

## Appendix C

### **Chemical Composition of Secondary Organic Aerosol Formed from the Photooxidation of Isoprene\***

---

\* This chapter is reproduced by permission from “Chemical Composition of Secondary Organic Aerosol Formed from the Photooxidation of Isoprene” by J. D. Surratt, S. M. Murphy, J. H. Kroll, N. L. Ng, L. Hildebrandt, A. Sorooshian, R. Szmigielski, R. Vermeylen, W. Maenhaut, M. Claeys, R. C. Flagan, J. H. Seinfeld, *Journal of Physical Chemistry A*, 110: 9665-9690, 2006. Copyright 2006, American Chemical Society.

# Chemical Composition of Secondary Organic Aerosol Formed from the Photooxidation of Isoprene

Jason D. Surratt,<sup>†</sup> Shane M. Murphy,<sup>‡</sup> Jesse H. Kroll,<sup>§</sup> Nga L. Ng,<sup>‡</sup> Lea Hildebrandt,<sup>‡</sup> Armin Sorooshian,<sup>‡</sup> Rafal Szmigielski,<sup>||</sup> Reinhilde Vermeylen,<sup>||</sup> Willy Maenhaut,<sup>⊥</sup> Magda Claeys,<sup>||</sup> Richard C. Flagan,<sup>§</sup> and John H. Seinfeld<sup>\*,§</sup>

Department of Chemistry, California Institute of Technology, Pasadena, California 91125, Department of Chemical Engineering, California Institute of Technology, Pasadena, California 91125, Departments of Environmental Science and Engineering and Chemical Engineering, California Institute of Technology, Pasadena, California 91125, Department of Pharmaceutical Sciences, University of Antwerp (Campus Drie Eiken), Universiteitsplein 1, BE-2610 Antwerp, Belgium, and Department of Analytical Chemistry, Institute for Nuclear Sciences, Ghent University, Proeftuinstraat 86, BE-9000 Gent, Belgium

Received: March 20, 2006; In Final Form: June 15, 2006

Recent work in our laboratory has shown that the photooxidation of isoprene (2-methyl-1,3-butadiene, C<sub>5</sub>H<sub>8</sub>) leads to the formation of secondary organic aerosol (SOA). In the current study, the chemical composition of SOA from the photooxidation of isoprene over the full range of NO<sub>x</sub> conditions is investigated through a series of controlled laboratory chamber experiments. SOA composition is studied using a wide range of experimental techniques: electrospray ionization–mass spectrometry, matrix-assisted laser desorption ionization–mass spectrometry, high-resolution mass spectrometry, online aerosol mass spectrometry, gas chromatography/mass spectrometry, and an iodometric-spectroscopic method. Oligomerization was observed to be an important SOA formation pathway in all cases; however, the nature of the oligomers depends strongly on the NO<sub>x</sub> level, with acidic products formed under high-NO<sub>x</sub> conditions only. We present, to our knowledge, the first evidence of particle-phase esterification reactions in SOA, where the further oxidation of the isoprene oxidation product methacrolein under high-NO<sub>x</sub> conditions produces polyesters involving 2-methylglyceric acid as a key monomeric unit. These oligomers comprise ~22–34% of the high-NO<sub>x</sub> SOA mass. Under low-NO<sub>x</sub> conditions, organic peroxides contribute significantly to the low-NO<sub>x</sub> SOA mass (~61% when SOA forms by nucleation and ~25–30% in the presence of seed particles). The contribution of organic peroxides in the SOA decreases with time, indicating photochemical aging. Hemiacetal dimers are found to form from C<sub>5</sub> alkene triols and 2-methyltetrols under low-NO<sub>x</sub> conditions; these compounds are also found in aerosol collected from the Amazonian rainforest, demonstrating the atmospheric relevance of these low-NO<sub>x</sub> chamber experiments.

## 1. Introduction

Secondary organic aerosol (SOA) is formed in the troposphere from the oxidation of volatile organic compounds (VOCs), where the resultant low-vapor-pressure oxidation products partition between the gas and aerosol phases. Recent laboratory experiments have established that SOA formation can also result from the heterogeneous reactions between particle-associated substances and relatively volatile species resulting in the formation of high-molecular-weight (MW) products via oligomerization (polymerization).<sup>1–5</sup> Until recently, the formation of SOA from the photooxidation of isoprene, the atmosphere's most abundant nonmethane hydrocarbon, was considered insignificant.<sup>6,7</sup> This was largely due to the known volatility of first-generation gas-phase oxidation products, such as meth-

acrolein (MACR), methyl vinyl ketone (MVK), and formaldehyde, from isoprene oxidation in the presence of NO<sub>x</sub>, and a previous chamber study that concluded that isoprene oxidation does not lead to SOA formation.<sup>8</sup> Recent field observations of certain organic aerosol compounds, diastereoisomeric 2-methyltetrols (2-methylerythritol and 2-methylthreitol), and 2-methylglyceric acid, attributable to isoprene oxidation, and the experimental observation that isoprene under highly acidic conditions can lead to the formation of polymeric, humic-like substances through heterogeneous reactions, re-opened the issue of SOA formation from isoprene.<sup>7,9–13</sup> After their ambient identification, Edney et al.<sup>14</sup> and Böge et al.<sup>15</sup> detected 2-methyltetrols in SOA formed from laboratory chamber studies of isoprene.

Recent work in our laboratory has shown that SOA formation from isoprene oxidation can be significant.<sup>16,17</sup> Extensive experiments were carried out under both low- and high-NO<sub>x</sub> conditions using either nitrous acid (HONO) or hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) as the OH radical source. Photooxidation experiments were also conducted using isoprene first-generation gas-phase oxidation products as the VOC precursor. Although no aerosol growth was observed from MVK oxidation, SOA

\* To whom correspondence should be addressed. Phone: (626) 395-4635. Fax: (626) 796-2591. E-mail seinfeld@caltech.edu.

<sup>†</sup> Department of Chemistry, California Institute of Technology.

<sup>‡</sup> Department of Chemical Engineering, California Institute of Technology.

<sup>§</sup> Departments of Environmental Science and Engineering and Chemical Engineering, California Institute of Technology.

<sup>||</sup> University of Antwerp.

<sup>⊥</sup> Ghent University.

formation was observed from MACR at high-NO<sub>x</sub> conditions. High-molecular-weight (MW) species were observed to form from isoprene oxidation under both low- and high-NO<sub>x</sub> conditions.<sup>17</sup> Moreover, SOA yields were observed to exhibit a dependence on the NO<sub>x</sub> level. This dependence appears to be attributed to differences in organic peroxy radical (RO<sub>2</sub>) chemistry. At high [NO] (i.e., high-NO<sub>x</sub> conditions), RO<sub>2</sub> radicals react mainly with NO to produce small alkoxy radicals (RO) that likely fragment into smaller organics, which are expected to be too volatile to partition appreciably to the aerosol phase, or form organic nitrate species (RONO<sub>2</sub>). In the absence of NO<sub>x</sub> (i.e., low-NO<sub>x</sub> conditions), RO<sub>2</sub> radicals instead react with HO<sub>2</sub> radicals (present in the chamber experiments in large quantities from the OH + H<sub>2</sub>O<sub>2</sub> reaction) to form organic hydroperoxides, which have been experimentally shown to be important SOA components from other VOC precursors.<sup>18,19</sup> Hydroperoxides have been suggested to be involved in polymerization in the aerosol phase via reactions with aldehydes to form peroxyhemiacetals.<sup>18,19</sup>

Although it is now established that OH-initiated oxidation of isoprene leads to SOA, a detailed understanding of the chemical reaction pathways leading to the production of isoprene SOA is lacking. Results from chamber studies have elucidated the importance of the further oxidation of MACR as a primary route for SOA formation from isoprene under high-NO<sub>x</sub> conditions. Known RO<sub>2</sub> chemistry at low-NO<sub>x</sub> conditions leads to the initial gas-phase oxidation products, likely hydroxyhydroperoxides, of isoprene, which upon further oxidation leads to SOA production. Nonetheless, detailed evaluation of the mechanism of SOA formation from the oxidation of isoprene has not yet been carried out.

In the present work, a suite of offline analytical techniques is used in conjunction with online aerosol mass spectrometry to investigate the detailed chemical composition of SOA from isoprene oxidation. SOA is produced from the photooxidation of isoprene under varying NO<sub>x</sub> conditions and is collected onto filters for offline chemical analyses. Offline mass spectrometry (MS) techniques are used to detect organic species from aerosol filter samples, including oligomeric components of isoprene SOA (as detected in prior studies only by online time-of-flight aerosol mass spectrometry (TOF-AMS) measurements). Tandem MS and gas chromatography (GC)/MS derivatization techniques are employed to structurally elucidate oligomeric components. Organic peroxides are detected and quantified from low-NO<sub>x</sub> isoprene SOA using a conventional iodometric-spectroscopic method. Tracer compounds for isoprene oxidation in the ambient atmosphere, as found in the Amazonian rainforest, are detected here for the first time in the low-NO<sub>x</sub> chamber experiments. The low-NO<sub>x</sub> conditions are most relevant to understanding SOA formation in highly vegetated, remote regions.<sup>7</sup> In some cases, such as the southeastern U.S., where atmospheric transport of pollutants from urban areas can influence SOA formation,<sup>20</sup> conditions closer to those of the high-NO<sub>x</sub> experiments may be applicable.

## 2. Experimental Section

**2.1. Chamber Experiments.** Experiments were carried out in Caltech's dual indoor 28 m<sup>3</sup> Teflon smog chambers.<sup>21,22</sup> Experimental protocols are similar to those described previously,<sup>16,17</sup> so they will be described only briefly here. Most experiments were carried out with hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) as the hydroxyl radical (OH) precursor; in some cases, HONO was used instead to demonstrate that the particular OH source has no effect on the outcome of the experiments. For some

experiments, ammonium sulfate seed particles were introduced into the chamber (at volume concentrations of 20–30 μm<sup>3</sup>/cm<sup>3</sup>) by atomization of a 0.015 M ammonium sulfate solution. A known concentration of isoprene (or any other precursor, such as MACR) was then introduced by sending air over a measured volume of the pure compound (Aldrich, 99.8%) into the chamber. For H<sub>2</sub>O<sub>2</sub>/high-NO<sub>x</sub> experiments, NO was also introduced into the chamber from a gas mixture (500 ppm gas cylinder in N<sub>2</sub>, Scott Specialty Gases). In low-NO<sub>x</sub> experiments, NO was not added and NO<sub>x</sub> concentrations were <1 ppb. When the isoprene (monitored by gas chromatography–flame ionization detection (GC-FID)), NO<sub>x</sub>, and seed concentrations became constant inside the chamber, irradiation by UV lights (centered at 354 nm) was started, initiating the reaction.

SOA volume growth (μm<sup>3</sup>/cm<sup>3</sup>) was monitored with a differential mobility analyzer (DMA). For quantification of SOA products collected on filter samples, the DMA volumes were used for each experiment to determine the total SOA mass collected. Filter sampling commenced when the particle growth had terminated, that is, when the aerosol volume had reached its maximum value. Depending on the total volume concentration of aerosol in the chamber, the filter sampling time was 2–4 h, which typically resulted in 3–7 m<sup>3</sup> of total chamber air sampled.

**2.2. Filter Extractions.** Collected Teflon filters (PALL Life Sciences, 47-mm diameter, 1.0-μm pore size, teflo membrane) were extracted in 5 mL of HPLC-grade methanol by 40 min of sonication. The filters were then removed from the methanol sample extracts and archived at –20 °C. Each extract was blown dry under a gentle N<sub>2</sub> stream (without added heat) and then reconstituted with 1 mL of a 50:50 (v/v) solvent mixture of HPLC-grade methanol and 0.1% aqueous acetic acid solution. The reconstituted extracts were then stored at –20 °C until analysis was performed. In most cases, filter extracts were chemically analyzed within 1–2 days after filter extraction. Lab control filters were extracted and treated in the same manner as the samples. Aliquots of each of these filter extracts were analyzed by the four mass spectrometry techniques to follow.

To ensure that H<sub>2</sub>O<sub>2</sub> was not condensing onto filter media and introducing artifacts in the chemical analyses, we collected several blank filters under dark conditions from the chamber containing typical experimental well-mixed concentrations of isoprene, NO, and ammonium sulfate seed aerosol, sampled for the same duration (~2–4 h) as a sample filter. No significant chemical artifacts or contaminants were observed in the analytical techniques from these blank filters, consistent with the lack of observed aerosol growth under dark conditions.

**2.3. Liquid Chromatography/Electrospray Ionization–Mass Spectrometry (LC/ESI-MS).** A Hewlett-Packard 1100 Series HPLC instrument, coupled with a single quadrupole mass analyzer and equipped with an electrospray ionization (ESI) source, was used to identify and quantify relatively polar, acidic SOA components. Data were collected in both positive (+) and negative (–) ionization modes; the quantitative analysis presented here is limited to the negative ionization mode. An Agilent Eclipse C<sub>18</sub> column (3.0 × 250 mm) was used to separate the organic species before detection. The eluents used were 0.1% aqueous acetic acid (A) and methanol (B). In the 40-min gradient elution program used, the concentration of eluent B increased from 5% to 90% in 35 min, and then decreased to 5% in 5 min. The total flow rate of the eluent used in the LC/MS analysis was 0.8 mL min<sup>–1</sup>. Optimum electrospray conditions were found using a 60 psig nebulizing pressure, 3.5 kV capillary voltage, 13 L min<sup>–1</sup> drying gas

flowrate, and a 330 °C drying gas temperature. During the full scan mode of analysis, the cone voltage was set at 60 V, avoiding fragmentation of most species and allowing their detection as deprotonated molecules ( $[M - H]^-$ ). During the upfront collision-induced dissociation (CID) mode of analysis, the cone voltage was set to 110 V, resulting in partial fragmentation of the  $[M - H]^-$  ions. By comparing these two sets of MS data (upfront CID mode to the full scan mode of analysis) and by examining the fragmentation patterns of the species, some structural information on the analyzed species was obtained. This was particularly useful in confirming results from other MS/MS techniques used and for the identification of oligomeric components.

Using a set of six acidic species (*meso*-erythritol, citramalic acid, 2-hydroxy-3-methylbutyric acid, pimelic acid, pinic acid, and suberic acid monomethyl ester) as surrogate standards, this method was also used to quantify the amount of polar acidic species. Filter extraction efficiency was established by standard additions of these surrogate standards to blank filters. On average, the extraction efficiency for each standard was ~60% with an estimated error bar of ca.  $\pm 15\%$  over the concentration range used to generate the LC/MS calibration curves. This average extraction efficiency was included in the calculations to quantify identified isoprene SOA products.

As we will note shortly, to investigate the probable importance of a  $C_4$  hydroxy dialdehyde species formed under high- $NO_x$  conditions, we derivatized selected sample extracts using Girard Reagent P (1-(carboxymethyl)pyridinium chloride hydrazide, MW = 187) to increase sensitivity for aldehydic species in the (+)LC/MS mode. Girard Reagent P (GirP) reacts with aldehydes and ketones to form water-soluble hydrazones with a permanently charged pyridine moiety, and water is eliminated in this reaction.<sup>23</sup> The organic unit that adds to aldehydes and ketones has a mass of 152 Da. A series of aldehyde standards, glyoxal (MW = 58), succinic semialdehyde (MW = 102), and glutaraldehyde (MW = 100), were derivatized using the GirP and analyzed with (+)LC/MS. These small polar aldehyde standards typically go undetected using (+)ESI techniques such as in LC/MS; however, upon derivatization they were detected as the singly charged  $[M - H_2O + 152(\text{GirP})]^+$  ions (glyoxal was also detected as doubly charged  $[M - 2H_2O + 152(\text{GirP})]^{2+}$  ion), where M is the MW of the aldehyde species. These compounds eluted between 1 and 2 min from the LC column, including a derivatized compound corresponding to the proposed  $C_4$  hydroxy dialdehyde species (MW = 102 and  $[M - H_2O + 152(\text{GirP})]^+ = 236$ ).

**2.4. ESI-Ion Trap Mass Spectrometry (ESI-ITMS).** Aliquots of the filter extracts were also analyzed by a ThermoElectron LCQ ion trap mass spectrometer equipped with an ESI source, via direct infusion. This instrument does not provide chromatographic separation, precluding quantification. Instead, the instrument was used for the qualitative detection of product species. In addition, specific ions of interest were isolated from the rest of the sample ion matrix and further fragmented to produce product ion mass spectra, aiding in structural elucidation.

Data were collected in both positive and negative ionization modes. Because the same species were detected in both modes ( $[M - H]^-$  and  $[M + Na]^+$  ions), we only present here the data collected under negative ionization; the data collected under positive ionization serve as confirmation of the negative ionization data.

**2.5. Matrix-Assisted Laser Desorption Ionization-Time-of-Flight Mass Spectrometer (MALDI-TOFMS).** Another

aliquot of the filter extract was analyzed on an Applied Biosystems Voyager-DE Pro MALDI-TOFMS instrument. After 6  $\mu\text{L}$  of each extract had been dried on the steel target plate, the plate was gently brushed with graphite particles, which served as the matrix. The samples were analyzed in the linear mode, in both positive and negative ionization modes. 400–500 laser shots were summed to obtain a representative mass spectrum of each sample. This method was mainly used to assess the molecular weight (MW) range of the aerosol, to detect oligomeric signatures, and to confirm the MWs of species identified by the ESI techniques.

**2.6. High-Resolution ESI-MS.** Extracts were also analyzed by a Waters LCT Premier Electrospray time-of-flight mass spectrometer with W geometry in the Department of Chemistry at the University of California, Irvine, operated in the negative ionization mode. Samples were analyzed by flow injection. The calibration was carried out using sodium formate clusters with co-injection of fmoc-amino acids of appropriate mass spiked into the analytical sample for lock-mass corrections to obtain accurate mass for the oligomeric ions with  $m/z$  266, 323, 365, 368, 467, and 470. These ions were only detected in the high- $NO_x$  experiments, and elemental compositions were determined with reasonable accuracy (within  $\pm 5$  ppm) and were consistent with other analytical observations (such as ESI-MS/MS and GC/MS derivatization data).

**2.7. Aerodyne Time-of-Flight Aerosol Mass Spectrometer (TOF-AMS).** During most chamber experiments, real-time particle mass spectra were collected continuously by an Aerodyne Time-of-Flight Aerosol Mass Spectrometer (TOF-AMS), and averaged spectra were saved every 5 min. The design and capabilities of the TOF-AMS instrument are described in detail elsewhere.<sup>24</sup> Briefly, chamber air enters the instrument through a 100- $\mu\text{m}$  critical orifice at a flowrate of 1.4  $\text{cm}^3/\text{s}$ . Particles with vacuum aerodynamic diameters between 50 and 800 nm are efficiently focused by an aerodynamic lens, passed through a chopper, and then impacted onto a tungsten vaporizer. The chopper can be operated in three modes: (1) completely blocking the beam to gather background mass spectra; (2) out of the beam's path to collect ensemble average mass spectra over all particles sizes; and (3) chopping the beam to create size-resolved mass spectra. The vaporizer is typically run at  $\sim 550$  °C to ensure complete volatilization of the SOA and the inorganic seed; during several runs the vaporizer temperature was lowered to  $\sim 160$  °C to reduce thermally induced fragmentation of oligomers. Once vaporized, molecules undergo electron ionization at 70 eV and are orthogonally pulsed every 19  $\mu\text{s}$  into the time-of-flight mass analyzer.

**2.8. Gas Chromatography/Mass Spectrometry (GC/MS).** Extracts of selected filters were analyzed for polar organic compounds by GC/MS using a method that was adapted from that reported by Pashynska et al.<sup>25</sup> The sample workup consisted of extraction of all or half of the filter with methanol under ultrasonic agitation and derivatization of carboxyl and hydroxyl functions into trimethylsilyl (TMS) derivatives. The extract was divided into two parts; one part was trimethylsilylated while the other part was stored in a refrigerator at 4 °C for eventual further analysis. GC/MS analyses were performed with a system comprising a TRACE GC2000 gas chromatograph, which was coupled to a Polaris Q ion trap mass spectrometer equipped with an external ionization source (ThermoElectron, San Jose, CA). A Heliflex AT-5MS fused-silica capillary column (5% phenyl, 95% methylpolysiloxane, 0.25  $\mu\text{m}$  film thickness, 30 m  $\times$  0.25 mm i.d.) preceded by a deactivated fused-silica precolumn (2 m  $\times$  0.25 mm i.d.) (Alltech, Deerfield, IL) was

used to separate the derivatized extracts. Helium was used as carrier gas at a flow rate of 1.2 mL min<sup>-1</sup>. The temperature program was as follows: isothermal hold at 50 °C for 5 min, temperature ramp of 3 °C min<sup>-1</sup> up to 200 °C, isothermal hold at 200 °C for 2 min, temperature ramp of 30 °C min<sup>-1</sup> up to 310 °C; and isothermal hold at 310 °C for 2 min. The analyses were performed in the full scan mode (mass range: *m/z* 50–800), and were first carried out in the electron ionization (EI) mode and subsequently in the chemical ionization (CI) mode. The ion source was operated at an electron energy of 70 eV and temperatures of 200 °C and 140 °C in the EI and CI modes, respectively. The temperatures of the GC injector and the GC/MS transfer line were 250 °C and 280 °C, respectively. For chemical ionization, methane was introduced as reagent gas at a flow rate of 1.8 mL min<sup>-1</sup>. We present here mainly the data collected in the EI mode; the data collected in the CI mode are used if insufficient MW information is obtained in the EI mode.

Selected extracts were also subjected to a hydrolysis/ethylation and/or a methoximation procedure prior to trimethylsilylation. The purpose of the hydrolysis/ethylation procedure was to confirm the presence of ester linkages, while that of the methoximation procedure was to evaluate the presence of aldehyde/keto groups, in oligomeric SOA. The hydrolysis/ethylation procedure involved reaction of the extract residues with 40 μL of analytical-grade ethanol and 8 μL of trimethylchlorosilane (Supelco, Bellefonte, PA) for 1 h at 60 °C. Details about the methoximation procedure can be found in Wang et al.<sup>12</sup>

**2.9. Gas Chromatography–Flame Ionization Detection (GC-FID).** Quantitative determination of the 2-methyltetrols (i.e., 2-methylthreitol and 2-methylerythritol), the C<sub>5</sub> alkene triols [i.e., 2-methyl-1,3,4-trihydroxy-1-butene (cis and trans) and 3-methyl-2,3,4-trihydroxy-1-butene], and 2-methylglyceric acid, in selected filters, was performed by GC-FID with a GC 8000 Top instrument (Carlo Erba, Milan, Italy). The sample workup was the same as that for GC/MS analysis except that filter parts were spiked with a known amount of erythritol (Sigma, St. Louis, MO) as an internal recovery standard; it was assumed that the GC-FID responses of the trimethylsilyl derivatives of the analytes and the internal recovery standard were similar. The GC column and conditions were comparable with those used for GC/MS; the column was a CP-Sil 8 CB capillary column (5% diphenyl, 95% methylpolysiloxane, 0.25 μm film thickness, 30 m × 0.25 mm i.d.) (Chrompack, Middelburg, The Netherlands) and the temperature program was as follows: isothermal hold at 45 °C for 3 min, temperature ramp of 20 °C min<sup>-1</sup> up to 100 °C, isothermal hold at 100 °C for 10 min, temperature ramp of 5 °C min<sup>-1</sup> up to 315 °C, and isothermal hold at 315 °C for 20 min. Measurement of the 2-methyltetrols in the low-NO<sub>x</sub> SOA samples was performed after the unstable products tentatively characterized as 2-methyltetrol performate derivatives had decayed to 2-methyltetrols, that is, after leaving the reaction mixture for 2 days at room temperature.

**2.10. Total Aerosol Peroxide Analysis.** The total amount of peroxides in the low-NO<sub>x</sub> isoprene SOA was quantified using an iodometric-spectrophotometric method adapted from that used by Docherty et al.<sup>18</sup> to analyze peroxides formed by α-pinene-ozonolysis. The method employed here differed only in the choice of extraction solvent: we used a 50:50 (v/v) mixture of methanol and ethyl acetate, rather than pure ethyl acetate. Calibration and measurements were performed at 470 nm on a Hewlett-Packard 8452A diode array spectrophotometer. A standard calibration curve was obtained from a series of

benzoyl peroxide solutions. Benzoyl peroxide was the standard used for quantification of organic peroxides formed from low-NO<sub>x</sub> experiments, because its MW is close to the average MW determined from the mass spectrometry techniques, in particular the MALDI-TOFMS measurements. The molar absorptivity determined from the standard curve was ~852, in excellent agreement with that determined by Docherty et al. and with the value of 845 determined with the original method development paper.<sup>18,26</sup> As a confirmation that the technique was reproducible, we extracted and analyzed in the same fashion, three α-pinene-ozonolysis filters collected from our laboratory chambers. We measured ~49% of the SOA mass, produced from α-pinene ozonolysis, to be organic peroxides, in excellent agreement to that of Docherty et al.'s measurement of ~47% for the same system. A few high-NO<sub>x</sub> isoprene filter samples were also analyzed by this method, but resulted in the detection of no organic peroxides (below detection limits of this technique).

**2.11. Particle-Into-Liquid Sampler Coupled to Ion Chromatography (PILS/IC).** The PILS/IC (particle-into-liquid sampler coupled to ion chromatography) is a quantitative technique for measuring water-soluble ions in aerosol particles. The PILS developed and used in this study<sup>27</sup> is based on the prototype design<sup>28</sup> with key modifications, including integration of a liquid sample fraction collector and real-time control of the steam injection tip temperature. Chamber air is sampled through a 1-μm cut-size impactor and a set of three denuders (URG and Sunset Laboratories) to remove inorganic and organic gases that may bias aerosol measurements. Sample air mixes with steam in a condensation chamber where rapid adiabatic mixing produces a high water supersaturation. Droplets grow sufficiently large to be collected by inertial impaction before being delivered to vials held on a rotating carousel. The contents of the vials are subsequently analyzed off-line using a dual IC system (ICS-2000 with 25 μL sample loop, Dionex Inc.) for simultaneous anion and cation analysis. The background levels of individual species (Na<sup>+</sup>, NH<sub>4</sub><sup>+</sup>, K<sup>+</sup>, Mg<sup>2+</sup>, Ca<sup>2+</sup>, SO<sub>4</sub><sup>2-</sup>, Cl<sup>-</sup>, NO<sub>2</sub><sup>-</sup>, NO<sub>3</sub><sup>-</sup>, oxalate, acetate, formate, methacrylate, pyruvate) concentrations for analyzed filter samples, presented as the average concentration plus three times the standard deviation ( $\sigma$ ), are less than 0.28 μg m<sup>-3</sup>.

### 3. Results

As noted, experiments were conducted at high- and low-NO<sub>x</sub> conditions. High-NO<sub>x</sub> conditions were achieved through the addition of substantial NO<sub>x</sub> (~800 to 900 ppb NO<sub>x</sub>) to the reaction chamber, leading to isoprene/NO<sub>x</sub> molar ratios of ~0.56 to 0.63. Under low-NO<sub>x</sub> conditions no NO<sub>x</sub> is added to the chamber, where NO<sub>x</sub> mixing ratios of <1 ppb (small amounts of NO<sub>x</sub> likely desorb from chamber walls) were observed. The low-NO<sub>x</sub> condition simulates a remote (NO<sub>x</sub>-free) atmosphere; for example, at typical isoprene and NO<sub>x</sub> mixing ratios observed in the Amazonian rainforest (~4 to 10 ppb and 0.02 to 0.08 ppb, respectively),<sup>7,29</sup> the isoprene/NO<sub>x</sub> ratios that result are ~50 to 500, comparable to the isoprene/NO<sub>x</sub> ratio of the present experiments (~500).

**3.1. High-NO<sub>x</sub> Condition.** Table 1 lists nine high-NO<sub>x</sub> chamber experiments that were conducted to generate SOA for aerosol filter sampling. All experiments were conducted with 500 ppb of isoprene or MACR in order to produce sufficient aerosol mass for all offline analytical measurements. In most of the experiments conducted, H<sub>2</sub>O<sub>2</sub> served as the OH radical precursor; in this manner, initial oxidation of isoprene is dominated by OH. It is estimated that ~3–5 ppm of H<sub>2</sub>O<sub>2</sub> was

TABLE 1: High-NO<sub>x</sub> Chamber Experiments Conducted

expt no.	VOC <sup>a</sup>	OH precursor <sup>b</sup>	seeded/ <sup>c</sup> nucleation	initial [NO] ppb	initial [NO <sub>2</sub> ] ppb	initial [NO <sub>x</sub> ] ppb	[O <sub>3</sub> ] <sup>d</sup> ppb	T, °C <sup>d</sup>	total SOA mass concentration <sup>d,e,f</sup> μg/m <sup>3</sup>
1	isoprene	H <sub>2</sub> O <sub>2</sub>	nucleation	827	34	860	498	28.5	74
2	isoprene	H <sub>2</sub> O <sub>2</sub>	dry AS	759	112	869	525	28.3	73
3	MACR	H <sub>2</sub> O <sub>2</sub>	dry AS	791	60	850	540	25.2	181
4	MACR	H <sub>2</sub> O <sub>2</sub>	nucleation	898	30	926	519	25.0	197
5	isoprene	H <sub>2</sub> O <sub>2</sub>	nucleation	805	87	891	294	24.3	104
6	isoprene	H <sub>2</sub> O <sub>2</sub>	AAS	825	80	904	450	24.6	111
7 <sup>g</sup>	isoprene	HONO	dry AS	50	333	382	132	20.1	68
8 <sup>g</sup>	isoprene	HONO	nucleation	89	279	366	134	21.4	73
9	isoprene	H <sub>2</sub> O <sub>2</sub>	dry AS	891	74	963	325	24.9	95

<sup>a</sup> All VOC gas phase mixing ratios were 500 ppb. MACR = methacrolein. <sup>b</sup> H<sub>2</sub>O<sub>2</sub> and HONO are not measured directly, but from isoprene decay during irradiation we estimate ~3 ppm of H<sub>2</sub>O<sub>2</sub>, and [HONO] is unlikely greater than measured [NO<sub>2</sub>]. <sup>c</sup> AS = ammonium sulfate seed, AAS = acidic ammonium sulfate seed. <sup>d</sup> Averaged over the course of the filter sampling. <sup>e</sup> Subtraction of seed aerosol taken into account when necessary. SOA volume derived from DMA wall loss uncorrected measurements for use in mass closure from filter sample analyses. <sup>f</sup> Assuming a SOA density of 1.35 g/cm<sup>3</sup>. This value is derived from comparison of DMA aerosol mass measurements. <sup>g</sup> 10% of light bank used and hence lower temperature observed. Also lower amounts of initial NO due to HONO as precursor.

used in each of these experiments based upon isoprene decay during irradiation.<sup>17</sup> All of these experiments were conducted at low relative humidity ( $RH < 5\%$ ) in order to limit the uptake of H<sub>2</sub>O<sub>2</sub> into the particle phase. In the high-NO<sub>x</sub> experiments using H<sub>2</sub>O<sub>2</sub> as an OH source, ~800 to 900 ppb of NO was injected into the chamber. With the HONO source, lower initial NO concentrations were achieved, because the source of NO was HONO photolysis and a NO<sub>x</sub> side-product from the HONO synthesis. Nucleation (seed-free) and ammonium sulfate seeded experiments were also conducted to examine if the presence of seed aerosol has an effect on the chemistry observed. In Experiment 6, acidified ammonium sulfate seed (0.015 M (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> + 0.015 M H<sub>2</sub>SO<sub>4</sub>) was used to investigate the possible effect of acid catalysis on oligomerization reactions, which has previously been observed to occur for other VOC precursors, such as α-pinene and 1,3,5-trimethylbenzene.<sup>1,3-5,30</sup> No discernible increase in SOA mass is observed for this acid-seeded experiment (Experiment 6) when comparing to its corresponding dry-seeded and nucleation (seed-free) experiments (Experiments 5 and 9).

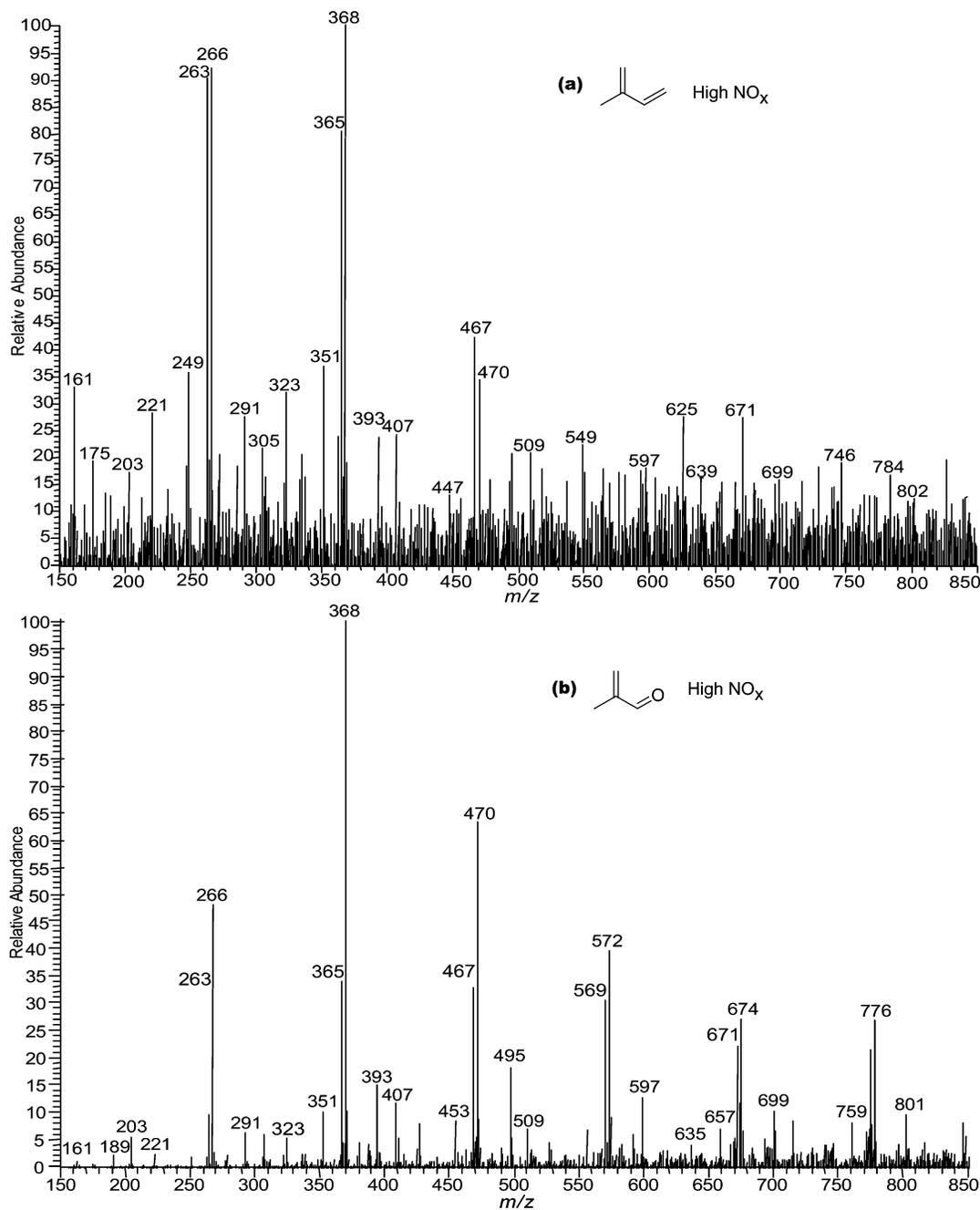
To illustrate the overall chemical composition typically observed under high-NO<sub>x</sub> conditions, shown in Figure 1a is a first-order (-)ESI-IT mass spectrum obtained via direct infusion analysis of an isoprene SOA sample collected from Experiment 1. Prior work in our laboratory has shown that most organics detected in the negative ion mode occur as the deprotonated molecules ( $[M - H]^-$  ions),<sup>2,3,20</sup> making (-)ESI sensitive for the detection of polar acidic species. As can be seen in Figure 1a, many such species are detected. Observable 102 Da differences between many of the  $[M - H]^-$  ions and the detection of high-MW species (up to MW ~470) indicate the presence of oligomeric species with more than the five carbons of the parent isoprene. Organic nitrate species are detected in this spectrum as even-mass  $[M - H]^-$  ions ( $m/z$  266, 368, and 470).

Figure 1b shows, by comparison, a first-order (-)ESI-IT spectrum, also obtained via direct infusion analysis, for a MACR high-NO<sub>x</sub> sample (Experiment 3). Many of the ions detected correspond exactly to those observed from isoprene oxidation (Figure 1a). It should be noted that when the MACR, H<sub>2</sub>O<sub>2</sub>, and dry ammonium sulfate seed aerosol are well-mixed in the chamber under dark conditions, no aerosol growth is observed, confirming that photooxidation is required to produce SOA. The SOA components formed in this MACR experiment (as shown in Figure 1b) extend out to higher MWs than those of isoprene, which is likely a result of the amount of MACR precursor

available in this experiment and also owing to the removal of one oxidation step (the oxidation of isoprene).

The SOA products detected in Figure 1a and b are confirmed by additional mass spectrometry techniques. Figure 2 shows a mass spectrum collected using the MALDI-TOFMS instrument in the positive ion mode for a high-NO<sub>x</sub>, seeded isoprene photooxidation experiment (Experiment 9). SOA components observed here are detected mainly as the sodiated molecules ( $[M + Na]^+$  ions), which is consistent with our experiences in analyzing polymeric standards, such as aqueous glyoxal, with a graphite matrix. In Figure 2, only species that correspond to ions detected in the (-)ESI-IT spectra are highlighted. For example, for the  $[M - H]^-$  ion series detected in (-)ESI-IT spectra at  $m/z$  161, 263, 365, and 467, a corresponding  $[M + Na]^+$  ion series is detected at  $m/z$  185, 287, 389, and 491, respectively, using MALDI-TOFMS. It should be noted that the (+)ESI-IT spectra also detected the same ions ( $[M + Na]^+$ ) as those of the MALDI technique, confirming that the species observed in Figures 1 and 2 are not a result of ionization artifacts specific to individual techniques.

The LC/MS results obtained in the negative ionization mode are used to quantify the SOA components common to all high-NO<sub>x</sub> isoprene SOA (as detected in Figures 1 and 2). Figure 3a and b show total ion chromatograms (TICs) for an isoprene photooxidation experiment (Experiment 1) and a MACR photooxidation experiment (Experiment 4), respectively, both carried out at high NO<sub>x</sub> in the absence of seed aerosol. These TICs show that many of the SOA products formed in each system are the same because the retention times (RTs) are comparable and the  $m/z$  values of the molecular ion species ( $[M - H]^-$ ) associated with each chromatographic peak are the same. Shown in Figure 3c-e are extracted ion chromatograms (EICs) for three organic nitrate species ( $[M - H]^-$  at  $m/z$  266, 368, and 470) common to both isoprene and MACR high-NO<sub>x</sub> photooxidation experiments. For each chamber experiment, EICs were used instead of TICs for the quantification of each  $[M - H]^-$  ion detected in order to deconvolute any coeluting species. Figure 4a shows a mass spectrum recorded for the largest chromatographic peak ( $RT = 15.7$  min) from the EIC of  $m/z$  368 (Figure 3d). The  $m/z$  759 ion that is also detected in this mass spectrum is a cluster ion corresponding to  $[2M + Na - 2H]^-$ ; such cluster ions are commonly observed in (-)LC/ESI-MS conditions. In Figure 4b is a resultant upfront CID mass spectrum taken for this same chromatographic peak, showing many product ions from the dissociation of  $m/z$  368. The product ion  $m/z$  305 corresponds to a neutral loss of 63 Da, which is

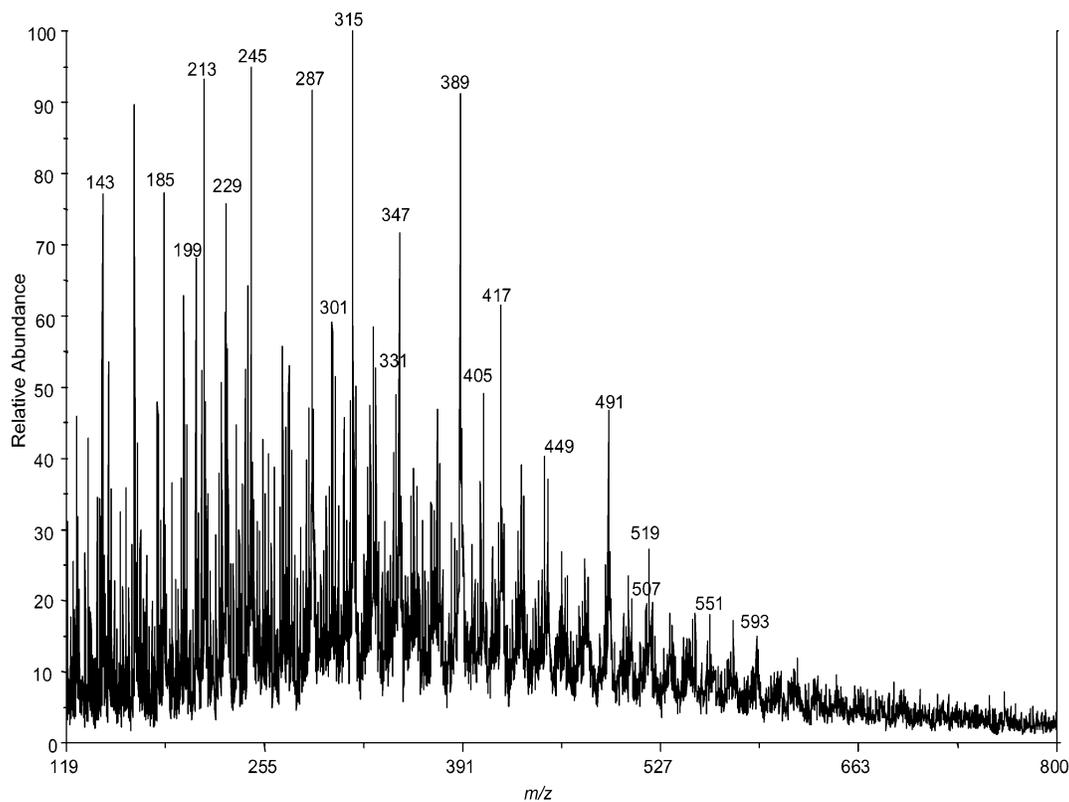


**Figure 1.** ESI-ITMS negative mode spectra collected via direct infusion analyses. (a) MS scan of a filter extract obtained from a 500 ppb isoprene, high- $\text{NO}_x$ , seeded experiment. (b) MS scan of a filter extract obtained from a 500 ppb MACR, high- $\text{NO}_x$ , seeded experiment. These mass spectra show that MACR oxidation produces many of the same SOA products as that of isoprene oxidation under high- $\text{NO}_x$  conditions. Common 102 Da differences between ions in both spectra are observed indicating the presence of oligomers.

likely nitric acid ( $\text{HNO}_3$ ). Another product ion  $m/z$  291 corresponds to neutral loss of 77 Da, likely from the combined losses of a methyl ( $\text{CH}_3$ ) radical and a nitrate ( $\text{NO}_3$ ) radical (or  $\text{CH}_3\text{ONO}_2$ ). The neutral loss of 102 Da results in the product ion  $m/z$  266; these types of product ions are used to aid in the structural elucidation of SOA components and will be discussed subsequently. Owing to the lack of available authentic oligomeric standards, quantification was carried out by using a series of calibration curves generated from surrogate standards (listed in the Experimental Section) covering the wide range of  $RT$ s for all detected species. Each surrogate standard contained a carboxylic acid group, the likely site of ionization for detected SOA components, except for the *meso*-erythritol standard. Because of the initial high percentage of aqueous buffer present in the LC/MS gradient, we were able to detect small polar

organics, such as 2-methylglyceric acid. To quantify this compound, the polyol *meso*-erythritol, detected as the  $[M - \text{H} + \text{acetic acid}]^-$  ion, was used. Unlike *meso*-erythritol, 2-methyltetrols (and other polyols) were not detected using the  $(-)$ -LC/MS technique. All surrogate standards were within ca.  $\pm 1.5$  min of the  $RT$ s of the detected SOA components. Table 2 shows the LC/MS quantification results for high- $\text{NO}_x$  SOA. Four types of oligomers are quantified here. For ease of comparison, experiments corresponding to the same VOC and OH precursor type are grouped together under the same column heading.

SOA components observed thus far are not artifacts formed on filters and are observed over varying isoprene concentrations, as confirmed by online particle mass spectrometry, Figure 5 shows mass spectra collected from three high- $\text{NO}_x$  chamber experiments using the Aerodyne TOF-AMS instrument. In these



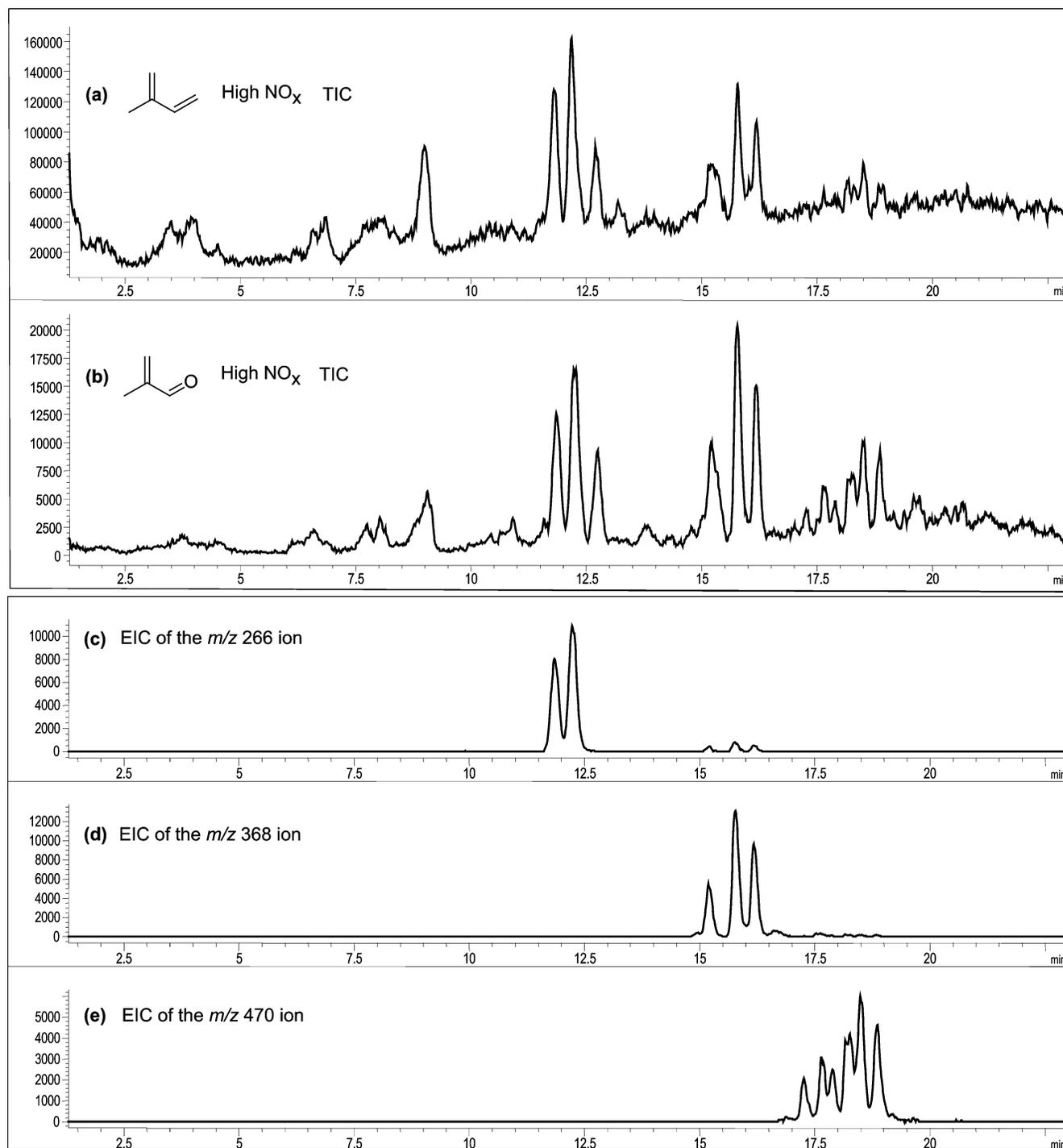
**Figure 2.** MALDI positive mode spectrum obtained with a graphite matrix for a 500 ppb isoprene, high- $\text{NO}_x$ , dry seeded experiment (Experiment 9). Highlighted  $\text{Na}^+$  adduct ions confirm the existence of the species detected by ESI.

experiments, the TOF-AMS instrument was operated at  $\sim 160$  °C to lessen the degree of thermal fragmentation of the high-MW SOA components. Figure 5a shows a TOF-AMS spectrum collected for a 50 ppb isoprene, high- $\text{NO}_x$  nucleation experiment (not included in Table 1 because of insufficient aerosol mass for offline chemical analysis techniques). Even at these isoprene concentrations, high-MW species are detected in the SOA produced. Differences of 102 Da are noted in this spectrum, again indicating the presence of oligomers. The oligomers present here confirm the species detected by the (-)ESI and (+)MALDI techniques (Figures 1 and 2, respectively), where the observed TOF-AMS ions result from a loss of a hydroxyl (OH) radical from the molecular ion (i.e.,  $\alpha$ -cleavage of a hydroxyl radical from a carboxylic acid group). ESI detects these oligomers as the  $[M - H]^-$  ion and MALDI as the  $[M + \text{Na}]^+$  ion, so ions measured in the TOF-AMS instrument are lower by 16 and 40 units, respectively. For example, ions of  $m/z$  145, 187, 247, and 289 measured by the TOF-AMS instrument (Figure 5), correspond to  $m/z$  161, 203, 263, and 305, respectively, using (-)ESI (Figure 1). Four different series of oligomers are highlighted in this spectrum, where ions of the same oligomeric series are indicated in a common color. Figure 5b corresponds to a MACR high- $\text{NO}_x$ , dry seeded experiment, in which a filter sample was collected (Experiment 3), showing the same oligomeric signature to that of the low concentration (50 ppb) isoprene experiment. Figure 5c corresponds to an isoprene high- $\text{NO}_x$ , HONO experiment (Experiment 8). Again, many ions at the same  $m/z$  values are detected, as those of Figures 5a and 5b, suggesting the chemical components of the SOA are the same in these samples. Though probably present, oligomeric compounds formed under conditions similar to those of Figure 5c were not detected in the original study of SOA formation from this laboratory<sup>16</sup> because a less-sensitive quadrupole AMS was used; such high-MW species were reported

in a subsequent study using the TOF-AMS.<sup>17</sup> These online chemical results confirm that the 102 Da differences observed in the offline analytical techniques (ESI and MALDI) are not a result of sample workup or ionization artifacts. Also, these online chemical results suggest that seeded versus nucleation experiments do not lead to significant differences in the chemistry observed, in agreement with the ESI analyses. The OH precursor (HONO or  $\text{H}_2\text{O}_2$ ) also does not have a substantial effect on the chemistry observed (i.e., similar products formed, however, abundances may vary), an observation that is also consistent with the offline mass spectrometry analyses.

PILS/IC measurements were carried out for Experiments 1 (nucleation) and 2 (dry seeded). In both experiments, the acetate anion was the most abundant organic anion detected ( $14.72 \mu\text{g}/\text{m}^3$  in Experiment 1 and  $23.47 \mu\text{g}/\text{m}^3$  in Experiment 2) followed by the formate anion ( $1.18 \mu\text{g}/\text{m}^3$  in Experiment 1 and  $2.90 \mu\text{g}/\text{m}^3$  in Experiment 2). It should be noted that these two ions elute off the IC column immediately after sample injection, and there is a possibility that other early-eluting monocarboxylic acid species coeluted with these two species leading to an overestimate of their mass. In addition, the extent to which the acetate and formate levels quantified here represent decay products from oligomers detected in the particle phase is uncertain. It is likely that a significant fraction of this mass results from the decomposition of oligomers at the sample collection conditions (high water concentrations and temperatures) in the PILS instrument and possibly by the use of potassium hydroxide (KOH) as the eluent for anion analyses in the IC instrument.

GC/MS with TMS derivatization (restricted to carboxyl and hydroxyl groups) was employed to determine the functional groups present within SOA components formed under high- $\text{NO}_x$  conditions. Figure 6a shows a GC/MS TIC of a high- $\text{NO}_x$  isoprene nucleation experiment (Experiment 5). 2-methylglyceric

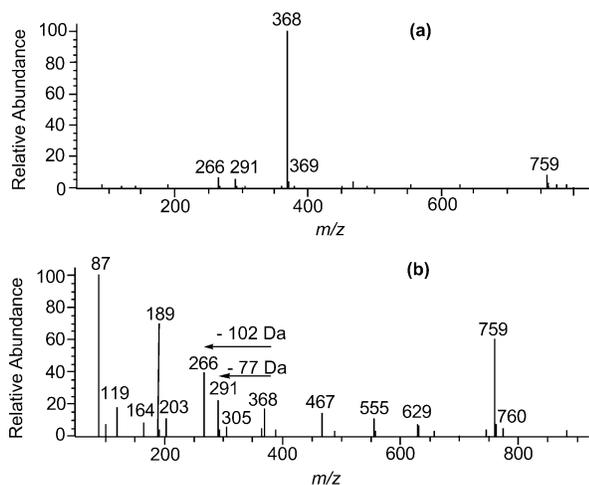


**Figure 3.** (a) LC/MS TIC of a filter extract from a 500 ppb isoprene, high- $\text{NO}_x$ , nucleation experiment. (b) LC/MS TIC of a filter extract from a 500 ppb MACR, high- $\text{NO}_x$ , nucleation experiment. The similar retention times and mass spectra associated with each chromatographic peak in these two TICs indicate that MACR is an important SOA precursor from isoprene oxidation under high- $\text{NO}_x$  conditions. c–e are LC/MS EICs of organic nitrate species common to both MACR and isoprene high- $\text{NO}_x$  samples. These organic nitrate ions are a part of the same oligomeric series confirmed by MS/MS analyses.

acid (2-MG), detected previously in ambient and laboratory filter samples,<sup>10,11,13,14</sup> was found to elute from the GC column at 29.08 min. The corresponding EI mass spectrum for this peak is shown in Figure 6b. The chemical structure of trimethylsilylated 2-MG, along with its respective MS fragmentation, is also shown in this mass spectrum. Using GC-FID to quantify the amount of 2-MG present in this same sample, it was found that  $3.8 \mu\text{g}/\text{m}^3$  was formed, which accounted for  $\sim 3.7\%$  of the SOA mass. This was consistent with LC/MS measurements of 2-MG from other high- $\text{NO}_x$  isoprene nucleation experiments

(such as 2.7% of the SOA mass for Experiment 1). A *di*-ester peak was observed to elute from the GC column at 51.59 min. The corresponding EI mass spectrum for this chromatographic peak is shown in Figure 6c along with its proposed chemical structure and MS fragmentation pattern.

**3.2. Low- $\text{NO}_x$  Condition.** Table 3 lists nine low- $\text{NO}_x$  chamber experiments. All experiments were conducted with  $\text{H}_2\text{O}_2$  as the OH radical precursor with no added  $\text{NO}_x$ . Ozone formation is attributed mainly to residual  $\text{NO}_x$  emitted by the chamber walls; these  $\text{O}_3$  concentrations observed likely have a



**Figure 4.** (a) Mass spectrum for the largest chromatographic peak ( $RT = 15.7$  min) from Figure 3d (EIC of  $m/z$  368 ion). (b) Upfront CID mass spectrum for the same chromatographic peak in Figure 3d (EIC of  $m/z$  368 ion). The neutral losses observed in the upfront CID mass spectrum are associated with a trimeric organic nitrate species. This fragmentation pattern of  $m/z$  368 is consistent with ion trap MS/MS results. The product ion  $m/z$  266 corresponds to a neutral loss of 102 Da (common to all MS techniques), the product ion  $m/z$  291 corresponds to a neutral loss of 77 Da (likely  $\text{CH}_3$  radical and  $\text{NO}_3$  radical,  $\text{CH}_3\text{-NO}_3$ ), the product ion  $m/z$  305 corresponds to a neutral loss of 63 Da (likely  $\text{HNO}_3$ ), the product ion  $m/z$  203 corresponds to a neutral loss of 165 Da, and the product ion  $m/z$  164 corresponds to a neutral loss of 204 Da (two losses of common monomer).

negligible effect on the gas-phase chemistry because of the slow reactivity of  $\text{O}_3$  toward isoprene. Experiments were conducted with 50% of the light banks in the chamber except for Experiments 10 and 11, in which 100% of the light banks were used and resulted in the higher temperatures observed. All experiments were conducted with 500 ppb of isoprene except for Experiment 17, in which 100 ppb of isoprene was used. As in the high- $\text{NO}_x$  experiments, these experiments were conducted at low relative humidity ( $RH < 9\%$ ) in order to limit the uptake of  $\text{H}_2\text{O}_2$  into the particle phase. Nucleation (seed-free) and seeded (ammonium sulfate and acidified ammonium sulfate) experiments were conducted in order to examine if the presence of seed aerosol has an effect on the chemistry observed. Assuming a density of  $\sim 1.25$  g/cm<sup>3</sup> (derived from the comparison of DMA aerosol volume and TOF-AMS aerosol mass measurements), acid seeded (0.015 M  $(\text{NH}_4)_2\text{SO}_4 + 0.015$  M  $\text{H}_2\text{SO}_4$ ) experiments formed the largest amounts of SOA mass ( $\sim 259$   $\mu\text{g}/\text{m}^3$  for Experiment 14) compared to the corresponding nucleation ( $\sim 72.5$   $\mu\text{g}/\text{m}^3$  for Experiment 12) and ammonium sulfate seeded experiments ( $\sim 72.8$   $\mu\text{g}/\text{m}^3$  for Experiment 15). Lower mixing ratios of isoprene (Experiment 17) in the presence of acid seed also resulted in larger amounts of SOA when compared to the nucleation and ammonium sulfate seeded experiments.

No particle-phase organics were detected using (–) and (+)-ESI techniques. Analysis of filter sample extracts using these techniques were nearly identical to the blank and control filters. This shows that SOA components at low- $\text{NO}_x$  conditions are not acidic in nature like those of the high- $\text{NO}_x$  SOA. Because of the expected presence of hydroperoxides and polyols, other analytical techniques, such as the iodometric-spectrophotometric method and GC/MS with TMS derivatization, were employed to understand the chemical nature of low- $\text{NO}_x$  SOA. The peroxide aerosol mass concentration was measured for all experiments except for Experiments 12, 13, and 16. The

iodometric-spectrophotometric method measures the total peroxide content (sum of ROOH, ROOR, and  $\text{H}_2\text{O}_2$ ) of the aerosol, but because no peroxides were measured from filters collected from air mixtures containing isoprene,  $\text{H}_2\text{O}_2$ , and seed aerosol, it is assumed that the peroxides measured are organic peroxides. The nucleation experiments (Experiments 10 and 18a) had the highest contribution of peroxides ( $\sim 61\%$  on average) to the SOA mass observed. Dry ammonium sulfate (Experiments 11 and 15) and acidified ammonium sulfate seeded experiments (Experiments 14 and 17) led to comparable contributions of organic peroxides to the overall SOA mass ( $\sim 25$  and 30%, respectively). Quality control tests were conducted by the addition of ammonium sulfate to standard solutions of benzoyl peroxide to test if the seed had an effect on the UV–vis measurement of total peroxides. The amount of ammonium sulfate added to the benzoyl peroxide standards was determined by the ratio of SOA volume growth to the typical ammonium sulfate seed volume employed ( $\sim 3:1$ ) as determined from the DMA. Little difference was observed ( $\sim 0.6\%$ ), showing that ammonium sulfate seed has a negligible effect on the measurement of peroxide content from seeded experiments. As observed previously,<sup>17</sup> the SOA mass was found to decrease rapidly in nucleation experiments after reaching peak growth, and as a result, the peroxide content of the SOA was measured at different times in Experiment 18. The iodometric-spectrophotometric measurement made at the peak growth in the aerosol volume, as determined from the DMA, for Experiment 18, showed that the peroxides accounted for  $\sim 59\%$  of the total SOA mass. Twelve hours later, once the aerosol volume decay reached its constant value, the peroxide contribution to the SOA mass was found to have dropped to 26%.

Figure 7 shows a (+)MALDI mass spectrum for a low- $\text{NO}_x$  acid-seed experiment (Experiment 14). The  $m/z$  range (49–620) of ion species observed was not significantly different from (+)-MALDI results obtained for nonacid-seeded experiments. The abundances of these ions were higher for the acid experiments, but quantification of these species is not possible because of uncertainties in the ionization efficiencies. In the absence of seed MALDI signal was low or nonexistent, likely due to very low ionization efficiencies in the absence of a sulfate matrix. Quantification is also difficult with MALDI because of inconsistencies and inhomogeneities of sample preparation and lack of understanding of sample matrix effects.<sup>31</sup> It is clear, however, that oligomerization occurs in low- $\text{NO}_x$  SOA. Common 14, 16, and 18 Da differences are observed between many peaks throughout this spectrum. Structural elucidation of these peaks in Figure 7 was not possible using the (+)MALDI technique owing to the inability of performing MS/MS experiments on selected ions from the sample matrix.

Figure 8 shows two TOF-AMS mass spectra for a 500 ppb, low- $\text{NO}_x$  nucleation experiment (Experiment 12) in the  $m/z$  range of 200–450. These mass spectra also indicate the existence of oligomeric components for low- $\text{NO}_x$  SOA. The mass spectrum in Figure 8a was collected at a low vaporizer temperature ( $\sim 150$  °C) while that in Figure 8b was collected at a higher temperature ( $\sim 600$  °C). The presence of more higher-mass peaks at high vaporizer temperatures (Figure 8b) may indicate that the low- $\text{NO}_x$  oligomers are heterogeneous, with some series of oligomers being easily volatilized below 200 °C while others are not volatile at these temperatures.

The chemical composition of the SOA formed under low- $\text{NO}_x$  conditions was found to change over the course of the experiment. The evolution of selected ions and of the total organic mass measured by the TOF-AMS instrument is shown

**TABLE 2: Quantified SOA Products (in ng/m<sup>3</sup>) from High-NO<sub>x</sub> Chamber Experiments**

	$[M - H]^-$ ion	surrogate standard used for quantification <sup>a</sup>	isoprene/high NO <sub>x</sub> /H <sub>2</sub> O <sub>2</sub>				MACR/high NO <sub>x</sub> /H <sub>2</sub> O <sub>2</sub>		isoprene/HONO	
			expt 1	expt 2	expt 6	expt 9	expt 3	expt 4	expt 7	expt 8
<i>mono</i> -nitrate oligomers	266	pimelic acid	1970	4170	3890	3910	9360	3860	1470	830
	368	pinic acid	1350	2450	3700	4440	20600	10100	830	750
	470	pinic acid	2330	2930	2300	2640	28900	16700		210
	572	pinic acid	<sup>b</sup>				2960	6810		
	674	suberic acid monomethyl ester					670	710		
	776	suberic acid monomethyl ester					220	450		
	878	suberic acid monomethyl ester						210		
total mass from <i>mono</i> -nitrate oligomers (μg/m <sup>3</sup> )			5.65	9.55	9.89	11.0	62.7	38.8	2.30	1.79
% contribution to the total SOA mass			8	13	9	12	35	20	3	2
2-MG <sup>c</sup> oligomers	119	<i>meso</i> -erythritol	2050	3170	9680	4500	1240	460	4170	11600
	221	citramalic acid	1170	2590	2330	2110	3840	1720	550	1000
	323	2-hydroxy-3-methylbutyric acid	630	970	430	470	2740	1320	70	160
	425	pimelic acid		280	290	260	1650	680		130
	527	pimelic acid					720	480		
total mass from 2-MG oligomers (μg/m <sup>3</sup> )			3.85	7.01	12.7	7.34	10.2	4.66	4.79	12.9
% contribution to the total SOA mass			5	10	11	8	6	2	7	18
<i>mono</i> -acetate oligomers	161	citramalic acid		40	100		90		110	70
	263	2-hydroxy-3-methylbutyric acid	680	1720	600	670	4070	1300	360	160
	365	pimelic acid	770	1890	820	1240	4830	1760	250	290
	467	pinic acid	340	450	180	420	3750	1310		130
	569	pinic acid		790			8600	2960		
	671	suberic acid monomethyl ester					450	360		
total mass from <i>mono</i> -acetate oligomers (μg/m <sup>3</sup> )			1.79	4.89	1.70	2.33	21.8	7.69	0.72	0.65
% contribution to the total SOA mass			2	7	2	2	12	4	1	1
<i>mono</i> -formate oligomers	147	<i>meso</i> -erythritol	200	380	11300		200			1370
	249	2-hydroxy-3-methylbutyric acid	460	1340		40	1970	810	60	
	351	2-hydroxy-3-methylbutyric acid	370	1000		60	2880	1390	30	
	453	pimelic acid	290	380			1800	710		
total mass from <i>mono</i> -formate oligomers (μg/m <sup>3</sup> )			1.32	3.10	11.3	0.10	6.85	2.91	0.09	1.37
% contribution to the total SOA mass			2	4	10	0.1	4	1	0.1	2
total mass identified (g/m <sup>3</sup> )			12.6	24.6	35.6	20.8	102	54.1	7.90	16.7
% of SOA identified			17	34	32	22	56	27	12	23

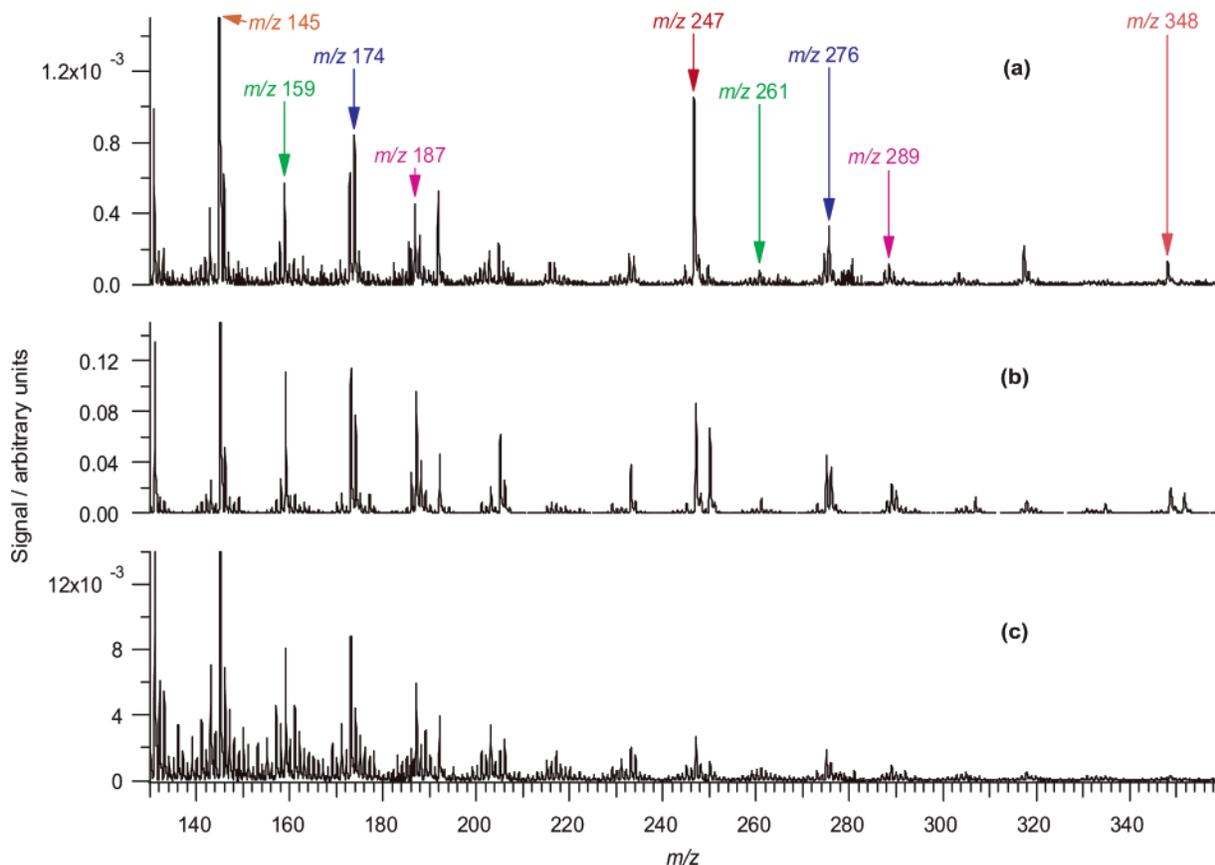
<sup>a</sup> Surrogate standards used covered the range of retention times for detected  $[M - H]^-$  ions. All standards used were within  $\pm 1.5$  minutes of retention times for sample  $[M - H]^-$  ions. <sup>b</sup> A blank cell indicates that the corresponding species was below the detection limit. <sup>c</sup> 2-MG = 2-methylglyceric acid.

in Figure 9. All ion signal intensities shown here are divided by the signal intensity of sulfate to correct for loss of particle mass to the chamber walls. Figure 9a shows the evolution of two prominent high-mass fragment ions  $m/z$  247 and 327. These high-mass fragment ions increase in abundance with time, with the increase in  $m/z$  327 being more significant. This increase is observed for all high-mass ( $m/z > 200$ ) fragment ions. Figure 9b shows the change in the intensity of the fragment ion  $m/z$  91, which is proposed to serve as a tracer ion for peroxides formed under low-NO<sub>x</sub> conditions, where the proposed formula for this fragment ion is C<sub>3</sub>H<sub>7</sub>O<sub>3</sub>, and the structure for one of its isomers is shown in Figure 9b. This peroxide tracer ion reaches its maximum signal after 7 h have elapsed in the experiment. Over the next 6 h, this ion decreases to a lower constant value; such a loss cannot be attributed to wall loss processes because the  $m/z$  91 signal has already been normalized to the sulfate signal. Figure 9c shows the time evolution of the organic mass from Experiment 13. The organic mass also decreases slightly after reaching its peak value; however, the decrease observed for the organic mass is much lower than that of the peroxide tracer ion ( $m/z$  91).

PILS/IC data were collected for some low-NO<sub>x</sub> experiments. Aerosol mass concentrations of acetate were much lower than those in the high-NO<sub>x</sub> case. For example, for Experiment 12, the acetate anion accounted for only 1.67 μg/m<sup>3</sup>, ~14–22 times lower than that of high-NO<sub>x</sub> levels. Formate anion was detected

at comparable mass concentrations to that of the high-NO<sub>x</sub> experiments (~1.51 μg/m<sup>3</sup>). Again, it should be noted that these two ions elute off the IC column immediately after sample injection and there is a possibility that other early-eluting monocarboxylic acid species coeluted with these two species, leading to an overestimate of their mass. No other organic anions were detected at significant levels from these low-NO<sub>x</sub> experiments.

Figure 10a shows a GC/MS TIC of a low-NO<sub>x</sub>, dry ammonium sulfate seeded experiment (Experiment 13). The chromatographic peaks at  $RT = 31.21$ , 32.25, and 32.61 min correspond to isomeric C<sub>5</sub> alkene triol species (*cis*-2-methyl-1,3,4-trihydroxy-1-butene, 3-methyl-2,3,4-trihydroxy-1-butene, *trans*-2-methyl-1,3,4-trihydroxy-1-butene, respectively), which have been measured previously in ambient aerosol from the Amazonian rainforest and Finnish boreal forests.<sup>11,12</sup> This is the first detection of these species in a controlled laboratory chamber experiment. The chromatographic peaks at  $RT$ s 38.22 and 38.97 min correspond to the 2-methyltetrols (2-methylthreitol and 2-methylerythritol, respectively), which also have been detected in ambient aerosol studies,<sup>7,10,11,13</sup> as well as in one previous photooxidation chamber study.<sup>14</sup> The C<sub>5</sub> alkene triols and 2-methyltetrols have received much attention in prior studies; the corresponding mass spectra for their respective chromatographic peaks can be found in Figure 1S (Supporting Information). GC-FID measurements were made to quantify the



**Figure 5.** TOF-AMS spectra collected at low vaporizer temperatures for the following high- $\text{NO}_x$  chamber experiments: (a) 50 ppb isoprene, 250 ppb  $\text{NO}_x$ ,  $\text{H}_2\text{O}_2$  as the OH precursor, no seed; (b) 500 ppb MACR, 800 ppb  $\text{NO}_x$ ,  $\text{H}_2\text{O}_2$  as the OH precursor, with seed; and (c) 500 ppb isoprene, HONO as the OH precursor, no seed. These spectra indicate that the OH precursor does not have a substantial effect on the chemistry observed, that MACR is an important SOA precursor from isoprene oxidation, and that the 102 Da differences observed in the offline mass spectrometry data are not a result of sample workup or ionization artifacts.

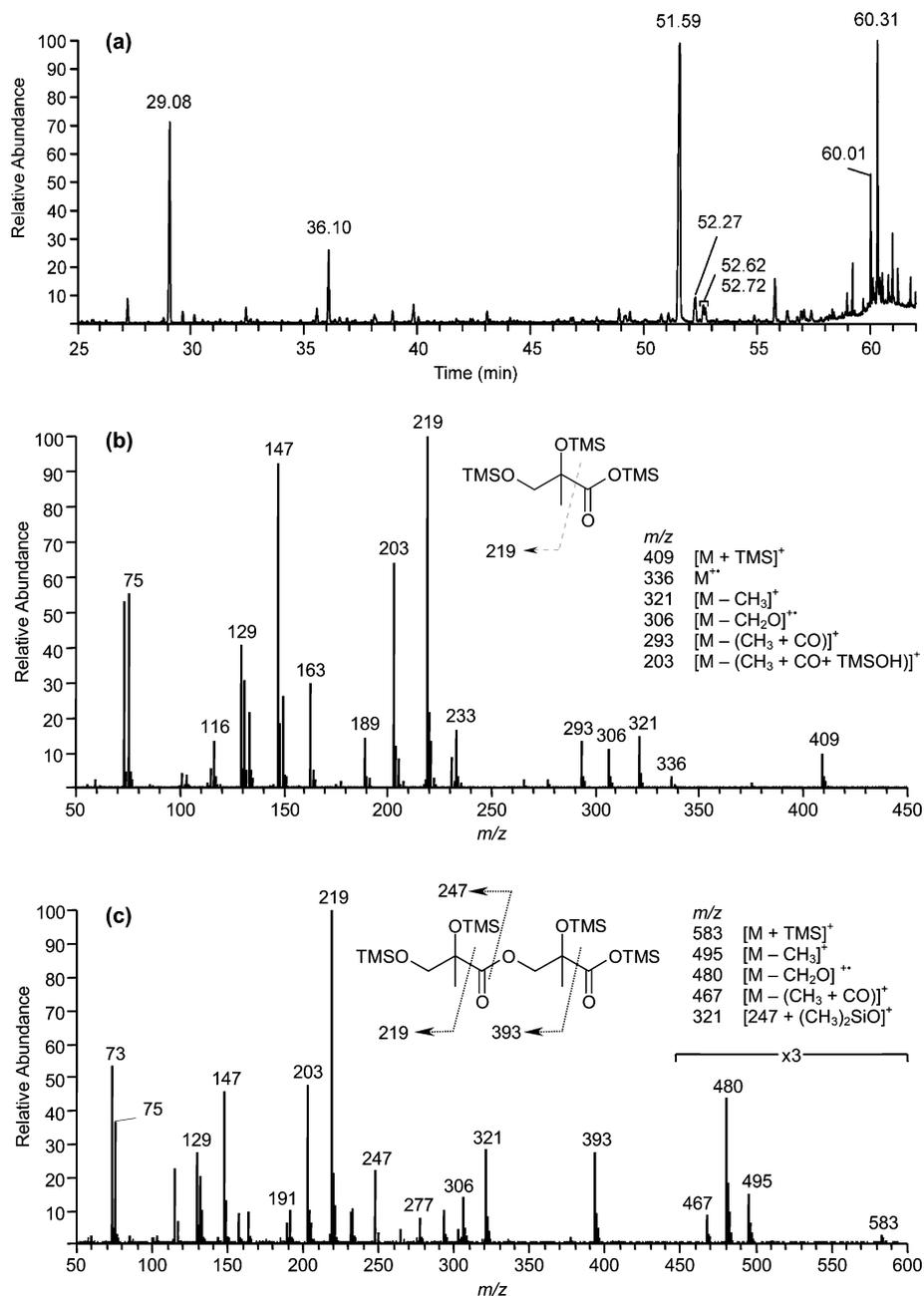
2-methyltetrols and  $\text{C}_5$  alkene triols for a low- $\text{NO}_x$  dry seeded experiment (Experiment 13—peaks in Figure 10a) and a low- $\text{NO}_x$  acid seeded experiment (Experiment 14). It was found that the 2-methyltetrols and  $\text{C}_5$  alkene triols accounted for 3.91% and 0.60% of the SOA mass, respectively, for the dry seeded experiment (Experiment 13), and decreased to 0.46% and 0.06% of the SOA mass, respectively, for the acid seeded experiment (Experiment 14). The inset shown in Figure 10a is the  $m/z$  219 EIC for six isomeric dimers (MW = 254) eluting between 58.8 and 59.2 min. The corresponding averaged EI mass spectrum for these chromatographic peaks is shown in Figure 10b. The general chemical structure of the trimethylsilylated dimer, along with its respective MS fragmentation, is also shown in this mass spectrum. The fragmentation pattern shown here indicates that the dimer forms by the reaction of a  $\text{C}_5$  alkene triol (indicated by the  $m/z$  335 fragment ion) with a 2-methyltetrol (indicated by the  $m/z$  219 fragment ion) to form the hemiacetal dimer shown. To confirm the MW of the isomeric hemiacetal dimers eluting between 58.8 and 59.2 min, an averaged CI( $\text{CH}_4$ ) mass spectrum was also collected and is shown in Figure 10c. The MW of the trimethylsilylated dimer (derivatized MW = 686) is confirmed by the  $[M + \text{H} - \text{CH}_4]^+$  ion at  $m/z$  671. The SOA products that elute at 34.91 and 35.47 min were tentatively characterized as diastereoisomeric 2-methyltetrol performate derivatives, which are unstable and upon reaction in the trimethylsilylation reagent mixture are converted into 2-methyltetrols. Their corresponding EI mass spectra can also be found in Figure 1S (Supporting Information). It should be noted that the peaks labeled \*1, \*2, and \*3 in the GC/MS TIC (Figure

10a) were also present in the laboratory controls and were identified as palmitic acid, stearic acid, and palmitoyl monoglyceride, respectively. Table 4 summarizes all low- $\text{NO}_x$  SOA components elucidated by GC/MS.

#### 4. Discussion

**4.1. Gas-Phase Chemistry.** Gas-phase oxidation of isoprene is dominated by the reaction with OH.<sup>16,17</sup> Under high- $\text{NO}_x$  conditions,  $\text{O}_3$  and  $\text{NO}_3$  radicals play only a minor role in the initial oxidation of isoprene because they form only once  $[\text{NO}]$  approaches zero, by which time most of the isoprene is consumed. Under low- $\text{NO}_x$  conditions,  $\text{O}_3$  and  $\text{NO}_3$  radicals also contribute negligibly to isoprene oxidation. Figure 11 shows the initial gas-phase reactions that occur under both low- and high- $\text{NO}_x$  conditions. In both cases, the initial oxidation of isoprene occurs by reaction with OH, followed by the immediate addition of  $\text{O}_2$  to form eight possible isomeric isoprene hydroperoxy ( $\text{RO}_2$ ) radicals (for simplicity, only three are shown).

Under high- $\text{NO}_x$  conditions, the isoprene hydroperoxy radicals react predominantly with  $\text{NO}$ ; however, they may also react with  $\text{NO}_2$  to form peroxy nitrates ( $\text{ROONO}_2$ , not shown in Figure 11), but these are likely unimportant to the formation of isoprene SOA because of their thermal instability.  $\text{RO}_2 + \text{NO}$  reactions result in the formation of either hydroxynitrates or hydroxyalkoxy (RO) radicals. Our observations of organic nitrates in high- $\text{NO}_x$  SOA as observed in Figure 1 ( $[M - \text{H}]^-$  ions with even  $m/z$  values) indicate that these hydroxynitrates



**Figure 6.** (a) TIC of a high- $\text{NO}_x$  isoprene nucleation experiment (Experiment 5) collected using GC/MS in the EI mode. (b) EI mass spectrum for the 2-MG residue ( $RT = 29.08$  min). (c) EI mass spectrum for a linear dimer made up of two 2-MG residues ( $RT = 51.59$  min). These two mass spectra confirm that 2-MG is present in high- $\text{NO}_x$  SOA and that it is involved in particle-phase esterification reactions resulting in polyesters (as shown by the dimer structure above).

are likely SOA precursors. Two of the hydroxyalkoxy radicals decompose into MVK and MACR, where their yields are 32–44% and 22–28%, respectively.<sup>32–35</sup> The remaining hydroxyalkoxy radical forms a 1,4-hydroxycarbonyl, which may isomerize and dehydrate to form 3-methylfuran.<sup>36</sup> SOA formation has been observed from the photooxidation of MACR and 3-methylfuran, indicating that these are SOA precursors (indicated by black boxes in Figure 11).<sup>17</sup> However, 3-methylfuran is not expected to contribute greatly to the SOA formed by isoprene oxidation because of its low gas-phase product yield (<2–5%).<sup>33–35</sup> The higher gas-phase product yields observed for MACR suggest that it is the most important SOA precursor from isoprene oxidation under high- $\text{NO}_x$  conditions; this is consistent with the similarities of the chemical products observed in isoprene and MACR SOA (Figure 1 and Table 2). Even though

MVK typically has the highest gas-phase product yield observed, it is not a contributor to SOA formation under high- $\text{NO}_x$  conditions because negligible amounts of aerosol were produced from the high- $\text{NO}_x$  photooxidation of 500 ppb MVK. Other products of isoprene oxidation under high- $\text{NO}_x$  conditions (not shown in Figure 11) include  $\text{C}_5$  hydroxycarbonyls,  $\text{C}_4$  hydroxycarbonyls, and  $\text{C}_5$  carbonyls; these may contribute to SOA formation, but experimental evidence is currently lacking.

Under low- $\text{NO}_x$  conditions, the isoprene hydroxyperoxy radicals react predominantly with  $\text{HO}_2$ . These reactions result in the formation of hydroxy hydroperoxides, which are highlighted in dotted boxes to indicate that these species are possible SOA precursors. Under similar reaction conditions, Miyoshi et al.<sup>32</sup> observed by IR spectroscopy that hydroperoxides are major gas-phase products from isoprene oxidation under  $\text{NO}_x$ -

TABLE 3: Low-NO<sub>x</sub> Chamber Experiments Conducted

expt no. <sup>a,b</sup>	seeded <sup>c</sup> / nucleation	[O <sub>3</sub> ] <sup>d</sup> ppb	T, °C <sup>d</sup>	total SOA mass concentration <sup>d,e,f</sup> μg/m <sup>3</sup>	peroxide aerosol mass concentration μg/m <sup>3</sup>	% contribution of peroxides to the SOA mass concentration observed
10 <sup>g</sup>	nucleation	32	29.1	186	116	62
11 <sup>g</sup>	dry AS	36	28.7	282	97	34
12	nucleation	b.d.l.	23.7	73	<sup>h</sup>	<sup>h</sup>
13	dry AS	12	24.1	69	<sup>h</sup>	<sup>h</sup>
14	AAS	b.d.l.	23.8	259	67	26
15	dry AS	11	23.9	73	19	25
16 <sup>i</sup>	dry AS	2	25.6	24	<sup>h</sup>	<sup>h</sup>
17	AAS	b.d.l.	23.6	93	23	24
18a	nucleation	7	26.2	55	32	59
18b <sup>j</sup>	nucleation	37	27.0	22	6	26

<sup>a</sup> All VOC gas-phase mixing ratios were 500 ppb, except for Experiment 17 (100 ppb). <sup>b</sup> H<sub>2</sub>O<sub>2</sub> was the OH precursor used for each low NO<sub>x</sub> isoprene experiment. H<sub>2</sub>O<sub>2</sub> is not measured directly, but from isoprene decay during irradiation we estimate ~3 ppm of H<sub>2</sub>O<sub>2</sub>. <sup>c</sup> AS = ammonium sulfate seed, AAS = acidic ammonium sulfate seed. <sup>d</sup> Averaged over the course of filter sampling. <sup>e</sup> Subtraction of seed aerosol taken into account when necessary. SOA volume derived from DMA wall loss uncorrected measurements for use in mass closure from filter sample analyses. <sup>f</sup> Assuming a SOA density of 1.25 g/cm<sup>3</sup>. This value is derived from comparison of DMA aerosol volume and AMS aerosol mass measurements. <sup>g</sup> 100% of light bank used and hence higher temperatures in chamber observed during sampling. <sup>h</sup> No peroxide measurement made for this sample. <sup>i</sup> Half of the typical [H<sub>2</sub>O<sub>2</sub>] used ~1.5 ppm. <sup>j</sup> Late sampling, after peak growth, during the rapid decay of the aerosol mass/volume typical of low NO<sub>x</sub> experiments.

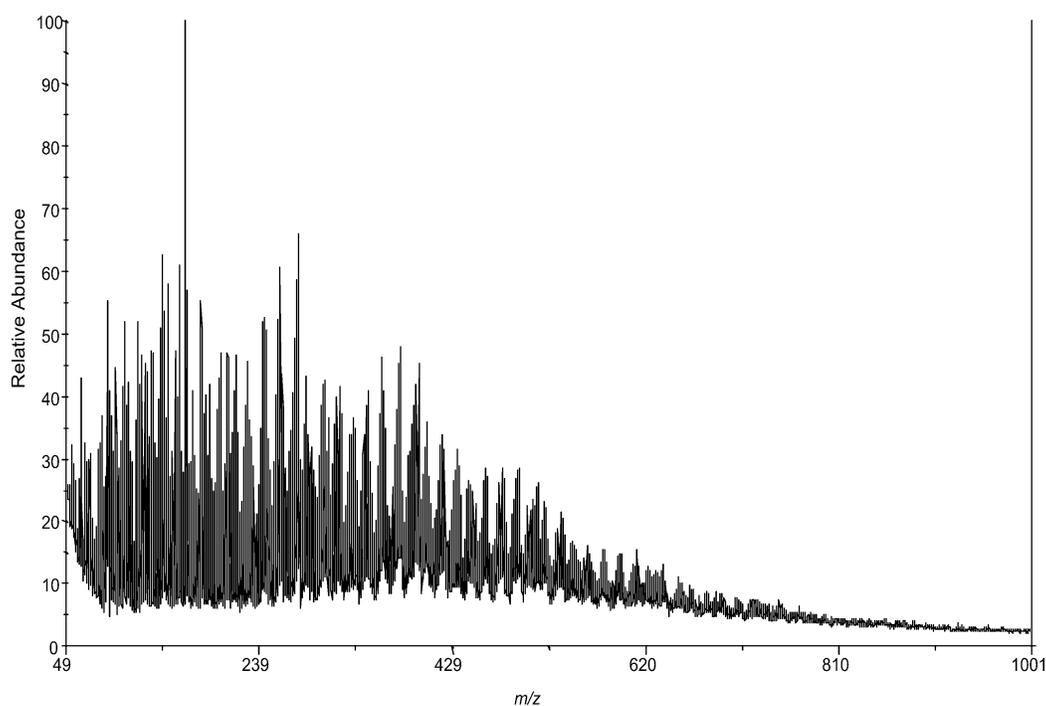
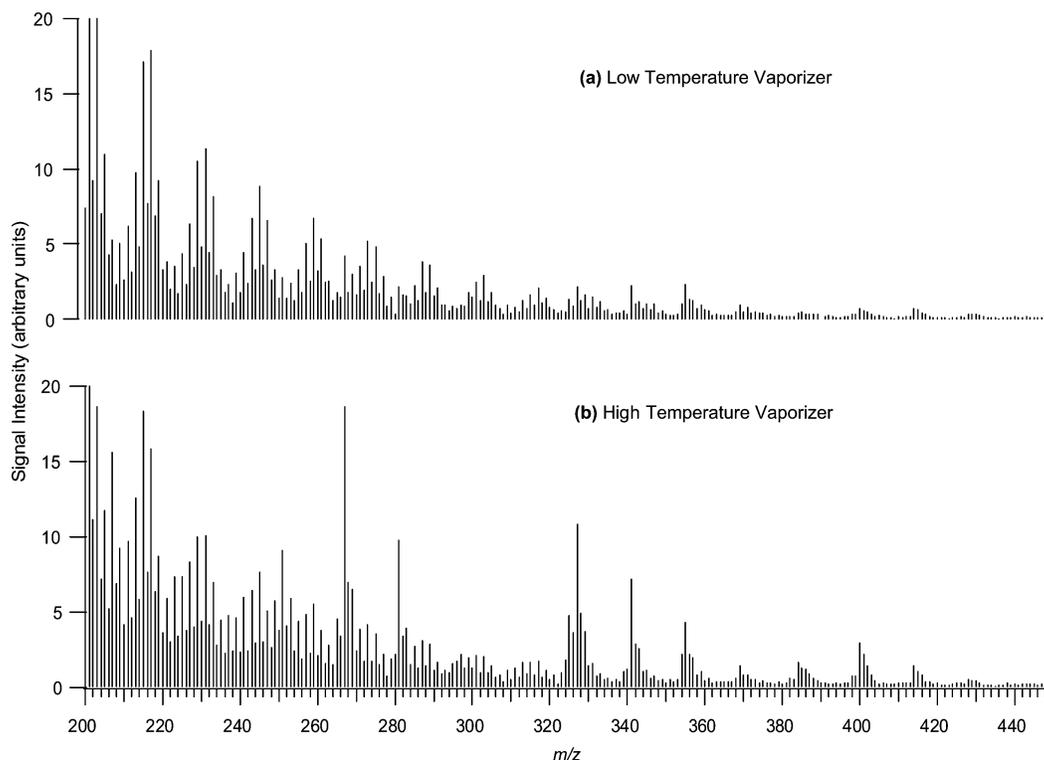


Figure 7. MALDI positive mode spectrum obtained with a graphite matrix for a 500 ppb isoprene, low-NO<sub>x</sub>, acid seeded experiment (Experiment 14). High-molecular mass species formed up to ~620 Da.

free conditions. Aerosol formation was also observed; however, the composition of the resultant aerosol was not investigated.

In contrast to Kroll et al.,<sup>16</sup> under the present conditions there may be some contribution (~10–30%) of RO<sub>2</sub> + RO<sub>2</sub> reactions under low-NO<sub>x</sub> conditions owing to the higher [isoprene]/[H<sub>2</sub>O<sub>2</sub>] ratios used in the current study.<sup>32</sup> For simplicity, only the RO<sub>2</sub> + RO<sub>2</sub> reactions that lead to hydroxyalkoxy radicals are shown in Figure 11. As in the high-NO<sub>x</sub> case, these hydroxyalkoxy radicals will likely form MVK, MACR, and hydroxycarbonyls. The RO<sub>2</sub> + RO<sub>2</sub> reactions not shown can lead to the formation of diols and other isomeric hydroxycarbonyls. As will be discussed subsequently, the diols that result from RO<sub>2</sub> + RO<sub>2</sub> reactions (not shown) may form SOA as well.<sup>15</sup>

**4.2. High-NO<sub>x</sub> SOA.** **4.2.1. Importance of MACR Oxidation.** MACR oxidation under high-NO<sub>x</sub> conditions produces significant amounts of SOA (Experiments 3 and 4). When comparing the SOA products from isoprene and MACR oxidation at high-NO<sub>x</sub> conditions, many of the same products are observed (Figure 1). Tandem MS data obtained for selected ions common to both isoprene and MACR samples, like the *m/z* 368 ion shown in Figure 4, produced similar product ion spectra, further indicating that these species are indeed the same. This observation is consistent with our previous proton-transfer reaction-mass spectrometry (PTR-MS) studies of isoprene oxidation, which demonstrate a strong correlation between the amount of SOA formed and the MACR reacted in the gas phase.<sup>37,38</sup> In these studies, aerosol growth continued well after isoprene was fully consumed, indicating the likely importance of second- (or later-)



**Figure 8.** TOF-AMS spectra for a 500 ppb isoprene low-NO<sub>x</sub> experiment (Experiment 12). (a) Mass spectrum obtained with a low temperature vaporizer (~150 °C). (b) Mass spectrum obtained with a high temperature vaporizer (~600 °C). The spectrum is richer at higher temperature with some prominent peaks at higher *m/z*, indicating that the high- MW oligomers that are not easily volatilized at <200 °C.

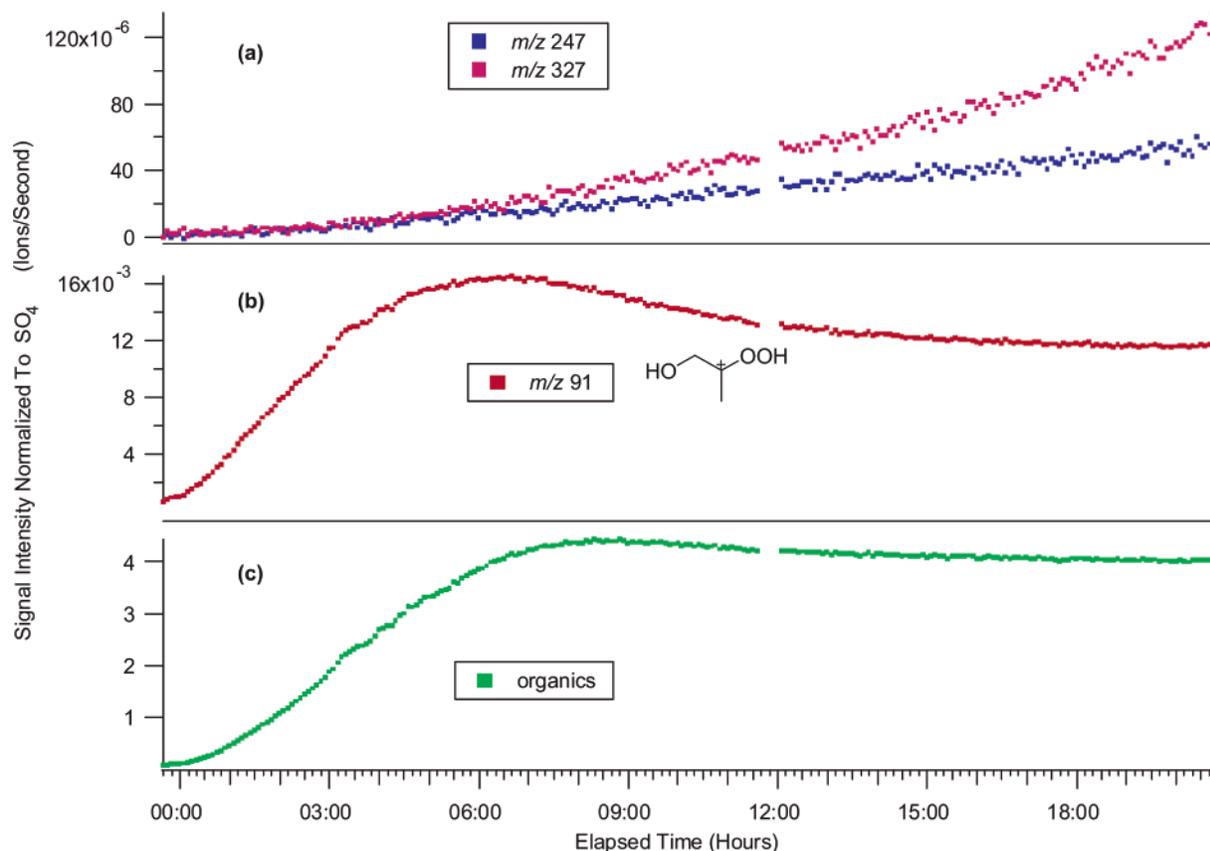
generation gas-phase products and/or heterogeneous (particle-phase) reactions. It should be noted that when the MACR, H<sub>2</sub>O<sub>2</sub>, and dry ammonium sulfate seed aerosol are well mixed in the chamber before irradiation begins, no aerosol growth is observed. This rules out the possibility of reactive uptake of MACR into the particle phase; instead the oxidation of MACR is a necessary step in SOA formation from the photooxidation of isoprene.

**4.2.2. Oligomers.** Oligomerization occurs in SOA formed under high-NO<sub>x</sub> conditions, where both offline and online mass spectrometry techniques (Figures 1, 2, and 5) measure species with much higher MWs than those of the parent isoprene, with characteristic 102 Da differences. Tandem MS techniques, such as upfront CID on the LC/MS instrument, confirm that oligomers are indeed formed from a common 102 Da monomeric unit. For example, when isolating the *m/z* 368 ion from the rest of the sample matrix and further fragmenting it to generate a product ion spectrum, two successive neutral losses of 102 Da were observed at *m/z* 266 and 164 (Figure 4b). Two isomeric compounds with *m/z* 266 in Figure 3c were found to elute off the LC column at ~2.5–3 min earlier than the compound with *m/z* 368 studied here. The fact that the compounds with *m/z* 266 ions elute off the LC column at earlier *RT*s, and that *m/z* 266 is a product ion of *m/z* 368, strongly suggests that these two ions are characteristic of the same oligomeric series. The compounds characterized by *m/z* 368 and 266 are likely a trimer and dimer, respectively. The other series of oligomers quantified in Table 2 also had 102 Da differences observed and similar LC/MS behaviors, with ions with lower mass eluting from the LC column at earlier *RT*s.

**4.2.3. Organic Nitrates.** Organic nitrates, detected as even-mass [*M* – H]<sup>–</sup> ions in (–)ESI spectra, were measured in all high-NO<sub>x</sub> experiments. All organic nitrates detected in high-NO<sub>x</sub> SOA samples had similar product ion spectra as *m/z* 368

(Figure 4b), with neutral losses of 63 (HNO<sub>3</sub>), 77 (CH<sub>3</sub> radical + NO<sub>3</sub> radical, possibly CH<sub>3</sub>NO<sub>3</sub>), and 102 Da, suggesting that all even-mass [*M* – H]<sup>–</sup> ions are oligomeric organic nitrate species. Unlike the (–)ESI techniques (Figures 1 and 3), the GC/MS technique did not allow for the detection of organic nitrate species, likely a result of their instability at the high temperature of the GC injector and/or derivatization techniques used during sample workup. Organic nitrates were also not clearly detected in the MALDI-TOFMS (Figure 2) and TOF-AMS (Figure 5) instruments. This is likely a result of the harsh ionization techniques employed by these instruments. Even with (–)ESI, these organic nitrates were not completely stable, as shown in Figure 4a for the *m/z* 368 ion. Organic nitrates found in the high-NO<sub>x</sub> SOA likely form from the further oxidation of the hydroxynitrate species found in the gas phase from RO<sub>2</sub> + NO reactions.

**4.2.4. 2-MG as Monomeric Units.** As shown in Table 2, other varieties of oligomers were observed as well. From further use of tandem MS techniques, it was found that one of these series of oligomers likely involved 2-MG (2-methylglyceric acid), a recently discovered SOA tracer compound for isoprene oxidation in the ambient atmosphere,<sup>10,13,14</sup> as an important monomer. Confirmation of the 2-MG monomer in high-NO<sub>x</sub> SOA was provided by GC/MS with TMS derivatization (Figure 6a and b). Because monomeric 2-MG is small and polar, it was not retained effectively by the LC reverse phase column (*RT* ≈ 1.3 min) and was detected in its deprotonated form at *m/z* 119. Figure 12 shows product ion spectra obtained with (–)ESI-ITMS for Experiment 9. In Figure 12a, the *m/z* 323 ion is isolated in the ion trap from the rest of the ion matrix and is collisionally activated to produce the MS<sup>2</sup> spectrum shown here. The *m/z* 221 ion is the base peak in this spectrum, and the *m/z* 119 ion also detected as the result of further fragmentation of the *m/z* 221 product ion. The fact that the *m/z* 119 ion was



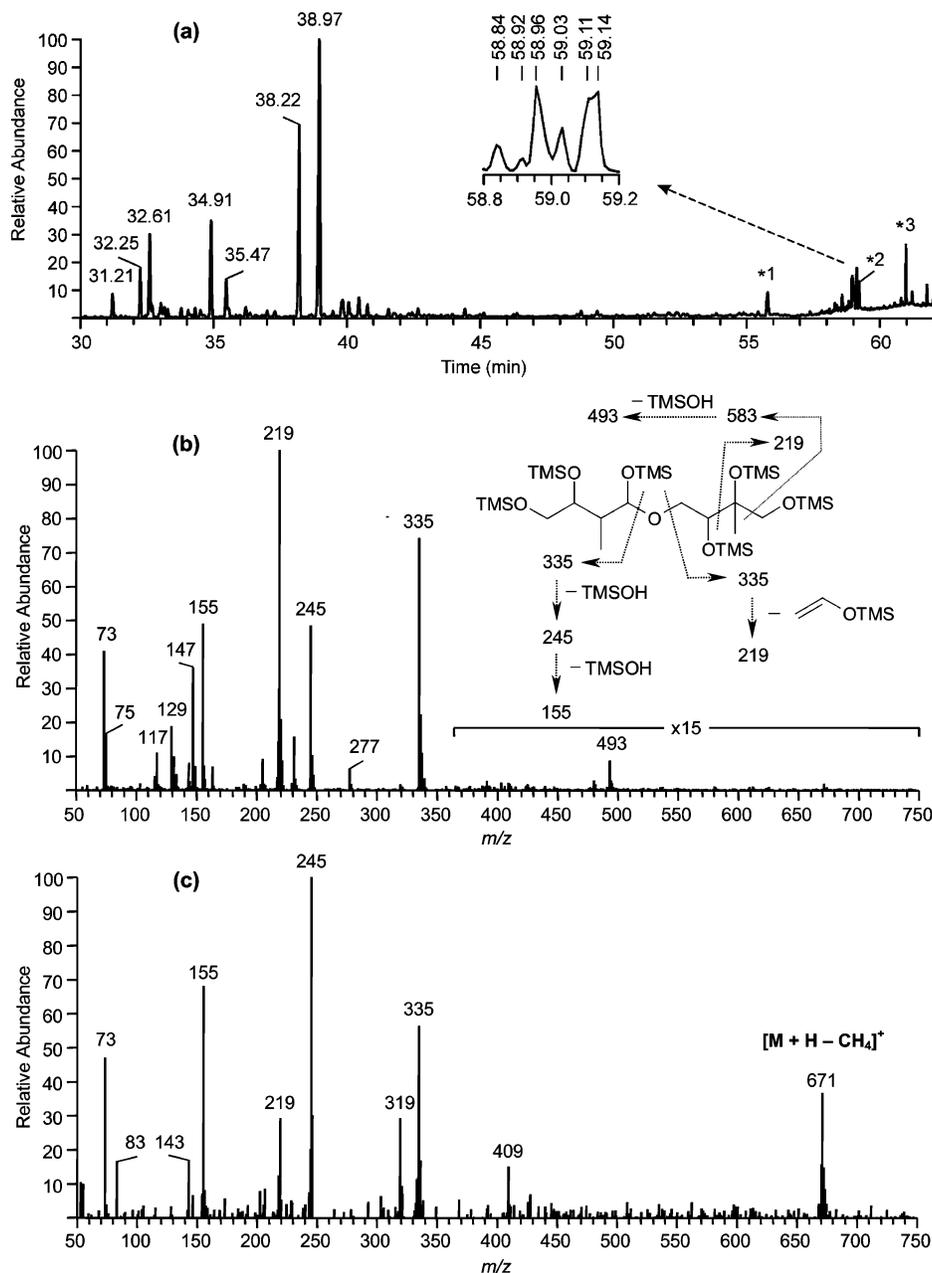
**Figure 9.** Time evolution plots produced from the TOF-AMS instrument for selected fragment ions and the total organic mass observed from a typical low- $\text{NO}_x$  experiment (Experiment 13). All ion signal intensities are divided by the signal intensity of sulfate. Because sulfate concentration is a tracer for wall loss (neither created nor removed during the run), the ratio of ion signal to sulfate signal should give an indication of the behavior without wall loss. (a) Time evolution plot for high-mass fragment ions  $m/z$  247 and 327. (b) Time evolution plot for the proposed peroxide fragment ion  $m/z$  91 ( $\text{C}_3\text{H}_7\text{O}_3$ ), where the structure of one isomer is shown. (c) Time evolution plot for the total organic mass. These plots indicate that the chemical composition changes with experimental time, where the decomposition of organic peroxides correlates to oligomerization within low- $\text{NO}_x$  SOA. The missing data points (11:30 to 12:00 hours) in these plots are due to the vaporizer in the TOF-AMS instrument being turned off.

detected as a product ion in the  $\text{MS}^2$  and  $\text{MS}^3$  spectra shown in Figure 12 strongly suggests that 2-MG is a monomer in this oligomeric series. It is important to note that  $m/z$  119 was also a fragment ion produced in the upfront CID spectrum for the  $m/z$  368 ion in Figure 4b. It was found that  $m/z$  119 was a common product ion to each oligomeric series, suggesting the importance of 2-MG in oligomerization reactions.

**4.2.5. Mono-Acetate and Mono-Formate Oligomers.** The PILS/IC measurements of high levels of particulate acetate and formate anions in both the seeded (Experiment 1) and nucleation (Experiment 2) experiments, coupled with the high volatilities of their acid forms produced in the gas phase from the oxidation of isoprene, suggests that these compounds resulted from the decomposition of oligomeric SOA. The formation of *mono*-acetate and *mono*-formate oligomers was observed by tandem (-)ESI-MS measurements. Figure 13 shows two product ion spectra for a *mono*-acetate dimer ( $[M - \text{H}]^-$  at  $m/z$  161) and *mono*-formate trimer ( $[M - \text{H}]^-$  at  $m/z$  249), respectively. The observation of a neutral loss of 42 Da (ketene,  $\text{H}_2\text{C}=\text{C}=\text{O}$ ) and a dominant product ion  $m/z$  59 (acetate anion) in the  $\text{MS}^2$  spectrum of the  $m/z$  161 ion (Figure 13a), provides strong evidence for acetylation. In the  $\text{MS}^2$  spectrum of the  $m/z$  249 ion (Figure 13b), the major product ion  $m/z$  147 results from the common neutral loss of 102 Da. The product ion  $m/z$  221 results from a neutral loss of 28 Da (CO), a rearrangement reaction that is characteristic of formates. The product ion  $m/z$  119 (deprotonated 2-MG) resulting from the combined neutral losses of 102 and 28 Da is also observed. *Mono*-acetate

oligomers were also detected by the GC/MS TMS derivatization method; the details of these findings will be discussed in a forthcoming GC/MS complementary paper.

**4.2.6. Heterogeneous Esterification Reactions.** Oligomer species containing the  $m/z$  119, 221, and 323 ions as detected by the (-)ESI techniques were also observed by GC/MS as their respective TMS derivatives as shown Figure 6a. As in previous measurements of 2-MG,<sup>14</sup> the EI mass spectrum shown in Figure 6b confirms the formation of monomeric 2-MG in high- $\text{NO}_x$  isoprene SOA. The dimer detected at  $m/z$  221 by (-)ESI techniques (as shown in Figure 12b) involving 2-MG as an important monomer, is detected at 51.59 min in Figure 6a. The chemical structure of this species likely contains one carboxyl and three hydroxyl groups, as shown in Figure 6c. The formation of an ester linkage is also denoted in this structure, which is the expected site of oligomerization. The ions  $m/z$  583 ( $[M + \text{TMS}]^+$ ) and  $m/z$  495 ( $[M - \text{CH}_3]^+$ ) confirm that the MW of this dimer species is 222 (which is also in agreement with the ESI results). The ion  $m/z$  467 ( $[M - (\text{CH}_3 + \text{CO})]^+$ ) is consistent with a terminal trimethylsilylated carboxylic group, while the ion  $m/z$  480 ( $[M - \text{CH}_2\text{O}]^+$ ) is explained by a rearrangement of a trimethylsilyl group and points to a terminal trimethylsilylated hydroxymethyl group. The elemental composition ( $\text{C}_8\text{H}_{14}\text{O}_7$ ) of the structure shown in Figure 6c was also confirmed by high-resolution ESI-TOFMS measurements. These results strongly suggest that particle-phase esterification reactions occurred between 2-MG molecules, where a hydroxyl group of one 2-MG molecule reacted with a

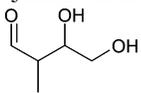
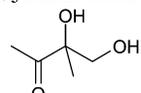
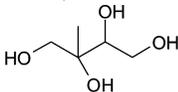
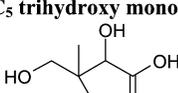
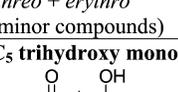
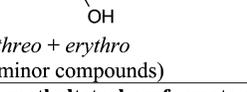
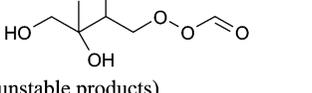


**Figure 10.** (a) GC/MS TIC of isoprene low- $\text{NO}_x$  SOA. The inset shows the  $m/z$  219 EIC for the dimeric products eluting between 58.8 and 59.2 min. Peak identifications:  $RT$ s 31.21, 32.25, and 32.61 min:  $\text{C}_5$  alkene triols;  $RT$ s 34.91 and 35.47 min: unstable products tentatively characterized as 2-methyltetrol performatate derivatives;  $RT$ s 38.22 and 38.97 min: 2-methyltetrols (2-methylthreitol and 2-methylerythritol, respectively). The EI spectra for the latter seven compounds are provided in Figure 1S (Supporting Information). The peaks labeled \*1, \*2, and \*3 were also present in the laboratory controls and were identified as palmitic acid, stearic acid, and palmitoyl monoglyceride, respectively. (b) Averaged EI spectrum for the dimeric products eluting between 58.8 and 59.2 min and fragmentation scheme; and (c) averaged  $\text{CI}(\text{CH}_4)$  spectrum for the latter products.

carboxylic acid group of another one. The products that result from this reaction would be the ester compound shown in Figure 6c and a water molecule. The neutral loss of 102 Da, likely corresponding to dehydrated 2-MG or a 2-MG residue in the form of a lactone (i.e., 2-hydroxy-2-methylpropiolactone), observed from the ESI-MS/MS techniques can be explained by the charge-directed nucleophilic reaction shown in Figure 14. To our knowledge, this is the first evidence of particle-phase esterification reactions in SOA. It should be noted that the mass spectra, not shown here, for the chromatographic peaks in Figure 6a at 60.01 and 60.31 min, correspond to branched and linear 2-MG acid trimers (corresponding to MW = 324), respectively. A detailed discussion of the EI mass spectral behavior of the TMS derivatives of 2-MG and 2-MG dimer and trimers will be presented in a complimentary GC/MS study.

Figure 15a and b compares the GC/MS EICs, using the  $m/z$  219 ion as the base peak, for a filter sample from Experiment 5 treated with trimethylsilylation only to that of a filter sample (also from Experiment 5) treated by hydrolysis/ethylation + trimethylsilylation, respectively, to show further confirmation of polyesters formed via esterification reactions between 2-MG molecules. When treating SOA from the same chamber experiment with the hydrolysis/ethylation procedure, a noticeable decrease in 2-MG and 2-MG oligomers is observed. For example, the peaks at 29.08, 51.59, and 60.31 min (Figure 15a) observed after trimethylsilylation appear as smaller peaks upon the hydrolysis/ethylation experiment, as shown in the second chromatogram (Figure 15b). This decrease is a result of the formation of ethyl esters of 2-MG and of linear dimer ( $RT$ s = 27.42 and 50.48 min, respectively). The mass spectra confirming

TABLE 4: Low-NO<sub>x</sub> Isoprene SOA Products Elucidated by GC/MS

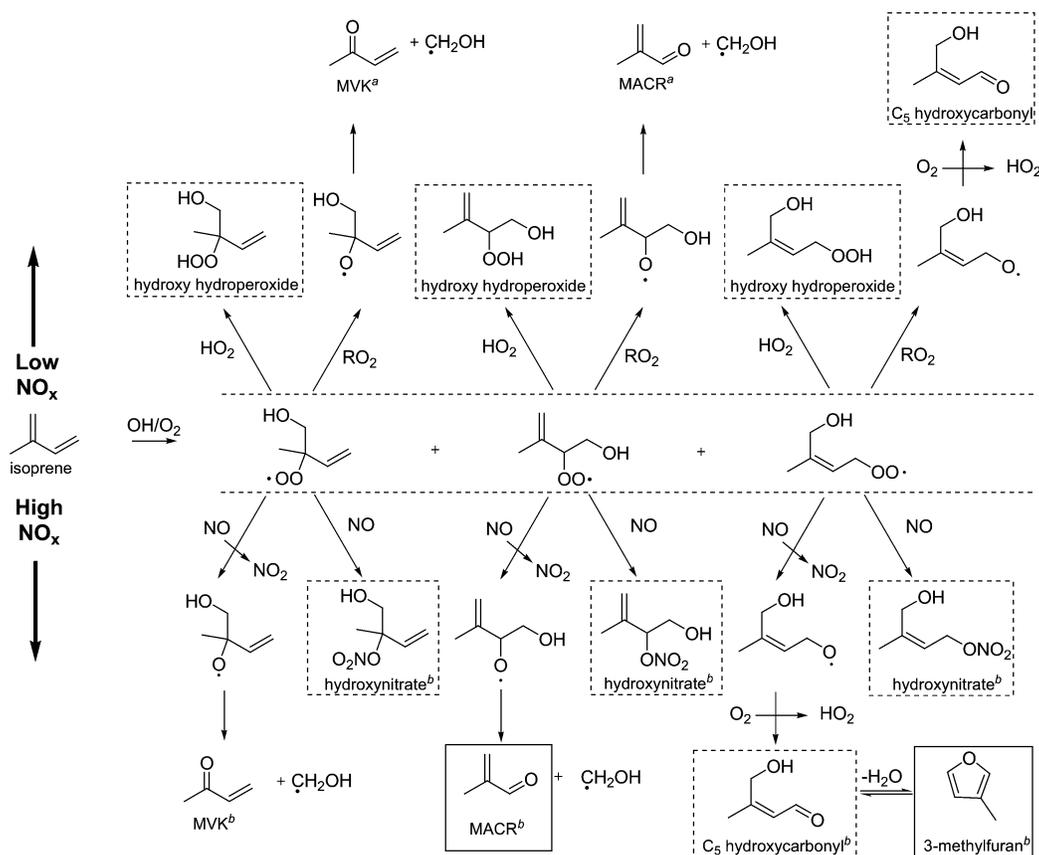
Compound / Structure	MW (MW TMS- derivative)	Elemental composition	Detection in Ambient Atmospheres
<b>C<sub>5</sub> alkene triols / ald form</b> 	118 (334)	C <sub>5</sub> H <sub>10</sub> O <sub>3</sub>	[Wang et al., 2005] <sup>12</sup> [Kourtchev et al., 2005] <sup>11</sup>
<b>C<sub>5</sub> alkene triols / keto form</b> 	118 (334)	C <sub>5</sub> H <sub>10</sub> O <sub>3</sub>	[Wang et al., 2005] <sup>12</sup> [Kourtchev et al., 2005] <sup>11</sup>
<b>2-methyltetrols</b>  <i>threo + erythro</i>	136 (424)	C <sub>5</sub> H <sub>12</sub> O <sub>4</sub>	[Claeys et al., 2004] <sup>7</sup> [Edney et al., 2005] <sup>14</sup> [Böge et al., 2006] <sup>15</sup> [Ion et al., 2005] <sup>10</sup> [Kourtchev et al., 2005] <sup>11</sup>
<b>C<sub>5</sub> trihydroxy monocarboxylic acid</b>  <i>threo + erythro</i> (minor compounds)	150 (438)	C <sub>5</sub> H <sub>10</sub> O <sub>5</sub>	Not yet detected in ambient aerosol
<b>C<sub>5</sub> trihydroxy monocarboxylic acid</b>  <i>threo + erythro</i> (minor compounds)	150 (438)	C <sub>5</sub> H <sub>10</sub> O <sub>5</sub>	Not yet detected in ambient aerosol
<b>2-methyltetrol performate derivatives</b>  (unstable products)	180 (396)	C <sub>6</sub> H <sub>12</sub> O <sub>6</sub>	Not yet detected in ambient aerosol
<b>Dimers (6 isomers)</b>  (minor compounds)	254 (686)	C <sub>10</sub> H <sub>22</sub> O <sub>7</sub>	Detected in ambient aerosol for the first time in this study

the formation of these ethyl ester species are shown in Figure 15c and d, respectively. The  $m/z$  365 and 277 ions in Figure 15c confirm the MW of the ethyl ester of 2-MG to be 148, where its formation is the result of polyesters decomposing into this derivatized monomer. The detection of  $m/z$  539 and 451 in Figure 15d confirm the MW of the ethyl ester of the linear 2-MG dimer, likely a result of the incomplete decomposition of larger polyesters (i.e., trimers, tetramers, pentamers, etc.) in high-NO<sub>x</sub> SOA.

Figure 16 shows the overall proposed reaction mechanism for SOA formation from the photooxidation of isoprene under high-NO<sub>x</sub> conditions. This figure denotes important initial gas-phase and particle-phase reactions that lead to the observed SOA products. As was discussed earlier, further gas-phase oxidation of MACR is required in order to form SOA from isoprene under high-NO<sub>x</sub> conditions. Oligomeric organic nitrates, such as the  $m/z$  368 ion, are comprised of an organic nitrate monomer, which is detected as the deprotonated  $m/z$  164 product ion (Figure 4b); therefore, it is possible that one gas-phase product of MACR oxidation is its hydroxynitrate form, as shown in Figure 16. Through further oxidation of the aldehyde group in this hydroxynitrate species, it is expected that the acid form of

this species results in the particle phase, thus being available for esterification reactions with 2-MG (Reaction 2 in Figure 16). To our knowledge, no organic nitrates have been measured in the gas phase from MACR oxidation (though nitrate formation has been inferred from OH-methacrolein reaction kinetics<sup>39</sup>); however, the detection of organic nitrates in the particle phase suggests that this is possibly a minor channel for SOA formation. However, the formation of 2-MG from the oxidation of MACR is still uncertain because of the unknown intermediates leading to its formation. Recently, it was proposed that 2-MG forms from the reaction of methacrylic acid or MACR with H<sub>2</sub>O<sub>2</sub> in the liquid aerosol phase under acidic conditions.<sup>13</sup> No aerosol growth was observed for MACR, H<sub>2</sub>O<sub>2</sub>, and dry ammonium sulfate seed aerosol under dark conditions; however, it is possible that other products such as formic and acetic acid, as well as oxidants formed during isoprene photooxidation, may promote the reactive uptake of MACR into the aerosol phase. Further measurements of MACR oxidation products are needed in order to better understand the formation of 2-MG, which might occur in either the particle or gas phase.

From our detailed analytical measurements discussed above, the importance of 2-MG to particle-phase reactions in high-

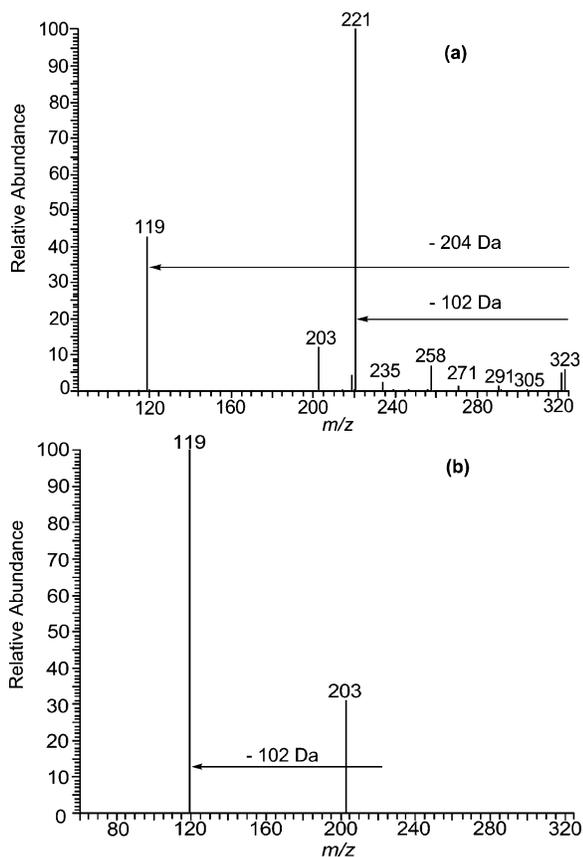


**Figure 11.** Reaction mechanism of isoprene oxidation under low- and high- $\text{NO}_x$  conditions. Dotted boxes indicate possible SOA precursors, whereas black boxes indicate known SOA precursors. For simplicity, only three of the eight initial isoprene hydroxyperoxy ( $\text{RO}_2$ ) radicals are shown.  $\text{RO}_2 + \text{RO}_2$  reactions leading to diols and other hydroxycarbonyls have been omitted for simplicity. <sup>a</sup>Miyoshi et al.<sup>31</sup> showed that  $[\text{isoprene}]/[\text{H}_2\text{O}_2]$  determines molar yields of MVK, MACR, and formaldehyde under low- $\text{NO}_x$  conditions. <sup>b</sup>Kroll et al.<sup>16</sup> summarized molar yields of gas-phase products from isoprene oxidation under high- $\text{NO}_x$  conditions reported in the literature.

$\text{NO}_x$  SOA is now well established. 2-MG monomers can react intermolecularly via esterification to produce 2-MG oligomers (Reaction 1), or react with *mono*-nitrate monomers to produce *mono*-nitrate oligomers (Reaction 2), or react with acetic or formic acid to produce *mono*-acetate and *mono*-formate oligomers, respectively (Reactions 3 and 4). These proposed esterification reactions are equilibrium reactions, and as a result, the addition of an acid or removal of water could promote the formation of these esters. As stated earlier, the high- $\text{NO}_x$  experiments were conducted at very low relative humidities ( $RH < 5\%$ ); therefore, this condition could allow for the ester formation we observe. We also observe high concentrations of organic acids (2-methylglyceric, acetic, and formic acid) at high- $\text{NO}_x$  conditions, which could provide the acidity needed to drive these reactions. It has been shown<sup>40</sup> that heterogeneous esterification of polyols by vapor-phase treatment with acetic acid and trifluoroacetic anhydride (used as an alternative to the sulfuric acid catalyst) will occur at room temperature without the use of liquids. Thus, it is reasonable to infer that esterification reactions may occur under the dry, room-temperature conditions of our chamber experiments. It should be noted that there is also evidence from the TOF-AMS that supports this reaction mechanism. The ratio of the TOF-AMS ion signals associated with the 2-MG dimer ( $m/z$  205) to that of the 2-MG monomer ( $m/z$  103) increases during the course of the high- $\text{NO}_x$  experiments, therefore providing additional confirmation of our proposed reaction mechanism in Figure 16. These results from the TOF-AMS, however, are not quantitative because of the majority of these molecules being fragmented (thermally or by the electron impact ionization) to smaller ions.

In comparison to MACR oxidation, the further oxidation of MVK likely does not produce SOA under high- $\text{NO}_x$  conditions because of its ketone moiety. The lack of an aldehydic hydrogen precludes the formation of acidic products (like that of 2-MG from MACR oxidation), which are necessary components needed for the particle-phase esterification reactions (Figure 16). One of the most abundant gas-phase products produced from MVK oxidation under high- $\text{NO}_x$  conditions is methylglyoxal. It was shown in a prior chamber study by Kroll et al.<sup>41</sup> that methylglyoxal does not reactively uptake onto inorganic seed aerosol; therefore, this could explain the lack of SOA growth from the further oxidation of MVK.

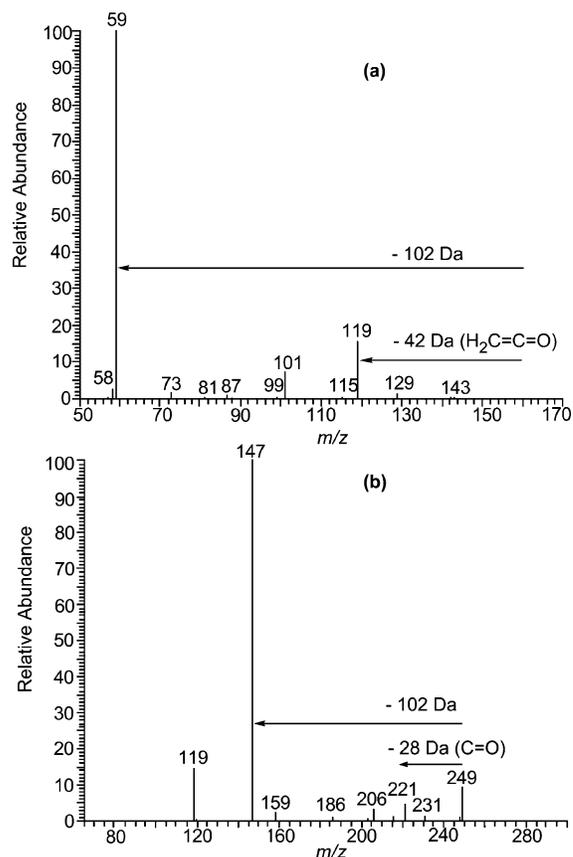
For the isoprene/ $\text{H}_2\text{O}_2$  experiments, except for Experiment 6, the most abundant oligomer series was the *mono*-nitrate oligomers (Table 2). The *mono*-nitrate oligomers accounted for  $\sim 8$ – $13\%$  of the SOA mass formed in these experiments. As for the isoprene/ $\text{H}_2\text{O}_2$  experiments, the *mono*-nitrate oligomers were the most abundant oligomers for the MACR/ $\text{H}_2\text{O}_2$  experiments ( $\sim 35\%$  of SOA mass for the seeded experiment versus  $\sim 20\%$  for the nucleation experiment). Even though most of the chemical products are the same in the  $\text{H}_2\text{O}_2$  and HONO experiments, the abundances of these products are different. In contrast to the  $\text{H}_2\text{O}_2$  experiments, the 2-MG oligomers are the most abundant oligomers for the HONO experiments. These differences could be due to different  $\text{NO}_x$  levels. SOA mass closure was observed to be the highest for the MACR/ $\text{H}_2\text{O}_2$ /seeded experiment ( $\sim 57\%$  of SOA identified) and the isoprene/ $\text{H}_2\text{O}_2$ /seeded experiments (22–34% identified). It is important to stress that the organic aerosol mass loadings formed in these isoprene high- $\text{NO}_x$  chamber experiments ( $\sim 50$ – $200 \mu\text{g m}^{-3}$ )



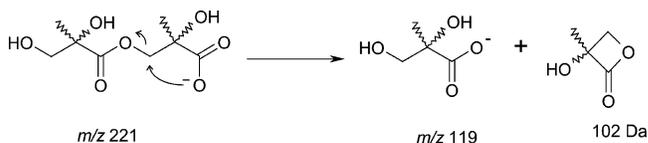
**Figure 12.** ESI-ITMS negative mode product ion spectra from a high- $\text{NO}_x$  isoprene SOA sample (Experiment 9). (a)  $\text{MS}^2$  spectrum for an isolated  $m/z$  323 ion. Two neutral losses of 102 Da are observed as shown by the product ions  $m/z$  221 and 119. (b)  $\text{MS}^3$  spectrum for an isolated  $m/z$  323 ion generated from the further fragmentation of the dominant daughter ion ( $= m/z$  221) in the  $\text{MS}^2$  spectrum. These spectra indicate that 2-MG ( $[M - \text{H}]^-$  ion  $= m/z$  119) is a monomer for the oligomeric  $m/z$  323 ion.

are much higher than those found in ambient aerosol where isoprene emissions are the highest ( $\sim 5 \mu\text{g m}^{-3}$ ). The amount of organic aerosol mass controls the gas-particle partitioning of semi-volatile species produced from the oxidation of hydrocarbons because more organic aerosol mass allows for more uptake of these species; therefore, the mass closure results presented apply only to the aerosol mass loadings produced in this current study and are not absolute for the isoprene system. The key insight from our analysis is the detection of these various oligomeric products formed from particle-phase esterification reactions.

**4.2.7. Additional Routes for SOA Formation.** As shown in Table 2, the polyester products from oligomerization of 2-MG and related components account only for a portion (22–34%) of the SOA formed from isoprene oxidation under high- $\text{NO}_x$  conditions. This lack of mass closure could result from the LC/MS technique underestimating the amount of polyesters, possibly related to the use of a  $\text{C}_{18}$  reverse phase column and the unavailability of authentic standards. The  $\text{C}_{18}$  reverse phase column could have degraded the oligomers into smaller units as they pass through the column, or very large oligomers could have permanently been retained onto the reverse phase material, and hence were not detected. Negative bias associated with filter sampling, such as evaporative losses during sampling or storage, could also be a source of incomplete mass closure. The presence of acetic acid in eluent mixture used for the LC/MS runs could also have caused an underestimation of the oligomers formed



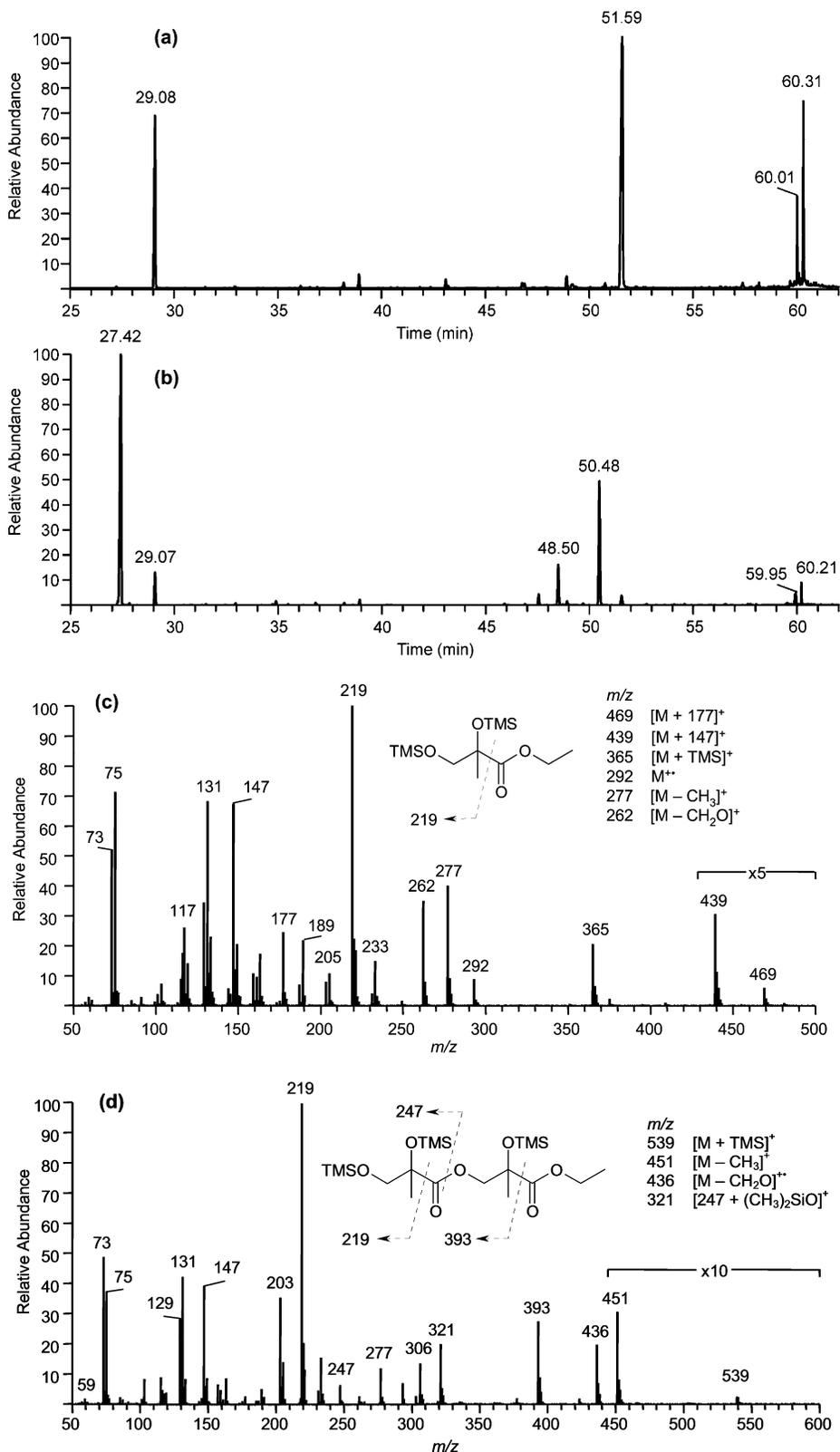
**Figure 13.** ESI-ITMS negative mode product ion mass spectra providing evidence for *mono*-acetate and *mono*-formate oligomers in high- $\text{NO}_x$  SOA. (a) Product ion mass spectrum for a *mono*-acetate dimer ( $m/z$  161). (b) Product ion mass spectrum for a *mono*-formate trimer ( $m/z$  249).



**Figure 14.** Proposed charge-directed nucleophilic reaction occurring during collisional activation in  $(-)$ ESI-ITMS, explaining the observation of 102 Da (2-hydroxy-2 methylpropiolactone) losses from oligomeric high- $\text{NO}_x$  SOA.

due to the possibility of acid-catalyzed hydrolysis during ESI, a process that would lead to a decrease in the detection of oligomeric compounds. Besides possible errors associated with quantifying esterification products identified by the LC/MS technique, the possibility still exists that other unidentified second- (or later-) generation gas- or particle-phase products from isoprene oxidation contribute to SOA formation, and as a result, would increase the mass closure significantly.

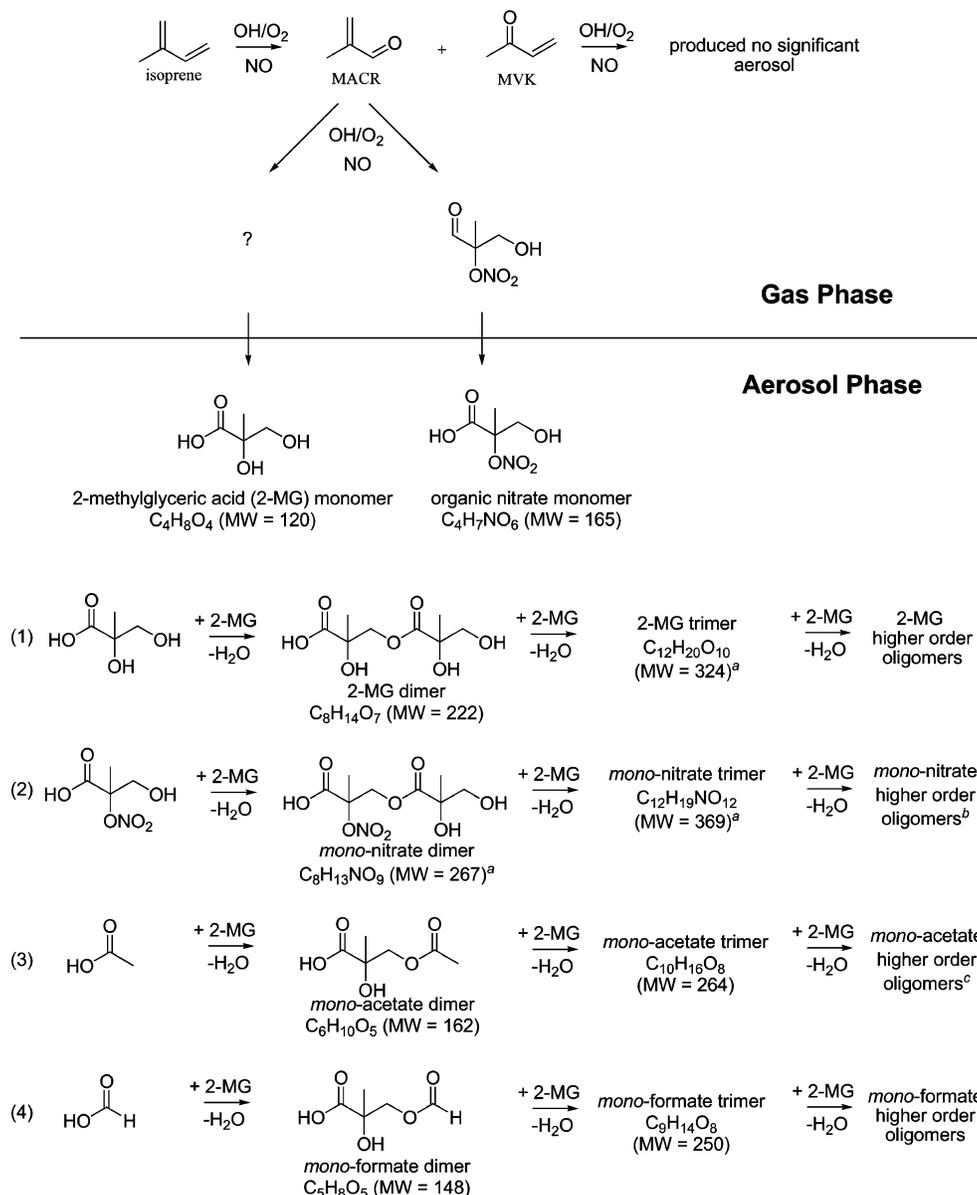
Glyoxal, a  $\text{C}_2$  dialdehyde, has been recently shown to be reactively taken up into particulate matter,<sup>41,42</sup> however, not at the low  $RH$ s employed in this study ( $RH < 5\%$ ). Theoretically, it has been shown that this reactive uptake of glyoxal results from thermodynamically favorable hydration and oligomerization.<sup>43,44</sup> When first interpreting the MS data from the ESI and MALDI techniques, it was considered that a dialdehyde species possibly corresponded to the 102 Da neutral losses observed from the oligomeric components. Figure 17 shows a proposed gas-phase reaction scheme for the formation of a  $\text{C}_4$  hydroxy dialdehyde species ( $\text{MW} = 102$ ) from the further oxidation of MACR. In contrast to glyoxal, dissolution may not be required



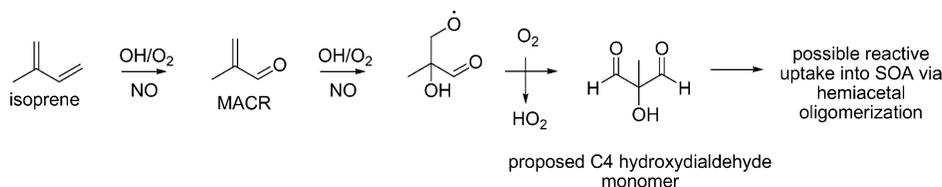
**Figure 15.** (a) GC/MS EIC ( $= m/z$  219) for high- $\text{NO}_x$  isoprene nucleation sample (Experiment 5) treated only with TMS derivatization. (b) GC/MS EIC ( $= m/z$  219) for a duplicate sample of the same experiment (Experiment 5) in part a, but treated this time by hydrolysis/ethylation followed by TMS derivatization. (c) EI mass spectrum for ethyl ester of 2-MG acid detected in part b ( $RT = 27.42$  min). (d) EI mass spectrum for ethyl ester of linear 2-MG acid dimer detected in part b ( $RT = 50.48$  min). The hydrolysis/ethylation followed by TMS derivatization results presented here confirm the existence of polyesters in high- $\text{NO}_x$  SOA.

for this proposed dialdehyde species to form SOA; therefore, other heterogeneous processes may occur. The detailed analysis of the GC/MS derivatization and the ESI tandem MS results, however, provides strong chemical evidence for the formation

of polyesters, where the neutral loss of 102 Da is explained by the dehydrated lactone form of 2-MG (Figure 14). In addition, a GC/MS derivatization analysis made for MACR high- $\text{NO}_x$  SOA (Experiment 3) that included a methoximation step prior



**Figure 16.** Proposed mechanism for SOA formation from isoprene photooxidation under high- $\text{NO}_x$  conditions. Symbol used: ?, further study needed in order to understand the formation (in gas/particle phase) of 2-MG. <sup>a</sup>Elemental compositions confirmed by high-resolution ESI-MS. <sup>b</sup>Elemental composition of *mono-nitrate* tetramer (MW = 471) confirmed by high-resolution ESI-MS. <sup>c</sup>Elemental compositions of *mono-acetate* tetramer and pentamer (MW = 366 and 468, respectively) confirmed by high-resolution ESI-MS.



**Figure 17.** Proposed gas-phase formation mechanism for a  $\text{C}_4$  hydroxydialdehyde monomer, possibly accounting for a fraction of the unidentified SOA mass in high- $\text{NO}_x$  experiments.

to trimethylsilylation to reveal aldehyde functions in the formed oligomers was negative.

To investigate further the probable importance of a  $\text{C}_4$  hydroxy dialdehyde species and its respective hemiacetal oligomers, selected sample extracts were derivatized using Girard Reagent P to increase sensitivity for aldehydic species in the (+)ESI mode. A high- $\text{NO}_x$  isoprene and a MACR sample were treated with this derivatizing agent, and as a result, the detection of the  $m/z$  236, 206, and 192 ions resulted for both

samples, which likely corresponds to the detection of the proposed  $\text{C}_4$  dialdehyde, glyoxal, and methylglyoxal, respectively. However, the proposed hemiacetal oligomers that would be produced from this  $\text{C}_4$  dialdehyde were not detected, consistent with observations made in the methoximation GC/MS experiment. It is possible that the detection of the proposed  $\text{C}_4$  dialdehyde resulted from the decomposition of oligomers during the derivatization step of the sample workup procedure (which is equivalent for the detection of glyoxal and methylgly-

lyoxal in the particle phase). As a confirmation that the observed ions were derivatized species of the proposed C<sub>4</sub> dialdehyde, glyoxal, and methylglyoxal, upfront CID LC/MS analysis was used to detect common neutral mass losses and fragment ions associated with derivatized aldehydes and ketones. The common neutral losses and fragment ions associated with the GirP derivatization detected were similar to those found by Lai et al.,<sup>23</sup> providing further evidence of the detection of these small aldehyde species in high-NO<sub>x</sub> isoprene and MACR SOA. The detection of these small dicarbonyls provides some evidence that aldehydes may account for a fraction of the unquantified (unidentified) SOA mass (Table 2) produced from isoprene oxidation under high-NO<sub>x</sub> conditions. The mechanism (reactive uptake and/or oligomerization) and the degree in which these aldehydes form SOA, however, remains unclear and bears further study.

**4.3. Low-NO<sub>x</sub> SOA.** **4.3.1. Hydroperoxides: Key Component to SOA Formation.** As discussed previously, in the absence of NO<sub>x</sub>, the RO<sub>2</sub> radical chemistry is dominated by RO<sub>2</sub> + HO<sub>2</sub> reactions, owing to the large amounts of HO<sub>2</sub> formed from the OH + H<sub>2</sub>O<sub>2</sub> reactions.<sup>17</sup> RO<sub>2</sub> + RO<sub>2</sub> reactions are expected to be less substantial (10–30% contribution) because of the high HO<sub>2</sub>/RO<sub>2</sub> ratios in these experiments, and as a result, hydroperoxides are expected to be the dominant gas-phase products. Because of their expected low volatilities, hydroperoxide species can partition to the aerosol phase and likely form high-MW species via peroxyhemiacetal formation with aldehydic species.<sup>18,19</sup> Hydroperoxides resulting from the oxidation of aromatic and biogenic VOCs have been observed and calculated to be important contributors to the overall SOA mass.<sup>18,45,46</sup> Indeed, as shown in Table 3, organic peroxides (i.e., hydroperoxides or ROOR) are also a significant component (~61% of the SOA mass for nucleation experiments and ~25% and 30% of the SOA mass for dry seeded and acid seeded experiments, respectively) of the low-NO<sub>x</sub> isoprene SOA. The large discrepancy in peroxide content observed between nucleation (seed-free) and seeded experiments is currently not understood. As discussed in the results section, there is no evidence of interference from ammonium sulfate on the peroxide content measurement. Owing to the neutral nature of the hydroperoxides (and ROOR) measured by the iodometric-spectrophotometric method, no tandem ESI-MS measurements could be made to structurally elucidate this fraction. Thus, it is difficult to explain the differences in the peroxide content observed between nucleation and seeded experiments. It is possible that in the seeded cases the hydroperoxide species are heterogeneously converted into neutral species other than peroxidic compounds, such as polyols. Further studies should be conducted to investigate the role of inorganic seed on the amount of peroxides formed in the aerosol phase. As noted in the high-NO<sub>x</sub> case, the mass closure results presented here apply only to the aerosol mass loadings produced in this current study and cannot be concluded as absolute for the isoprene system at low-NO<sub>x</sub> conditions.

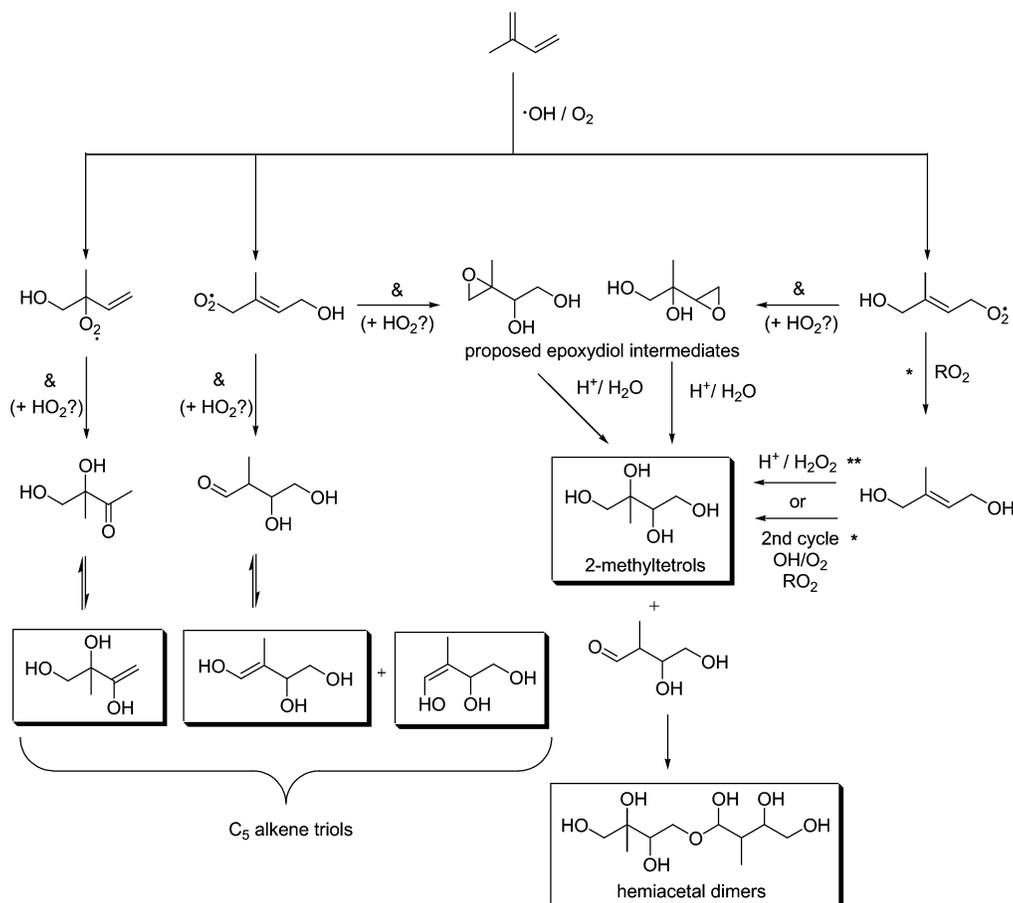
**4.3.2 Oligomerization.** Oligomers were found to form under low-NO<sub>x</sub> conditions, as shown in the (+)MALDI (Figure 7), GC/MS with TMS derivatization (Figure 10), and TOF-AMS (Figures 8 and 9) data. In contrast to high-NO<sub>x</sub> conditions, no distinct pattern or obvious monomeric unit, like the 102 Da differences observed in the high-NO<sub>x</sub> oligomeric SOA (Figures 1, 2, and 5), was observed in the low-NO<sub>x</sub> oligomers. The oligomers formed in the low-NO<sub>x</sub> case are not acidic in nature like those in the high-NO<sub>x</sub> case. Structural elucidation of these oligomers is limited because these neutral products are not

ionizable using ESI-MS. MALDI (Figure 7) was able to provide some indication of the MW ranges of the oligomeric SOA, but structural elucidation was not possible. The large mass contribution of organic peroxides to the low-NO<sub>x</sub> SOA (Table 3) provides some insight into the oligomerization reactions occurring. It is possible that some fraction of the oligomeric SOA is formed by peroxyhemiacetals, which result from heterogeneous reactions of hydroperoxides and aldehydes.

Because of the neutral nature of the oligomeric SOA produced under low-NO<sub>x</sub> conditions, only the GC/MS derivatization technique provides structural elucidation of the oligomers formed owing to the presence of polyols. Hemiacetal formation reactions between C<sub>5</sub> alkene triols (Table 4) and 2-methyltetrols (Table 4) were found to occur using this technique (Figure 10b). The reaction involves a terminal hydroxyl group of a 2-methyltetrol, which serves as a nucleophile, reacting with the tautomeric keto form of one C<sub>5</sub> alkene triol (Table 4) to form the hemiacetal dimer shown in Figure 10b. As was observed by the GC/MS *m/z* 219 EIC, six isomeric forms of this hemiacetal dimer could be partially resolved. However, further elucidation of higher-order hemiacetal (acetal) oligomers could not be conducted owing to their likely thermal decomposition in the GC injector of the GC/MS instrument, their high MW preventing their elution from the GC column, and lack of ionization when using ESI-MS techniques. As for the confirmation of peroxyhemiacetal oligomers, analytical techniques need to be developed in order to further elucidate the neutral higher-order hemiacetal (acetal) oligomers likely present in low-NO<sub>x</sub> SOA.

**4.3.3. Acid Catalysis.** The SOA mass for the acid seed experiment (Experiment 14) is significantly larger (~3.6 times) than that of the dry seeded/nucleation experiments (Experiments 15/12), in contrast to high-NO<sub>x</sub> conditions, in which acid seed had no such observable effect. Note that the SOA mass concentration was virtually identical in experiments using dry (nonacid) seed aerosol and in those in the absence of seed aerosol, where particle formation takes place by nucleation (Experiments 12, 13, and 15). GC-FID measurements made for selected low-NO<sub>x</sub> experiments also provide evidence for acid-catalyzed particle-phase reactions. The C<sub>5</sub> alkene triols and 2-methyltetrols decreased in their contributions to the overall SOA mass when acid seed was present. For example, the 2-methyltetrols and C<sub>5</sub> alkene triols contributed ~3.91% and 0.6%, respectively, to the SOA mass for Experiment 13 (nonacid case), whereas in Experiment 14 (acid case), the 2-methyltetrols and C<sub>5</sub> alkene triols were found to decrease to ~0.46% and 0.06%, respectively, of the SOA mass. This result is in contrast to that observed by Edney et al.<sup>14</sup> in which isoprene tracer compounds were observed to increase in concentration, and is possibly due to the differing isoprene/NO<sub>x</sub> ratios employed. In conjunction with the above GC-FID results, the fact that C<sub>5</sub> alkene triols and 2-methyltetrols were found to form hemiacetal dimers (and likely higher order oligomers) suggests that the presence of acidified aerosol catalyzes hemiacetal (and likely acetal) oligomer formation under low-NO<sub>x</sub> conditions. The same may be the case for peroxyhemiacetal formation reactions.

**4.3.4. Formation Mechanism of Low-NO<sub>x</sub> SOA products Observed by GC/MS.** The detection of organic peroxides in the particle phase (Table 3) by the iodometric-spectrophotometric method provides strong evidence that the hydroperoxides that result from the gas-phase RO<sub>2</sub> + HO<sub>2</sub> reactions are sufficiently polar (nonvolatile) to partition to the aerosol phase, thereby elucidating one major reaction pathway leading to SOA formation under low-NO<sub>x</sub> conditions. The detection of 2-methyltetrols,



**Figure 18.** Low- $\text{NO}_x$  SOA formation pathways as elucidated by GC/MS. Boxes indicate products detected in low- $\text{NO}_x$  SOA. Symbols used: &, further study needed for the formations of the hypothetical carbonyl diol and epoxydiol intermediates which may result from the rearrangements of  $\text{RO}_2$  radicals and/or hydroperoxides; \*, for details about this pathway leading to 2-methyltetrols and also holding for isomeric products, see reference 7; \*\*, for details about this alternative pathway, see ref 14. 2-methyltetrol performate derivatives (shown in Table 4) were omitted for simplicity; however, these could serve as precursors for 2-methyltetrols if in the presence of acid and water.

$\text{C}_5$  alkene triols, 2-methyltetrol performate derivatives, and hemiacetal dimers (Table 4) suggests that the  $\text{RO}_2$  radicals that form from the initial oxidation ( $\text{OH}/\text{O}_2$ ) of isoprene follow some other route. The formation of 2-methyltetrols has been explained by self- and cross-reactions of the  $\text{RO}_2$  radicals formed from the initial oxidation ( $\text{OH}/\text{O}_2$ ) of isoprene, leading to intermediate 1,2-diols, which may undergo a second cycle of oxidation ( $\text{OH}/\text{O}_2$ ) reactions followed by self- and cross-reactions of the  $\text{RO}_2$  radicals.<sup>7</sup>

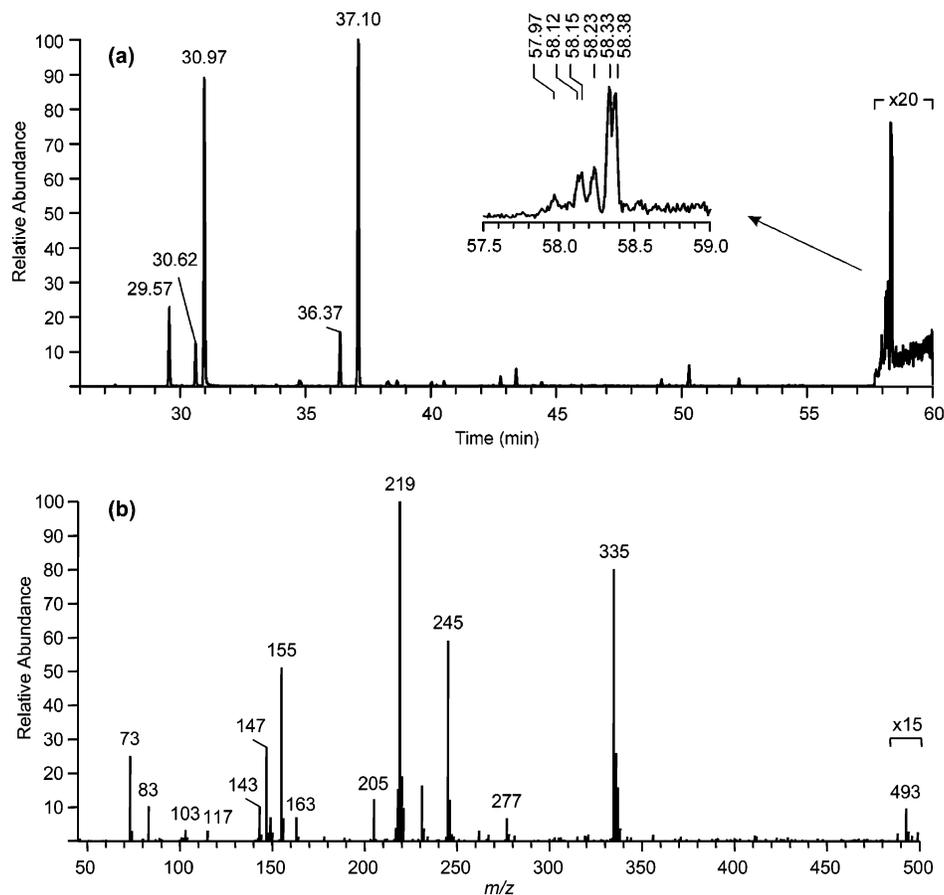
The detection of  $\text{C}_5$  alkene triols in ambient aerosol may indicate the importance of intermediate epoxydiol derivatives of isoprene, which may also be intermediates in the formation of 2-methyltetrols.<sup>11,12</sup> Wang et al.<sup>12</sup> hypothesized from MS evidence that these epoxydiol intermediates could be trapped in the aerosol phase and subsequently converted into  $\text{C}_5$  alkene triols and 2-methyltetrols through acid-catalyzed reactions. Acid-catalyzed reactions of epoxydiols may be a formation pathway for 2-methyltetrols and  $\text{C}_5$  alkene triols, but these monomers may also form from other pathways.

Shown in Figure 18 is a proposed mechanism for the formation of key SOA components from the oxidation of isoprene under low- $\text{NO}_x$  conditions. As suggested by Böge et al.,<sup>15</sup> 2-methyltetrols may form by several possible pathways. The formation of the 2-methyltetrols through two cycles of oxidation ( $\text{OH}/\text{O}_2$ ) reactions followed by self- and cross-reactions of the  $\text{RO}_2$  radicals is only briefly included in this figure. It is possible that epoxydiols may form from rearrangements of hydroxyhydroperoxides or hydroxyperoxy radicals.

Once formed, these epoxydiols could be taken up into the particulate phase, and through hydrolysis form 2-methyltetrols. In addition, an alternative pathway leading to the formation of 2-methyltetrols has been reported in a recent study by Böge et al.<sup>15</sup> That study proposed that intermediates in the formation of 2-methyltetrols (i.e., 2-methyl-3-butene-1,2-diol and 2-methyl-2-vinylloxirane) are converted to 2-methyltetrols through reaction with hydrogen peroxide on acidic particles. The latter pathway is also included in the scheme in Figure 18. Further gas and particle-phase studies are needed in order to fully elucidate the pathways leading to the formation of 2-methyltetrols, the  $\text{C}_5$  alkene triols, and related dimeric products.

**4.3.5. Evolution of SOA Composition.** As in Kroll et al.,<sup>17</sup> a rapid decay of the SOA mass was observed after the initial SOA growth reached its maximum for all low- $\text{NO}_x$  nucleation experiments. This loss is not attributable to wall removal processes because the particles shrink in size rather than reduce in number (as measured by the DMA). The loss of SOA mass was observed to stop immediately after chamber lights were turned off, and resume once the lights were turned back on, indicating a photochemical effect.

Indeed, when comparing the peroxide measurements made at (or around) the initial SOA growth maximum to some later experimental time after SOA mass decay, it was found that the organic peroxide content of the aerosol decreased significantly (~59% to 26% of SOA mass, respectively, for Experiment 18). This observation provides strong evidence that organic peroxides decompose in the particle phase due to photolysis



**Figure 19.** (a) GC/MS EIC using specific ions for the TMS derivatives of 2-methyltetrols ( $m/z$  219),  $C_5$  alkene triols ( $m/z$  231), and hemiacetal dimers ( $m/z$  219 and 335) for a PM<sub>2.5</sub> aerosol sample collected in Rondônia, Brazil, during the onset of the wet season from 10–12 November 2002 (39 h collection time). The inset shows a detail of the isomeric hemiacetal dimers, formed between 2-methyltetrols and  $C_5$  dihydroxycarbonyls, which elute between 57 and 59 min; (b) averaged EI mass spectrum (only limited mass range  $m/z$  50–500 available) for the TMS derivatives of the isomeric hemiacetal dimers.

and/or subsequent particle-phase reactions, or they are driven out of the particle as a result of gas-phase compounds being reacted away, shifting the equilibrium back to the gas phase. TOF-AMS measurements also confirmed that the peroxide content of low- $NO_x$  SOA decreases with time as shown in Figure 9b. This decrease in peroxide content as a function of time also coincided with high-mass fragment ions ( $m/z > 200$ ) increasing in their abundance (in Figure 9a only  $m/z$  247 and 327 are shown), suggesting the possibility that peroxide decomposition causes oligomerization reactions. These oligomerization reactions likely lead to hemiacetals (as elucidated by GC/MS).

**4.3.6. Tracer Compounds for Isoprene Oxidation in the Remote Atmosphere.** The low- $NO_x$  chamber experiments conducted in this study confirm that 2-methyltetrols indeed serve as tracer compounds for isoprene oxidation in the ambient atmosphere, especially in remote regions such as the Amazonian rainforest. The detection of  $C_5$  alkene triols and hemiacetal dimers in the present low- $NO_x$  experiments corresponds well to their observation in ambient aerosol collected from the Amazonian rainforest<sup>12</sup> and Finnish boreal forests (note that hemiacetal dimers in aerosol collected from the Finnish boreal forests is not yet confirmed).<sup>11</sup> From these field studies,  $C_5$  alkene triols were postulated to form by acid-catalyzed ring-opening reactions of epoxydiol derivatives of isoprene in low  $RH$  environments. However, hemiacetal dimers were not recognized in ambient samples; this current study elucidates their formation under low- $NO_x$  conditions. Once it was realized that

hemiacetal dimers form from  $C_5$  alkene triols and 2-methyltetrols, we referred back to data collected from the Amazonian rainforest.<sup>47</sup> When investigating the GC/MS data carefully, it was found that the hemiacetal dimers were indeed detected, suggesting the atmospheric relevance of these low- $NO_x$  chamber experiments. Shown in Figure 19 is a GC/MS EIC of an Amazonian fine aerosol sample (i.e., PM<sub>2.5</sub>; particulate matter with an aerodynamic diameter  $< 2.5 \mu m$ ) collected during the wet season (low- $NO_x$  conditions) using multiple ions, that is,  $m/z$  231 (to show the  $C_5$  alkene triols),  $m/z$  219 (to show 2-methyltetrols as well as the dimers), and  $m/z$  335 (characteristic of the dimers). An averaged EI mass spectrum for the hemiacetal dimers is also included in this Figure to further confirm their presence in ambient aerosol.

## 5. Conclusions

The composition of SOA from the photooxidation of isoprene under both high- and low- $NO_x$  conditions has been thoroughly investigated through a series of controlled laboratory chamber experiments. It is found that the chemical nature of the resultant SOA is significantly different in the two  $NO_x$  regimes. Under high- $NO_x$  conditions, the SOA components are acidic and form upon the further oxidation of MACR. SOA components formed under low- $NO_x$  conditions, by contrast, are not acidic, with primary species identified being polyols and organic peroxides. On the basis of SOA growth, acid-catalysis seems to play a larger role under low- $NO_x$  conditions. Organic peroxides (likely

dominated by hydroperoxides) contribute significantly to the low-NO<sub>x</sub> SOA mass (~61% for nucleation experiments and ~25% and 30% for dry seeded and acid seeded experiments, respectively). However, differences in the organic peroxide contribution and the rate of loss in SOA mass for nucleation (seed-free) and seeded experiments are not well understood and require further investigation. The chemical composition changes with time in the low-NO<sub>x</sub> case, showing evidence of chemical aging.

Oligomerization is an important SOA formation pathway for both low- and high-NO<sub>x</sub> conditions because oligomers were observed in both cases. The nature of the oligomers, however, is distinctly different in each NO<sub>x</sub> regime. Under high-NO<sub>x</sub> conditions, the oligomers have clear monomeric units, with observable 102 Da differences using both online and offline mass spectrometry techniques. Using tandem ESI-MS techniques and GC/MS with trimethylsilylation, it is found that polyesters account for these high-NO<sub>x</sub> oligomers, with 2-MG as the key monomeric unit. These polyesters account only for a fraction (~22–34%) of the SOA mass formed from isoprene oxidation. This lack of mass closure could result from an underestimate of the amount of polyesters formed or additional, unidentified MACR or isoprene oxidation products that contribute to the SOA mass. One key unresolved question is the path by which 2-MG is formed, which at present is not understood. Further gas- and particle-phase studies on isoprene oxidation under high-NO<sub>x</sub> conditions are needed in order to elucidate the 2-MG formation pathway.

Previously detected tracer compounds for isoprene oxidation in the ambient atmosphere were detected in the low-NO<sub>x</sub> experiments. C<sub>5</sub> alkene triols and hemiacetal dimers are reported here for the first time in a controlled laboratory experiment, suggesting that the oxidative conditions used in these experiments are relevant to remote regions. The GC/MS results suggest that hemiacetal dimers formed in these low-NO<sub>x</sub> chamber experiments result from the reactions of 2-methyltetrols and C<sub>5</sub> alkene triols (a reaction that is likely relevant to the real atmosphere). Besides the formation of hemiacetal (acetal) oligomers in low-NO<sub>x</sub> SOA, it is speculated that peroxyhemiacetal oligomers could also form, because of the large amounts of peroxides measured in the particle phase. The formation of low-NO<sub>x</sub> oligomers may correlate to the decomposition of peroxides with experimental time, providing some insight into the mechanism of oligomerization. Additional analytical techniques need to be developed in order to elucidate the neutral/unstable products found in SOA produced from the photooxidation of isoprene.

**Acknowledgment.** Research at Caltech was funded by the U.S. Environmental Protection Agency to Achieve Results (STAR) Program grant no. RD-83107501-0, managed by EPA's Office of Research and Development (ORD), National Center for Environmental Research (NCER), and by the U.S. Department of Energy, Biological, and Environmental Research Program DE-FG02-05ER63983; this work has not been subjected to the EPA's required peer and policy review and therefore does not necessarily reflect the views of the Agency and no official endorsement should be inferred. Jason Surratt was supported in part by the United States Environmental Protection Agency (EPA) under the Science to Achieve Results (STAR) Graduate Fellowship Program. Research at the Universities of Antwerp and Ghent was supported by the Belgian Federal Science Policy Office through the BIOSOL project (contract SD/AT/02A) and a visiting postdoctoral fellowship to Rafal Szmigielski, and by the Research Foundation –

Flanders (FWO). We would like to thank John Greaves at the University of California, Irvine for the accurate mass measurements on the ESI-TOF instrument. We would like to also thank Paul Ziemann at the University of California, Riverside for his useful communications regarding peroxide measurements in SOA.

**Supporting Information Available:** Electron impact (EI) mass spectra for 2-methyltetrols, C<sub>5</sub> alkene triols, and 2-methyltetrol performate derivatives for their respective chromatographic peaks found in Figure 10. This material is available free of charge via the Internet at <http://pubs.acs.org>.

## References and Notes

- (1) Inuma, Y.; Böge, O.; Gnauk, T.; Herrmann, H. *Atmos. Environ.* **2004**, *38*, 761.
- (2) Gao, S.; Keywood, M.; Ng, N.; Surratt, J. D.; Varutbangkul, V.; Bahreini, R.; Flagan, R. C.; Seinfeld, J. H. *J. Phys. Chem. A* **2004**, *108*, 10147.
- (3) Gao, S.; Ng, N.; Keywood, M.; Varutbangkul, V.; Bahreini, R.; Nenes, A.; He, J.; Yoo, K.; Beauchamp, J.; Hodyss, R.; Flagan, R.; Seinfeld, J. *Environ. Sci. Technol.* **2004**, *38*, 6582.
- (4) Tolocka, M.; Jang, M.; Ginter, J.; Cox, F.; Kamens, R.; Johnston, M. *Environ. Sci. Technol.* **2004**, *38*, 1428.
- (5) Kalberer, M.; Paulsen, D.; Sax, M.; Steinbacher, M.; Dommen, J.; Prevot, A.; Fisseha, R.; Weingartner, E.; Frankevich, V.; Zenobi, R.; Baltensperger, U. *Science* **2004**, *303*, 1659.
- (6) Kanakidou, M.; Seinfeld, J.; Pandis, S.; Barnes, I.; Dentener, F.; Facchini, M.; Van Dingenen, R.; Ervens, B.; Nenes, A.; Nielsen, C.; Swietlicki, E.; Putaud, J.; Balkanski, Y.; Fuzzi, S.; Horth, J.; Moortgat, G.; Winterhalter, R.; Myhre, C.; Tsigaridis, K.; Vignati, E.; Stephanou, E.; Wilson, J. *Atmos. Chem. Phys.* **2005**, *5*, 1053.
- (7) Claeys, M.; Graham, B.; Vas, G.; Wang, W.; Vermeylen, R.; Pashynska, V.; Cafmeyer, J.; Guyon, P.; Andreae, M. O.; Artaxo, P.; Maenhaut, W. *Science* **2004**, *303*, 1173.
- (8) Pandis, S.; Paulson, S.; Seinfeld, J. H.; Flagan, R. C. *Atmos. Environ.* **1991**, *25*, 997.
- (9) Limbeck, A.; Kulmala, M.; Puxbaum, H. *Geophys. Res. Lett.* **2003**, *30*.
- (10) Ion, A. C.; Vermeylen, R.; Kourtchev, I.; Cafmeyer, J.; Chi, X.; Gelencsér, A.; Maenhaut, W.; Claeys, M. *Atmos. Chem. Phys.* **2005**, *5*, 1805.
- (11) Kourtchev, I.; Ruuskanen, T.; Maenhaut, W.; Kulmala, M.; Claeys, M. *Atmos. Chem. Phys.* **2005**, *5*, 2761.
- (12) Wang, W.; Kourtchev, I.; Graham, B.; Cafmeyer, J.; Maenhaut, W.; Claeys, M. *Rapid Commun. Mass Spectrom.* **2005**, *19*, 1343.
- (13) Claeys, M.; Wang, W.; Ion, A.; Kourtchev, I.; Gelencsér, A.; Maenhaut, W. *Atmos. Environ.* **2004**, *38*, 4093.
- (14) Edney, E. O.; Kleindienst, T. E.; Jaoui, M.; Lewandowski, M.; Offenberg, J. H.; Wang, W.; Claeys, M. *Atmos. Environ.* **2005**, *39*, 5281.
- (15) Böge, O.; Miao, Y.; Plewka, A.; Herrmann, H. *Atmos. Environ.* **2006**, *40*, 2501.
- (16) Kroll, J. H.; Ng, N. L.; Murphy, S. M.; Flagan, R. C.; Seinfeld, J. H. *Geophys. Res. Lett.* **2005**, *32*.
- (17) Kroll, J. H.; Ng, N. L.; Murphy, S. M.; Flagan, R. C.; Seinfeld, J. H. *Environ. Sci. Technol.* **2006**, *40*, 1869.
- (18) Docherty, K.; Wu, W.; Lim, Y.; Ziemann, P. *Environ. Sci. Technol.* **2005**, *39*, 4049.
- (19) Johnson, D.; Jenkin, M. E.; Wirtz, K.; Martin-Reviejo, M. *Environ. Chem.* **2004**, *1*, 150.
- (20) Gao, S.; Surratt, J. D.; Knipping, E. M.; Edgerton, E. S.; Shahgholi, M.; Seinfeld, J. H. *J. Geophys. Res.*, in press, 2006.
- (21) Cocker, D.; Flagan, R. C.; Seinfeld, J. H. *Environ. Sci. Technol.* **2001**, *35*, 2594.
- (22) Keywood, M.; Varutbangkul, V.; Bahreini, R.; Flagan, R.; Seinfeld, J. *Environ. Sci. Technol.* **2004**, *38*, 4157.
- (23) Lai, C.; Tsai, C.; Tsai, F.; Lee, C.; Lin, W. *Rapid Commun. Mass Spectrom.* **2001**, *15*, 2145.
- (24) Drewnick, F.; Hings, S.; DeCarlo, P.; Jayne, J.; Gonin, M.; Fuhrer, K.; Weimer, S.; Jimenez, J.; Demerjian, K.; Borrmann, S.; Worsnop, D. *Aerosol Sci. Technol.* **2005**, *39*, 637.
- (25) Pashynska, V.; Vermeylen, R.; Vas, G.; Maenhaut, W.; Claeys, M. *J. Mass Spectrom.* **2002**, *37*, 1249.
- (26) Banerjee, D.; Budke, C. *Anal. Chem.* **1964**, *36*, 792.
- (27) Sorooshian, A.; Brechtel, F. J.; Ma, Y.; Weber, R. J.; Corless, A.; Flagan, R. C.; Seinfeld, J. H. *Aerosol Sci. Technol.* **2006**, *40*, 396.
- (28) Weber, R.; Orsini, D.; Daun, Y.; Lee, Y.; Klotz, P.; Brechtel, F. *Aerosol Sci. Technol.* **2001**, *35*, 718.

- (29) Seinfeld, J. H.; Pandis, S. N. *Atmospheric Chemistry and Physics: From Air Pollution to Climate Change*; Wiley: New York, 1998.
- (30) Iinuma, Y.; Böge, O.; Miao, Y.; Sierau, B.; Gnauk, T.; Herrmann, H. *Faraday Discuss.* **2005**, *130*, 279.
- (31) Knochenmuss, R.; Zenobi, R. *Chem. Rev.* **2003**, *103*, 441.
- (32) Miyoshi, A.; Hatakeyama, S.; Washida, N. *J. Geophys. Res.* **1994**, *99*, 18779.
- (33) Tuazon, E.; Atkinson, R. *Int. J. Chem. Kinet.* **1990**, *22*, 1221.
- (34) Paulson, S.; Flagan, R. C.; Seinfeld, J. H. *Int. J. Chem. Kinet.* **1992**, *24*, 79.
- (35) Sprengnether, M.; Demerjian, K.; Donahue, N.; Anderson, J. J. *Geophys. Res.* **2002**, *107*.
- (36) Baker, J.; Arey, J.; Atkinson, R. *Environ. Sci. Technol.* **2005**, *39*, 4091.
- (37) Ng, N. L.; Kroll, J. H.; Keywood, M. D.; Bahreini, R.; Varutbangkul, V.; Lee, A.; Goldstein, A. H.; Flagan, R. C.; Seinfeld, J. H. *Environ. Sci. Technol.* **2006**, *40*, 2283.
- (38) Lee, A.; Goldstein, A. H.; Ng, N. L.; Kroll, J. H.; Varutbangkul, V.; Flagan, R. C.; Seinfeld, J. H. *J. Geophys. Res.* **2006**, *111*.
- (39) Chuong, B.; Stevens, P. S. *Int. J. Chem. Kinet.* **2004**, *36*, 12.
- (40) Yuan, H.; Nishiyama, Y.; Kuga, S. *Cellulose* **2005**, *12*, 543.
- (41) Kroll, J. H.; Ng, N. L.; Murphy, S. M.; Varutbangkul, V.; Flagan, R. C.; Seinfeld, J. H. *J. Geophys. Res.* **2005**, *110*.
- (42) Liggio, J.; Li, S.; McLaren, R. *Environ. Sci. Technol.* **2005**, *39*, 1532.
- (43) Barsanti, K.; Pankow, J. *Atmos. Environ.* **2005**, *39*, 6597.
- (44) Tong, C.; Blanco, M.; Goddard, W. A., III; Seinfeld, J. H. *Environ. Sci. Technol.* **2006**, *40*, 2333.
- (45) Johnson, D.; Jenkin, M. E.; Wirtz, K.; Martin-Reviejo, M. *Environ. Chem.* **2005**, *2*, 35.
- (46) Bonn, B.; von Kuhlmann, R.; Lawrence, M. *Geophys. Res. Lett.* **2004**, *31*.
- (47) Decesari, S.; Fuzzi, S.; Facchini, M.; Mircea, M.; Emblico, L.; Cavalli, F.; Maenhaut, W.; Chi, X.; Schkolnik, G.; Falkovich, A.; Rudich, Y.; Claeys, M.; Pashynska, V.; Vas, G.; Kourtev, I.; Vermeylen, R.; Hoffer, A.; Andreae, M. O.; Tagliavini, E.; Moretti, F.; Artaxo, P. *Atmos. Chem. Phys.* **2006**, *6*, 375.