mixed Anhydride And Benzoic Acid At 100°C In Chloroform

Wave length (microns)		Sample	Blank	<u>%T</u>	Average Io	Log ave Io
8.690	97.3 97.5 97.2	56.7 56.8 56.7	97.2 97.3 97.4	58.303 58.316 58.273	1.7153	0.234338
8.888	96.8 96.7 96.4	53.4 53.2 53.1		55.136 55.072 55.082	1.8149	0.258859
8.224	96.3 96.1 96.1	51.4 51.4 51.4	96.1	53.402 53.485 53.458	1.8709	0.272047
5.823	96.5 96.4 96.5	44.7 44.7 44.8	96.4 96.4 96.5	46.345 46.369 46.424	2.1561	0.333670
7.570	95.8 96.0 95.9	80.4 80.5 80.4	96.0 95.9 95.8	83.837 83.897 83.881	1.1922	0.076352
Concentration	on benz	oic-ace	etic mi	ixed anl	nydride	0.0829080
Concentration	on benz	oic anh	nydride	·		0.0663150
Concentration	on acet	ic anhy	/dride			0.0517563
Concentration		0.0408189				
Concentration	0.0946918					
	2 00					

Equilibrium Constant (anhydride) -- 2.00

Equilibrium Constant (acid-I) ---- 1.86

Equilibrium Constant (acid-II) ---- 2.04

Equilibrium Mixture Derived From Benzoic Anhydride And Acetic Acid At 100°C In Chloroform

Wave length (microns)	Blank	Sample	Blank	<u>%T</u>	Average Io	Log ave. Io		
8.690	96.7 96.8	45.2 45.2	96.8 96.9	46.718 46.670	2.1409	0.330590		
	96.9	45.2	96.7	46.743		0.000000		
8.888	95.9 95.9 95.8	47.2 47.1 47.1		49.192 49.139 49.164	2.0339	0.308329		
8.224	95.4 95.6 95.6	42.2 42.2 42.2		44.188 44.165 44.119	2.2651	0.355079		
5.823	95.7 95.8 96.0	43.5 43.6 43.7	95.7 96.1 96.0	45.454 45.393 45.520	2.2001	0.342439		
7.570	96.1 96.3 96.5	79.3 79.6 79.6		82.475 82.572 82.487	1.2119	0.083464		
Concentration	on henz	oic-ace	atic mi	ived anl	nydride (1.22560		
					(
Concentration			J					
Concentration	on acet	ic anhy	rdride		(0.058464		
Concentration benzoic acid 0.029669								
Concentration acetic acid 0.098309								
Equilibrium Constant (anhydride) 3.12								

Equilibrium Constant (acid-I) ---- 2.23

Equilibrium Constant (acid-II) ---- 6.95

Equilibrium Mixture Derived From Benzoic-acetic Mixed Anhydride And Acetic Acid At 1500C In Chloroform

Wave length (microns)	Blank	Sample	Blank	<u>%T</u>	Average Io	Log ave. Io		
8.690	97.8 97.7 97.4	65.4 65.2 65.1	97.4	66.871 66.837 66.872	1.4956	0.174814		
8.888	97.5 97.3 97.5	63.3 63.2 63.3	97.4 97.5 97.5	64.959 64.887 64.923	1.5403	0.187604		
8.224	97.4 97.1 97.3	71.2 71.1 71.1		73.138 73.148 73.185	1.3675	0.13583		
5.823	97.0 96.9 96.7	56.3 56.2 56.1		58.071 58.057 57.984	1.7230	0.23629		
7.570	97.3 97.2 97.3			89.357 89.403 89.311	1.1191	0.048860		
Concentratio	n benz	oic-ace	tic mi	ixed anh	nydride	0.06296500		
Concentratio	n benz	oic anh	ydr i de	·	·	0.02781174		
Concentratio	n acet	ic anhy	dride			0.03813330		
Concentratio	n benz	oic aci	.d			0.02406000		
Concentration acetic acid 0.07184750								
Equilibrium Constant (anhydride) 3.74								
Equilibrium Constant (acid I) 1.27								

Equilibrium Constant (acid II) ---- 4.75

Equilibrium Mixture Derived From Benzoic And Acetic Anhydrides At 70° In Chloroform

Wave Length	<u>Blank</u>	Sample	Blank	<u>%T</u>	Average %T	Log ave. Io
8.224	96.8 96.7 96.7	73.9 73.9 73.9	96.7 96.8 96.7	76.382 76.382 76.421	76.395	0.116907
8.888	97.7 97.5 97.6	59.7 59.6 59.6	97.8 97.7 97.5	61.074 61.056 61.096	61.078	0.214102
8.690	96.5 96.4 96.3	57.5 57.4 57.4	96.4 96.3 96.3	59.512 59.574 59.605	59.564	0.224998
Concentration Concentration	on of i	benzoic	anhydi	ride	anhydride -	

Equilibrium Constant = 1.04

Equilibrium Mixture Derived From Benzoic-Acetic Mixed Anhydride At 70° C In Chloroform

Wave Length	Blank	Sample	Blank	<u>%T</u>	Average %T	Log ave Io	
8.224	95.4 95.6 95.4	66.6 66.8 66.5	95.5 95.7 95.4	69.774 69.837 69.706	69.772	0.156340	
8.888	94.8 94.7 94.5	58.2 58.1 58.0	95.0 94.6 94.5	61.327 61.384 61.375	61.362	0.212182	
8.690	95.8 96.1 96.0	52.7 52.8 52.8	96.0 96.1 96.1	54.953 54.942 54.971	54.955	0.259974	
Concentration of benzoic-acetic mixed anhydride 0.0972660 Concentration of benzoic anhydride 0.0281644 Concentration of acetic anhydride 0.0313660							

Equilibrium Mixture Derived From Benzoic And Acetic Anhydrides At 100°C In Chloroform

Wave Length	Blank	Sample	Blank	ZT	Average %T	Log ave. Io
8.224	94.8 95.0 95.2	68.1 68.3 68.3	94.9 95.2 95.1	71.797 71.819 71.781	71.799	0.143733
8.888	95.4 95.5 95.6	67.0 67.0 67.1	95.5 95.5 95.5	70.193 70.157 70.225	70.192	0.153660
8.690	96.0 96.0 96.0	55.9 55.9 55.9	96.1 95.9 96.0	58.198 58.259 58.229	58.229	0.234845
Concentration Concentration Concentration	on of	benzoic	anhyd:	ride	<i>D</i>	0.0911480 0.0265109 0.0157940

Equilibrium Constant = 1.98

Equilibrium Mixture Derived From Benzoic-Acetic Mixed Anhydride At 100°C In Chloroform

Wave Length	<u>Blank</u>	Sample	Blank	<u>%T</u>	Average %T	Log ave Io
8.224	96.0 96.3 96.3	38.3 38.4 38.3	96.2 96.4 96.3	39.854 39.853 39.771	39,826	0.399836
8.888	97.8 97.6 97.8	35.8 35.8 35.8		36.605 36.661 36.624	36.630	0.4361600
8.690	96.5 96.5 96.6	43.0 43.1 43.1	96.4 96.6 96.7	44.582 44.640 44.593	44.605	0.350620
Concentration	on of 1	penzoic	anhydi	ride	anhydride -	0.088116

Equilibrium Mixture Derived From Benzoic And Acetic Anhydrides At 150°C In Chloroform

Wave Length	Blank	Sample	Blank	<u>%T</u>	Average %T	$\underline{\text{Log ave}}.\underline{\underline{\text{Io}}}$
8.224	95.6 95.4 95.5	63.4 63.3 63.3	95.7 95.6 95.4	66.283 66.282 66.317	66.294	0.178516
8.888	94.8 94.7 95.0	52.5 52.5 52.6	94.7 94.9 94.9	55.408 55.379 55.397	55.391	0.256552
8.690	96.7 96.9 96.8	56.4 56.4 56.4	96.8 96.9 96.9	58.191 58.204 58.234	58.210	0.234995
Concentration Concentration	on of	benzoic	anhydi	cide	anhydride -	0.0826050 0.0339594 0.0466070

Equilibrium Constant = 4.31

Equilibrium Mixture Derived From Benzoic-Acetic Mixed Anhydride At 150°C In Chloroform

Wave Length	Blank	Sample	Blank	%T	Average %T	Log ave. Io
8.224	96.4 96.3 96.3	57.5 57.5 57.5	96.3 96.3 96.2	59.678 59.709 59.740	59.709	0.223935
8.888	95.7 96.1 96.3	51.6 51.9 52.0	95.8 96.2 96.4	53.890 53.978 53.969	53.946	0.268041
8.690	95.4 95.5 95.6	52.0 52.1 52.1	95.4 95.5 95.5	54.507 54.554 54.526	54.529	0.263352
Concentration Concentration	on of l	benzoic	anhydi	cide	anhydride -	0.045713

Equilibrium Mixture Derived From p-Methoxybenzoic And Acetic Anhydrides At 75°C In Chloroform

Wave Length	Blank	Sample	Blank	<u>%T</u>	Average %T	Log ave. Io
8.711	97.0 97.2 97.1	62.6 62.7 62.7	97.1	64.502 64.539 64.572	64.537	0.190162
8.888	98.1 98.0 97.9			72.134 72.179 72.151	72.155	0.141729
8.177	97.5 97.7 97.7	72.0 72.1 72.0	97.8	73.808 73.759 73.694	73.720	0.132068
Concentratio	on of p	ethox	ybenzo	ic-acet	cic mixed and	nyd - 0.0342403

Concentration of <u>p</u>-methoxybenzoic-acetic mixed anhyd - 0.0342403 Concentration of <u>p</u>-methoxybenzoic anhydride ----- 0.0223059 Concentration of acetic anhydride ----- 0.0220160

Equilibrium Constant = 2.26

Equilibrium Mixture Derived From p-Methoxybenzoic Acetic Mixed Anhydride At 75°C In Chloroform

Wave Length	Blank	$\underline{\mathtt{Sample}}$	Blank	<u>%T</u>	Average %T	Log ave. Io
8.711	95.8 95.9 95.7	46.8 46.7 46.7	95.9 95.7 95.7	48.826 48.747 48.798	48.790	0.311666
8.888	94.9 95.2 95.0	57.0 57.1 57.0	95.0 95.1 95.1	60.031 60.010 59.968	60.003	0.221805
8.177	96.0 96.2 96.2	58.2 58.3 58.3	96.2 96.3 96.4	60.561 60.571 60.539	60.554	0.217854

Concentration of <u>p</u>-methoxybenzoic-acetic mixed anhyd - 0.058300 Concentration of <u>p</u>-methoxybenzoic anhydride ----- 0.039276 Concentration of acetic anhydride ----- 0.037472

Equilibrium Mixture Derived From <u>o-Methoxybenzoic</u> And Acetic Anhydrides At 75°C in Chloroform

Ī	lave Length	Blank	Sample	Blank	<u>%T</u>	Average %T	Log ave. Io		
8	3.711	97.2 97.5 97.6	33.5 33.6 33.6	97.3 97.5 97.5	34.465 34.461 34.443	34.456	0.462730		
3	3.888	97.9 97.8 97.7	35.0 35.0 35.0	97.8	35.769 35.787 35.805	35.787	0.446275		
8	3.177	96.0 96.1 96.2	57.8 57.8 57.9	96.1 96.1 96.1	60.176 60.145 60.218	60.179	0.220452		
	Concentration of p-methoxybenzoic-acetic mixed anhyd - 0.083015 Concentration of p-methoxybenzoic anhydride 0.032177								

Concentration of \overline{p} -methoxybenzoic anhydride ----- 0.032177 Concentration of acetic anhydride ----- 0.095341

Equilibrium Constant = 2.25

Equilibrium Mixture Derived From p-Methoxybenzoic And Acetic Anhydrides At 100°C In Chloroform

Wave Length	Blank	Sample	Blank	%T	Average %T	Log ave. Io	
8.711	98.0 98.2 98.1	57.4 57.4	98.1 98.1 98.1	58.439 58.481 58.511	58.477	0.233000	
8.888	97.5 97.4 97.9	66.8 66.7 67.0	97.6 97.5 97.8	68.477 68.445 68.472	68.465	0.164530	
8.177	96.0 96.3 96.4	73.7 73.9 73.9	96.2 96.4 96.4	76.690 76.699 76.659	76,683	0.115280	
Concentration of p-methoxybenzoic-acetic mixed anhyd - 0.0440581 Concentration of p-methoxybenzoic anhydride 0.0175770 Concentration of acetic anhydride 0.0261400							

Equilibrium Mixture Derived From p-Methoxybenzoic And Acetic Anhydrides At 100°C In Chloroform

Wave Length	<u>Blank</u>	Sample	Blank	<u>%T</u>	Average %T	Log ave Io		
8.711	94.8 95.0 95.2	60.7	94.9 95.1 95.2	63.890 63.861 63.865	63.872	0.194678		
8.888	95.8 96.0 96.1	65.2	96.0	67.918 67.916 67.985	67.940	0.167850		
8.177	97.0 97.0 97.1		97.1	83.711 83.771 83.771	83.751	0.076892		
Concentration of p-methoxybenzoic-acetic mixed anhyd - 0.0349620 Concentration of p-methoxybenzoic anhydride 0.0086781 Concentration of acetic anhydride 0.0300000								

Equilibrium Constant = 4.69

Equilibrium Mixture Derived From p-Methoxybenzoic-Acetic Mixed Anhydride At 100°C In Chloroform

Wave Length	Blank	Sample	Blank	<u>%</u> T	Average %T	Log ave. Io		
8.711	96.4 95.8 95.9	56.3 56.0 56.1	95.8	58.432 58.455 58.467	58.451	0.233200		
8.888	94.8 94.9 94.9	66.9 67.0 67.0		70.569 70.600 70.600	70.590	0.151246		
8.177	95.3 94.9 95.0		95.0	74.540 74.565 74.592	74.566	0.127430		
Concentration of <u>p</u> -methoxybenzoic-acetic mixed anhyd - 0.044280 Concentration of <u>p</u> -methoxybenzoic anhydride 0.020830 Concentration of acetic anhydride 0.022085								

Equilibrium Mixture Derived From p-Methoxybenzoic And Acetic Anhydrides At 150°C In Chloroform

Wave Length	<u>Blank</u>	<u>Sample</u>	Blank	<u>%T</u>	Average %T	Log ave. Io		
8.711	98.0 98.4 97.9	72.7 72.9 72.6	98.3	74.108 74.123 74.157	74.329	0.128819		
8.888	96.3 96.4 96.5	74.4 74.6 74.6	96.5	77.298 77.345 77.305	77.319	0.111699		
8.177	95.8 95.9 96.1	69.1 69.2 69.4	95.9	72.280 72.158 72.178	72.205	0.141574		
Concentration of p-methoxybenzoic-acetic mixed anhyd - 0.022783 Concentration of p-methoxybenzoic anhydride 0.025524								

Concentration of \overline{p} -methoxybenzoic anhydride ----- 0.025524 Concentration of acetic anhydride ----- 0.017042

Equilibrium Constant = 11.93

Equilibrium Mixture Derived From p-Methoxybenzoic-Acetic Mixed Anhydride At 150°C In Chloroform

Wave Length	Blank	Sample	Blank	<u>%T</u>	Average %T	Log ave. Io		
8.711	97.5 97.3 97.4	61.5 61.3 61.4	97.2	63.044 63.033 63.071	63.043	0.200356		
8.888	98.0 97.5 97.7	75.5 75.1 75.3	97.5	77.080 77.025 77.112	77.072	0.113032		
8.177	96.4 96.7 96.4	78.6 78.8 78.6	96.6	81.535 81.531 81.577	81.548	0.088560		
Concentration of p-methoxybenzoic-acetic mixed anhyd - 0.039340 Concentration of p-methoxybenzoic anhydride 0.013158 Concentration of acetic anhydride 0.012894								

Equilibrium Mixture Derived From <u>p</u>-Nitrobenzoic And Acetic Anhydrides At 75°C In Chloroform

Wave Length	Blank	Sample	Blank	<u>%T</u>	Average %T	Log ave. To		
7.897	96.8 96.7 96.7	65.5 65.4 65.4	96.9 96.7 96.8	67.630 67.422 67.596	67.549	0.170380		
8.888	98.4 98.5 98.7	18.8 18.8 18.8	98.4 98.6 98.7	19.105 19.076 19.047	19.076	0.720168		
6.486	97.8 96.9 96.8	42.2 41.8 41.8	97.9 96.8 96.8	43.127 43.159 43.181	43.156	0.364948		
Concentration of <u>p</u> -nitrobenzoic-acetic mixed anhyd - 0.0354960 Concentration of <u>p</u> -nitrobenzoic anhydride 0.0057885 Concentration of acetic anhydride 0.1816900								

Equilibrium Constant = 1.20

Equilibrium Mixture Derived From <u>p</u>-Nitrobenzoic-Acetic Mixed Anhydride At 75°C In Chloroform

Wave Length	Blank	Sample	Blank	<u>%T</u>	Average %T	$\frac{\text{Log ave}}{\text{I}}$		
7.897	95.2 95.1 95.1	76.3 76.2 76.2	95.3 95.0 95.2		80.119	0.096244		
8.888	97.4 97.2 97.5	70.8 70.7 70.9	97.6 97.3 97.6	72.615 72.699 72.680	72.665	0.138651		
6.486	98.1 97.5 98.0	41.6 41.4 41.6	97.9 97.7 98.0	42.448 42.418 42.448	42.438	0.372026		
Concentration of <u>p</u> -nitrobenzoic-acetic mixed anhyd - 0.0230540 Concentration of <u>p</u> -nitrobenzoic anhydride 0.0209410 Concentration of acetic anhydride 0.0200378								

Equilibrium Mixture Derived From <u>p</u>-Nitrobenzoic And Acetic Anhydrides At 100°C In Chloroform

Wave Length	Blank	<u>Sample</u>	Blank	<u>%T</u>	Average %T	Log ave. To
7.897	97.1 96.8 96.9	85.3 85.0 85.2	97.2 96.8 97.1	87.802 87.809 87.835	87.815	0.0564060
8.888	95.8 96.0 96.1	78.8 79.0 79.1	95.9 96.2 96.3	82.211 82.206 82.224	82.214	0.0845058
6.486	97.8 97.8 97.8	58.0 58.1 58.0	97.8 98.0 97.6	59.304 59.346 59.365	59.338	0.226652

Concentration of p-nitrobenzoic-acetic mixed anhyd - 0.0129194 Concentration of p-nitrobenzoic anhydride ----- 0.0130697 Concentration of acetic anhydride ----- 0.0098700

Equilibrium Constant = 1.29

Equilibrium Mixture Derived From p-Nitrobenzoic-Acetic Mixed Anhydride At 100°C In Chloroform

Wave Length	<u>Blank</u>	Sample	Blank	<u>%T</u>	Average %T	Log ave. Io		
7.897	95.6 95.7 95.5	64.9 65.0 64.9	95.7 95.7 95.6	67.851 67.920 67.922	67.898	0.168113		
8.888	95.6 95.4 95.4		95.6 95.6 95.5	60.146 60.421 60.136	60.234	0.220136		
6.486	94.8 95.0 95.1	21.3 21.3 21.3	94.9 95.2 95.1	22.456 22.397 22.397	22.417	0.649470		
Concentration of <u>p</u> -nitrobenzoic-acetic mixed anhyd - 0.040816 Concentration of <u>p</u> -nitrobenzoic anhydride 0.036704 Concentration of acetic anhydride 0.034427								

Equilibrium Mixture Derived From p-Nitrobenzoic

And Acetic Anhydrides At 100°C In Chloroform

Wave Length	Blank	Sample	Blank	<u>%T</u>	Average %T	Log ave. Io		
7.897	97.6 97.4 97.7	79.8	97.5 97.4 97.6	81.906 81.930 81.925	81.920	0.086612		
8.888	95.6 95.8 95.5		95.7	72.879 72.898 72.831	72.869	0.137446		
6.486	94.8 94.3 96.0	47.6 47.4 48.2	94.7 94.5 95.9	50.237 50.211 50.234	50.227	0.299048		
Concentration of p-nitrobenzoic-acetic mixed annual - 0.0210147								

Concentration of <u>p</u>-nitrobenzoic-acetic mixed anhyd - 0.0210147 Concentration of <u>p</u>-nitrobenzoic anhydride ----- 0.0152745 Concentration of acetic anhydride ----- 0.0209212

Equilibrium Mixture Derived From p-Nitrobenzoic And Acetic Anhydrides At 150°C In Chloroform

Wave Length	Blank	Sample	Blank	<u>%T</u>	Average %T	Log ave. Io
7.897	96.8 96.8 96.8	71.6 71.6 71.6	96.8 96.7 96.8	73.966 74.005 73.966	73.979	0.130884
8.888	95.7 95.6 95.7			64.016 64.087 64.020	64.041	0.193512
6.486	95.2 95.3 95.4	36.0 36.0 35.7	95.3 95.3 95.5	37.795 37.775 37.777	37.749	0.423080

Concentration of <u>p</u>-nitrobenzoic-acetic mixed anhyd - 0.033568 Concentration of <u>p</u>-nitrobenzoic anhydride ----- 0.020507 Concentration of acetic anhydride ----- 0.030951

Equilibrium Constant = 1.77

Equilibrium Mixture Derived From p-Nitrobenzoic-Acetic Mixed Anhydride At 150°C In Chloroform

Wave Length	Blank	Sample	Blank	<u>%T</u>	Average %T	Log ave. Io			
7.897	95.4 95.7 95.8	80.3	95.3 95.8 95.7	83.796 83.864 83.864	83.841	0.078748			
8.888	96.4 96.5 96.3	74.4	96.4 96.4 96.2	77.178 77.138 77.090	77.135	0.112742			
6.486	97.3 97.7 97.8	51.3	97.5 97.6 97.7	52.464 52.534 52.348	52.449	0.281076			
Concentration of <u>p</u> -nitrobenzoic-acetic mixed anhyd - 0.019224 Concentration of <u>p</u> -nitrobenzoic anhydride 0.014840									

Concentration of acetic anhydride ----- 0.014690

 $\frac{\text{Rate Of Disproportionation Of }\underline{p}\text{-Methoxybenzoic-acetic Mixed}}{\frac{\text{Anhydride At }100^{\circ}\text{C Reaction Followed By Measurement Of}}{\underline{p}\text{-Methoxybenzoic Anhydride}}$

Time (min.)	Blank	Sample	Blank	<u>%T</u>	Ave Io	Log ave. Io	Concentration ⁻¹ (liters.mole ⁻¹)
. 0	98.2 98.0 97.9	91.8 91.6 91.6	98.3 98.1 98.2	93.435 93.421 93.421	1.0703	0.029503	8.8261
5	98.3 98.6 98.5	89.5 89.4 89.4	98.6 98.5 98.6	90.909 90.715 90.715	1.1015	0.041995	9.0334
10	98.5 98.3 98.1	85.3 85.1 85.1	98.4 98.0 98.2	86.642 86.704 86.704	1.1536	0.06206	9.4073
15	98.7 98.0 97.7	81.8 81.2 81.1		82.961 82.899 82.924	1.2058	0.08127	9.7656
20	97.5 97.6 97.7	77.4 77.6 77.5	97.4 97.7 97.3	79.425 79.467 79.527	1.2582	2 0.09785	10.1210
25	97.4 97.2 97.3	76.3 76.3 76.3	97.3 97.4 97.3	78.376 78.417 78.417	7 1.2754	0.105646	10.5150
30		72.2 71.7 71.8	97.9 97.1 97.3	73.684 73.803 73.868	1.3552	2 0.132004	10.928
35	96.9 97.0 96.8	69.3 69.4 69.3	96.7 97.1 96.9	71.590 71.509 71.553	1.3976	c.145386	11.286
40	96.8 96.8 96.7	65.7 65.7 65.8		67.942 67.942 68.151	2 1.4703	0.167407	11.904

Specific Rate Constant = $1.29 \times 10^{-3} \text{ moles}^{-1} \text{ min.}^{-1} \text{ Liters}$

Rate Of Disproportionation Of <u>p</u>-Methoxybenzoic-acetic Mixed Anhydride At 100°C. Reaction Followed By Measurement Of <u>p</u>-Methoxybenzoic-acetic Mixed Anhydride

Time (min.) <u>Blank</u>	Sample	Blank	<u>%T</u>	Ave. Io	Log ave. Io	Concentration ⁻¹ ($\frac{\text{liters.mole}^{-1}}{}$
. 0	98.1 98.0	21.7 21.6 21.7	98.2 97.9	22,109	4.5285		6.3973
5		23.7 23.7 23.7		24.171 24.171 24.183	4.1438	0.617398	6.8351
10	97.8 97.9 97.8	25.5 25.6 25.5		26.060 26.135 26.073	3.8328	0.583518	7.2319
15	97.8 97.6 98.1	27.2 27.1 27.3		27.826 27.780 27.814	3.5963	0,555856	7.5918
20		28.9 29.0 28.8	97.3 97.4 97.1	29.686 29.774 29.644	; - 3.3668 -	0.527214	8.0043
25		30.8 30.7 30.8	97.5 97.4 97.8	31.589 31.503 31.508	3.1672	0.500678	8.4286
35	92.3 92.0 92.2	31.8 31.8 31.9	91.8	34.434 34.602 34.579	2.8953	0.461695	9.1402

Specific Rate Constant = $1.38 \times 10^{-3} \text{ moles}^{-1} \text{ min.}^{-1} \text{ Liters}$

 $\frac{\text{Rate Of Disproportionation Of } \underline{p\text{-Methoxybenzoic-acetic Mixed}}{\underline{\text{Anhydride At 100°C Reaction Followed By Measurement Of}}_{\underline{p\text{-Methoxybenzoic Anhydride}}}$

Time) <u>Blank</u>	Sample	Blank	<u> 27 </u>	Ave.Io	Log ave. Io	Concentration (liters.moles)
. 0		91.7 91.6 91.7	98.0	93.428 93.469 93.428	1.0701	0.029411	8.7108
5	98.2	90.1 90.2 90.0	98.3	91.891 91.806 91.930	1.0884	0.036790	8.9206
10	97.9 97.9 97.9	88.0 88.0 88.1	97.9	89.887 89.887 89.943	1.1122	0.050068	9.1827
15	97.8 97.8 97.7	85.5 85.4	97.7	87.378 87.468 87.365	1.1441	0.058468	9.3370
20	98.0 97.9 97.8	83.9 83.8 83.8	97.9	85.612 85.597 85.597	1.1681	0.067477	9 . 5 057
25	97.9 97.9 98.0	80.3 80.2 80.3	97.9	82.022 81.920 81.855	1.2205	0.086540	9.8814
35	97.7 97.8 97.6	74.9 75.0 74.8	97.7 97.9 97.5	76.663 76.647 76.678	1.3044	0.115412	10.515
				~ -	3	. 1 _1 .	_1

Specific Rate Constant = $1.60 \times 10^{-3} \text{ moles}^{-1} \text{ min.}^{-1}$ Liters

Rate Of Disproportionation Of p-Methoxybenzoic-acetic Mixed

Anhydride At 100°C Reaction Followed By Measurement Of
p-Methoxybenzoic Anhydride

Time (<u>min.</u>)]	Blank	Sample	<u>Blank</u>	<u>%T</u>	Ave. To	Log ave. Io	Concentration $^{-1}$ (liters.moles $^{-1}$)
. 0	97.7 97.7 97.5	91.3 91.2 91.3	97.7	93.497 93.346 93.545	1.0699	0.02934	8.8183
5	97.9 97.7 97.8	87.1 87.0 86.9	97.9 97.8 97.9	88.968 89.000 88.809	1.1245	0.050960	9.1996
10	97.8 97.7 97.8	84.0 83.9 84.1	97.9 97.9 97.9	85.845 85.787 85.947	1.1646	0.066178	9.4696
15	97.8 97.8 97.6	79.5 79.4 79.4	97.8 97.8 97.5	81.288 81.186 81.394	3 1.2301	0.089947	9.9502
20	97.8 97.9 97.5	76.5 76.6 76.4	97.8 98.0 97.6	78.220 78.203 78.318	1.2780	0.106530	10.309
25	97.8 97.6 97.9		98.0	75.140	1.3303	0.123949	
30	97.8 97.7 97.6	70.5 70.5 70.4	97.7 97.9 97.6	72.122 72.085 72.131	2 5 1.3867	0.141984	11.185
35	97.8 97.8 97.6	68.1 68.2 68.0	97.7 97.9 97.5	69.667 69.698 69.703	3 1.4349 3	0.156820	11.600
40.5	97.8 97.8 97.7	66.0 66.0 66.0	97.8 97.7 97.7	67.484 67.519 67.553	9 1.4810 3	0.170560	12.019
45	97.8 97.7 97.9	64.1 64.0 64.2	97.5	65.541 65.573 65.543	3 1.5255	5 0.183310	12.422

Specific Rate Constant = $1.34 \times 10^{-3} \text{ moles}^{-1} \text{ min.}^{-1}$ Liters

Rate Of Disproportionation Of Benzoic-acetic Mixed Anhydride

At 100°C. Reaction Followed By Measurement Of

Benzoic-acetic Mixed Anhydride

Time		Sample	Blank	<u>%T</u>	Ave.Io	Log ave. Io	Concentration ⁻¹ (liters.mole ⁻¹)
0	90.0 90.2 89.8	43.0 42.7 42.9	90.0 89.9 89.9	47.778 47.418 47.746	3 2.0987	0.321980	7.1184
5	86.9 86.8 86.7	43.9 44.0 43.9	86.8 86.8 86.7	50.546 50.691 50.634	1.9753	0.295636	7.7527
10	87.2 87.6 87.3	45.0 45.0 45.1	87.5 87.7 87.2	51.516 51.340 51.690) 1.9411	0.288052	7.9568
15	86.3 85.8 85.3	46.8 46.9 46.8	85.7	54.323 54.693 54.961	1.8295	0.262325	8.7372
20	87.6 87.0 87.1	49.1 48.9 49.0	87.9 87.0 87.1	55.954 56.206 56.257	1.7812	0.25071	9.1420
25	89.3	52.2 52.1 52.1	89.3	58.651 58.342 58.310	1.7113	0.23325	10.1767
30	88.4 88.3 88.6	52.8 53.0 52.9		59.694 59.920 59.672	1.6733	0.223578	10.2514
35	88.2 88.2 88.3	54.5 54.5 54.2	87.8	61.791 61.931 61.486	1.6198	0.209466	10.9421

Specific Rate Constant = $2.28 \times 10^{-3} \text{ moles}^{-1} \text{ min.}^{-1}$ Liters

Rate Of Disproportionation Of Benzoic-acetic Mixed Anhydride

At 100°C. Reaction Followed By Measurement Of

Benzoic-acetic Mixed Anhydride

Time (min.)	Blank	Sample	Blank	<u> </u>	Ave. <u>Io</u>	Log	ave. <u>Io</u>	Concentration -1 (liters.mole -1)
0		42.2 42.3 42.5	90.0	46.888 47.000 46.961		0.	328260	6.9822
5		45.4 45.7 45.4	90.2	50.110 50.581 50.249	1.9875	5 0.	29831	7.6832
10	89.5 89.5 89.4	47.2 47.3 47.2	89.6	52.737 52.819 52.766	1.8948	3 0.	277564	8.2575
15	89.9 90.1 90.2	50.7 50.3 50.5	90.2	56.022 55.795 56.048	1.7871	. 0.	252145	9.0900
20	90.9 91.3 91.0		91.1	58.784 58.223 58.209	1.7121	. 0.	233526	9.8148
25	90.0 89.6 90.0	54.4 54.5 54.0	89.5	60.545 60.859 60.033		2 0.	219112	10.4604
30	91.2 91.3 91.2	57.0 57.0 56.9		62.465 62.500 62.458	1.6006	ō 0.	204282	11.2197

Specific Rate Constant = $2.82 \times 10^{-3} \text{ moles}^{-1} \text{ min.}^{-1}$ Liters

Rate Of Disproportionation Of Benzoic-acetic Mixed Anhydride

At 100°C. Reaction Followed By Measurement Of

Benzoic-acetic Mixed Anhydride

		40.1.3.1.1.1.1.1.1.1.1.1.1.1.1.1.1.1.1.1.					
<u>Time</u>	<u>Blank</u>	Sample	Blank	<u>%T</u>	Ave Io	$\underline{\text{Log ave.}}_{\underline{\text{I}}}$	Concentration-1
. 0	92.0 91.5 91.4	43.5 43.2 43.2	91.7 91.6 91.5	47.359 47.187 47.238	2.1159	0.325499	7.0414
5	91.8 92.0 91.7	45.8 45.9 45.8	91.6 91.8 91.6	49.836 49.945 49.972	5 2.0032	2 0.301724	7.5963
10	92.0 91.0 92.0	47.4 47.0 47.2	90.6	51.521 51.790 51.388	1.9392	2 0.287624	7.9687
15	90.4 90.2 90.5	48.7 49.0 48.9	90.5	53.931 54.233 54.092	3 1.8487	7 0.266868	8.2485
20	91.3 91.2 91.3	50.4 50.8 50.6	91.3 91.5 91.2	55.202 55.610 55.452	1.8043	3 0.256312	8.9422
25	91.8 92.0 92.1	53.4 53.5 53.4	92.0	58.106 58.152 58.106	2 1.720	5 0.235655	9.7260
30	91.2 91.1 91.3	54.3	91.2 91.4 91.2	60.087 59.505 59.726	5 1.6729	0.223448	10.2574
35	92.0 92.4 92.1	57.1 57.0 57.0	92.3 92.0 92.2	61.964 61.822 61.855	⊇ 1. 6160	0.208440	10.9959
60	91.5 91.8 91.9	60.5 60.4 60.5		66.012 65.795 65.868	5 1.5176	5 0.181058	12.6589

Specific Rate Constant = $2.16 \times 10^{-3} \text{ moles}^{-1} \text{ min.}^{-1}$ Liters

Rate Of Disproportionation Of Benzoic-acetic Mixed Anhydride

At 100°C. Reaction Followed By Measurement Of

Benzoic-acetic Mixed Anhydride

	Time	Blank	<u>Sample</u>	Blank	<u>%T</u>	Ave.Io	Log ave. I	Concentration-1
	0	85.0 85.2 85.1	39.7 39.8 39.7	85.0 85.3 85.1	46.705 46.686 46.650	2.1422	0.33086	6.9274
	5	83.7 84.0 84.3	41.7 41.7 41.8	83.7 84.0 84.2	49.820 49.642 49.614	2.0123	0.303693	7.5470
	10	84.3 85.0 85.1	44.6 44.8 44.9	84.6 84.9 84.9	52.812 52.736 52.823	1.8942	0.277416	8.2619
	15	85.6 83.5 83.5	46.2 45.6 45.7	85.5 83.5 83.4	54.000 54.610 54.763)) 1.8362 }	0.263918	8.6945
. '	20 ,	85.7 85.6 85.6	48.5 48.4 48.3	85.7 85.8 85.5	56.592 56.476 56.458	5 1.7696	0.247874	9.2466
	25	85.6 85.8 85.8	50.2 50.4 50.3		58.644 58.707 58.658	1.7044	0.231570	9.8976
	30	86.0 86.0 85.9	52.1 52.3 52.4	86.2 86.1 86.0	60.511 60.778 60.965	3 1.6460	0.216430	10.5900
	45	83.0 83.3 83.4	53.2 53.3 53.4	83.2 83.4 83.2	65.222 65.146 65.306	i.5331	0.185569	12.3512
1	20	90.8 92.0 92.3	60.0 60.1 60.0	90.5 92.4 92.2	67.217 67.353 67.208	3 1.4867	0.172203	13.3098

Specific Rate Constant = $2.44 \times 10^{-3} \text{ moles}^{-1} \text{ min.}^{-1}$ Liters

Effect Of Added Substances On The Rate Of Disproportionation Of Benzoic-acetic Mixed Anhydride. Reaction Followed By Measurement Of Benzoic-acetic Mixed Anhydride At 100°C

Substance Added	Concentration Moles Liters	Time	Blank	Sample	Blank Ave. To	Concen- tration-1
Benzoic Acid	0.20	0	90.1 90.7 91.0	42.8 43.1 43.3	90.8 2.1049	7.0697
n n	0.20	5	89.8 89.6 90.0	45.7 45.6 45.8	89.9 1.9650	7.8129
11	0.20	10	89.3 89.6 89.3	48.7	89.6 1.8392	8.6614
II.	0.20	15	90.0 90.0 89.8	51.3 51.2 51.1	89.9 90.0 1.7555 89.6	9.3784
11	0.20	20	89.7 90.0 89.7	53.0 53.2 53.1	89.6 89.9 1.6896 89.6	10.063
Specific I	Rate Constant =	3.25	50 x 10	0^{-3} min	\cdot^{-1} moles ⁻¹ L	iters
Sodium Acetate	0.1 gms Suspension		90.2 90.7 90.9	43.2	90.7 2.1029	7.1001
11	· · ·	ō	90.4 90.6 90.8	47.0 47.1 47.2	90.5 1.9367	7.9844
t)	п	10	90.5 90.4 90.4	50.2 50.1 50.1	90.4 1.8031	8.9522
и	11	15	90.6 90.8 91.0	52.7 52.8 52.9	90.7 90.9 1.7206 91.1	9.7246
7]	11	20	91.2 91.4 91.7	56.0 56.1 56.3	91.3 91.5 1.6297 91.8	10.8067
Specific :	Rate Constant =	3.6	4 mole	s ^{-l} min	·-l Liters	

Substance Added	Concentration Moles Liters	Time min.	Blank	Sample	Blank Ave. Io	Concen- tration-1
Powdered Glass	Suspension C.10 gm	0	92.1	43.6	91.8 91.7 2.1154	
a a	и	5	83.7 84.1 84.3	41.8 41.6 41.9	84.0 2.0119	7.5494
n	н	10	87.1 87.7 87.3	45.1 44.9 45.1	87.6 1.9403	7,9619
11		15	85.4 83.6 83.6	46.3 45.7 45.7	85.3 83.6 1.8422 83.7	8,6379
11	it	20	87.8	49.2	88.0 87.4 1.7899	
Specific	Rate Constant =	= 2.01	lO x 10	0^{-3} mole	es ^{-l} min. ^{-l} I	iters
			0= 3	20 5	0= 1	
-		0	85.1 85.3 85.2	39.7 39.9 39.6	85.3 2.1434 85.0	6.9223
		5	85.2 85.3 85.6	42.6 42.8 42.9	85.0 85.5 1.9965 85.7	7.5693
satisfication of a property of the satisfication of	manifesting in a	10	84.4 85.3 85.2	44.7 44.5 44.5	84.8 85.1 1.9065 85.0	8.1788
	MATERIAL PROPERTY AND ASSESSMENT OF THE PROPERTY AND ASSESSMENT OF THE PROPERTY AND ASSESSMENT OF THE PROPERTY OF THE PROPERTY ASSESSMENT OF THE PROPERTY ASSESSMENT OF THE PROPERTY ASSESSMENT OF THE PROPERTY OF THE PROPERTY ASSESSMENT OF THE PROPERTY ASSESSMENT OF THE PROPERTY OF THE P	15	86.3 86.8 87.2	46.4 46.6 46.9	87.0 1.8627	8.4844
Name and American		20	85.8 85.9 85.7	48.7 48.6 48.7	85.0 85.5 1.7627 85.6	9.3103
				- 3		_

Specific Rate Constant = $2.44 \times 10^{-3} \text{ moles}^{-1} \text{ min.}^{-1} \text{ Liters}$

Substance Added	Concentration Moles Liters	Time min.	Blank	Sample	<u> 31ank</u>	Ave. To	Concentration-1
Acetic Acid	0.2	0	92.1 91.6 91.4	43.6 43.1 43.2	91.7 91.5 92.5	2.1117	7.0601
Tf .	0.2	5	93.2 93.3 93.4	47.7 47.8	93.4 93.7		7.8722
ff	0.2	10	93.9 94.1 94.0	51.4 51.6 51.5	93.7 94.3 94.4	1.8265	8.7607
11	0.2		93.6	53.3	93.9	1.7590	
11	0.2	20	93.6 93.5 93.3	55.8 55.8 55.7	93.4 93.4 93.5	1.6757	10.223
Specific 1	Rate Constant :	= 3.00	666 x :	10 ⁻³ mo	les ^{-l}	min1	Liters
Acetic Acid	0.3	0	90.0 90.1 90.2			2.1096	7.0931
11	0.3	5	89.7 89.6 89.8	45.9 46.0 46.1	89.8	1.9503	7.8994
11	0.3	10	89.9 89.8 90.1	49.3 49.4 49.6	90.0	1.8216	8.8002
ff .	0.3	15	90.8 90.6 90.2	52.1 52.0 51.7	90.7 90.8 89.9	1.7426	9.5107
H	0.3	20	91.2 91.0 91.3	54.9 54.7 55.0	91.1 90.8 91.5	1.6591	10.424
				2	7	٦	

Specific Rate Constant = $3.33 \times 10^{-3} \text{ moles}^{-1} \text{ min.}^{-1}$ Liters

Summary Of Data For N-Acyl Bara-Toluidine Systems

- A. Principal Wave lengths
 - 1. N-(acetyl)-p-toluidine --- 7.269 microns
 - 2. N-(benzoyl)-p-toluidine --- 6.676 microns
 - 3. N-(p-methoxybenzoyl)-p-toluidine --- 8.534 microns
 - 4. N-(p-nitrobenzoy1)-p-toluidine --- 7.393 microns
- B. Beer's Law Curves
 - 1. N-(acetyl)-p-toluidine
 - a. At N-acetyl wave length $Log \frac{IO}{T} = 1.271 \text{ (N-acetyl)} + 0.000$
 - b. At N-benzoyl wave length $Log \frac{Io}{I} = 1.211 \text{ (N-acetyl)} + 0.00025$
 - c. At N-(p-methoxybenzoyl) wave length $Log \frac{Io}{I} = 0.0858 \text{ (N-acetyl)} + 0.000$
 - d. At N-(<u>p</u>-nitrobenzoyl) wave length $Log \frac{Io}{I} = 0.3150 \text{ (N-acetyl)} + 0.000$
 - 2. $N-(benzoyl)-\underline{p}-toluidine$
 - a. At N-acetyl wave length $Log \frac{Io}{T} = 0.196 \text{ (N-benzoyl)} + 0.000$
 - b. At N-benzoyl wave length $Log \frac{Io}{I} = 1.814 \text{ (N-benzoyl)} + 0.008$
 - 3. $N-(\underline{p}-methoxybenzoyl)-\underline{p}-toluidine$
 - a. At N-acetyl wave length $\log \frac{Io}{T} = 1.184(N-p-Methoxybenzoyl) + 0.000$
 - b. At N-(p-methoxybenzoyl)-p-toluidine wave length $\log \frac{Io}{T} = 10.810 \text{ (N-p-methoxybenzoyl)} + 0.000$

- 4. N-(p-nitrobenzoyl) p-toluidine
 - a. At N-acetyl wave length $Log \frac{Io}{T} = 0.7442 \text{ (N-p-nitrobenzoyl)} + 0.000$
 - b. At N-p-nitrobenzoyl p-toluidine $\label{eq:log_log_p} \text{Log} \ \frac{\text{Io}}{\text{I}} = 6.560 \ (\text{N-p-nitrobenzoyl}) + 0.000$

Simultaneous Equations Derived From Above Data

- A. For the benzoic-acetic mixed anhydride at 7.269 microns; 0.196 (benzoyl) + 1.271 (acetyl) = $\log \frac{Io}{I}$ at 6.676 microns; 1.814 (benzoyl) + 1.211 (acetyl) = $\log \frac{Io}{I}$ 0.0083
- B. For <u>p</u>-methoxybenzoic-acetic mixed anhydride at 7.269 microns; 1.184 (methoxybenzoyl) + 1.271 (acetyl) = $\frac{Io}{I}$ at 8.534 microns; 10.81 (methoxybenzoyl) + 0.0858 (acetyl) = $\frac{Io}{I}$
- C. For p-nitrobenzoic-acetic mixed anhydride at 7.269 microns; 0.7442 (nitrobenzoyl) + 1.271 (acetyl) = $\frac{Io}{I}$ at 7.393 microns; 6.565 (nitrobenzoyl) + 0.3150 (acetyl) = $\frac{Io}{I}$

Beer's Law Curve For N-Benzoyl p-toluidine At N-Acetyl-p-toluidine Wave Length 7.269 Microns In Chloroform

Concentration	Blank	Sample	Blank	<u>%T</u>	Ave. Io	Log Io
0.1908	98.2 98.0 98.3	90.1 90.0 90.2	98.1 98.2 98.2	91.798 91.743 91.806	1.0895	0.03720
0.1526	98.5 98.3 98.2	91.8 91.7 91.7	98.4 98.2 98.1	93.245 93.333 93.428	1.0712	0.029870
0.1145	98.5 98.2 98.3	93.5 93.5 93.4	98.4 98.4 98.4	94.972 95.116 94.966	1.0524	0.022184
0.07632	98.5 98.5 98.4	95.2 95.3 95.2	98.5 98.4 98.4	96.649 96.800 96.747	1.0337	0.014394

 $Log \frac{Io}{I} = 0.1960 (N-Benzoyl tolu) + 0.000$

Beer's Law Curve For N-Benzoyl-p-toluidine At Its Peak 6.676 Microns In Chloroform

Concentration	Blank	Sample	Blank	%T	Ave Io	Log Io
0.1908	98.2 98.1 98.2	43.7 43.6 43.7	98.2 98.3 98.3	44.501 44.399 44.478	2.2492	0.352028
0.1526	98.2 98.1 98.1	51.0 51.1 51.0	98.0 98.2 98.0	51.987 52.063 52.014	1.9219	0.283728
0.1145	98.6 98.5 98.7	60.4 60.5 60.6	98.5 98.4 98.8	61.288 61.452 61.367	1.6294	0.212028
0,7632	97.0 97.0 96.9	67.8 67.6 67.7	97.1 96.9 96.9	69.860 69.726 69.865	1.4323	0.156033
0.5724	98.3 98.3 98.4	76.0 76.0 75.9	98.4 98.5 98.5	77.275 77.235 77.094	1.2953	0.112372
0.3816	98.6 98.6 98.5	82.5 82.4 82.5	98.7 98.5 98.6	83.628 83.612 83.713	1.1954	0.077514
0.1908	98.6 98.5 98.5	89.6 89.6 89.7	98.5 98.5 98.4	90.826 90.965 91.112	1.0992	0.041078
Los	g <u>Io</u> =	1.814 (Benzov	rl tolu)	+ 0.008	

 $Log \frac{Io}{I} = 1.814 \text{ (Benzoyl tolu)} + 0.008$

Beer's Law Curve For N-Acetyl-p-toluidine At N-(p-Nitro-benzoyl)-p-toluidine Wave Length 7.393 Microns

Concentration	Blank	Sample	Blank	<u>%T</u>	Ave. Io	$\frac{\text{Log}}{\text{I}}$
0.2121	97.8 97.8 97.7	84.0 84.1 84.2	97.8 97.6 97.8	85.889 86.079 86.138	1.1623	0.065328
0.1697	97.7 97.8 97.8	86.4 86.3 86.4	97.8 97.7 97.8	88.388 88.286 88.343	1.1320	0.053850
0.1273	97.7 97.6 97.5	90.1 90.0 90.0	97.6 97.5 97.4	92.268 92.260 92.355	1.0834	0.034790
0.08495	97.7 97.5 97.7	92.2 92.1 92.2	97.6 97.6 97.7	94.418 94.413 94.370	1.0588	0.024818
Log	$\frac{Io}{I} =$	0.3150	(Acety	71 tolu)	+ 0.000	

Beer's Law Curve For N-(acetyl)-p-toluidine At N-(Benzoyl)-p-toluidine Peak 6.677 Microns In Chloroform

Concentration	Blank	Sample	Blank	<u>ZT</u>	Ave. Io	Log Io
0.2176	98.2 98.1 98.2	64.2 63.8 64.0	98.1 98.1 98.0	65.410 65.035 65.239	1.5330	0.18554
0.17408	98.2 98.3 98.2	70.5 70.3 70.4	98.2 98.0 98.1	71.792 71.625 71.726	1.3944	0.144384
0.13056	98.2 98.0 98.1	76.0 76.4 76.2	98.0 98.1 98.1	77.471 77.919 77.675	1.2872	0.109648
0.08704	98.0 98.2 98.0	83.0 82.9 83.0	98.1 98.2 98.2	84.650 84.419 84.607	1.1826	0.072836
0.04352	98.3 98.3 98.5	89.9 89.8 90.0	98.4 98.2 98.4	91.408 91.399 91.416	1.0939	0.038980
0.02176	98.3 98.0 98.2	98.9 93.7 93.8	98.2 98.1 98.4	95.572 95.563 95.422	1.0469	0.019908
Lo	g <u>Io</u> =	1.211	(Acety)	l tolu)	+ 0.00025	

Beer's Law Curve For N-Acetyl-p-toluidine At Its Wave Length 7.269 Microns In Chloroform

Concentration moles Liters-1	<u>Blank</u>	Sample	Blank	<u>%T</u>	Ave. Io	$\frac{\text{Log}}{\text{T}}$
0.2677	97.6 97.8 97.6	44.3 44.7 44.6	97.5 98.0 97.5	45.412 45.658 45.720	2.1931	0.341060
0.2176	98.1 98.1 98.1	53.0 53.0 53.1	98.0 98.1 98.0	54.054 54.026 54.156	1.8491	0.266964
0.1741	97.7 98.0 98.1	59.0 59.2 59.1	98.0 98.1 98.1	60.296 60.377 60.244	1.6589	0.219823
0.1306	98.1 98.2 98.4	67.6 67.6 67.6	98.2 98.3 98.3	68.874 68.804 68.734	1.4640	c.16554
0.0870	97•9 98.0 97•9	76.3 76.2 76.3	98.0 97.9 97.9	77.896 77.794 77.936	1.2841	0.108603
0.0435	98.0 98.0 98.2	86.4 86.5 86.4	98.0 98.1 98.1	88.163 88.220 88.028	1.1345	0.050860
0.02176	97.9 98.0 97.7	92.2 92.0 92.1	97.9 97.7 97.9	94.177 94.069 94.171	1.0622	0.026202
Т	Io	1 007 /	^ ^ -	/\	. 0 000	

 $Log \frac{Io}{I} = 1.271 \text{ (Acetyl tolu)} + 0.000$

Beer's Law Curve For N-Acetyl-p-toluidine At (p-Methoxy-benzoyl)-p-toluidine Wave Length 8.534 Microns In Chloroform

Concentration moles liters	<u>Blank</u>	Sample	Blank	<u>%T</u>	Ave. <u>Io</u>	$\frac{\text{Log}}{\text{I}}$
0.262	95.4 95.8 95.2	90.5 90.8 90.1	95.4 95.6 95.0	94.863 94.879 94.742	1.0545	0.023045
0.2096		92.3 92.2 92.3	96.3 96.1 96.3	95.746 95.891 95.945	1.0431	0.018322
0.1834		92.7 92.6 92.7	96.0 95.8 95.9	96.663 96.609 96.764	1.0343	0.014646
0.1332	96.2 96.1 96.4	93.9 93.6 94.0	96.3 96.0 96.3	97.558 97.449 97.560	1.0254	0.010892
	To					

 $Log \frac{Io}{I} = 0.0858 \text{ (Acetyl tolu)} + 0.000$

Beer's Law Curve For N-(p-methoxybenzoyl)-p-toluidine At N-acetyl-p-toluidine 7,269 Microns In Chloroform

Concentration	Blank	Sample	Blank	<u>%T</u>	Ave. Io	Log Io
0.0692	97.1 96.8 96.7	80.2 79.9 80.0	-	82.637 82.626 82.644	1.2101	0.082825
0.0554	96.8 97.2 97.4		97.0 97.4 97.5	86.171 86.125 86.095	1.1610	0.06483
0.0415	96.9 97.0 96.9	86.5 86.6 86.3	97.1 97.2 96.7	89.129 89.186 89.152	1.1216	0.049828
0.0277	95.6 95.8 95.6	88.6 88.7 88.5	95.7 95.7 95.4	92.629 92.637 92.670	1.0793	0.03314
0.0138	95.8 95.8 96.0	92.2 92.3 92.5	95.7 95.9 96.2	96.292 96.296 96.253	1.0386	0.016452
Lo	$g \frac{Io}{I} =$	1.184	(<u>p</u> -MeO	tolu) +	0.000	

Beer's Law Curve For N-(p-Methoxybenzoyl)-p-toluidine

At Its Wave Length 8.534 Microns

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Concentration	Blank	Sample	Blank	<u>%T</u>	Ave. Io	Log Io
0.02995	97.5 97.7 97.6	46.8 46.7 46.6	97.6 97.6 97.5	47.975 47.823 47.770	2.0881	0.319752
0.02395	98.4 98.3 98.4	55.2 55.1 55.2	98.5 98.4 98.4	56.069 56.024 56.097	1.7837	0.251318
0.01796	98.4 98.3 98.4	63.2 63.2 63.1	98.4 98.2 98.4	64.227 64.325 64.126	1.5543	0.191628
0.01198	98.4 98.4 98.3	73.1 73.2 73.1	98.4 98.4 98.4	74.288 74.390 74.326	1.3452	0.128786
0.00598	98.2 98.3 98.3	85.4 85.3 85.2	98.3 98.4 98.3	86.921 86.731 86.673	1.1524	0.061502
	Т 🙃					

 $Log \frac{Io}{I} 10.81 (pMe0tolu) + 0.000$

Beer's Law Curve For N-(p-Nitrobenzoyl)-p-toluidine At

N-Acetyl-p-toluidine Wave Length

7.269 Microns In Chloroform

Concentrati moles Liter		Sample	Blank	%T	Ave. Io	Log Io
0.0940	96.8 96.2 96.3	82.3 82.0 81.9	96.9 96.4 96.1	84.977 85.150 85.135	1.1757	0.07030
0.0752	95.8 96.1 96.4	84.1 84.4 84.6	95.9 96.0 96.2	87.741 87.870 87.850	1.1386	0.056368
0.0564	95.9 95.7 95.8	86.9 86.9 86.8	95.8 95.9 95.7	90.662 90.709 90.652	1,1032	0.042658
0.0376	96.2 96.3 96.2	90.2 90.3 90.4	96.1 96.2 96.4	93.811 93.818 93.873	1.0657	0.027596
0.01880	95.9 95.8 95.6	92.9 92.7 92.5	95.8 95.7 95.7	96.922 96.814 96.706	1.0329	0.014058
	$Log \frac{Io}{I} =$	0.7442	(p-Nit	ro tolu)	+ 0.000	

Beer's Law Curve For N(p-Nitrobenzoyl)-p-toluidine At Its
Wave Length 7.393 Microns In Chloroform

,		- 1 - 2 2 2				_
Concentration	<u>Blank</u>	Sample	Blank	<u>%T</u>	$\frac{\text{Ave.}}{\text{I}}$	$\frac{\text{Log}}{\text{T}}$
0.03484	97.8 97.9 97.8		97.9 97.9 98.0	60.091 59.959 60.061	1.6656	0.221666
0.02788	97.8 97.7 97.8		97.8 97.7 97.7	65.848 65.916 65.984	1.5170	0.18099
0.02093	97.6 97.6 97.5	70.9		72.606 72.643 72.717	1.3776	0.139116
0.01394	97.6 97.6 97.5	79.0	97.6 97.6 97.6	80.942 80.942 81.086	1.2347	0.091565
0.006974	97.5 97.6 97.6	87.7		89.810 89.856 89.754	1.1134	0.046656
Log	$S = \frac{IO}{I} =$	6 <u>.5</u> 60	(p-Nit	ro tolu)	+ 0.000	

Cleavage Of Benzoic-acetic Mixed Anhydride With p-Toluidine

Reaction Medium	Wave Length	Blank	Sampl	e Blan	<u>k %T</u>	Ave. <u>Io</u>	Log Io				
Benzene	7.269	93.4 93.5 93.6	26.2 26.3 26.4	93.5 93.6 93.4	28.036 28.113 28.220	3 . 5558.	0.550936				
	6.676	93.3 93.3 93.2	26.4 26.5 26.3	93.2 93.3 93.1	28.289 28.403 28.234	3.5324	0.548068				
	Melting p	oint o	f mixt	ure =	149 - 150 ⁰	C					
	Concentra	tion N	-acety	1 = 0.	431956						
	Concentra	Concentration N-benzoyl = 0.0098277									
	Acetylati	Acetylation ratio = 44.0									
50 w/w Acetone- Benzene	7.269	93.5 93.2 93.1	50.0 49.8 49.7	93.5 93.3 93.2	53.475 53.404 53.354	1.8722	0.272356				
	6.676	93.7 94.0 93.8	50.2 50.3 50.1	94.0 94.1 94.0	53.489 53.482 53.354	1.8718	0.272262				
	Melting p	oint o	f mixt	ure =	148.6-15	o ^o c					
	Concentra	tion N	-acety	1 = 0.	213414						
	Concentra	tion N	-benzo	y1 = 0	.005665						
	Acetylation	on rat:	io = 3'	7.7							

Reaction Medium	Wave Length					Ave. Io	
Acetone	7.269	94.5 94.0 94.1	36.4 36.2 36.3	94.4 94.2 94.3	38.538 38.469 38.535	2.5964	0.414368
	6.676	95.3 95.1 95.0	36.1 36.1 36.0	95.2 95.0 94.8	37.900 37.980 37.934	2.6358	0.420916
	Melting p	oint o	f mixt	ure =	148.4-15	0 ⁰ C	
	Concentra	tion N	-acety	1 = 0.	32425		
	Concentra	tion N	-benzo	yl = 0	.0109437		
	Acetylati	on rat:	io = 2	9.6		,	
50 w/w Acetone- Water	7.269	94.5 94.6 94.5	29.9 29.8 30.0	94.7 94.3 94.7	31.606 31.551 31.712	3.1606	0.499768
	6.676	94.1	29.4	94.2 94.2 94.4	31.299 31.226 31.372	3.1949	0.504456
	Melting p	oint o	f mixt	ure =	149 - 150 ⁰	C	
	Concentra	tion N-	-acety	1 = 0.	49747		
	Concentra	tion N-	-benzo	yl = 0	.0117124		

Acetylation ratio = 42.5

$\frac{\texttt{Cleavage Of } \underline{p}\text{-}\texttt{Methoxybenzoic-acetic Mixed}}{\texttt{Anhydride With } \underline{p}\text{-}\texttt{Toluidine}}$

Reaction Medium	Wave Length	Blank	Sampl	e Blani	<u>k %T</u>	Ave Io	Log Io
Bezene	7.269	97.8 97.9 98.0	64.9 64.8 65.0	97.9 98.1 98.0	66.326 66.088 66.326	1.5095	0.178835
	8.534	97.8	91.9	97.8 97.7 97.9	94.069 94.015 94.021	1.0634	0.026694
	Melting po	oint of	C mixt	ure = 3	143°C-14	4°c	
	Concentra	tion N	-acety	1 = 0.	139427		
	Concentra	tion N	-p-metl	noxybei	nzoyl = (0.0013620	02
	Acetylatio	on rati	io = 10	02			
50 w/w Acetone-	.7.269	97.0 97.1 97.2	45.6 45.7 45.5	97.2 97.3 96.9	46.961 47.016 46.883	2.1297	0.328320
	8.534	96.8 97.1 97.2	88.3	97.0 97.3 97.0	90.815 90.843 90.834	1.1009	0.041750
	Melting point of mixture = 143.5°C=144.6°C						
	Concentrat	tion N-	-acety:	1 = 0.2	256616		
	Concentrat	tion N-	-p-meti	noxybei	nzoyl = (0.0018241	17
	Acetylatio	on rati	lo = 1	40			

Reaction Medium	Wave Length	Blank	Sampl	e Blan	k %T	Ave. Io	Log Io
Acetone	7. 269	98.9 98.6 98.9	54.5	98.7 98.6 98.9	55.263 55.273 55.308	1.8089	0.257396
	8.534	98.9 98.9 98.7	91.2	98.9	92.159 92.214 92.206	1.0846	0.035270

Melting point of mixture = $143^{\circ}\text{C}-144^{\circ}\text{C}$ Concentration N-acetyl = 0.200965 Concentration N-p-methoxybenzoyl = 0.0016676 Acetylation ratio = 120

50 w/w Acetone- Water	7.269		35.7 35.8 35.8	98.0	36.521 36.549 36.511	2.7379	0.437414
	8.534	98.2 98.3 98.5	83.8 83.8 84.0	98.1	85.379 85.492 85.322	1.1709	0.068523

Melting point of mixture = $144^{\circ}\text{C}=145^{\circ}\text{C}$ Concentration N-acetyl = 0.340763Concentration N-p-methoxybenzoyl = 0.00363418Acetylation ratio = 93.7

Cleavage Of p-Nitrobenzoic-acetic Mixed Anhydride With p-toluidine

Reaction Medium	Wave Length	Blank	Samp1	e <u>Blan</u>	<u>k %T</u>	Ave. Io	Log Io
Benzene	7 . 269	95.3 95.6 95.3	61.6 61.7 61.5	95.6 95.4 95.2	64.536 64.607 64.566	1.5487	0.189966
	7.393	95.8 95.9 96.1	44.1 44.2 44.2	95.9 96.0 96.0	46.009 46.065 46.017	2,1724	0.336940
	Melting p	oint o	f mixt	ure =	125 - 131 ⁰	C ,	
	Concentra	tion N	-acety	1 = 0.	12286		
	Concentra	tion N	<u>-p-ni</u> t	robenz	oyl = 0.	045428	
	Acetylati	on rat:	io = 2	.70			
50 w/w Acetone-	7.269	94.9	47.0	95.0 94.8 94.6	49.525 49.551 49.445	2.0212	0.305612
	7.394	94.5 94.6 94.6	40.5 40.5 40.4	94.4 94.6 94.5	42.879 42.811 42.728	2.3470	0.37051
	Melting p	oint o	f mixt	ure = :	126-132 ⁰	3	
	Concentra	tion N	-acety	1 = 0.2	213477		

Concentration N-p-nitrobenzoy1 = 0.046194

Acetylation ratio = 4.66

Reaction Medium	Wave Length	Blank	Sample	e <u>Blan</u> l	<u>k %T</u>	Ave Io	Log Io
Acetone	7.269	93.1	39.2	93.3 93.0 93.1		2.3718	0.375020
	7.394	93.2 93.0 93.5	26.2 26.2 26.1	93.1 93.4 93.4	28.126 28.111 27.929	3.5644	0.551988
•	Melting po	oint o	f mixt	ure = 1	126,5-13	1°C	
	Concentra	tion N	-acety	1 = 0.2	25308		
	Concentra	tion N	- <u>p</u> -nit	robenz	oyl = C.	071937	
	Acetylatio	on rat:	io = 3	•52		,	
50 w/w Acetone-	7.269	92.8 93.0 93.1	48.0 48.1 48.2	93.0 93.1 93.2	51.668 51.692 51.744	1.9341	0.286482
	7.394	93.1	33.5	93.2	36.040 35.963 36.032	2.7768	0.443548
	Melting po	oint of	f mixt	ure = 1	124.8-13	3.1°C	
	Concentra	tion N	-acety	1 = 0.1	19121		
	Concentra	tion N	- <u>p</u> -nit	robenz	oy1 = 0.	058387	

Acetylation ratio = 3.28

Apparent First Order Rate Constants For The Hydrolysis Of Para Substituted Benzoic-acetic Mixed Anhydrides

A. p-Methoxybenzoic-acetic Mixed Anhydride

x = 9.630 milliliters

t(min)	$\frac{x_{\infty} - x_{t}}{}$	t(min)	$\frac{\mathbf{x}_{\infty} - \mathbf{x}_{t}}{\mathbf{x}_{t}}$
19.0	5.09	234.5	3.40
39.2 59.0	4.59 4.06	255.0 295.0	3.25 3.08
79.0 99.0	4.03 4.10	324.0 360.0	2.95 2.89
119.0	3.97	395.0	2.72
159.0 179.0	3.83 3.83	455.0 485.0	3.09 2.40
202.5	3.56	515.0	2.21

Corrected first order rate constant = 1.41 x 10^{-5} Sec.⁻¹

B. <u>p</u>-Nitrobenzoic-acetic Mixed Anhydride

x **∞** = 22.35 Milliliters

t(min)	x on -xt	t(min)	x o - xt
1 2 3 4 6 8 10 12	10.77 10.35 10.33 10.25 9.93 9.83 9.73 9.53	15 18 21 24 28 32 37 43 51	9.37 9.15 9.00 8.82 8.77 8.26 8.04 7.74 7.33

Corrected first order rate constant = $23.7 \times 10^{-5} \text{ Sec.}^{-1}$

C. Benzoic-acetic Mixed Anhydride

x = 14.885 milliliters

t(min)	x x _t	t(min)	x - xt
1.2 4.0 8.5 12.5 16.0 20.0 24.0 28.0	7.311 7.204 7.085 6.985 6.898 6.845 6.715 6.655	32.0 36.0 39.0 43.5 47.5 54.0 60.0 70.0	6.525 6.406 6.335 6.265 5.035 5.765 5.527

Corrected first order rate constant = $3.77 \times 10^{-5} \text{ Sec.}^{-1}$

PART II. STUDIES OF THE ALPHA-CHYMOTRYPSIN CATALYZED HYDROLYSIS OF SPECIFIC SUBSTRATES

THE KINETICS OF THE C CHYMOTRYPSIN CATALYZED HYDROLYSIS OF THREE N-ACYL-L-TYROSIN HYDRAZIDES

It has been known for a long time that the enzyme dechymotrypsin can catalyze the hydrolysis of certain acylated and unacylated amino acid esters, amides and hydrazides.

The work which is reported here is concerned with the kinetics of the hydrolysis of three N-acyl-L-tyrosinhydra-zides; the N-trimethylacetyl, the N-dichloroacetyl and the N-formyl-L-tyrosinhydrazides. The results are shown in Table 1.

Table 1

The Kinetic Constants for Three N-Acyl-L-tyrosinhydrazides

THE BITTE OF COMPONING TOT THEE IN	-MGAT-D-CALOSTILLA	uraziues
	$k_3(\min.^{-1})x10^{-3}$	K _s x10-3
N-Formyl-L-tyrosinhydrazide	0.068	11.8
N-Trimethylacetyl-L-tyrosinhydrazi	de 0.30	29.2
N-Dichloroacetyl-L-tyrosinhydrazid	e 0.53	3.84

At the time that this work was undertaken it was believed that the course of the hydrolysis of the N-acyl-Ltyrosinhydrazides could be predicted upon the basis of the classical Michaelis-Menten formulation. However, it has subsequently been shown by other investigators in these laboratories that this belief may not be valid. The values of the various kinetic constants which are reported here are calculated on the basis of the classical mechanism and must be regarded as provisional. Therefore no attempt has been made to correlate any of the results either among themselves or with respect to data determined by other investigators.

The classical approach, which was found to be valid for the hydrolysis of the amides, assumes that the enzyme and substrate react reversibly with one another to form an enzyme-substrate complex. This complex may then undergo a transformation resulting in the formation of the free enzyme and the products of hydrolysis.

$$E+S \stackrel{K_S}{=} [ES] \stackrel{k_3}{\longrightarrow} E+P_1+P_2$$

$$K_S = \frac{[E][S]}{[ES]}$$
 $\frac{d[P]}{dt} = k_3[ES]$

On the basis of this reaction mechanism it is possible to derive the following relationship:

$$\frac{-dS}{dt} = V_t = k_3 [ES] = \frac{k_3 [E][S]}{K_S + [S]} - A$$

Where [S] = Substrate concentration at time t

[E] = Total enzyme concentration

[ES] = Enzyme-substrate complex concentration

 V_{t} = Velocity at time t

By taking the reciprocal of the above equation, the following equation is obtained:

$$V = \frac{K_s + [s]}{k_s [e][s]} = \frac{K_s}{k_s [s]} (V_s) + \frac{1}{k_s [e]} - B$$

When t=0, then 5=50 and V=Vo equation B becomes

$$\frac{1}{V_0} = \frac{K_S}{k_3 [E]} \left(\frac{1}{[S]} \right) + \frac{1}{k_3 [E]} - C$$

Equation C establishes a relationship which enables one to estimate the kinetic constants $K_{\rm S}$ and $k_{\rm 3}$ from a knowledge of the initial velocities at various substrate concentrations.

A complicating factor arises when the products of

hydrolysis combine with the enzyme and hence prevent its further interaction with the substrate. The effect of product inhibition may be included in the basic formulation by defining the following equilibrium:

This results in the modification of equation B to include $K_{\mbox{\scriptsize m}}$:

$$V = \frac{K_S}{k_3[E]} \left(\frac{1}{S} \right) + \frac{1}{k_3[E]} + \frac{K_S}{[E][KP]} \left(\frac{S_0 - S}{S} \right) - B$$

It is possible to neglect the effect of product inhibition if the experiment is designed so that only a small percentage of hydrolysis is followed and if it is assumed that $K_S > K_O$.

This is the basis upon which the experiments which led to the results shown in Tables 3, 5 and 7 were carried out. It is predicated upon the validity of equation B.

By integration of equation B' it is possible to derive equation D.

From this relationship it is seen that a plot of $\frac{\ln S_0/S}{t}$ versus $\frac{S_0-S}{t}$ should be a straight line whose slope is a function of S_0 , K_p , and K_s . For the hydrolysis of several N-acyl-L-tyrosinamides this linear result has been obtained.

W. Lands of these laboratories has recently observed the hydrolysis of N-acetyl-L-tyrosinhydrazide over a relatively large extent of reaction. When the data for this reaction were examined according to equation D a decided deviation from linearity was obtained. The data indicate that the interaction of the enzyme and product may be greater than any that has been observed so far, with the apparent K_{p} much greater than K_{s} .

The data for the hydrolysis of N-formyl-L-tyrosinhydrazide and N-trimethylacetyl-L-tyrosinhydrazides (Tables 3 and 5) do not show any influence of product interaction. This is presumably due to the small extent of reaction which was observed and the tendency of any random error in the data to be magnified when plotted according to equation D.

N-Dichloroacetyl-L-tyrosinhydrazide hydrolyzes at a somewhat more rapid rate. When the data for its hydrolysis were examined by equation D a classical K_p of approximately 3.8 x 10^{-3} was indicated (Figure 8). The effect of this apparent classical product inhibition was corrected for by application of the Lands modification of the Jennings correction (recorded as f_0 in Table 7).

N-Trimethylacetyl-L-tyrosinhydrazide was found to be an extremely soluble substrate. It was possible to work with solutions which were as concentrated as 0.24F. This substrate concentration was much higher than had ever been obtained before.

The Ammonium Salt Of N-Acetyl-L-Cysteic Acid Carboxyl-amide As A Substrate

During his work on the structure of insulin, Sanger observed that **c**chymotrypsin catalyzed the hydrolysis of a peptide bond which involved the carboxyl group of cysteic acid. This was the first time it was suggested that **c**chymotrypsin would act upon cysteic acid peptides.

It was hoped that this would prove to be a general observation and that simple cysteic acid derivatives would serve as suitable substrates. N-Acetyl-L-cysteic acid carboxylamide (I) was synthesized to serve as a model substrate.

It was found, however, that it was not possible to demonstrate (Table 8) that any appreciable hydrolysis had

occurred when **Q** chymotrypsin was allowed to act upon (I). In like manner it was not possible to demonstrate that (I) has any appreciable ability to act as a competitive inhibitor (Table 9). Under these conditions the ammonium salt of N-acetyl-L-cysteic carboxylamide has no ability to influence the behavior of **Q**chymotrypsin.

There is an accumulation of evidence to indicate that a negative charge is associated with the active site of chymotrypsin. In view of this evidence it is not surprising that the negatively charged cysteyl ion fails to interact with the enzyme.

The cysteic acid peptide link in insulin which was hydrolyzed is a bond in which the amino group of serine is involved. It is suggested (2) that at some time, perhaps during the preliminary treatment of the insulin molecule with performic acid, an "N" to "O" shift had occurred. In this process the carboxyl group of the cysteic acid residue had migrated to the adjacent hydroxyl group converting the peptide link into an ester bond. It is well known that a chymotrypsin will catalyze the hydrolysis of a wide variety of esters.

As a corollary to this work an experiment was run to determine the effect of sulfate ion on the activity of a chymotrypsin. The results are shown in Table 10. It is evident that under these conditions sulfate ion does not affect the enzyme.

The Synthesis Of The N-Acyl-L-tyrosinhydrazides

The synthetic route to N-trimethylacetyl-L and N-dichloroacetyl-L-tyrosinhydrazides is outlined below.

$$HO = CH_{2} - CH_{-}C - O + C_{2}H_{5}OH \xrightarrow{C_{2}H_{5}OH} HO = CH_{2} - CH_{-}C - OC_{2}H_{5}(I)$$

$$KHCO_{3} = R - C - CI$$

$$HO = CH_{2} - CH_{-}C - OH_{-}OH_{2}$$

$$HO = CH_{2} - CH_{-}C - OH_{-}OH_{2}$$

$$HO = CH_{2} - CH_{-}C - OC_{2}H_{5}$$

$$HO = CH_{2} - CH_{-}C - OC_{2}H_{5}$$

$$HO = CH_{2} - CH_{-}C - OC_{2}H_{5}$$

$$H_{2}N - NH_{2}$$

$$HO = CH_{2} - CH_{-}C - OC_{2}H_{5}$$

$$HO = CH_{2} - CH_{2} - CH_{-}C - OC_{2}H_{5}$$

$$HO = CH_{2} - CH_{2} - CH_{2} - CH_{2}$$

$$HO = CH_{2} - CH_{2} - CH_{2} - CH_{2}$$

$$HO = CH_{2} - CH_{2} - CH_{2} - CH_{2}$$

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$$HO = CH_{2} - CH_{2} - CH_{2} - CH_{2}$$

$$HO = CH_{2} - CH_{2} - CH_{2} - CH_{2}$$

$$HO = CH_{2} - CH_{2} - CH_{2} - CH_{2}$$

$$HO =$$

L-Tyrosine Ethyl Ester (I)

One mole of L-tyrosine (Merck & Co.) was suspended in $2\frac{1}{2}$ liters of absolute ethanol⁽³⁾ contained in a five liter three neck round bottom flask. The center neck contained a mechanically sealed electrically driven stirrer. One of the side necks contained a drying tube filled with calcium chloride. Into the remaining neck was inserted a delivery tube for the admission of anhydrous hydrogen chloride. The whole assembly was packed in ice and maintained at 0° C.

The cold ethanolic suspension was saturated with anhydrous hydrogen chloride, the process taking $2\frac{1}{2}$ hours. A clear solution was obtained and allowed to stand over night at room temperature.

The amber colored ethanolic solution was then evaporated to dryness under reduced pressure (25 mm.) on a steam bath.

The crude L-tyrosine ethyl ester hydrochloride was ground to a fine powder and suspended, with the aid of a mechanical stirrer, in one liter of dry chloroform cooled to 0° C. Two hundred ml. of a cold saturated solution of ammonia in chloroform was added over a period of 30 minutes. After an additional $\frac{1}{2}$ hour the precipitated ammonium chloride was filtered off and the chloroform evaporated to dryness under reduced pressure.

The crude L-tyrosine ethyl ester was recrystallized twice from dry ethyl acetate.

Final M.P =
$$108.5-109^{\circ}$$
C
[α] = + 20.5 (methanol)

Yield =
$$69\%$$

The material was kept in the dark and stored at 5° C until used.

Alkyl Acid Chlorides

Trimethyl, dichloro and trichloroacetyl chlorides were synthesized by the method of H. C. Brown. $^{(4)}$ This method is

based upon the reaction of the desired acid with benzoyl chloride. The method is applicable only to acid chlorides which have a boiling point lower than benzoyl chloride.

	B.P.	<u>Yield</u>
Trimethylacetyl Chloride	104-105°C	85%
Dichloroacetyl Chloride	106-107.5°C	74%
Trichloroacetyl Chloride	116-118°C	53%

Acylation of L-Tyrosine Ethyl Ester

Five one-hundredths mole of L-tyrosine ethyl ester was dissolved in 300 ml. of dry ethyl acetate contained in one liter Erlenmeyer flask. The solution was cooled to -5 C in an ice salt bath. The cold solution was agitated with an emulsifying, vibrating, electrically driven stirrer. Twenty-five thousandths mole of the acid chloride was then added. After two minutes 50 ml. of 0.05 N potassium carbonate solution was added, two minutes later another 0.025 mole portion of the acid chloride was added followed two minutes later by another 50 ml. portion of potassium carbonate. The process was repeated until four samples of acid chloride and the potassium carbonate solution had been added. During the process of addition the entire system was completely emulsified.

The cold solution was allowed to separate into two phases. The ethyl acetate phase was washed with 6 N HCl and then twice with cold distilled water. The ethyl acetate solution was dried by being shaken with calcium chloride and then with anhydrous potassium carbonate.

The ethyl acetate was removed by evaporation under reduced pressure. The product that was obtained was not treated further but converted directly to the hydrazide.

	Crude Yield	Physical State
N-Trimethylacetyl-L-Tyrosine Ethyl Ester	55%	oil
N-Dichloroacetyl-L-Tyrosine Ethyl Ester	48%	gummy solid

The Conversion of the N-Acyl-L-tyrosine Ethyl Esters to the Corresponding Hydrazides.

The crude esters were dissolved in $2\frac{1}{2}$ times their weight of anhydrous methanol. The solutions were heated to boiling and a 50% excess of hydrazine hydrate was quickly added. The cool mixtures were allowed to stand over night.

The volumes of the methanolic solutions were reduced and the hydrazides allowed to crystallize. The yield of the crude materials averaged 80%.

The compounds were recrystallized from methanol-water mixtures.

N-Trimethylacetyl-L-tyrosinhydrazide

M.P. =
$$179-180^{\circ}$$
 C
[α]_D²⁵ = 29.0° (water)

N-Dichloroacetyl-Ltyrosinhydrazide

$$M.P. = 193-193.5^{\circ} C$$

$$[\alpha]_{D}^{26} = +12.5^{\circ}$$
 (40% ethanol)

Elementary Analysis		Elementary Analysis	
С	Calc $= 60.19$	С	Calc = 43.20
	Found $= 60.17$		Found = 43.17
Н	Calc = 7.53	Н	Calc = 4.25
	Found = 7.52		Found = 4.24
N	Calc = 15.05	N	Calc = 13.70
	Found = 15.12		Found = 13.75
Overall yield from tyrosine ethyl ester = 18%		C1	Calc = 23.23
CALORI	ne ecuyi escer = 10%	O.I	Found $= 23.31$
		Overall yield from tyrosine ethyl ester = 13%	

N-Formyl-L-tyrosinhydrazide

The synthesis of N-formyl-L-tyrosinhydrazide was similar to the above sequence except that the acylation was carried out in a different manner.

Five one-hundredths mole of L-tyrosine ethyl ester was dissolved in 250 ml. of 80% formic acid and placed in a 500 ml. three neck, round bottom flask equipped with a mechanically sealed electrically driven stirrer. The flask was protected from the atmosphere by a calcium chloride drying tube. 43 ml. (0.5 mole) of acetic anhydride was slowly added to the agitated mixture in the flask. The rate of addition was such that the temperature did not rise above 70°C. One half hour was required for the addition.

The mixture was allowed to stand over night at room temperature and then concentrated to a syrup under vacuum.

The syrup was dissolved in 200 mls. of ethyl acetate and thoroughly washed with 3 N sodium hydroxide solution, then with 6 N HCl and finally with distilled water. The ethyl acetate was removed by evaporation under reduced pressure.

The syrup obtained (60% yield) was then treated with hydrazine in the manner indicated above.

N-Formyl-L-tyrosinhydrazide
M.P. =
$$218^{\circ}$$
C (Decomp.)
[\propto]_b²⁶ = + 21.4 (30% ethanol)

Elementary Analysis Calc =
$$53.75$$
 Calc = 5.95 H Found = 53.70 Found = 6.04

Calc = 18.85 N Found = 18.80

Over all yield from L-Tyrosine Ethyl Ester = 15%

The Synthesis of the Ammonium Salt of N-Acetyl-L-cysteic Acid Carboxylamide

The synthetic route used to obtain this compound is outlined below.

L-Cystine Dimethyl Ester Dihydrochloride (I)

This compound was synthesized from commercially available L-cystine in a manner exactly analogous to the synthesis of L-tyrosine ethyl ester (p. 164). The product was recrystallized from methanol. Yield 89%

N,N' Diacetyl-L-cystine Dimethyl Ester (II)

The free base, cystine dimethyl ester, is an unstable syrupy substance, which slowly decomposes to free sulfur, alamine and other substances. (5) Therefore it is necessary to carry out the acetylation of this compound without first preparing the free base.

One tenth mole of the finely powdered diester dihydrochloride was suspended in two liters of dry tetrahydrofuran contained in a five liter three neck round bottom flask protected from the atmosphere by a drying tube filled with calcium chloride. The suspension was cooled to -5°C. One mole of dry redistilled triethylamine was added. The suspension was maintained for one half hour. Twenty-two hundredths mole of freshly redistilled acetyl chloride was slowly added. The mixture was allowed to stir for four hours.

The triethylamine hydrochloride was removed by filtration and the solvent removed by distillation under reduced pressure. The yield of the crude crystalline material was 71%.

N-Acetyl-L-cysteic Acid Carboxylmethyl Ester (III)

Five-hundredths mole of the diacetyl dimethyl cystine derivative was placed in a 500 ml. three neck flask, and 250 ml. of distilled water containing 60 ml. of concentrated hydrochloric acid was added. The mixture was slowly stirred with a mechanical agitator. After solution was complete, liquid bromine was slowly added. The addition was continued until the bromine color was no longer discharged. (about 0.25 mole required.)

The resulting solution was evaporated under reduced pressure. The product crystallized from the residue in a 75% yield. M.P. = 186 (With Decomposition.)

The Ammonium Salt of N-Acetyl-L-cysteic Acid Carboxylamide (IV)

Four hundredths mole of the crude N-acetyl methyl ester derivative was dissolved in one liter of dry methanol contained in a two liter single neck round bottom blask and cooled to 0°C. The solution was saturated with anhydrous ammonia and while still at 0°C the glass stopper was wired in place. The flask was placed in a heavy walled crock and allowed to stand at room temperature for five days.

The solvent was stripped off at reduced pressure and the product recrystallized from anhydrous ethanol.

M.P. =
$$201-202^{\circ}C$$

[α]_D²⁵ = -9.27° (water)

Elementary Composition C
$$\begin{array}{c} \text{Calc} = 26.39 \\ \text{Found} = 26.35 \end{array}$$
 $\begin{array}{c} \text{Calc} = 5.74 \\ \text{Found} = 26.35 \end{array}$ Found $\begin{array}{c} \text{Calc} = 5.74 \\ \text{Found} = 5.77 \end{array}$

Overall yield from Amino Acid = 18%

Analytical Procedure

In order to get the information necessary to calculate $k_{\mbox{\scriptsize 3}}$ and $K_{\mbox{\scriptsize g}}$ the following general procedure was adopted.

A buffer solution was made up by dissolving enough tris hydroxymethylaminomethane (Tham) to make a 0.2F solution. Concentrated hydrochloric acid was added drop wise to the solution, which was maintained at 25°C until the desired pH was approached. The pH of the stock buffer solution was read on a Beckman pH meter. The stock buffer solution was then diluted ten fold and its pH observed. Final adjustments were made to the stock solution until the diluted solution had the desired pH value.

All the runs were carried out at a buffer concentration of 0.02 F (except where noted) and at a pH of 7.8. The pH optimum curves (Figures 1, 3 and 5) of the three substrates

seem to indicate a pH optimum which is slightly lower than this. However, since the previous work was done at 7.8 it was decided to carry out these investigations at this value also.

The reaction between hydrazine and \underline{p} -dimethylamino benzaldehyde is the basis of the colorimetric procedure used to follow the course of the hydrolysis. (6)

$$H_2N-NH_2$$
 + 2 $\frac{QH}{CH_3}$ + QH_2OH_2 + QH_2OH_3 + QH_3OH_3 + QH_3OH_3 + QH_3OH_3 + QH_3OH_3

To start the procedure of a typical run, eight ml. of the substrate solution, equilibrated to 25°C were pipetted into a 10 ml. volumetric flask. One ml. of the stock 25°C buffer solution was then added. At zero time, one ml. of 25°C enzyme solution was pipetted into the volumetric flask, the pipette allowed to drain for ten seconds, the flask stoppered and inverted 12 times and placed in a $25^{\circ}\pm0.02^{\circ}\text{C}$ constant temperature bath.

At various times aliquot samples were pipetted into a 10 ml. volumetric flask which contained one ml. of 1.60 F HCl

and one ml. of a 10% solution of p-dimethylamino benzaldehyde in 100% ethanol. The solution was then diluted to the calibration mark and the optical density determined at 455 millimicrons with a Beckman model D spectrophotometer. The color samples were allowed to stand about 15 minutes at room temperature before the optical density was determined. Subsequently it was found that it was advantageous that the azine solution should be incubated at 25°C before the optical density was determined.

At the conclusion of each run the pH was checked to be certain that it had remained constant.

The enzyme concentration was one milligram of protein per ml. of solution. The enzyme sample analyzed 14.44% protein nitrogen.

Often it was noted that the enzyme solutions were quite turbid, which was presumably due to the precipitation of the enzyme from the acidic solution. This causes the plot of optical density versus time to intersect the optical density axis rather than pass through the origin (Figure 6). This necessitates that a blank correction be applied to each optical density reading. This blank correction must be obtained for each run by extrapolation of the optical density versus time curve until it intersects the optical density axis. Subsequently it was found that the turbidity could be greatly reduced by adding seven ml. of water to the 10 ml. volumetric flask which contained the standard

acidified p-dimethylaminobenzaldehyde solution. In this way the enzyme solution was pipetted into a solution whose initial acid concentration was lower and precipitation did not occur.

The observed optical densities were corrected by subtraction of the blank values and recorded in Tables 3 to 10 as $D_t(corr.)$. From these values the first order term was calculated. V_0 was estimated both from a first order plot and a zero order plot. The average of the two values was used in the calculation of K_s and k_3 . The final results were obtained by a "least squares" calculation on the data as expressed by equation B.

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Table 2

The Relative Rates Of The X Chymotrypsin Catalyzed Hydrolysis Of N-Trimethylacetyl-L-tyrosinhydra-zide At Different pH Values

Time of Hydrolysis = 122 min.

 $So = 2.56 \times 10^{-3} \text{ molar}$

6.25 0.706 6.23 6.65 1.142 6.63 7.13 1.399 7.12 7.25 7.25	Initial pH	Observed Optical Density	Final pH
7.30 7.40 1.495 7.37 7.75 1.519 7.72 8.14 1.352 8.12 8.30 1.350 8.27 8.42 1.226 8.37 7.36 1.472 7.33 7.50 1.505 7.48 7.63 7.63 1.521 7.62 7.71 1.491 7.84 8.00 1.478 7.96	6.65 7.13 7.30 7.40 7.75 8.14 8.30 8.42 7.50 7.63 7.65 7.71 7.84	1.142 1.399 1.475 1.495 1.519 1.350 1.226 1.472 1.505 1.534 1.521 1.480	6.63 7.12 7.25 7.37 7.72 8.12 8.27 8.37 7.48 7.62 7.62 7.83

Figure 1
pH Optimum Curve for
Trimethylacetyl-L-tyrosinhydrazide
Optical Density vs. pH

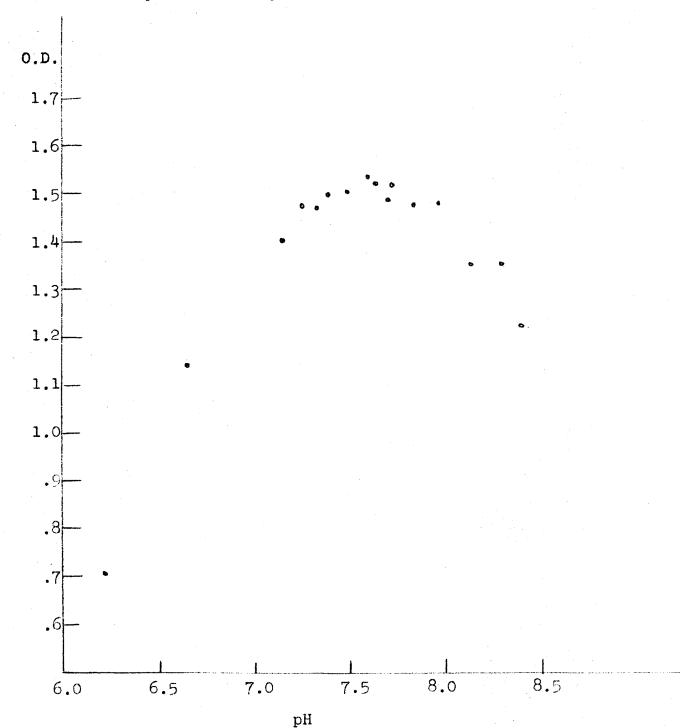


Table 3

The \triangleleft Chymotrypsin Catalyzed Hydrolysis Of N-Trimethylacetyl-L-tyrosinhydrazide

Initial Substrate Concentration = 0.240 Molar

Extent of Hydrolysis = 0.48% Blank = 1.65

Time (Min.)	D _t (corr)*	ln(D∞ -D _t)	$\frac{\ln \frac{D_{\infty}}{D_{\infty} - D_{t}}}{(D_{\infty} - D_{t})}$
2 4 6 8 10 12 14 16	0.33 0.89 1.19 1.65 2.17 2.55 2.99 3.31	6.81872 6.81811 6.81778 6.81729 6.81672 6.81620 6.81577 6.81535	0.00203 0.00264 0.00297 0.00346 0.00403 0.00455 0.00540

 $V_0 = 5.68 \times 10^{-5} \text{ moles.min.}^{-1}$

Initial Substrate Concentration = 0.20 Molar

Extent of Hydrolysis = 0.22% Blank = 0.830

Time (min.)	D _t (corr)*	$\ln(D_{\infty} - D_{t})$	$\frac{\ln \frac{D_{\infty}}{D_{\infty} - D_{t}}}$
2	0.194	6.63724	0.00133
4	0.444	6.63691	0.00166
6	0.860	6.63662	0.00195
8	1.036	6.63612	0.00245
10	1.016	6.63615	0.00242
12	1.322	6.63575	0.00285
14	1.532	6.63547	0.00310
16	1.706	6.63524	0.00330

 $V_{o} = 4.34 \times 10^{-5} \text{ moles min.}^{-1}$

^{*}This column was obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 1.21 x 10⁻¹ Molar

Extent of Hydrolysis = 1.19% Blank = 1.35

Time (Min.)	D _t (corr)*	$\frac{\ln(D_{\infty} - D_{t})}{}$	$\frac{\ln \frac{D \omega}{(D_{\infty} - D_{t})}$
4 8 12 16 20 24 28 32	0.73 1.39 2.11 2.69 3.41 4.09 4.85 5.53	6.12962 6.12818 6.12661 6.12534 6.12376 6.12259 6.12068 6.11911	0.00234 0.00378 0.00535 0.00062 0.00820 0.00937 0.0128
	_5	7	•

 $V_0 = 4.28 \times 10^{-5} \text{ moles} \cdot \text{min}^{-1}$

Initial Substrate Concentration = 3.02×10^{-1} Molar

Extent of Hydrolysis = 1.27% Blank = 0.360

Time (min.)	D _t (corr)*	$\frac{\ln(D_{\infty} - D_{t})}{\ln(D_{\infty} - D_{t})}$	$ \begin{array}{ccc} & & D_{\infty} \\ & & (D_{\infty} & -D_{t}) \end{array} $
2 4 6 8 10 12 14 16	0.105 0.270 0.365 0.565 0.768 0.933 1.113 1.253 1.470	4.74405 4.74261 4.74091 4.73916 4.73739 4.73673 4.73489 4.73401 4.73209	0.00403 0.00537 0.00717 0.00982 0.01169 0.01225 0.01319 0.01407 0.01599
	5	T.	

 $V_0 = 2.18 \times 10^{-5} \text{ moles min.}^{-1}$

^{*}This column was obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 1.048×10^{-2}

Extent of Hydrolysis = 2.10% Blank = 0.033

Time (Min.)	D _t (corr)*	$\frac{\ln(D_{\infty} - D_{t})}{}$	$\frac{\operatorname{In} \operatorname{D}_{\boldsymbol{\omega}}}{(\operatorname{D}_{\boldsymbol{\omega}} - \operatorname{D}_{\operatorname{t}})}$
2½ 4 6 8 10 12 14	0.027 0.217 0.310 0.414 0.514 0.610 0.742 0.840	3.68541 3.68345 3.68083 3.67819 3.67567 3.67304 3.66996 3.66751	0.00421 0.00617 0.00879 0.01143 0.01495 0.01658 0.01966

 $V_0 = 1.38 \times 10^{-5} \text{ moles · min.}^{-1}$

Initial Substrate Concentration = 7.56×10^{-3}

Extent of Hydrolysis = 2.70% Blank = 0.195

III ?	*		ln D∞
Time (min.)	D _t (corr)	$\frac{\ln(D_{\infty} - D_{t})}{}$	(D ∞ -D _t)
4 8 12 16 20 24 28 32 36	0.103 0.248 0.412 0.490 0.654 0.761 0.885 1.006 1.107	5.65533 5.65124 5.6446 5.64134 5.63586 5.63203 5.62758 5.62322 5.61957	0.01057 0.01446 0.02124 0.02436 0.02984 0.03367 0.03812 0.04248 0.04613
V =	= 8.33 x 10 ⁻⁶ n	noles·min1	

^{*}This column was obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 7.49×10^{-3}

Extent of Hydrolysis = 3.80% Blank = 0.114

Time (min.)	D _t (corr)*	$ln(D_{\infty} - D_{t})$	$\frac{\operatorname{D}_{\boldsymbol{\omega}}}{(\operatorname{D}_{\boldsymbol{\omega}}-\operatorname{D}_{\dot{\mathbf{t}}})}$
3 6 9 12 17 20 24 28	0.061 0.163 0.283 0.488 0.581 0.683 0.836 0.996	3.34686 3.34361 3.33905 3.33535 3.32848 3.32472 3.31921 3.31339	0.00614 0.00939 0.01395 0.01763 0.02452 0.02828 0.03379
\mathbb{V}_{\circ}	$= 9.76 \times 10^{-6}$	moles·min1	

Initial Substrate Concentration - 5.99×10^{-3}

Extent of Hydrolysis = 3.67% Blank = 0.172

Time (min.)	D _t (corr)*	ln(D∞ -D _t)	$\frac{\ln \frac{D\omega}{(D_{\infty} - D_{t})}$
2 4 6 8 10 12 14	0.030 0.098 0.150 0.211 0.276 0.309 0.404 0.426	3.12047 3.11745 3.11514 3.11242 3.10954 3.10807 3.08823 3.07921	0.00848 0.01200 0.01431 0.01703 0.01991 0.02138 0.02557
$V_{o} = 7$.41 x 10 ⁻⁶ mc	oles·min1	

^{*}This column was obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 4.49×10^{-3}

Extent of Hydrolysis = 1.87% Blank = 0.150

Time (min.)	D _t (corr)*	ln(D∞ -D _t)	$\frac{\operatorname{D}\omega}{\left(\operatorname{D}\omega-\operatorname{D}_{t}\right)}$
4 8 14 16 20 24 28 32	0.088 0.185 0.340 0.393 0.476 0.560 0.653 0.749	2.82821 2.82233 2.81294 2.80973 2.80569 2.79954 2.79396 2.78815	0.01389 0.02077 0.02916 0.03237 0.03641 0.04256 0.04814
V_{O}	$= 6.23 \times 10^{-6}$	moles·min.	

Initial Substrate Concentration = 2.99×10^{-3}

Extent of Hydrolysis = 4.35% Blank = 0.152

Time (min.)	D _t (corr)*	$\ln(D_{\infty} - D_{t})$	$\frac{\ln D_{\infty}}{(D_{\infty} - D_{t})}$
10 15 20 25 30 35	0.018 0.154 0.203 0.256 0.272 0.365 0.398	1.70992 1.70339 1.69442 1.68649 1.68074 1.66421 1.65794	0.03379 0.04032 0.04929 0.05722 0.06297 0.07950 0.08577
V =	= 2.38 x 10 ⁻⁶ m	noles·minl	

^{*}This column was obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 2.99×10^{-3}

Extent of Hydrolysis = 4.35% Blank 0.152

Time (min.)	D _t (corr)*	ln(D∞ -D _t)	$\frac{1n D_{\bullet \bullet}}{(D_{\bullet \bullet} - D_{t})}$
4 8 10 15 20 24 28 32	0.043 0.114 0.144 0.247 0.316 0.372 0.439 0.512 = 4.12 x 10 ⁻⁶ m	2.41859 2.41235 2.40971 2.40078 2.35330 2.38006 2.37434 2.36812	0.01691 0.02315 0.02579 0.03472 0.05220 0.05544 0.06126 0.6723
V	$= 4.12 \times 10^{-6}$ n	moles·min.	

Initial Substrate Concentration = 1.08×10^{-3}

Extent of Hydrolysis = 7.20% Blank = 0.067

Time (min.)	D _t (corr)*	ln(Dm -Dt)	$\frac{\operatorname{D} \circ \circ}{(\operatorname{D} \circ \circ -\operatorname{D}_{t})}$
4 8 12 16 20 24 28 32	0.038 0.063 0.084 0.113 0.152 0.159 0.174 0.204	1.39158 1.38604 1.38003 1.37270 1.36533 1.36098 1.35196 1.34937	0.02568 0.03117 0.03718 0.04451 0.05188 0.05623 0.06525 0.06784
, O = T		Tep. IIITII.	

^{*}This column was obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 7.49×10^{-4}

Extent of Hydrolysis = 6.15% Blank = 0.107

Time (min.)	D _t (corr)*	$\ln(D_{\infty} - D_{t})$	$ \begin{array}{ccc} & & D_{\infty} \\ & & (D_{\infty} & -D_{\hat{t}}) \end{array} $
5 10 15 20 25 30 35 40	0.023 0.040 0.066 0.081 0.103 0.125 0.145 0.168	1.00404 0.99780 0.98816 0.98257 0.97340 0.96595 0.95831	0.04654 0.5278 0.06242 0.06801 0.07628 0.08463 0.09227 0.10113
V_{o}	$= 1.12 \times 10^{-6} \text{ m}$	oles·minl	

Initial Substrate Concentration = 6.59×10^{-4}

Extent of Hydrolysis = 6.15% Blank = 0.09

Time (min.)	D _t (corr)*	ln(D∞ -D _t)	$ \begin{array}{ccc} & D\infty \\ \hline & (D\infty & -D_{t}) \end{array} $
5 15 15 25 25 35 40	0.021 0.041 0.060 0.074 0.096 0.115 0.136 0.154	0.87705 0.86919 0.86120 0.85527 0.84587 0.83768 0.82855 0.82066	0.04562 0.05348 0.06147 0.06740 0.07680 0.08499 0.09412 0.10201
V	o = 1.02 x 10 ⁻⁶ r	moles·min. ⁻¹	

^{*}This column was obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 5.76×10^{-4}

Extent of Hydrolysis = 7.15% Blank = 0.082

Time (min.)	D _t (corr)*	ln(D∞ -D _t)	$\begin{array}{ccc} & & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & \\ & & & \\ & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & &$
5 10 15 20 25 30 35 40	0.003 0.020 0.038 0.057 0.077 0.094 0.111	0.74953 0.74146 0.73237 0.72368 0.71393 0.70557 0.69350 0.69014	0.03963 0.04770 0.05679 0.06548 0.07523 0.08359 0.09566
77 -	- 8 50 - 10-7 -		• 💉

 $V_0 = 8.50 \times 10^{-7} \text{ moles min.}^{-1}$

Initial Substrate Concentration = 4.91×10^{-4}

Extent of Hydrolysis = 8.55% Blank = 0.083

Time (min.)	D _t (corr)*	ln(D∞ -D _t)	$ \begin{array}{ccc} & D\infty \\ & (D_{\infty} & -D_{t}) \end{array} $
4 12 20 28 36 44 52 60	0.012 0.035 0.052 0.078 0.097 0.119 0.142 0.162	0.57672 0.56588 0.55446 0.54582 0.52886 0.51581 0.50197 0.48980	0.05285 0.06374 0.07521 0.08385 0.10081 0.11386 0.12768 0.13987
V -	7 27 × 10-7 m	olog.min -l	

 $V_0 = 7.27 \times 10^{-7} \text{ moles·min.}^{-1}$

^{*}This column was obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 4.94×10^{-4}

Extent of Hydrolysis = 8.10% Blank = 0.050

Time (Min.)	D _t (corr)*	$\ln(D_{\infty} - D_{t})$	$ \begin{array}{c c} \text{In} & D_{\infty} \\ \hline (D_{\infty} & -D_{t}) \end{array} $
6 12 18 24 30 36 42 48 54	0.011 0.030 0.070 0.067 0.081 0.100 0.114 0.124 0.152	0.60213 0.59167 0.57987 0.57098 0.56531 0.55158 0.53606 0.52176 0.5406	0.03391 0.04437 0.05517 0.06406 0.06973 0.08446 0.09098 0.09797

 $V_0 = 7.12 \times 10^{-7} \text{ moles} \cdot \text{min}^{-1}$

^{*}This column was obtained by subtraction of the blank value from the original data

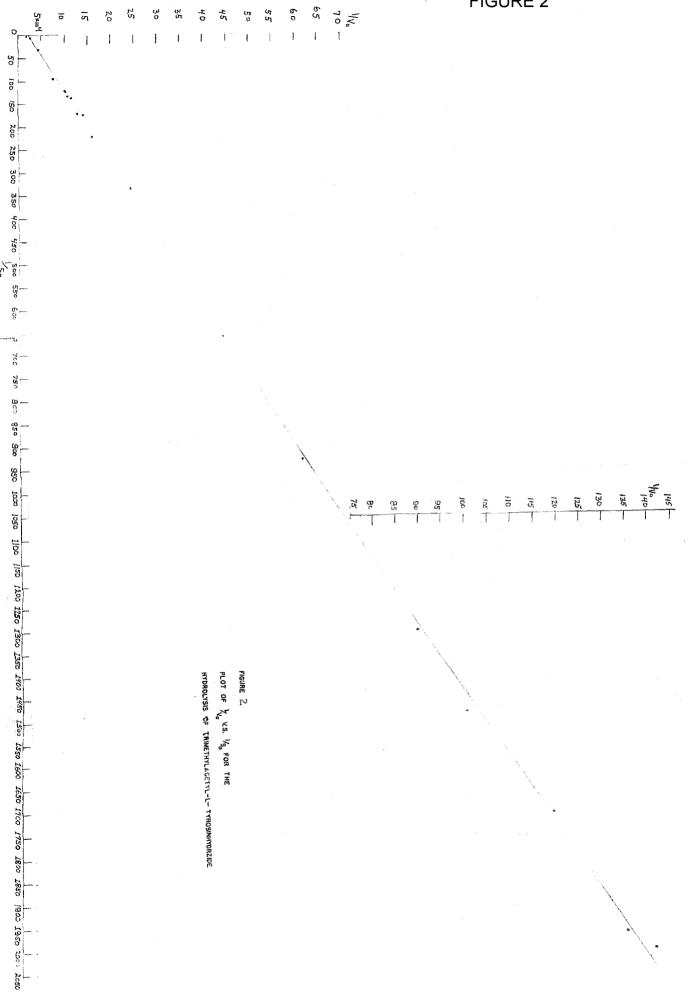


Table 4

The Relative Rates Of The ${\color{red} \boldsymbol{\alpha}}$ Chymotrypsin Catalyzed Hydrolysis Of N-Formyl-L-tyrosinhydrazide At Different pH Values

Time of Hydrolysis = 135 min. [S_0] = 1.2 x 10⁻⁴ molar

Initial pH	Observed Optical Density	Final pH
6.20 6.70 7.00 7.250 7.250 7.250 8.42 8.363 7.45 8.88 7.75 7.760 7.60 7.58	0.252 0.318 0.382 0.411 0.420 0.422 0.360 0.324 0.365 0.322 0.390 0.420 0.434 0.434 0.438 0.438 0.439 0.439 0.429	6.65 6.65 7.47 7.47 7.48 8.45 8.35 8.35 7.66 7.66 7.61 7.55

Figure 3
pH Optimum Curve for
N-Formyl-L-tyrosinhydrazide
Optical Density vs. pH

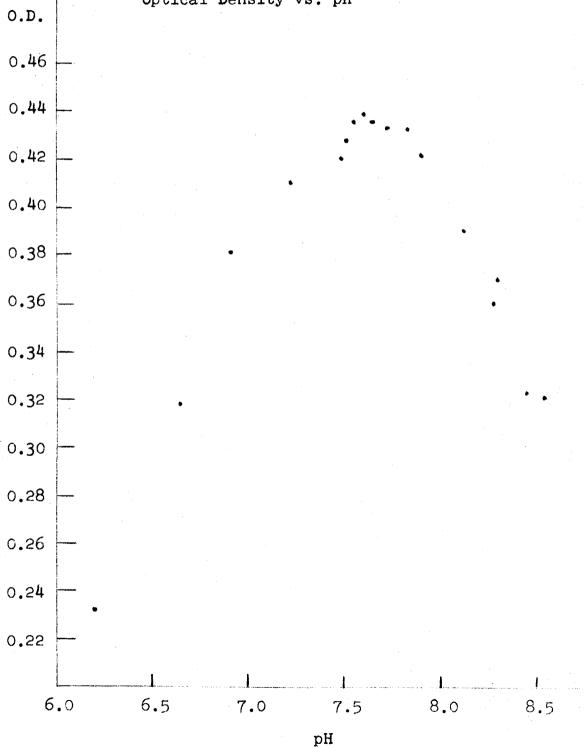


Table 5

The Chymotrypsin Catalyzed Hydrolysis Of N-Formyl-L-tyrosinhydrazide

Initial Substrate Concentration = 13.47×10^{-3} Molar

Extent of Hydrolysis = 1.23% Blank = 0.220

Time (Min.)	D _t (corr)*	ln(D∞ -D _t)	$ \begin{array}{c c} \text{ln} & D\infty \\ \hline & (D\infty - D_t) \end{array} $
4 8 12 16 20 24 28 32 36	0.040 0.112 0.194 0.270 0.334 0.406 0.497 0.561 0.622	6.23830 6.23690 6.23529 6.23380 6.23255 6.23113 6.22933 6.22808 6.22687	0.00501 0.00641 0.00802 0.00957 0.01076 0.01218 0.01398 0.01523 0.01644
	•7	Doing	

 $V_0 = 49.40 \times 10^{-7} \text{ moles · min.}^{-1}$

Initial Substrate Concentration = 11.23×10^{-3} Molar

Extent of Hydrolysis = 1.37% Blank = 0.210

Time (Min.)	D _t (corr)*	ln(D∞ -D _t)	ln Deo (Deo -D _t)
4 8 12 16 20 24 28 32 36	0.029 0.106 0.174 0.241 0.308 0.375 0.439 0.530 0.586	6.05547 6.05367 6.05216 6.05049 6.04890 6.04732 6.04581 6.04365 6.04232	0.00559 0.00741 0.00890 0.01057 0.01216 0.01374 0.01525 0.01741
		7	

 $V_0 = 45.50 \times 10^{-7} \text{ moles min.}^{-1}$

^{*}This column was obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 8.98×10^{-3} molar

Extent of Hydrolysis = 1.46% Blank = 0.200

Time (min.)	D _t (corr)*	$\ln(D_{\infty} - D_{t})$	$ \begin{array}{ccc} & & D & \bullet \\ & & (D \bullet \bullet & -D_t) \end{array} $
4 8 12 16 20 24 28 32 36	0.030 0.085 0.141 0.216 0.278 0.324 0.375 0.440 0.502	3.52862 3.52701 3.52536 3.52316 3.52132 3.51996 3.51845 3.51652	0.00668 0.00829 0.00994 0.01215 0.01398 0.01534 0.01685 0.01878
V	$= 38.90 \times 10^{-7}$	moles.min -1	

 $V_0 = 38.90 \times 10$ ' moles min.

Initial Substrate Concentration = 8.00×10^{-3} Molar

Extent of Hydrolysis = 1.52% Blank = 0.155

Time (min.)	Dt(corr)*	$\ln(D_{\bullet} - D_{t})$	$ \begin{array}{c c} \text{In} & D & \bullet \\ & (D & \bullet & -D_{t}) \end{array} $
4	0.026	3.41375	0.00581
8	0.075	3.41214	0.00742
12	0.143	3.40939	0.00967
16	0.195	3.40517	0.01139
20	0.247	3.40695	0.01311
24	0.312	3.40151	0.01526
28	0.371	3.40131	0.01725
32	0.427	3.40054	0.01902

 $V_0 = 37.82 \times 10^{-7} \text{ moles·min.}^{-1}$

^{*}This column was obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 6.74×10^{-3} Molar

Extent of Hydrolysis = 1.66% Blank = 0.173

Time (Min.) D _t (co	rr)* ln(D ∞ -	D_{t}) $\frac{D_{\infty}}{D_{\infty}-D_{t}}$
4 8 12 16 20 24 28 36 36	0.027 0.077 0.126 0.178 0.227 0.275 0.337 0.386 0.428	3.23985 3.23789 3.23557 3.23392 3.23175 3.22969 3.22771 3.22577	0.00976 0.01208 0.01373 0.01590 0.01796 0.01994 0.02188
	$V_0 = 33.74 x$	10^{-7} moles min.	

Initial Substrate Concentration = 5.56×10^{-3} Molar

Extent of Hydrolysis = 1.72% Blank = 0.120

Time (min.)	D _t (corr)*	$ln(D_{\infty} - D_{t})$	$ \begin{array}{c c} \text{ln} & D^{\infty} \\ \hline (D_{\infty} & -D_{t}) \end{array} $
4 8 12 16 20 24 28 32	0.068 0.093 0.142 0.189 0.231 0.279 0.317 0.370	3.05055 3.04937 3.04704 3.04481 3.04280 3.04051 3.03869 3.03572	0.00886 0.01003 0.01236 0.01459 0.01660 0.01889 0.02071 0.02368
V	$= 30.05 \times 10^{-7}$	moles·min1	

 * This column was obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 4.63×10^{-3} Molar

Extent of Hydrolysis = 1.9% Blank = 0.111

Time (Min.)	D _t (corr)*	ln(D∞ -D _t)	$\frac{\operatorname{D} \infty}{(\operatorname{D} \infty - \operatorname{D}_{t})}$
4	0.048	2.86340	0.00982
8	0.074	2.86271	0.01051
12	0.136	2.85916	0.01406
16	0.170	2.85721	0.01601
20	0.203	2.85532	0.01795
24	0.249	2.85233	0.02089
28	0.269	2.85152	0.02175
32	0.326	2.84822	0.02505

 $V_0 = 26.78 \times 10^{-7} \text{ moles·min.}^{-1}$

Initial Substrate Concentration = 3.90×10^{-3} Molar

Extent of Hydrolysis = 2.66% Blank = 0.150

Time (Min.)	D _t (corr)*	ln(D∞ -D _t)	ln Dœ (Dæ -Dt)
4 8 12 16 20 24 28 32 36	0.040 0.077 0.110 0.159 0.192 0.236 0.265 0.308 0.372	2.68847 2.68590 2.68233 2.68115 2.67856 2.67509 2.67297 2.67000 2.66556	0.01276 0.01533 0.01840 0.02005 0.02267 0.02614 0.02826 0.03123 0.03567
* 7	00 0 70-1		

 $V_0 = 23.3 \times 10^{-7} \text{ moles.min.}^{-1}$

^{*}This column was obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 3.71×10^{-3} Molar

Extent of Hydrolysis = 2.37% Blank = 0.120

Time (Min.)	D _t (corr)*	ln(D ∞ -D _t)	$ \begin{array}{ccc} & D_{\infty} \\ & (D_{\infty} - D_{t}) \end{array} $
4 8 12 16 20 24 28 30 38	0.048 0.066 0.112 0.148 0.185 0.218 0.242 0.279	2.63813 2.63684 2.63354 2.63095 2.62828 2.62590 2.62416 2.62147 2.61645	0.001193 0.01322 0.01652 0.01901 0.02178 0.02416 0.02590 0.02859
V	$_{0} = 23.55 \times 10^{-7}$	moles·min1	

Initial Substrate Concentration = 2.08×10^{-3} Molar

Extent of Hydrolysis = 2.43% Blank = 0.130

Time (Min.)	D _t (corr)*	$\frac{\ln(D_{\infty} - D_{t})}{}$	$ \begin{array}{c c} \ln & D & \\ \hline & (D & -D_{t}) \end{array} $
4	0.029	2.05386	0.02019
8	0.038	2.05271	0.02134
12	0.062	2.04953	0.02414
16	0.086	2.04653	0.02752
20	0.104	2.04440	0.02965
24	0.127	2.04122	0.03283
28	0.153	2.03780	0.03625
32	0.171	2.03680	0.03725
36	0.192	2.03275	0.04130

 $V_0 = 23.40 \times 10^{-7} \text{ moles min.}^{-1}$

^{*}This column was obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = $0.926 \times 10^{-3} \text{ Molar}$

Extent of Hydrolysis = 3.27% Blank = 0.110

Time (Min.)	D _t (corr)*	ln(D ∞ -D _t)	$ \begin{array}{ccc} & D_{\infty} \\ \hline & (D_{\infty} - D_{t}) \end{array} $
5 10 15 20 25 30 45 50	0.017 0.015 0.027 0.024 0.060 0.046 0.090 0.100 0.113	1.22730 1.22788 1.22434 1.22515 1.21461 1.21877 1.20567 1.20267	0.03655 0.03597 0.03948 0.03870 0.04925 0.04508 0.05718 0.06118 0.06509
7.7	- 6 FF 30-(

 $V_o = 6.55 \times 10^{-7} \text{ moles min.}^{-1}$

^{*}This column was obtained by subtraction of the blank value from the original data

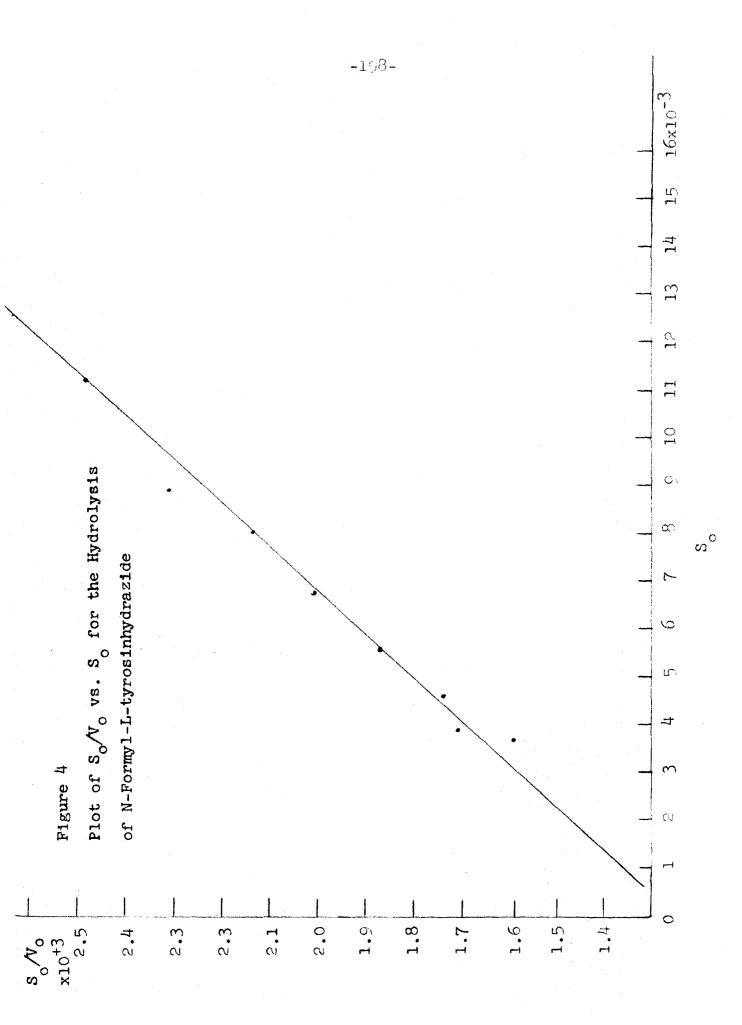


Table 6

The Relative Rates Of The ${\color{red} \alpha}$ Chymotrypsin Catalyzed Hydrolysis Of N-Dichloroacetyl-L-tyrosinhydrazide At Different pH Values

Time of Hydrolysis = 40 min. [S_{\bullet}] = $6 \times 10^{-4} \text{ molar}$

Initial pH	Observed Optical Density	<u>Final pH</u>
6.20 6.70 7.09 7.25 7.83 8.32 8.40 8.40 7.76 7.65 7.65 7.65 7.40 7.30	0.370 0.571 0.702 0.852 0.869 0.685 0.750 0.660 0.833 0.883 0.887 0.895 0.894 0.905 0.909 0.907 0.892 0.875	6.68 6.69 6.69 6.69 6.69 7.78 6.69 7.78 6.69 7.78 6.69 7.79 7.99 7.99 7.99 7.99 7.99 7.99 7.99 7.99 7.99 7.99 7.99

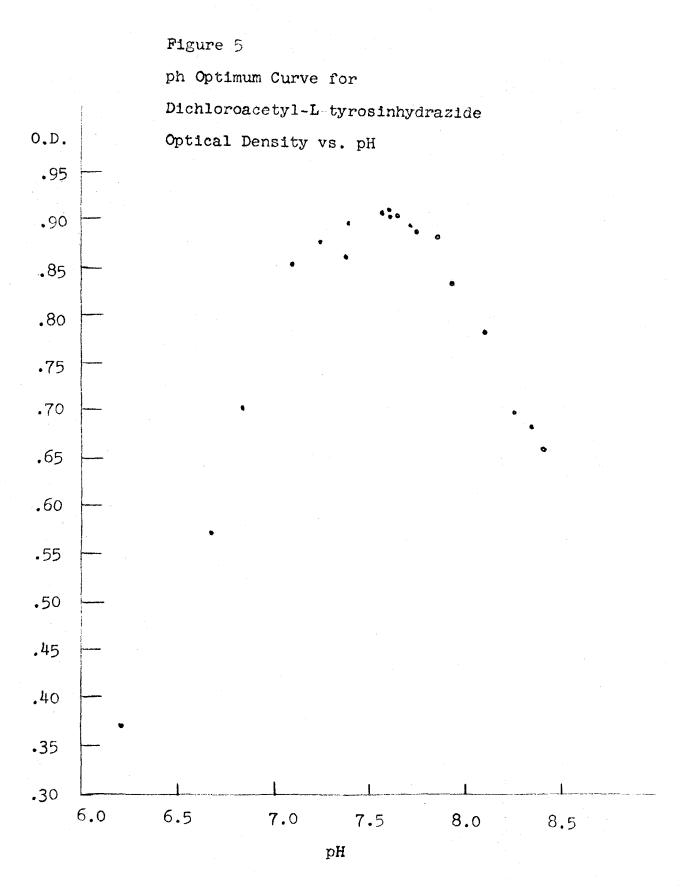


Table 7

The Kinetics of the Chymotrypsin Catalyzed Hydrolysis of N-Dichloroacetyl-L-tyrosinhydrazide

Initial Substrate Concentration = 3.82×10^{-3} Molar

Extent of Hydrolysis = 9.96% Blank = 0.150

<u>t</u>	D _t (corr)*	ln Doo -Dt	fo
2 3 4 5 6 7 9	0.298 0.458 0.580 0.758 0.905 1.032 1.338 1.454	0.01952 0.03113 0.4227 0.05285 0.06344 0.07401 0.09482 0.10499	0.9893 0.9854 0.9790 0.9739 0.9689 0.9637 0.9542
		_	

 $V_0 = 4.20 \times 10^{-5} \text{ moles min.}^{-1}$

Initial Substrate Concentration = 3.26×10^{-3} Molar

Extent of Hydrolysis = 11.52% Blank = 0.120

<u>t</u>	D _t (corr)*	$\frac{\ln \frac{D_{\infty}}{D_{\infty} - D_{\mathbf{t}}}}{\frac{D_{\infty}}{D_{\infty}} - \frac{D_{\mathbf{t}}}{D_{\mathbf{t}}}}$	f _o
2 3 5 6 7 8 9 10	0.298 0.440 0.730 0.872 1.039 1.170 1.300 1.435	0.02562 0.03748 0.06216 0.07448 0.08915 0.10083 0.11254 0.12584	0.9879 0.9865 0.9753 0.9644 0.9576 0.9522 0.9468

 $V_0 = 4.04 \times 10^{-5} \text{ moles · min.}^{-1}$

^{*}The data in this column were obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 3.234×10^{-3} Molar

Extent of Hydrolysis = 10.80% Blank = 0.125

t	D _t (corr)*	$\frac{\ln \frac{D_{co}}{D_{co} - D_{t}}}{\frac{D_{co}}{D_{co}} - D_{t}}$	fo
2345678910	0.275 0.421 0.545 0.708 0.819 0.933 1.103 1.197	0.02400 0.03410 0.04566 0.05728 0.06960 0.08013 0.09163 0.10303 0.11422	0.9886 0.9833 0.9773 0.9718 0.9659 0.9607 0.9554 0.9502
	$V_{\rm c} = 3.84 \times 10^{-5}$	moles•min1	

Initial Substrate Concentration = 2.855×10^{-3} Molar

Extent of Hydrolysis = 12.24% Blank = 130

<u>t</u>	D _t (corr)*	$\frac{\ln \frac{D_{\infty}}{D_{\infty} - D_{t}}}{\frac{D_{\infty}}{D_{t}}}$	$\frac{\mathbf{f_o}}{\mathbf{o}}$
2 3 4 5 6 7 8 1/4 9 1/4 10 1/2	0.268 0.398 0.510 0.602 0.770 0.902 1.055 1.192 1.339	0.03874 0.05112 0.06812 0.07038 0.08621 0.10949 0.11544 0.12976 0.14522	0.9876 0.9815 0.9762 0.9701 0.9661 0.9579 0.9508 0.9396
77	2 kg - 10-5 mg	Jogamin -1	

 $V_0 = 3.47 \times 10^{-9} \text{ moles · min.}^{-1}$

^{*}The data in this column were obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 2.45×10^{-3} Molar

Extent of Hydrolysis = 11.13% Blank = 0.100

<u>t</u> .	D _t (corr)*	$\frac{\ln \frac{D_{\infty}}{D_{\infty} - D_{\mathbf{t}}}}{\frac{1}{2}}$	f _o
3 L	0.362 0.471	0.04000 0.05098	0.9801 0.9747
5	0.574 0.730	0.07341	0.9640 0.9588
7 8	0.830 . 0.940	0.09577 0.10802	0.9536 0.9478
9	1.062	0.11790	0.9433

 $V_0 = 3.39 \times 10^{-5} \text{ moles · min.}^{-1}$

Initial Substrate Concentration = 2.31×10^{-3} Molar

Extent of Hydrolysis = 11.31% Blank = 0.130

<u>t</u>	D _t (corr)*	$\frac{\ln \frac{D \infty}{D_{\infty} - D_{\mathbf{t}}}}{\ln \frac{D \infty}{D_{\infty}} + \frac{D \infty}{D_{\infty}}}$	f _o
2 3 4 5 6 7 8 9	0.229 0.286 0.396 0.515 0.608 0.702 0.805 0.925 0.971	0.02351 0.03531 0.04711 0.05868 0.07078 0.08295 0.09514 0.10761 0.12000	0.9882 0.9827 0.9765 0.9711 0.9602 0.9595 0.9538 0.9480

 $v_0 = 2.86 \times 10^{-5} \text{ moles · min.}^{-1}$

^{*}The data in this column were obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 1.85×10^{-3} Molar

Extent of Hydrolysis = 12.54% Blank = 0.062

t	D _t (corr)*	ln D∞ D∞ -D _t	fo
2 34 56 7 8 90	0.183 0.288 0.349 0.469 0.603 0.624 0.719 0.787 0.888	0.025134 0.03922 0.05309 0.06697 0.08068 0.09409 0.10760 0.12087 0.13402	0.9810 0.9809 0.9737 0.9672 0.9605 0.9543 0.9419 0.9359
	W = 0.54 × 10 ⁻⁵ mo	log.min -l	

 $V_0 = 2.54 \times 10^{-7} \text{ moles·min.}$

Initial Substrate Concentration = 1.50×10^{-3} Molar

Extent of Hydrolysis = 12.16% Blank = 0.130

t	D _t (corr)*	ln D∞ D∞ -Dt	fo
2 3 4 5 6 7 8 9 10	0.160 0.220 0.280 0.360 0.435 0.502 0.565 0.608 0.697	0.05187 0.06285 0.07040 0.08931 0.10176 0.11675 0.14717 0.15674 0.17372	0.9861 0.9809 0.9851 0.9682 0.9614 0.9558 0.9489 0.9459
	r	-	

 $V_0 = 2.23 \times 10^{-5} \text{ moles·min.}^{-1}$

^{*}This column was obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 9.24 x 10-4 Molar

Extent of Hydrolysis = 12.97% Blank = 0.095

<u>t</u>	D _t (corr)*	$\frac{\ln \frac{D_{\infty}}{D_{\infty} - D_{t}}}{\frac{D_{\infty}}{D_{t}}}$	<u>f</u> o
23456789	0.073 0.175 0.198 0.214 0.327 0.348 0.409 0.458	0.04876 0.07041 0.08666 0.09472 0.12732 0.13410 0.15406 0.17039	0.9892 0.9792 0.9715 0.9689 0.9530 0.9499 0.9395

 $V_0 = 1.45 \times 10^{-5} \text{ moles · min.}^{-1}$

Initial Substrate Concentration = 9.09 x 10-4 Molar

Extent of Hydrolysis = 15.016% Blank = 0.100

t	D _t (corr)*	$\frac{\ln \frac{D_{\infty}}{D_{\infty} - D_{t}}}{\frac{1}{2}}$	<u>f</u> o
2	0.134	0.07611	0.9831
3	0.148	0.08044	0.9783
3 4	0.202	0.09733	0.9704
5	0.259	0.11548	0.9620
6	0.318	0.13461	0.9558
7	0.356	0.14713	0.9477
8	0.405	0.16351	0.9405
9	0.456	0.18085	0.9327
10	0.523	0.23982	0.9228

 $V_0 = 1.54 \times 10^{-5} \text{ moles min.}^{-1}$

^{*}The data in this column were obtained by subtraction of the blank value from the original data

Table 7 Continued

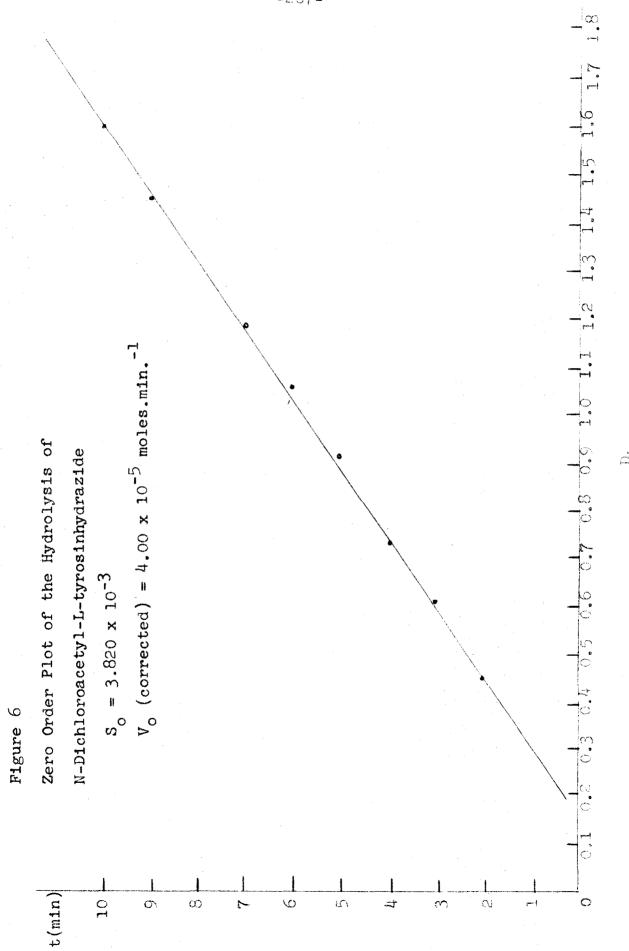
Initial Substrate Concentration = 3.02×10^{-4} Molar

Extent of Hydrolysis = 16.03% Blank = 0.085

<u>t</u>	D _t (corr)*	ln Dec -Dt	fo
2	0.039	0.10369	0.9831
3	0.071	0.13989	0.9689
3 4	c.076	0.14991	0.9666
5	0.095	0.16891	0.9581
5	0.115	0.18991	0.9489
7	0.143	0.21991	0.9367
8	0.149	0.22491	0.9339
9	0.167	0.24591	0.9257
10	0.185	0.26691	0.9129

 $V_0 = 5.83 \times 10^{-6} \text{ moles · min.}^{-1}$

^{*}The data in this column were obtained by subtraction of the blank value from the original data



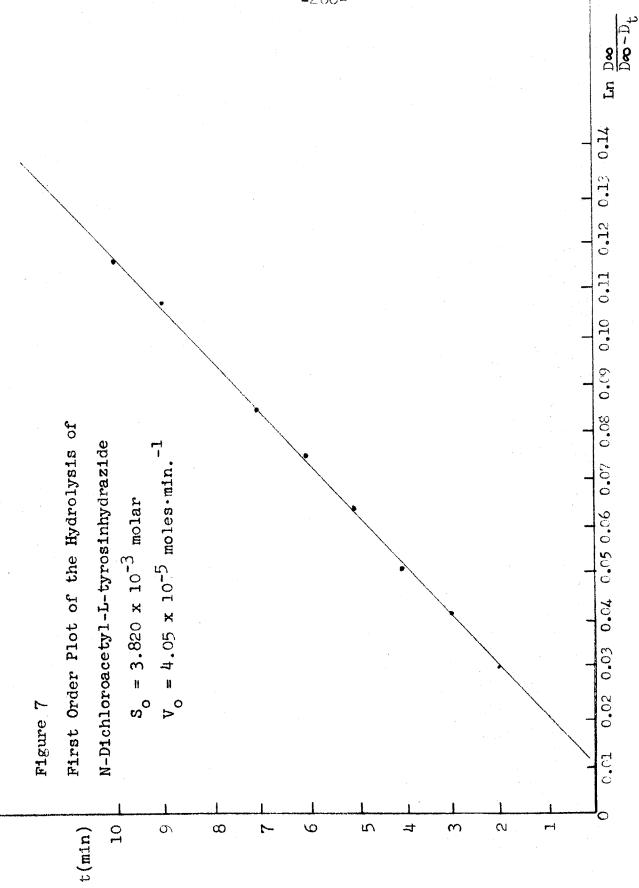
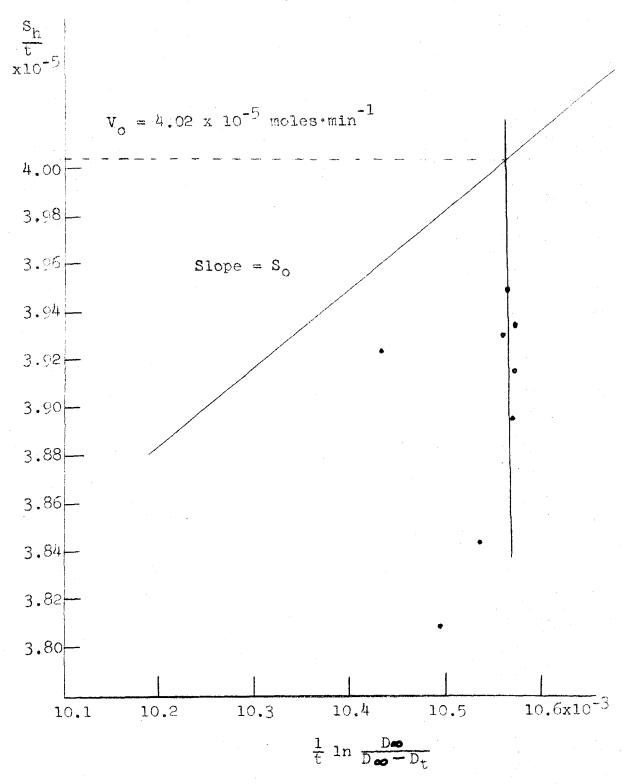


Figure 8

Plot According to Integrated Equation - D



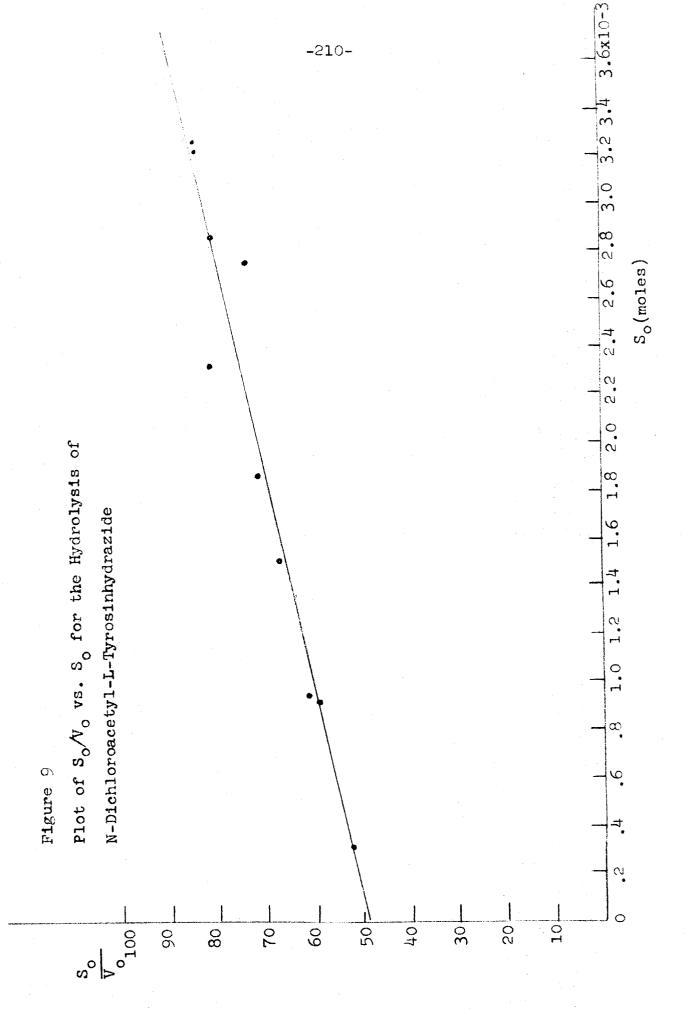


Table 8

$\frac{\text{The } \bowtie \text{Chymotrypsin Catalyzed Hydrolysis Of The Ammonium Salt}}{\text{Of N-Acetyl-L-cysteic Acid Carboxylamide}}$

Substrate Concentration = 200 u moles·milliliter $^{-1}$

Normality of Base used in titration * = 0.10 N

Milliliters of Base equivalent to total substrate in titration = 2.0

Initial ph = 6.20 Final pH = 6.18 T=0	_01ba	m_81hr	Ш_00ha	T-07 ¹ hn
End Point(ml.) $\frac{1=0}{0.339}$	0.340	$\frac{T=8\frac{1}{2}hr.}{0.350}$	0.350	0.350
Initial pH = 6.85 Final pH = 6.79	M01 bm	m_01bm	-00hr	M-071ha
End Point(ml.) $\frac{T=0}{0.398}$	$\frac{1=2\pi r}{0.414}$	$\frac{T=8\frac{1}{2}hr.}{0.420}$	0.425	0.420
Initial pH = 7.25 Final pH = 7.20	∏_0 <u>1</u> ba	T-81hr		∏-27 1 hn
End Point(ml.) $\frac{T=0}{0.255}$	0.200	$\frac{T=8\frac{1}{2}hr}{0.385}$		0.390
Initial pH = 7.32 Final pH = 7.30 T=0	$T=2\frac{1}{2}hr$.	$\frac{T=8\frac{1}{2}hr}{0.400}$	T=22nr.	$T=27\frac{1}{2}hr$.
End Point(ml.) $\overline{0.390}$	0.400	0.400	0.420	0.420
Initial pH = 7.95 Final pH = 7.94 T=0	Т=2 1 hr	Т <u>=8±</u> hr	Т=22hr	$T=27\frac{1}{5}hr$
End Point(m1.) $\frac{100}{0.320}$	0.335	$\frac{T=8\frac{1}{2}hr}{0.345}$	0.370	0.380
Initial pH = 8.30 Final pH = 8.25 T=0	T=2½hr.	T=8=hr.	T=22hr.	$T=27\frac{1}{2}hr$.
End Point(ml.) $\frac{1}{0.290}$	0.310	$\frac{T=8\frac{1}{2}hr.}{0.324}$	0.350	0.355

H. T. Huang and Carl Niemann - J.A.C.S. 73 1541 (1951)

Table 9

The Kinetics Of \upomega Chymotrypsin Catalyzed Hydrolysis Of N-Acetyl-L-tyrosinhydrazide In The Presence of N-Acetyl-L-cysteic Acid-carboxylamide As An Inhibitor

(Tham Concentration = 0.5F for this set of runs)

Initial Substrate Concentration = 6.67×10^{-3} Molar

Blank = 0.100 Extent of Hydrolysis = 6.23% Concentration of Inhibitor = 0.000 Molar

<u>t</u>	D _t (corr)*	ln(D∞ -D _t)	$ \begin{array}{c c} \operatorname{In} & \operatorname{D} \infty \\ \hline & (\operatorname{D} \infty & -\operatorname{D}_{t}) \end{array} $
2 4 6 8 10 12 14 16	0.212 0.380 0.568 0.728 0.899 1.095 1.258 1.428 1.582	25.162 24.994 24.806 24.646 24.474 24.279 24.116 23.946 23.784	0.01247 0.01916 0.02672 0.03317 0.04015 0.04819 0.05493 0.06392 0.07232
7.7	- 0 Hz = 20-5 ma	-1	

 $V_0 = 2.41 \times 10^{-5} \text{ moles · min.}^{-1}$

Initial Substrate Concentration = 6.67×10^{-3} Molar

Blank = 0.100 Extent of Hydrolysis = 7.04% Concentration of Inhibitor = 10.26×10^{-3} Molar

<u>t</u>	D _t (corr)*	ln(D -D _t)	$\frac{\ln \left(D - D_{t}\right)}{\left(D - D_{t}\right)}$
2 4 6 8 10 12 14 ¹ / ₄ 16	0.209 0.390 0.618 0.809 0.906 1.172 1.382 1.560	3.22515 3.21844 3.20426 3.20165 3.19405 3.17835 3.17755 3.17049 3.16248	0.00122 0.01941 0.02859 0.03820 0.04350 0.05951 0.05993 0.06736 0.07535
77	0 (0 = 30=5	a	

 $V_{o} = 2.63 \times 10^{-5} \text{ moles} \cdot \text{min.}^{-1}$

^{*}This column obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 4.23×10^{-3} Molar

Blank = 0.090 Extent of Hydrolysis = 7.40% Concentration of Inhibitor = 0.000 Molar

<u>t</u>	D _t (corr)*	ln(D∞ -D _t)	$\frac{\ln D\infty}{(D\infty - D_t)}$
2 4 6 8 10 12 14 16 18	0.153 0.280 0.440 0.550 0.669 0.810 0.916 1.086	2.76642 2.75931 2.74912 2.74206 2.73437 2.72516 2.71819 2.70689 2.69948	0.01605 0.02316 0.03315 0.04041 0.04810 0.05731 0.06428 0.07558

 $V_0 = 1.79 \times 10^{-5} \text{ moles.min.}^{-1}$

Initial Substrate Concentration = 4.23×10^{-3} Molar

Blank = 0.090Extent of Hydrolysis = 7.50%Concentration of Inhibitor = 10.26×10^{-3} Molar

<u>t</u>	D _t (corr)*	ln(D w -D _t)	$ \begin{array}{ccc} & & D\infty \\ & & (D\infty & -D_{t}) \end{array} $
2	0.148	2.76764	0.01553
4	0.395	2.75200	0.03047
6	0.415	2.75072	0.03175
8	0.535	2.74337	0.03910
12	0.830	2.72485	0.05762
14	0.930	2.71727	0.06520
16	1.064	2.70838	0.07409
18	1.220	2.69793	0.08454

 $V_0 = 1.76 \times 10^{-5} \text{ moles·min.}^{-1}$

^{*}This column obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 2.53×10^{-3} Molar

Blank = 0.105 Extent of Hydrolysis = 6.75% Concentration of Inhibitor = 0.000 Molar

. <u>t</u>	D _t (corr)*	ln(Do -Dt)	$ \begin{array}{ccc} \ln & D\infty \\ & (D\infty & -D_{t}) \end{array} $
2 4 6 8 10 12 14 16	0.075 0.128 0.237 0.396 0.375 0.468 0.530 0.582 0.650	2.24918 2.24411 2.23248 2.22613 2.21758 2.20629 2.20055 2.19477 2.18718	0.01931 0.02438 0.03601 0.04236 0.05091 0.06220 0.06794 0.07372

 $V_0 = 1.05 \times 10^{-5} \text{ moles.min.}^{-1}$

Initial Substrate Concentration = 2.53×10^{-3} Molar

Blank = 0.075Extent of Hydrolysis = 8.94%Concentration of Inhibitor = 10.26×10^{-3} Molar

<u>t</u>	D _t (corr)*	ln(D∞ -D _t)	$ \begin{array}{ccc} & & & & & & & \\ & & & & & & & \\ & & & & $
2	0.119	2.24887	0.01962
4	0.161	2.24379	0.02470
6	0.280	2.23109	0.03740
8	0.363	2.22215	0.04635
10	0.416	2.21638	0.05211
12	0.519	2.20508	0.06341
14	0.601	2.19600	0.07249
16	0.675	2.18774	0.08075
18 ¹ / ₂	0.777	2.17623	0.09226

 $V_0 = 1.16 \times 10^{-5} \text{ moles.min.}^{-1}$

^{*}This column obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = $1.69 \times 10^{-3} \text{ Molar}$

Blank = 0.105 Extent of Hydrolysis = 7.13% Concentration of Inhibitor = 0.000 Molar

t	D _t (corr)*	$\ln(D\boldsymbol{\infty} - D_t)$	$ \begin{array}{ccc} & D \infty \\ & (D \infty - D_{t}) \end{array} $
2 4 6 8 10 12 14 16 18	0.048 0.097 0.155 0.204 0.245 0.304 0.317 0.376 0.461 $V_{o} = 6.85 \times 10^{-6} \text{ mo}$	1.84130 1.83322 1.82391 1.81596 1.81091 1.79956 1.79741 1.78759 1.77326	0.02429 0.03210 0.04140 0.04936 0.05441 0.06576 0.06791 0.06873 0.08306

Initial Substrate Concentration = 1.69×10^{-3} Molar

Blank = 0.100 Extent of Hydrolysis = 6.4% Concentration of Inhibitor = 10.26×10^{-3} Molar

t	D _t (co	rr)* ln(D∞	$-D_{t}$) $\frac{\ln D \omega}{(D \omega - D_{t})}$
2 4 6 8 10 12 14 16 18	0.04 0.09 0.13 0.17 0.24 0.27 0.31 0.38 0.41	4 1.834 8 1.827 5 1.821 3 1.810 5 1.805 3 1.798 6 1.786	150 0.03082 747 0.03785 148 0.04384 042 0.05490 016 0.06016 088 0.06644 0.07857
	V - 6 08 v	10-6 mologimin -	-1

 $V_0 = 6.08 \times 10^{-6} \text{ moles·min.}^{-1}$

^{*}This column obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 1.27×10^{-3} Molar

Blank = 0.080 Extent of Hydrolysis = 7.41% Concentration of Inhibitor = 0.000

t	D _t (corr)*	ln(D∞ -D _t)	$ \begin{array}{c c} \text{ln} & D\infty \\ \hline & (D\infty - D_{t}) \end{array} $
2 4 6 8 10 12 14 16 18	0.058 0.078 0.120 0.173 0.193 0.258 0.298 0.305 0.358	1.55033 1.54607 1.53708 1.52562 1.52176 1.50696 1.49806 1.49649	0.02886 0.03312 0.04211 0.05357 0.05793 0.07223 0.08113 0.08270 0.09464

 $V_0 = 5.25 \times 10^{-6} \text{ moles·min.}^{-1}$

Initial Substrate Concentration = 1.27×10^{-3} Molar

Blank = 0.110 Extent of Hydrolysis = 7.58% Concentration of Inhibitor = 10.26×10^{-3} Molar

-	D _t (corr)*	ln(D∞ -D _t)	ln <u>D</u> ∞ (D∞ -D _t)
2 4 6 8 10 12 14 16	0.033 0.083 0.145 0.178 0.212 0.253 0.272 0.325 0.368	1.54773 1.53704 1.52401 1.51798 1.51050 1.50170 1.49717 1.48523 1.47545	0.03154 0.04223 0.05526 0.06129 0.06877 0.7757 0.08210 0.09404 0.10382

 $V_0 = 5.61 \times 10^{-6} \text{ moles.min.}^{-1}$

^{*}This column obtained by subtraction of the blank value from the original data

Table 10

The Kinetics Of Chymotrypsin Catalyzed Hydrolysis Of N-Acetyl-L-tyrosinhydrazide In The Presence Of Added Sodium Sulfate

(Tham Concentration = 0.82F for this set of runs)

Initial Substrate Concentration = 1.82×10^{-3} Molar

Blank = 0.250 Concentration of Sodium Sulfate = 0.000

<u>t</u> .	D _t (corr)*	ln(D -D _t)	$\frac{\ln D}{(D - D_{t})}$
2 1/15 4 6 8 10 12 14 16 18	0.165 0.325 0.508 0.711 0.898 1.082 1.227 1.420 1.600	1.92161 1.89295 1.86191 1.82986 1.79940 1.76849 1.74344 1.70729	0.01627 0.04493 0.07597 0.10802 0.13848 0.16939 0.19449 0.22336 0.23069
	ļ		

 $V_0 = 2.28 \times 10^{-5} \text{ moles · min.}^{-1}$

Initial Substrate Concentration = 1.82×10^{-3} Molar

Blank = 0.190 Concentration of Sodium Sulfate = 0.000

<u>t</u>	D _t (corr)*	In(D -D _t)	$\frac{\ln \frac{D}{(D - D_{t})}$
2 4 6 8 10 12 14 ¹ / ₄ 16	0.170 0.383 0.559 0.774 0.955 1.130 1.324 1.463 1.632	1.91513 1.88114 1.85395 1.81970 1.78983 1.76027 1.72633 1.70129 1.66997	0.02275 0.05674 0.08393 0.11918 0.14805 0.17761 0.21155 0.23659 0.26791

 $V_0 = 2.23 \times 10^{-5} \text{ moles · min.}^{-1}$

^{*}This column obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 1.82×10^{-3} Molar

Blank = 0.171 Concentration of Sodium Sulfate = 1×10^{-3} Molar

<u>t</u>	D _t (corr)*	ln(D∞ -D _t)	$\frac{1 \text{n} D \cdot \infty}{(D \cdot \infty - D_{t})}$
2 4 6 8 10 12 14 16 18	0.179 0.393 0.594 0.776 1.012 1.168 1.217 1.482 1.565 $V_0 = 2.52 \times 10^{-5} \text{ mg}$	1.83780 1.80105 1.76730 1.73572 1.69323 1.66412 1.65479 1.60282 1.56716	0.10210 0.13683 0.17058 0.20216 0.24465 0.27376 0.28309 0.33506 0.37072

Initial Substrate Concentration = 1.82×10^{-3} Molar

Blank = 0.190 Concentration of Sodium Sulfate = 5×10^{-3} Molar

<u>t</u>	D,	(corr)*	ln(D ∞	-D _t)	ln (D ∞ D ∞ -D _t)
2 4 6 8 10 12 1 14 16 18	C C C 1 1	0.169 0.371 0.582 0.793 0.971 0.169 0.311 0.421	1.913 1.882 1.850 1.816 1.787 1.753 1.728 1.708	97 34 61 25 54 64 92	0. 0. 0. 0.	02464 05491 08754 12127 15063 18434 20924 22896 26626

 $V_0 = 2.56 \times 10^{-5} \text{ moles.min.}^{-1}$

^{*}This column obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 1.82×10^{-3} Molar

Blank = 0.200 Concentration of Sodium Sulfate = 10×10^{-3} Molar

t	D _t (corr)*	ln(D ~ -D _t)	$ \begin{array}{c c} \text{ln} & D \infty \\ \hline (D \infty - D_{C}) \end{array} $
2 4 6 8 10 12 14 16	0.182 0.374 0.582 0.778 0.943 1.112 1.312 1.494	1.91132 1.88251 1.85034 1.80270 1.79193 1.76336 1.72846 1.69562	0.02656 0.05537 0.08754 0.13518 0.14595 0.17452 0.20942 0.24226 0.26585

 $V_0 = 2.43 \times 10^{-3} \text{ moles · min.}^{-1}$

Initial Substrate Concentration = 1.82×10^{-3} Molar

Blank = 0.170 Concentration of Sodium Sulfate = 40×10^{-3} Molar

<u>t</u>	D _t (corr)*	ln(D ∞ -D _t)	$\frac{\operatorname{ln} \operatorname{D}^{\bullet \bullet}}{(\operatorname{D}^{\bullet \bullet} - \operatorname{D}_{t})}$
2 4 6 8 10 12 14 16	0.188 0.373 0.586 0.750 0.952 1.118 1.287 1.448	1.91043 1.88266 1.84971 1.80730 1.79076 1.76140 1.73289 1.70402 1.67635	0.02831 0.05608 0.08853 0.12094 0.14798 0.17784 0.20535 0.23422 0.26189

 $V_0 = 2.51 \times 10^{-5} \text{ moles.min.}^{-1}$

^{*}This column obtained by subtraction of the blank value from the original data

Initial Substrate Concentration = 1.82×10^{-3} Molar

Blank = 0.200 Concentration of Sodium Sulfate = 50×10^{-3} Molar

t	D _t (corr)*	ln(D∞ -D _t)	ln D o (D o -D _t)
2 4 6 8 10 12 14 16 ¹ / ₄	0.209 0.438 0.623 0.846 1.072 1.247 1.410 1.009	1.90733 1.87272 1.84388 1.80796 1.74432 1.73994 1.71091 1.67429 1.64384	0.03055 0.06516 0.09400 0.12992 0.19356 0.19794 0.22703 0.26365 0.29410

 $V_0 = 2.60 \times 10^{-5} \text{ moles.min.}^{-1}$

Initial Substrate Concentration = 1.82×10^{-3} Molar

Blank = 0.185Concentration of Sodium Sulfate = 100×10^{-3} Molar

<u>t</u>	D _t (corr)*	ln(D∞ -D _t)	$\frac{\operatorname{D}_{\infty}}{(\operatorname{D}_{\infty}-\operatorname{D}_{t})}$
2½ 4 6 8 10 12 14 16 18	0.195 0.353 0.515 0.688 0.866 1.046 1.224 1.356 1.468	1.90939 1.88570 1.86081 1.83354 1.80467 1.77461 1.74397 1.72060	0.02846 0.05215 0.07707 0.10434 0.13321 0.16327 0.19391 0.21728 0.22568
	<u></u>		

 $V_0 = 2.25 \times 10^{-5} \text{ moles.min.}^{-1}$

^{*}This column obtained by subtraction of the blank value from the original data

APPENDIX

THE SYNTHESIS OF N-TRICHLOROACETYL-L-TYROSINAMIDE

This compound is a desirable substrate to study because of its usefulness in providing evidence pertaining to the theory which relates the magnitude of k_3 and the molar volume of the N-acyl function. It is predicted that the compound will have the largest k_3 of any amide yet studied in this laboratory.

At first it was thought that this compound could be formed by the acylation of tyrosinamide with trichloro-acetyl chloride. The reaction was attempted under the same conditions that tyrosine ethyl ester was acylated (p.166). However the products which were obtained by this procedure had a low chlorine content. This was presumably due to the fact that water very rapidly attacked trichloroacetyl chloride and prevented it from reacting with tyrosinamide. Under these conditions the trichloroacetyl amide bond was also probably hydrolyzed.

Next a nonaqueous acylation was tried. The conditions were similar to those employed for the acetylation of cystine dimethyl ester dihydrochloride (p.171). The product from the reaction of one mole of acid chloride and one mole of amino acid derivative was analyzed and found to have a high chlorine content. This indicates that some O-N diacylation

had taken place. Attempts to selectively hydrolyze the phenyl ester without cleavage of the trichloroacetyl amide linkage resulted in failure.

It was next proposed to protect the phenyl hydroxyl group by forming the carbobenzoxyl ester and then to acetylate under anhydrous conditions. The free phenyl hydroxyl group could then be regenerated by catalytic hydrogenolysis of the carbobenzoxy function. This idea was abandoned when it was found that under the conditions of the hydrogenolysis the chlorine atoms were cleaved from the trichloroacetyl rest. Thus when trichloroacetyl anilide was treated with hydrogen in the presence of a catalysis at atmospheric pressure, mono and dichloroacetyl anilide were formed.

Finally it was determined to attempt the synthesis by the use of an amide-ester interchange. To this end ethylthio trichloroacetate was synthesized. The reaction was carried out in a Soxhlet Extractor. L-Tyrosinamide was placed in an extraction thimble, which was then placed in the upper portion of the extractor. A 20% solution containing 1.2 equivalents of ethylthio trichloroacetate and acetone was placed in the boiler. After one hour of operation all of the tyrosinamide was transferred to the boiler. The solution was a very dark brown color. Upon cooling, a crystalline mass settled out. This material was identified as L-tyrosinamide hydrochloride. No other material could be identified from

reaction mixture.

The experiment was then repeated using ethyl trichloroacetate in place of the thioester. Midway through
the extraction a crystalline solid began separating out.

At the conclusion of the run, the solution above the
crystalline solid was clear with no evidence of decomposition. The crystalline material was recrystallized to a
melting point of 184-185°C. It contained no chlorine.

On the basis of the elementary composition it is suggested
that this material is 2,2 dimethyl-5-(p-hydroxybenzyl)tetrahydroimidazole-4-one. This material presumably
arises by a condensation of acetone with the tyrosinamide.

Elementary composition

$$[\alpha]_{\rm p}^{25} = -18.2 \quad (30\% \text{ ethanol})$$

The experiment was repeated a third time using an-hydrous ethyl acetate as the solvent. When the solvent was allowed to cool a new crystalline substance separated out of a colorless solution. This material contained chlorine and proved to be the desired product. The yield of the crude material was 65% M.P. 158-160°C. The material was recrystallized using chlorobenzene, methanol, water-methanol and anhydrous ethanol-benzene as solvents.

M.P. =
$$160-161^{\circ}$$
 C
$$[\alpha]_{D}^{25} = +30.5^{\circ} (30\% \text{ ethanol})$$
Elementary composition
$$\begin{array}{c} \text{Calc} = 41.89 \\ \text{Found} = 41.83 \end{array} \begin{array}{c} \text{Calc} = 8.87 \\ \text{N} \end{array}$$
Found = 8.91
$$\begin{array}{c} \text{Calc} = 3.38 \\ \text{H} \end{array} \begin{array}{c} \text{Calc} = 33.79 \\ \text{Found} = 3.39 \end{array}$$
Found = 33.68

PROPOSITIONS

1. Dialkylcarbodiimines have been shown to be active agents in the synthesis of anhydrides. (1) It is therefore proposed that they be used in the synthesis of tetra-alkyl pyrophosphites.

$$2 \frac{RO}{RO} P-OH + R'N=C=N-R' \longrightarrow \frac{RO}{RO} P-O-P \stackrel{OR}{\circ} + \frac{R!}{N!} \frac{RO}{RO} P-O-P \stackrel{OR}{\circ} + \frac{RO}{RO} P-O$$

Tetra-alkyl pyrophosphites have been shown to be useful as intermediates in peptide synthesis, (2) but hereto-fore have been prepared only in low yields.(3)

2. Thiol esters have been shown to possess a greater reactivity than the corresponding oxygen esters. (4) In particular their carbonyl group appears to be more susceptible to nucleophilic attack. It is therefore proposed that a synthetic route to alkyl acetylenic ketones lies in the reaction of thiol esters and sodium acetylide. (5)

ch = cNa +
$$c=0$$
 \rightarrow $CH=C-C-C-O^{\circ}$ \rightarrow ch = cCR + NqSC2H5

3. Dihaloisonitriles (6) are compounds which can be considered to be structurally analogous to phosgene. It is therefore proposed that these compounds offer a synthetic approach to dialkylcarbodiimines.

- 4. Carbazole is an amine which displays acidic properties.

 The carbonyl group of N-acyl carbazoles (7) should then
 be subject to nucleophilic attack. It is therefore proposed that N-acyl carbazoles offer possibilities as
 mild, selective acylating agents.
- 5. Heretofore the reaction of anhydrides with amines of alcohols has been followed by titrametric procedures. (8)

 It is desirable that a procedure be developed which is more convenient and of greater accuracy than the titrametric procedure. It is proposed that hydroxylamine (9) offers the basis of such a colorimetric procedure. Concentrated solutions of hydroxylamine may effectively quench the reaction and convert the anhydride to a hydroxamic acid.
- 6. It has been shown (10) that, under mild acidic conditions, the acyl group of N-acyl- hydroxyl alkyl compounds easily migrates to the adjacent hydroxyl group.

It is proposed that these "N to 0" shifts are important phenomena in the acid denaturation of proteins which contain serine or threonine.

7. N-phthalamido-L-amino acid hydrazides could be of important use in the field of amino acid chemistry. The reaction between the phthalamido group and hydrazine make these compounds difficult to prepare. (11) It has been indicated (12) that by treating the phthalamido moiety with ammonia a reversible cleavage of the imide group is possible.

It is proposed that the application of this reaction

could greatly simplify the synthesis of N-phthalamido-L-amino acid hydrazides.

8. It has been shown that chymotrypsin can cause the hydrolysis of **p** keto acids. Cyclopentadiene reacts with aldehydes and ketones in the presence of base to form highly colored fulvenes. (13) It is therefore proposed that cyclopentadiene can form the basis of a colorimetric procedure which can be used to follow the chymotrypsin catalyzed hydrolysis of the compounds listed below.

$$R-C-CH_{2}COH$$
 $R-C-CH_{2}COH$
 $R-C-CH_{2}CH_{3}$
 $R-C-CH_{2}C=N$
 $R-C-CH_{2}-CH_{3}$
 $R-C-CH_{2}-CH_{3}$
 $R-C-CH_{2}-CH_{3}$

9. Acid chlorides are capable of interacting with other acids to form a mixture of acid chlorides and acids. (14)

It is therefore proposed that under conditions of high temperature and pressure a new industrial synthesis of acid chlorides could be developed using formyl chloride as an active intermediate. (15)

10. It is proposed that the makers of "Sta Cold" are guilty of misleading advertising.

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