Development of Photoinduced Copper-Catalyzed Amination of Alkyl Electrophiles: Synthesis and Mechanism

Thesis by Hyungdo Cho

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ABSTRACT

The formation of carbon-nitrogen (C–N) bonds is crucial in organic chemistry due to the importance of nitrogen-containing functional groups. While traditional nucleophilic substitution reactions, such as S_N1 , S_N2 , and S_NAr , are limited in scope and efficiency, transition metal-catalyzed versions of these reactions, particularly involving copper, offer a more versatile approach by activating electrophiles and facilitating C–N bond formation via oxidative addition and reductive elimination.

Copper-catalyzed C–N couplings have been extensively developed but are primarily effective for aryl electrophiles rather than alkyl electrophiles due to the need for thermal activation, which often leads to undesired side reactions in alkyl electrophiles. The development of photoinduced copper-catalyzed reactions by Fu and Peters addresses these challenges, enabling the activation of alkyl electrophiles without thermal activation.

Over the past decade, the Fu group has focused on expanding the scope of this novel approach. The research detailed in this thesis focuses on developing photoinduced coppercatalyzed C–N coupling reactions for more challenging substrates, such as sterically hindered alkyl electrophiles and amines.

Chapter 2 discusses the photoinduced, enantio-convergent coupling of racemic tertiary alkyl electrophiles with aniline nucleophiles, catalyzed by bisphosphine-copper complexes. The mechanism of this reaction was elucidated using various tools, identifying key copperbased intermediates, including a chiral copper(II)–anilido complex that couples with a tertiary organic radical to form the C–N bond with good enantioselectivity.

Chapter 3 presents the photoinduced, copper-catalyzed coupling of secondary alkyl amines with secondary or tertiary alkyl bromides to synthesize N-tertiary alkyl amines under mild conditions. This novel reaction provides unique stereoselectivity and compatibility with strained electrophiles, contributing valuable methodologies to the synthesis of bioisosteres and other complex amine structures.

Overall, this work broadens the understanding and application of photoinduced coppercatalyzed reactions, offering new pathways for the synthesis of sterically hindered amines.

PUBLISHED CONTENT AND CONTRIBUTIONS

This dissertation contains materials adapted with permission from the following publications:

1. Cho, H.; Suematsu, H.; Oyala, P. H.; Peters, J. C.; Fu, G. C. Photoinduced, Copper-Catalyzed Enantioconvergent Alkylations of Anilines by Racemic Tertiary Electrophiles: Synthesis and Mechanism. *J. Am. Chem. Soc.* **2022**, *144*, 4550–4558. DOI: 10.1021/jacs.1c12749.

H.C. optimized the reaction, explored the substrate scope, and performed mechanistic studies. H.C. participated in the writing of the manuscript.

2. Cho, H.; Tong, X.; Zuccarello, G.; Anderson, R. L.; Fu, G. C. Synthesis of Tertiary Alkyl Amines via Photoinduced, Copper-Catalyzed Nucleophilic Substitution of Unactivated Alkyl Halides by Secondary Alkyl Amines. *Submitted*.

H.C. discovered and optimized the reaction, explored the substrate scope, and performed mechanistic studies. H.C. participated in the writing of the manuscript.

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LIST OF ABBREVIATIONS

C h a p t e r 1

INTRODUCTION

1.1. Transition metal-catalyzed nucleophilic substitution for carbon–nitrogen bond formation.

Due to the ubiquity and significance of nitrogen-containing functional groups in a wide range of both naturally occurring and synthesized organic compounds, developing strategies to form carbon-nitrogen bonds has become a fundamental area of research in organic chemistry. Nucleophilic substitution, a reaction in which a leaving group attached to a carbon atom in an electrophile (C–X bond, $X =$ the leaving group) is displaced by the attack of a nucleophile (e.g., amines), offers a straightforward and modular approach to constructing the desired C–N bonds (Figure 1.1a). However, classical nucleophilic substitution reactions, such as S_N1 , S_N2 , and S_NAr , are only effective for building a limited range of structures (Figure [1](#page-24-0).1b).¹ For instance, in S_N1 reactions, the use of either a Brønsted or Lewis acid to promote carbocation formation often leads to the deactivation of the amine as a nucleophile. In S_N2 reactions, either the elimination of HX by a base or a nucleophile (with unreactive electrophiles) or excessive alkylation (with reactive electrophiles) frequently results in failure to efficiently deliver the desired product. Additionally, S_NAr reactions, which occur when the leaving group is attached to aromatic carbons (such as in benzenes), require strong electron-withdrawing groups on the aromatic ring to proceed effectively.^{[2](#page-24-1)}

Transition metal-catalyzed cross-coupling reactions offer a powerful strategy to overcome the limitations of classical nucleophilic substitution (**Figure 1.2**). [3](#page-24-2) In these reactions, the electrophiles are "activated" by the catalyst, which facilitates the cleavage of the carbon-leaving group bond, generating alkyl-derived intermediates. This cleavage increases the formal oxidation state of the metal catalyst, a step known as oxidative addition.^{[4](#page-24-3)} Upon ligand exchange on the transition metal catalyst, a process called transmetalation, the C–N bond is formed between the alkyl intermediate and a nitrogen nucleophile by the catalyst, yielding the intended product. This bond formation step decreases the formal oxidation state of the metal catalyst, a process known as reductive elimination. The catalyst a. Nucleophilic substitution reaction

$$
R-X \xrightarrow{\text{Nu}^{\ominus}} R-\text{Nu}
$$

b. Representative types of nucleophilic substitution reaction

Figure 1.1. Classical nucleophilic substitution reactions.

Figure 1.2. Transitional metal-catalyzed cross-coupling: representative mechanisms.

1.2. Thermally-induced copper-catalyzed cross-coupling reactions.

Among the transition metals frequently utilized for cross-coupling reactions, copper exhibits privileged reactivity toward C–N bond formation. Initially discovered by Ullmann and Goldberg in 1905, copper-catalyzed C–N couplings have been extensively developed over the past few decades.^{[5](#page-24-4)} Under these conditions, electrophiles and nitrogen nucleophiles typically undergo coupling reactions in the presence of a catalytic amount of copper salt/ligand and a stoichiometric amount of external base to absorb the proton equivalent from the N–H bond (**eq 1.1**). While the copper catalyst generally enables a facile C–N reductive

elimination, the oxidative addition of the $C-X$ bond of the electrophile to copper is often sluggish, necessitating thermal activation to drive the transformation.^{[6](#page-24-5)}

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X
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NH_2R
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X = \text{halides}
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X = \text{halides}
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M = \text{base-H} \oplus
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M = \text{base-H} \oplus
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(1.2)
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While the broad reactivity of coupling between aryl halides and various nitrogen-based nucleophiles (such as azoles, amides, and alkyl amines) has been demonstrated, effectively providing a wide range of aryl amines that classical S_NAr reactions cannot access, its application toward alkyl electrophiles to address the limitations of their S_N1 and S_N2 reactions had been unsuccessful for a prolonged period.^{[7](#page-24-6)} Unlike aryl halides, in the presence of a nucleophile and base, alkyl halides easily undergo undesired elimination of HX to form olefins, and the thermal activation required to facilitate oxidative addition also promotes these side reactions. Due to these challenges, no copper-catalyzed couplings between unactivated secondary and tertiary alkyl electrophiles and nitrogen nucleophiles were reported before 2013.

1.3. Photoinduced copper-catalyzed C–N bond formation.

Exploiting alternative pathways to activate alkyl electrophiles without the need for thermal input could circumvent this problem and broaden the potential of copper-catalyzed cross-coupling of alkyl electrophiles. In 2012, Fu and Peters reported a solution to this challenge, demonstrating copper-catalyzed C–N coupling under light irradiation (**Figure** 1.3).^{[8](#page-24-7)} In this system, copper(I) species decorated with phosphine and carbazolide ligands undergo photoinduced excitation. The excited copper species possessing a high-energy electron can undergo single electron transfer (SET) to the aryl halide, resulting in the formation of an aryl radical and copper(II) species. This aryl radical is proposed to recombine with the copper(II)-carbazolide complex to form an arylated carbazole through reductive elimination. Because this photoinduced activation process can occur at unusually low temperatures, where thermally induced side reactions are suppressed, applying it to the activation of alkyl electrophiles could enable efficient cross-coupling with nucleophiles.

Figure 1.3. Photoinduced copper-catalyzed C–N bond formation: Fu and Peters' report and proposed mechanism.

b. Enantioconvergent alkylation of carbazoles/indoles by tertiary alkyl electrophiles.

It was soon found that this strategy is indeed effective for alkyl halides, leading to the development of a range of photoinduced, copper-catalyzed cross-coupling reactions between unactivated secondary alkyl halides and various nitrogen-based nucleophiles (**Figure 1.4a**).^{[9–](#page-24-8)} ^{[13](#page-25-0)} Additionally, the photoinduced activation of copper allows both enantiomers of racemic

alkyl halides to be converted into a prochiral alkyl radical that asymmetrically couples with a nucleophile in a chiral environment, achieving enantio-convergent synthesis of chiral amines (**Figure 1.4b**). [14](#page-25-1) In light of these earlier advancements, current research in the Fu group on copper catalysis focuses on developing: 1) various asymmetric C–N coupling reactions to deliver diverse chiral amine motifs, and 2) C–N coupling with more challenging substrates.

1.4. Mechanistic hypotheses and challenges in photoinduced, copper-catalyzed C–N bond formation.

Mechanistically, two key steps are hypothesized to contribute to the success of these reactions. First, the photoinduced excitation of copper species and subsequent activation of the alkyl electrophile should be efficient (**eq 1.2**). Earlier studies often relied on the reactivities of the copper-nucleophile complex without an additional ligand. The photochemical properties of these species are significantly influenced by the nucleophile and frequently required high-energy UV light to function. Developing a system that can robustly provide photochemical reactivity regardless of the type of nucleophile under milder light irradiation would be more desirable. Recently, the use of certain bisphosphine ligands or naphthol derived ligands has shown potential in this regard, enabling alkyl halides to couple with nucleophiles under blue light irradiation (**Figure 1.5**).^{[12,](#page-18-0)[15,](#page-25-2)[16](#page-25-3)} However, studies on the identification of the photo-reductants, origin of their photochemical reactivity, and the effect of substituents are lacking, resulting in a poor understanding of these species.

Figure 1.5. BINOL and BINAP derived ligands for photoinduced copper-catalyzed C**–**N couplings developed in Fu group.

Another important mechanistic step is the capture and coupling of alkyl radicals (**eq 1.3**). Although now considered key intermediates in many reactions, ^{[17](#page-25-4)} alkyl radicals are highly unstable species with short lifetimes that were long regarded as intractable.^{[18](#page-25-5)} Failure to efficiently capture the alkyl radical by copper intermediates and subsequently form the C– N bond could result in an unsuccessful cross-coupling reaction. While many reported coppercatalyzed reactions propose alkyl radical intermediates, the unified character of the electronics and sterics of the copper intermediates that allow efficient capture of the alkyl radical, as well as the detailed mechanism of the alkyl radical capture processes, are still poorly understood. For example, few computational evaluations^{[19–](#page-25-6)[21](#page-25-7)} and experimental model studies^{[22,](#page-25-8)[23](#page-25-9)} have proposed two possible mechanisms of radical capture (**Figure 1.6a**). The first pathway involves direct addition of the alkyl radical to a copper(II) center, forming a copper alkyl (or aryl) species with a formal oxidation state of +3. Reductive elimination from the copper center results in C–N bond formation. The alternative pathway involves direct radical-radical coupling between the nitrogen of the nucleophile attached to copper and the alkyl radical. Due to the higher electronegativity of copper, the SOMO of the copper (II) metalloradical can have more ligand character (ligand field inversion), 24 24 24 rendering certain copper(II)-nucleophile species be better viewed as copper(I)-nucleophile radicals that favor radical-radical coupling. The feasibility of both mechanisms has been demonstrated experimentally under designed model systems, but not within the context of photoinduced copper-catalyzed C–N coupling reactions (**Figure 1.6b**).

a. Two computationally proposed mechanisms of radical capture and C-Nbond formation.

Figure 1.6. Proposed mechanisms for reductive elimination in copper-catalyzed C–N coupling.

1.5. Overview of individual chapters.

Building on these hypotheses and challenges, the goal of my research in the Fu group was to contribute to or achieve three specific objectives: 1) developing novel C–N coupling reactions that address current limitations in organic synthesis, 2) identifying the working photoreductants in the developed reactions and studying the mechanisms of their photoexcitation and alkyl electrophile activation processes, and 3) identifying the copper intermediates that capture alkyl radicals and broadening the understanding of alkyl radical capture and subsequent C–N bond formation by these copper species.

In Chapter 2, we describe photoinduced, enantio-convergent coupling of a variety of racemic tertiary alkyl electrophiles with aniline nucleophiles catalyzed by bisphosphinecopper complexes. The reaction can generate a new C–N bond with good ee at the fully

substituted stereocenter of the product (**Figure 1.7a**). The mechanism of this new reaction has been interrogated with the aid of a wide array of tools, including the independent synthesis of proposed intermediates and reactivity studies, spectroscopic investigations featuring photophysical and EPR data, and DFT calculations. These studies led to the identification of three copper-based intermediates in the proposed catalytic cycle, including a chiral three-coordinate formally copper(II)–anilido (DFT analysis points to its formulation as a copper(I)–anilidyl radical) complex that serves as a persistent radical that couples with a tertiary organic radical to generate the desired C–N bond with good enantioselectivity (**Figure 1.7b**).

a. Enantioconvergent alkylations of anilines by racemic tertiary electrophiles.

Figure 1.7. Overview of the key concepts and findings presented in chapter 2.

[Intentionally Redacted]

1.6. Notes and references.

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C h a p t e r 2

PHOTOINDUCED, COPPER-CATALYZED ENANTIOCONVERGENT ALKYLATIONS OF ANILINES BY RACEMIC TERTIARY ELECTROPHILES: SYNTHESIS AND MECHANISM

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2.1. Introduction

Amines play an important role in a wide array of disciplines, including biology, materials science, organic chemistry, and pharmaceutical chemistry. $1 - 4$ $1 - 4$ $1 - 4$ Although the nucleophilic substitution of an alkyl halide by an amine is an attractive approach to the synthesis of higher-order amines, traditional S_N2 reactions have limited applicability in the case of less reactive (e.g., hindered and unactivated) electrophiles, which instead engage in undesired side reactions, such as the elimination of $H-X$, $5,6$ $5,6$ Furthermore, conventional substitution pathways almost never enable the control of stereochemistry at the carbon of the new C–N bond, starting with a readily available racemic electrophile.

To address these shortcomings with respect to reactivity and enantioselectivity, a number of laboratories have pursued the use of transition metals to catalyze substitution reactions of secondary and tertiary alkyl electrophiles by nitrogen nucleophiles.^{[7](#page-198-4)-13} To date, catalytic enantioconvergent substitutions have only been described for a few families of (mostly secondary) alkyl electrophiles, e.g., allylic electrophiles, 14 14 14 α-halocarbonyl compounds, $15-18$ $15-18$ $15-18$ α-cyano-α-halocarbonyl compounds, $18, 19$ $18, 19$ $18, 19$ propargylic electrophiles, $18, 20$ $18, 20$ benzylic electrophiles, 18 18 18 and unactivated alkyl halides that bear a directing group.^{[21](#page-199-6)}

Whereas a variety of bioactive molecules include as a subunit an arylamine (aniline) wherein the nitrogen is attached to a stereocenter (**Figure 2.1**), $22,23$ $22,23$ to our knowledge there has been only one report of the synthesis of this motif via the enantioconvergent alkylation of an aniline by a racemic alkyl electrophile.^{1[5,24,](#page-200-2)[25](#page-200-3)} In the present study, we establish that, with the aid of a chiral copper catalyst and light, anilines can be coupled with a variety of racemic tertiary alkyl electrophiles to generate N-alkylanilines that bear a fully substituted stereocenter with good enantiomeric excess (**Figure 2.2**). Mechanistic studies are consistent with [**P**CuCl] acting as a photoreductant and with [**P**Cu(NHAr)]Cl serving as a key intermediate in the catalytic cycle.

Figure 2.2. This study: photoinduced, copper-catalyzed enantioconvergent alkylations of anilines by racemic tertiary electrophiles.

2.2. Result and discussion

2.2.1. Reaction development

Seeking to expand the rather limited scope of enantioconvergent substitutions of alkyl electrophiles by nitrogen nucleophiles, we chose to explore reactions of α -halonitriles with anilines to generate α -aminonitriles. In particular, we decided to examine the use of tertiary α-halonitriles as electrophiles so as to produce α-disubstituted α-aminonitriles. Whereas catalytic enantioselective Strecker reactions of aldimines provide a versatile approach to the synthesis of enantioenriched α -monosubstituted α -aminonitrile families, $26,27$ $26,27$ corresponding reactions of ketimines to afford α-disubstituted α-aminonitriles are less well-developed.^{[28](#page-200-6)-32}

Table 2.1. Effect of reaction parameters. All data are the average of two runs.

Upon irradiation (blue LED) in the presence of CuCl and DTBM-SEGPHOS (**P**), a racemic tertiary α-chloronitrile undergoes substitution by *p*-toluidine (1.2 equiv) to furnish the desired α-disubstituted α-aminonitrile in good yield and ee (**Table 2.1**, entry 1: 77% yield, 92% ee; CuCl, **P**, and BTPP are all commercially available). Control experiments establish that CuCl, **P**, and light are critical for coupling under these conditions (entries $2-5$; $>95\%$ recovery of the alkyl halide). At a higher reaction temperature (e.g., $-10\degree C$), a lower yield and ee are observed (entry 6). The yield of the reaction is not particularly sensitive to small amounts of water or of air, which have no impact on enantioselectivity (entries 7 and 8). When a lower catalyst loading is employed $(2.0 \text{ mol\% copper})$, a turnover number greater than 25 is observed (entry 9). A discrete, isolable bisphosphine copper(I) complex, [**P**CuCl] (Cu_A) , ^{[33,](#page-201-1)[34](#page-201-2)} may be used in place of CuCl/**P** (entry 10). Whereas the photoinduced, coppercatalyzed coupling occurs in good yield at -78 °C, the corresponding S_N2 reaction does not proceed to a significant extent even at 80 °C (entry 11).

An array of racemic tertiary α -halonitriles serve as suitable electrophiles in these enantioconvergent N-alkylations. For example, consistently good enantioselectivity is observed as the R^1 group varies from Me to *i*-Bu (**Figure 2.3**, entry 1–3; on a gram scale of entry 2 (1.28 g of product), the coupling proceeds in 64% yield, 92% ee), although the yield is sensitive to the size of R^1 . Functional groups such as an ester, olefin, unactivated alkyl chloride, aryl fluoride, aryl chloride, aryl bromide, amide, and phosphonate are compatible with the method (entry 5–12; also, an acetal, alcohol, alkylboronate ester, alkyl bromide, alkyl iodide, aryl iodide, aryl triflate, benzofuran, epoxide, ketone, sulfide, and tertiary amine: see the **Section 2.4.5**). Not only α-aryl-α-halonitriles (entry 1–10), but also α-acyl- and αphosphonyl-substituted (entry 11 and 12; Br as the leaving group) α-halonitriles, serve as suitable coupling partners.

Figure 2.3. Photoinduced, copper-catalyzed enantioconvergent alkylations of anilines by racemic tertiary α-halonitrile: scope of electrophiles. Reactions were conducted on a 0.8 mmol scale, and yields are for purified compounds. All data are the average of two runs. X $=$ Cl, unless otherwise noted. $aX = Br$.

The scope of this photoinduced, copper-catalyzed asymmetric N-alkylation is also reasonably broad with respect to the arylamine nucleophile. In the case of aniline itself, a substantial amount of addition of the electrophile to the para position is observed (C–C coupling: 24% yield, 27% ee; C–N coupling: 18% yield, 86% ee). However, if the para position bears a substituent, moderate-to-good yields and good enantioselectivities are obtained with either an electron-poor or an electron-rich aniline as the nucleophile (**Figure 2.4**, entry 13–18). Furthermore, meta substitution of the aniline can sufficiently impede addition to the para position such that the desired N-alkylation proceeds in fair yield as well as good enantioselectivity (entry 19–21).

Figure 2.4. Photoinduced, copper-catalyzed enantioconvergent alkylations of anilines by racemic tertiary α-halonitrile: scope of nucleophiles. Reactions were conducted on a 0.8 mmol scale, and yields are for purified compounds. All data are the average of two runs. X = Cl, unless otherwise noted. Bpin, pinacolboryl.

It is noteworthy that enantioconvergent N-alkylation can be achieved with electrophiles that bear a leaving group other than chloride or bromide. In the case of a carbonate or a fluoride, although substitution does not occur in good yield and ee under the standard conditions, the addition of TBACl/B(OMe)₃ enables the desired C–N bond formation to proceed smoothly with good enantioselectivity (**eq 2.1**). To our knowledge, this is the first example of an alkyl fluoride serving as a suitable electrophile in a metal-catalyzed enantioconvergent substitution by a nitrogen nucleophile.

The standard reaction conditions can be applied to the enantioconvergent N-alkylation of anilines by racemic tertiary electrophiles that lack a cyano group. Thus, α-chloroamides are also suitable substrates, leading to coupling with good ee for both N-alkyl and N-aryl secondary amides (**Figure 2.5**, entry 22–26). Furthermore, the chiral catalyst can provide promising enantioselectivity when it is necessary to distinguish between two alkyl substituents on the electrophilic carbon, such as secondary vs. methyl (entry 25) or branched primary vs. methyl (entry 26).

Figure 2.5. Photoinduced, copper-catalyzed enantioconvergent alkylations of anilines by tertiary *α*-haloamides as electrophiles. Reactions were conducted on a 0.8-mmol scale, and yields are for purified compounds. All data are the average of two runs. $X = Cl$, unless otherwise noted. $a X = Br$.

2.2.2. Mechanistic studies: overview

Our current working hypothesis is that these photoinduced, copper-catalyzed enantioconvergent C–N couplings may be proceeding through the pathway outlined in **Figure 2.6**. Thus, copper(I) complex **CuA**, which is a competent catalyst for the coupling (**Table 2.1**, entry 8; 10 mol% **CuA**, rather than 10 mol% CuCl/12 mol% **P**, was utilized in all catalyzed reactions for our mechanistic studies) undergoes excitation upon blue-LED irradiation to generate **CuA***, which reacts with electrophile R–Cl to afford organic radical

R• and copper(II) complex Cu_B. Complex Cu_B then undergoes base-induced substitution by the aniline to furnish copper complex Cu , which combines with $R \cdot$ to provide the coupling product and regenerate copper(I) complex **CuA**.

Figure 2.6. Outline of a possible mechanism for the photoinduced, copper-catalyzed enantioconvergent alkylation of anilines by racemic tertiary electrophiles. $P^{\frown}P = P$.

2.2.3. Mechanistic studies: Cu^A

A solution of **Cu^A** in toluene at –78 °C shows a characteristic absorption at 376 nm (ε $= 3890$ M⁻¹ cm⁻¹), as does the reaction mixture (the coupling depicted in Table 1) prior to irradiation (**Figure 2.7a**); because the concentrations of copper for the two UV–vis spectra are the same, virtually all of the copper in the reaction mixture appears to be present as **Cu^A** prior to irradiation (see the **Section 2.4.10.5**). As illustrated in **Figure 2.7b**, roomtemperature ${}^{1}H$ and ${}^{31}P$ NMR spectroscopic analysis is consistent with this conclusion $(>95\%)$.^{[35](#page-201-3)} During a catalytic C–N coupling, the concentration of **Cu**A decreases somewhat as the reaction progresses, to 54% of total copper at 64% conversion, according to ${}^{1}H$ NMR spectroscopy at –78 °C (see the **Section 2.4.10.5**).

a. Absorption and emission spectra

Figure 2.7. Cu^A as a major component of the reaction mixture. a) Absorption and emission spectra of Cu_A (0.5 mM), reaction mixture ($|Cu_A|$ = 0.5 mM, $[2$ -chloro-2phenylbutanenitrile] = 5 mM, $[p$ -toluidine] = 6 mM, and $[BTPP] = 6$ mM), blue-LED lamp. b) ¹H NMR and ³¹P NMR spectroscopy: comparison of a reaction mixture prior to irradiation versus **CuA**.

Having identified Cu_A as a major component of the reaction mixture, we carried out an investigation of its photophysical properties. The steady-state emission spectrum of **CuA*** shows a single emission band (λ_{max} = 480 nm) at temperatures ranging from 77 K to room temperature (see the **Section 2.4.10.6**). The luminescence decay of **CuA*** observed at 480

nm at room temperature shows two sets of decay curves, with lifetimes of 3.8 ns and 0.36 μ s (**Figure 2.8**).

Figure 2.8. Luminescence lifetime of **CuA*** showing two sets of decay curves.

To avoid complications arising from the two luminescence-decay pathways, we measured the lifetime of a non-emissive excited state of **Cu^A** by transient absorption spectroscopy (λ_{pump} = 355 nm, λ_{probe} = 580 nm) as a function of electrophile concentration at room temperature (**Figure 2.9**). Addition of 2-chloro-2-phenylbutane-nitrile results in a decrease of the excited-state lifetime of **CuA*** that allows for the determination of the secondorder rate constant, $k_q = 3.7 \times 10^8 \text{ M}^{-1} \text{ s}^{-1}$, for the quenching process. The excited-state potential of CuA^* is estimated to be $E_{1/2}$ (Cu^{III^*}) ~ -2.6 V (vs Fc/Fc⁺), based on E^{00} (3.0 eV) and $E_{1/2}$ (Cu^{II/I}; ~ 0.4 V vs Fc/Fc⁺; irreversible), which are derived from emission spectroscopy and electrochemical characterization by cyclic voltammetry (CV); a CV of the model electrophile shows an irreversible feature at $E_P \sim -2.2$ V (see the **Section 2.4.10.8**).

Figure 2.9. Stern-Volmer quenching of CuA^* by 2-chloro-2-phenylbutanenitrile ($\lambda_{\text{pump}} =$ 355 nm, $\lambda_{\text{probe}} = 580$ nm).

2.2.4. Mechanistic studies: steps 1 and 2 of the catalytic cycle

No reaction between Cu_A and 2-chloro-2-phenylbutanenitrile occurs in toluene at – 78 °C after 30 min in the dark, as determined by ¹H NMR spectroscopy. However, when this mixture is irradiated at -78 °C for 30 min, the initially colorless solution turns dark purple, and analysis by ${}^{1}H$ NMR spectroscopy shows that a C–C coupled dimer derived from the electrophile is formed (**Figure 2.10**: 43% yield based on **CuA**; ~ 1:1 mixture of diastereomers). Formation of the dimer is readily accommodated by the reaction mechanism illustrated in **Figure 2.6**; specifically, excitation of **Cu^A** (step 1), followed by chlorine atom transfer, furnishes R• (step 2), which engages in radical–radical coupling in the absence of aniline (this R–R dimer is observed as a minor side product in the photoinduced, coppercatalyzed C–N coupling of this electrophile under the standard conditions; see **Section 2.4.10.2**).

Stoichiometric reactivity

Figure 2.10. Steps 1 and 2 of the catalytic cycle: X-band CW-EPR spectra of **Cu^B** from a photoinduced reduction of 2-chloro-2-phenylbutanenitrile by Cu_A ($|Cu_A| = 5$ mM and $[2$ chloro-2-phenylbutanenitrile] = 50 mM; black), independent synthesis of Cu **B** (red; X-ray crystal structure: thermal ellipsoids at 50% probability (solvents and hydrogen atoms are omitted for clarity)), and a catalyzed reaction after 30 min at -78 °C (blue); acquisition parameters: MW frequency = 9.37 GHz, MW power = 140 μ W, modulation amplitude = 0.4 mT, conversion time = 5.02 ms, and temperature = 77 K.

2.2.5. Mechanistic studies: Cu^B

X-band continuous-wave (CW) EPR spectroscopy of the dark-purple solution obtained from this photoinduced reduction of 2-chloro-2-phenylbutanenitrile by **Cu^A** shows a signal characterized by large ³¹P hyperfine couplings from two phosphorous atoms (**Figure 2.10**, black spectrum). We hypothesized that this paramagnet is $PCuCl₂$ (Cu_B), which we then independently synthesized from P and CuCl₂ (toluene, r.t. to -78 °C) and crystallographically characterized (**Figure 2.10**; see the **Section 2.4.10.10**); the CW-EPR spectrum of this **Cu^B** at 77 K matches that of the photoinduced reduction of 2-chloro-2-phenylbutanen-itrile by **Cu^A** (**Figure 2.10**, black and red spectra). Upon monitoring the standard catalyzed C–N coupling reaction (**Table 2.1**), we determined that **CuB** is the predominant, but not the sole, paramagnetic species after 30 min of irradiation (**Figure 2.10**, blue spectrum; 82% conversion of the electrophile, 66% yield of the coupling product).

2.2.6. Mechanistic studies: Cu^C and step 3 of the catalytic cycle

X-band CW-EPR spectroscopic analysis of a catalytic reaction after 10 min of irradiation shows that a different copper(II) species is dominant at the beginning of the coupling process (**Figure 2.11**, red spectrum; 32% conversion of the electrophile, 22% yield of the coupling product; see the **Section 2.4.10.11**). We have determined that a matching EPR spectrum is observed upon treating Cu_B with *p*-toluidine in the presence of BTPP at – 90 °C (**Figure 2.11**, black spectrum; if either *p*-toluidine or BTPP is absent, the black EPR spectrum is not observed). Under the same conditions, use of a different copper halide complex (Br instead of Cl) or a different base (2-*tert*-butyl-1,1,3,3-tetramethylguanidine (BTMG) instead of BTPP) leads to no noticeable change in the black EPR spectrum (including superhyperfine structures resolved in their second derivatives; see **Section 2.4.10.13**), consistent with the paramagnet being a copper(II) complex in which neither the halide nor the Brønsted base is bound. We therefore postulated that this complex might be three-coordinate $[PCu(NHAr)]X (Cuc)$, $[PCu=NAr]$, or four-coordinate $[PCu(NHAr)_2]$.

Figure 2.**11.** X-band CW-EPR spectra of a catalyzed reaction after 5 min at –78 °C (red) and of Cu_C prepared independently (black); acquisition parameters: MW frequency = 9.36 GHz, MW power = 140 μ W, modulation amplitude = 0.4 mT, conversion time = 5.02 ms, and temperature $= 77$ K.

This paramagnetic copper complex (Cu _C) is unstable at -78 °C, decomposing over 30 min (as observed by CW-EPR spectroscopy, see the **Section 2.4.10.12**) and forming (*E*)-1,2 di-*p*-tolyldiazene (this dimer is observed as a minor side product in photoinduced, coppercatalyzed N-alkylations of *p*-toluidine under the standard coupling conditions). Because the thermal instability of **Cu^C** frustrated our attempts at crystallographic characterization, we investigated its structure through EPR spectroscopy (**Figure 2.12**). The X-band EPR signal of **Cu**c is dominated by hyperfine coupling to ^{63/65}Cu ($I = 3/2$) and two inequivalent ³¹P ($I =$ 1/2) nuclei, without any additional hyperfine splitting being clearly resolved (see **Section 2.4.10.14**).

Figure 2.12. X-band CW-EPR spectra (left panel) and $2nd$ derivative (right panel) of Cu_C in toluene and isotopologues of p -toluidine; acquisition parameters: MW frequency $=$ 9.372–9.374 GHz, MW power = 140 μ W, modulation amplitude = 0.1 mT, conversion time $= 5.3$ ms, and temperature $= 77$ K.

Figure 2.13. Q-band HYSCORE of Cu_c in toluene of isotopologues of *p*-toluidine (left) measured at 1206 mT ($g = 2.015$) with overlay of ^{14/15}N simulation contours (right, red) with experimental contours (right, gray); experimental conditions: MW frequency $=$ 34.005 GHz, $\tau = 128$ ns, $t_1 = t_2 = 100$ ns, $\Delta t_1 = \Delta t_2 = 12$ ns, shot repetition time (srt) = 1.5 ms, and temperature $= 30$ K.

We investigated the incorporation of *p*-toluidine in the spin system using isotopically labeled *p*-toluidine (*p*-toluidine-¹⁵*N* and *p*-toluidine-*ND2*). The X-band CW-EPR spectra of these isotopologues show very subtle differences in the second-derivative plots (**Figure 2.12**, bottom panel). For quantitative determination of the natural abundance ^{14}N and introduced 2 H and ${}^{15}N$ couplings, we turned to pulsed EPR spectroscopy. Field-dependent Q-band hyperfine sublevel correlation (HYSCORE) spectra of ¹⁵N labeled **Cu**_C show a single set of elongated correlation ridges in the $(-,+)$ quadrant, indicating that the coupling falls into the strongly coupled regime where $A > 2v_I$ (Figure 2.13, bottom panel).^{[36](#page-201-0)} These features are well simulated by a *single class* of highly anisotropic ¹⁵N hyperfine tensor with $A(^{15}N) = \pm [6, 90, 10]$ 6] MHz, $a_{\text{iso}}(^{15}N) = \pm 24.3$ MHz. The ¹⁴N signals present in the analogous HYSCORE spectra of natural-abundance **Cu^C** also fall into the strong-coupling regime but are additionally complicated by the influence of **¹⁴**N nuclear quadrupole interaction (**Figure 2.13**, top panel). These spectra are well simulated by scaling the ^{15}N hyperfine tensor determined from ^{15}N HYSCORE by the proportion of ¹⁵N/¹⁴N gyromagnetic ratios ($|\gamma^{15}N|/|\gamma^{14}N| = 1.403$), with further variation of the ¹⁴N nuclear quadrupole parameters. The simulated **¹⁴**N quadrupole parameters are identical to those determined for the amide nitrogen of structurally relevant Ni(III)-amide and -anilido species using similar HYSCORE spectroscopic methods,^{[37](#page-201-1)} and they are consistent with the presence of a trisubstituted nitrogen with a lone pair oriented roughly orthogonal to the trigonal plane, as expected for a Cu–NHAr moiety. Q-band HYSCORE spectroscopy of ²H-labeled **Cu^C** shows a single set of intense cross-peaks in the $(+,+)$ quadrant that thus fall into the weakly coupled regime where $A < 2v_I$ (**Figure 2.14**), which are well-simulated by a *single class* of relatively anisotropic ²H hyperfine tensor with $A(^{2}H) = \pm [8.9, 4.3, 1.5] \text{ MHz}, a_{iso}(^{2}H) = \pm 4.9 \text{ MHz}.$

The presence of multiple equivalent NHAr ligands could be ruled out by simulations that showed that the equivalent ^{14/15}N nuclei would generate combination peaks^{[38](#page-201-2)[,39](#page-201-3)} that are absent in the observed spectra (see the **Section 2.4.10.14**). Additionally, multiple equivalent **¹**H couplings of the class detected directly from the **²**H HYSCORE could not be accommodated by simulations of the X-band CW-EPR spectra. Collectively, the spectroscopic data are consistent with the presence of a single trisubstituted nitrogen with a single N–H, i.e., **Cu^C** as [**P**Cu(NHAr)]Cl.

Figure 2.14. Q-band HYSCORE of **Cu^C** in toluene of isotopologues of *p-*toluidine (left) measured at 1206 mT ($g = 2.015$) with overlay of ²H simulation contours (right, red) with experimental contours (right, gray); experimental conditions: MW frequency = 34.005 GHz, $\tau = 128$ ns, $t_1 = t_2 = 100$ ns, $\Delta t_1 = \Delta t_2 = 12$ ns, shot repetition time (srt) = 1.5 ms, and temperature $= 30$ K.

Further support for the proposed structure of **Cu**_{**C**} has been obtained by comparing the experimentally derived CW-EPR parameters to values predicted by DFT calculations for the various possible structures. Only three-coordinate [**P**Cu(NHAr)]Cl is predicted to have EPR parameters similar to those observed (see the **Section 2.4.11**). Spin-density calculations of **Cu** c indicate that there is considerable spin density on the aniline ligand (0.33 e^- on NH, 0.32 e – on the aromatic ring) and less spin density on copper (0.15 e–) (**Figure 2.15**), indicating that **Cu^C** is more accurately viewed as a copper(I)–(anilidyl radical) complex, rather than as its formal assignment as a copper (II) –anilido complex.^{[40](#page-201-4)}

Figure 2.15. Calculated spin-density plot of **Cu**_c viewed perpendicular to the NHAr plane and at a 45° angle (bp86 def2-TZVP; contour value = 0.005).

2.2.7. Mechanistic studies: step 4 of the catalytic cycle

We have shown that **Cu**_c can couple with an organic radical to afford a new C–N bond. Thus, generation of **Cu**_C at low temperature, followed by the addition of a persistent tritylderived radical, leads to N-alkylation in 41% yield (**Figure 2.16a**). Based on the spin density at the nitrogen center and the sterically congested copper center of **CuC**, we propose that the tertiary alkyl radical derived from the electrophile undergoes C–N bond formation through direct radical-radical coupling between the carbon of the electrophile and the nitrogen of **CuC**, rather than through carbon-copper bond formation (**Figure 2.16b**). Similarly, the substantial spin density at the para carbon of the aniline $(0.17 e⁻)$ aligns with our observation of significant C–C bond formation at that position when it is not blocked, through the radicalradical coupling. [41](#page-201-5) The lower ee of the C−C coupling process (**section 2.2.1**) may be due to the greater distance of the site of bond formation from the chiral environment of the copper complex.

a. Step 4 of the catalytic cycle

b. Proposed mechanism of C-N coupling

Figure 2.16. Step 4 of the catalytic cycle a) Stoichiometric reaction of Cu_c with persistent alkyl radical. b) Proposed mechanism of C–N bond formation.

2.3. Conclusions

We have developed a photoinduced, copper-catalyzed method for the enantioconvergent N-alkylation of anilines by an array of racemic tertiary alkyl electrophiles that generates fully-substituted stereocenters with good ee. The catalyst, composed of commercially available components, effects asymmetric C–N bond formation at –78 °C, whereas the corresponding uncatalyzed coupling does not proceed at a significant rate even at 80 °C. Although the use of alkyl chlorides as electrophiles is the primary focus of this study, examples have been presented of the use of other electrophiles, specifically, an alkyl fluoride and an alkyl carbonate, under similar conditions. Mechanistic studies have provided support for key intermediates and elementary steps in the proposed catalytic cycle. Future investigations will explore the development of other enantioselective bond-forming processes that employ catalysts based on earth-abundant copper.

2.4. Experimental section

2.4.1. General information

Chemicals. Unless otherwise noted, reagents purchased from commercial suppliers were used as received. CuCl, (S) -**P**, and (R) -**P** were purchased from Strem. BTPP was purchased from Sigma-Aldrich and stored inside a glovebox freezer $(-30 \degree C)$; as needed, BTPP was filtered through a PTFE syringe filter $(0.45 \mu m, ImoSep^{TM})$ into a vial, the vial was capped with a PTFE-lined cap, and the vial was removed from the glovebox. Tetrabutylammonium chloride was purchased from Sigma Aldrich and gently ground under a nitrogen atmosphere before use. Unless otherwise noted, anilines purchased from commercial sources were purified by distillation or sublimation before use. Anhydrous toluene (99.8%) was purchased from Sigma-Aldrich, stored under nitrogen, and used with standard air-free techniques; other solvents were purified by passage through activated aluminum oxide in a solvent-purification system. All solvents for mechanistic studies were further degassed by freeze-pump-thaw cycles (five cycles) and stored over molecular sieves under a nitrogen atmosphere; the quality of the solvents was tested before use, using benzophenone-ketyl radical in THF. All manipulations of air‐sensitive materials were carried out in oven-dried glassware under a nitrogen atmosphere using standard Schlenk or glovebox techniques. Flash column chromatography was performed using silica gel (SiliaFlash® P60, particle size 40-63 μm, Silicycle). Analytical thin-layer chromatography was conducted with glass TLC plates (silica gel 60 F254), and spots were visualized under UV light or after treatment with standard TLC stains.

NMR spectroscopy. NMR spectra were collected on a Bruker 400 MHz or a Varian 500 MHz spectrometer at ambient temperature; chemical shifts (δ) are reported in ppm downfield from tetramethylsilane, using the solvent resonance as an internal standard.

Other analyses. HPLC analyses were carried out on an Agilent 1100 series system with Daicel CHIRALPAK[®] or Daicel CHIRALCEL[®] columns $(4.6 \times 250$ mm, particle size 5 μm). GC analyses were obtained on an Agilent 6890N GC. HRMS were acquired on an Agilent 1260 Infinity II HPLC-MS system in electrospray ionization (ESI+) mode or a JEOL AccuTOF GCx Plus system in field ionization (FI+) mode. FT-IR measurements were carried out on a VERTEX 80 FT-IR purge spectrometer. Absorbance spectra were acquired on a Cary 50 UV-Vis spectrophotometer. Optical-rotation data were obtained with a Jasco P-2000 polarimeter at 589 nm, using a 100 mm pathlength cell in the solvent and at the concentration indicated. Elemental analyses were carried out in the Beckman Institute Crystallography Facility on a PerkinElmer 2400 Series II CHN Elemental Analyzer.

X-ray crystallography. X-ray crystallographic studies were carried out in the Beckman Institute Crystallography Facility on a Bruker APEX-II CCD diffractometer with filtered Cu-Kα radiation or Mo-Kα radiation. Suitable single crystals for X-ray structure determination were selected from the mother liquor and mounted in a nylon loop in immersion oil. The structures were solved and refined using APEX 3 software. Structure solution was performed using the SHELXT structure solution program using direct methods. Refinement was performed using the SHELXL refinement package using least squares minimization. All non-hydrogen atoms were refined with anisotropic displacement parameters. All C–H hydrogen atoms were refined isotropically on calculated positions by using a riding model with their U_{iso} values constrained to 1.5 U_{eq} of their pivot atoms for terminal sp³ carbon atoms and 1.2 times for all other atoms.

Photoinduced reactions. Photoinduced reactions were performed using Kessil[®] PR160-440 nm lamps. The light source was placed approximately 5 cm above the sample, and the reaction mixture was stirred at 500 rpm using a magnetic stir bar. Temperature control was maintained with a methanol bath cooled by an SP Scientific cryostat. For reactions longer than 24 h, the methanol bath was replaced with a fresh bath after 24 h to prevent the accumulation of ice.

Photophysical measurements. Steady-state fluorimetry and time-resolved transient absorption and luminescence measurements were performed in the Beckman Institute Laser Research Center. Room-temperature samples for transient absorption and luminescence measurements were prepared with dry, degassed solvents inside a nitrogen-filled glovebox and transferred to a 1 cm pathlength fused quartz or glass cuvette (Starna Cells) which was sealed with a high-vacuum Teflon valve (Kontes) or a PTFE-lined open-top screw cap (Starna Cells). Samples for low-temperature measurements were prepared in a 4 mm EPR tube (Wilmad or Norell) sealed with a rubber stopper. Steady-state emission spectra were collected on a Jobin S4 Yvon Spec Fluorolog-3-11 with a Hamamatsu R928P photomultiplier tube detector with photon counting.

For luminescence and transient absorption at the nanosecond to microsecond time scale, a Q-switched Nd:YAG laser (Spectra-Physics Quanta-Ray PRO-Series; 355 nm; pulse duration 8 ns, operating at 10 Hz) was used as the source of the excitation pulse, with laser power at 0.5 mJ/pulse. Probe light for transient absorption kinetics measurements was provided by a 75-W arc lamp (PTI Model A 1010) that was operated in continuous wave or pulsed modes. The laser light was aligned so as to be collinear with the arc lamp beam, and the scattered excitation light was rejected with appropriate long pass and short pass filters. Transmitted light from the sample was detected with a photomultiplier tube (Hamamatsu R928). All instruments and electronics in these systems were controlled by software written in LabVIEW (National Instruments).

Transient absorption difference spectra were collected using the same excitation source $(\lambda_{\rm ex} = 355 \text{ nm})$ and a white light flash lamp source with nanosecond durations. All instruments and electronics in these systems were controlled by software written in LabVIEW (National Instruments). Data processing was performed with MatlabR2018a and OriginPro 8.

Luminescence decay measurements at the picosecond time scale were performed as previously described.[42,](#page-201-6)[43](#page-201-7) A mode-locked Nd:YAG laser (Vanguard 2000-HM532; Spectra-Physics) provided 10 ps pulses that were regeneratively amplified (Continuum) and frequency tripled (355 nm excitation). Laser power was reduced to 0.5 mJ/pulse. Fluorescence from the sample was focused onto the entrance slit of a spectrograph (Acton Research Corp SpectraPro 275) through a 355 nm dielectric mirror to reject scattered excitation light. Fluorescence decays were obtained at a spectrograph center wavelength of 420 nm. Decays were collected using a streak camera (C5680; Hamamatsu Photonics) in photon counting mode over a 50 ns window.

CW EPR spectroscopy*.* 77 K X-band CW EPR spectra were obtained on a Bruker EMX spectrometer using a quartz liquid nitrogen immersion Dewar on solutions prepared as frozen glasses in toluene, unless otherwise noted.

Pulse EPR spectroscopy*.* All pulse EPR and electron nuclear double resonance (ENDOR) experiments were carried out using a Bruker ELEXSYS E580 pulse EPR spectrometer. All Q-band data was acquired using a Bruker D2 resonator. Temperature control was achieved using an ER 4118HV-CF5-L Flexline Cryogen-Free VT cryostat (ColdEdge) equipped with an Oxford Instruments Mercury ITC.

Pulse Q-band ENDOR was acquired using the Davies pulse sequence $(\pi - T_{RF} \pi_{RF} - T_{RF} - \pi/2 - \tau - \pi$ – echo), where T_{RF} is the delay between MW pulses and RF pulses, π_{RF} is the length of the RF pulse, and the RF frequency is randomly sampled during each pulse sequence.

Q-band HYSCORE spectra were acquired using the 4-pulse sequence $(\pi/2 - \tau \pi/2 - t_1 - \pi - t_2 - \pi/2$ – echo), where τ is a fixed delay, while t_1 and t_2 are independently incremented by Δt_1 and Δt_2 , respectively. The time domain data was baseline-corrected (third-order polynomial) to eliminate the exponential decay in the echo intensity, apodized with a Hamming window function, zero-filled to eight-fold points, and fast Fourier-transformed to yield the 2-dimensional frequency domain.

In general, the ENDOR spectrum for a given nucleus with spin $I = \frac{1}{2}$ (${}^{1}H$, ${}^{31}P$) coupled to the $S = \frac{1}{2}$ electron spin exhibits a doublet at frequencies (eq. 2.2)

$$
\nu_{\pm} = \left| \frac{A}{2} \pm \nu_N \right| \tag{2.2}
$$

where v_N is the nuclear Larmor frequency and A is the hyperfine coupling. For nuclei with $I \ge 1$ (¹⁴N, ²H), an additional splitting of the v_{\pm} manifolds is produced by the nuclear quadrupole interaction (P) (**eq. 2.3**)

$$
\nu_{\pm,m_I} = \left| \nu_N \pm \frac{3P(2m_I - 1)}{2} \right| \tag{2.3}
$$

In HYSCORE spectra, these signals manifest as cross-peaks or ridges in the 2-D frequency spectrum that are generally symmetric about the diagonal of a given quadrant. This technique allows hyperfine levels corresponding to the same electron-nuclear submanifold to be differentiated, as well as separating features from hyperfine couplings in the weak-coupling regime $(|A| < 2|\nu_I|)$ in the $(+,+)$ quadrant from those in the strong coupling regime ($|A| > 2|\nu_I|$) in the (-,+) quadrant. Because the (-,-) and (+,-) quadrants of these frequency spectra are symmetric to the $(+,+)$ and $(-,+)$ quadrants, only two of the quadrants are typically displayed in the literature.

Figure 2.17. a) HYSCORE powder patterns for an $S = \frac{1}{2}$, $I = \frac{1}{2}$ spin system with an isotropic hyperfine tensor A. b) HYSCORE powder patterns for an $S = \frac{1}{2}$, $I = \frac{1}{2}$ spin system with an axial hyperfine tensor that contains isotropic (a_{iso}) and dipolar (T) contributions. Blue correlation ridges represent the strong coupling case; red correlation ridges represent the weak coupling case.

For systems with appreciable hyperfine anisotropy in frozen solutions or solids, HYSCORE spectra typically do not exhibit sharp cross peaks, but show ridges that represent the sum of cross peaks from selected orientations within the excitation bandwidth of the MW pulses at the magnetic field position at which the spectrum is collected. The length and curvature of these correlation ridges can allow for the separation and estimation of the magnitude of the isotropic and dipolar components of the hyperfine tensor (**Figure 2.17**).

EPR simulations. Simulations of all CW and pulse EPR data were achieved using the EasySpin[44](#page-202-0) simulation toolbox (release 5.2.33) with Matlab 2020b using the following Hamiltonian (**eq. 2.4**):

$$
\widehat{H} = \mu_B \overrightarrow{B}_0 g \hat{S} + \mu_N g_N \overrightarrow{B}_0 \hat{I} + h \hat{S} \cdot A \cdot \hat{I} + h \hat{I} \cdot P \cdot \hat{I}
$$
\n(2.4)

In this expression, the first term corresponds to the electron Zeeman interaction term where μ_B is the Bohr magneton, g is the electron spin g-value matrix with principal components $g = [g_{xx} g_{yy} g_{zz}]$, and \hat{S} is the electron spin operator; the second term corresponds to the nuclear Zeeman interaction term where μ_N is the nuclear magneton, g_N is the characteristic nuclear g-value for each nucleus (e.g. ^{1}H , ^{2}H , ^{31}P) and \hat{I} is the nuclear spin operator; the third term corresponds to the electron-nuclear hyperfine term, where \bm{A} is the hyperfine coupling tensor with principal components $\boldsymbol{A} = [A_{xx}, A_{yy}, A_{zz}]$; and for nuclei with $I \geq 1$, the final term corresponds to the nuclear quadrupole (NQI) term which arises from the interaction of the nuclear quadrupole moment with the local electric field gradient (efg) at the nucleus, where \bm{P} is the quadrupole coupling tensor. In the principal axis system (PAS), **P** is traceless and parametrized by the quadrupole coupling constant e^2Qq/h and the asymmetry parameter η such that (**eq. 2.5**):

$$
\boldsymbol{P} = \begin{pmatrix} P_{xx} & 0 & 0 \\ 0 & P_{yy} & 0 \\ 0 & 0 & P_{zz} \end{pmatrix} = \frac{e^2 Q q / h}{4I(2I - 1)} \begin{pmatrix} -(1 - \eta) & 0 & 0 \\ 0 & -(1 + \eta) & 0 \\ 0 & 0 & 2 \end{pmatrix} \tag{2.5}
$$

where $\frac{e^2 Q q}{h}$ $\frac{d^2 Q q}{h} = 2I(2I - 1)P_{zz}$ and $\eta = \frac{P_{xx} - P_{yy}}{P_{zz}}$ $\frac{x-y_y}{P_{zz}}$. The asymmetry parameter may have values between 0 and 1, with 0 corresponding to an electric field gradient (EFG) with axial symmetry and 1 corresponding to a fully rhombic EFG.

The orientations between the hyperfine and NQI tensor principal axis systems and the g-matrix reference frame are defined by the Euler angles $(α, β, γ)$, with rotations performed within the zyz convention where α rotates xyz counterclockwise about z-axis to give x'y'z', β rotates x'y'z counterclockwise about y'-axis to give x",y",z", and γ rotates xyz counterclockwise about z"-axis to give final frame orientation.

2.4.2. Preparation of electrophiles

The yields have not been optimized.

General Procedure 1 (GP-1): Preparation of *α***-chloronitriles.** An oven-dried 250 mL round-bottom flask charged with a magnetic stir bar and capped with a rubber septum was placed under a nitrogen atmosphere by evacuating and backfilling the flask (three cycles). Anhydrous tetrahydrofuran (to form a 0.4 M solution of LDA) and anhydrous diisopropylamine (1 equiv) were added to the flask, and then the solution was cooled to $-$ 78 °C. Next, a solution of *n*-butyllithium (1 equiv; in hexane) was added into the flask over 1 min, and the resulting solution was stirred at –78 °C for 15 min. The substrate was added dropwise over 2 min, and then the reaction mixture stirred at -78 °C for an additional 40 min. Next, the alkyl bromide or iodide (1.1-1.2 equiv) was added dropwise into the reaction mixture over 5 min. The reaction mixture was slowly warmed to room temperature and stirred overnight. Then, the reaction mixture was acidified with 2 N aqueous HCl, and the organic layer was separated and dried over Na2SO4. The solid was removed via filtration, and the filtrate was concentrated in vacuo. The product was purified by flash column chromatography on silica gel (hexanes/dichloromethane = $10/1 \rightarrow 5/1$).

Next, an oven-dried 100 mL round-bottom flask was charged with the 2-alkyl-2-phenyl acetonitrile, PCl₅ (1.1-2.0 equiv), anhydrous 1,2-dichloroethane (1 equiv), and a magnetic stir bar. The flask was then equipped with a reflux condenser and a CaCl₂ drying tube. The reaction mixture was heated to 80 °C, and the reaction mixture was monitored by TLC. After the starting material had been consumed, the reaction mixture was cooled to room temperature and poured into ice (3 g/mmol of substrate). Dichloromethane (30 mL) was added, and then the organic layer was separated, dried over Na2SO4, filtered, and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (hexanes/dichloromethane = $10/1 \rightarrow 5/1$, unless otherwise noted).

2-Chloro-2-phenylpropanenitrile. The title compound was synthesized according to **GP-1** from 2-phenylpropanenitrile (3.28 g, 25.0 mmol), PCl⁵ (6.25 g, 30.0 mmol), and 1,2 dichloroethane (2.0 mL, 25 mmol). The product was purified by flash column chromatography on silica gel. 1.81 g (10.9 mmol, 44% yield). Colorless oil.

¹H NMR (500 MHz, CDCl3) δ 7.68 (d, *J =* 7.8 Hz, 2H), 7.49 – 7.39 (m, 3H), 2.29 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 138.3, 129.9, 129.2, 125.5, 119.4, 57.1, 33.0.

FT-IR (film): 3065, 3039, 2994, 2937, 2243, 1492, 1452, 1380, 1229, 1145, 1074, 1038, 785, 745, 694, 650 cm⁻¹.

HRMS (FI+) m/z [M]⁺ calcd for C₉H₈ClN⁺: 165.0340, found: 165.0337.

2-Chloro-2-phenylbutanenitrile. The title compound was synthesized according to **GP-1** from 2-phenylbutanenitrile (9.73 g, 67.0 mmol), PCl⁵ (14.6 g, 70.0 mmol), and 1,2 dichloroethane (5.3 mL, 67 mmol). The product was purified by flash column chromatography on silica gel. 7.69 g (42.8 mmol, 64% yield). Colorless oil.

¹H NMR (500 MHz, CDCl3) δ 7.64 (d, *J =* 7.3 Hz, 2H), 7.47 – 7.38 (m, 3H), 2.47 (dq, *J =* 14.5, 7.3 Hz, 1H), 2.38 (dq, *J =* 14.4, 7.3 Hz, 1H), 1.16 (t, *J =* 7.3 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 137.3, 129.8, 129.1, 125.9, 118.3, 63.4, 39.0, 10.4.

FT-IR (film): 3064, 2982, 2941, 2881, 2242, 1492, 1450, 1207, 1080, 1002, 955, 908, 833, 751, 694 cm⁻¹.

HRMS (FI+) m/z [M]⁺ calcd for C₁₀H₁₀ClN⁺: 179.0496, found: 179.0494.

2-Chloro-4-methyl-2-phenylpentanenitrile. The title compound was synthesized according to **GP-1** from 2-phenylacetonitrile (5.86 g, 50.0 mmol), diisopropylamine (7.1 mL, 50 mmol), *n*-butyllithium (20.0 mL of 2.5 M solution, 50.0 mmol), 1-bromo-2 methylpropane (6.0 mL, 55 mmol), and THF (120 mL) for the first step, and from the

intermediate (53.1 mmol), $PCl₅$ (16.2 g, 78 mmol), and 1,2-dichloroethane (10 mL, 120 mmol) for the second step. The product was purified by flash column chromatography on silica gel. 3.86 g (18.6 mmol, 18% yield over 2 steps). Colorless oil.

¹H NMR (500 MHz, CDCl3) δ 7.66 (d, *J =* 7.1 Hz, 2H), 7.48 – 7.37 (m, 3H), 2.39 (dd, *J =* 14.4, 6.2 Hz, 1H), 2.32 (dd, *J =* 14.4, 5.9 Hz, 1H), 1.91 – 1.78 (m, 1H), 1.05 (d, *J =* 6.7 Hz, 3H), 0.86 (d, *J =* 6.7 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 137.7, 129.8, 129.1, 126.0, 118.8, 61.6, 53.6, 26.6, 23.40, 23.37.

FT-IR (film): 3065, 2962, 2874, 2242, 1492, 1469, 1450, 1390, 1370, 1199, 754, 697 cm^{-1} .

HRMS (FI+) m/z [M]⁺ calcd for C₁₂H₁₄ClN⁺: 207.0809, found: 207.0808.

2-Chloro-2,3-diphenylpropanenitrile. The title compound was synthesized according to **GP-1** from 2-phenylacetonitrile (4.69 g, 40.0 mmol), diisopropylamine (5.7 mL, 40 mmol), *n*-butyllithium (16.0 mL of 2.5 M solution, 40.0 mmol), benzyl bromide (5.2 mL, 44 mmol), and THF (100 mL) for the first step, and from the intermediate (10.0 mmol), PCl₅ (3.33 g, 16.0 mmol), and 1,2-dichloroethane (2.0 mL, 24 mmol) for the second step. The product was purified by flash column chromatography on silica gel. 1.15 g (4.77 mmol, 38% yield over 2 steps). Pale-yellow solid.

¹H NMR (500 MHz, CDCl₃) δ 7.59 – 7.52 (m, 2H), 7.44 – 7.38 (m, 3H), 7.27 (dt, *J* = 14.4, 7.0 Hz, 3H), 7.06 (d, *J =* 7.2 Hz, 2H), 3.68 (d, *J =* 13.7 Hz, 1H), 3.58 (d, *J =* 13.7 Hz, 1H).

¹³C NMR (126 MHz, CDCl3) δ 136.7, 132.9, 130.9, 129.8, 128.9, 128.4, 128.3, 126.4, 118.0, 62.6, 51.7.

FT-IR (film): 3065, 3033, 2934, 2243, 1497, 1455, 1450, 1253, 1079, 1031, 764, 748, 699, 629 cm⁻¹.

HRMS (FI+) m/z [M]⁺ calcd for C₁₅H₁₂ClN⁺: 241.0653, found: 241.0626.

3-Chloro-3-cyano-3-phenylpropyl acetate. The title compound was synthesized according to GP-1 from 3-cyano-3-phenylpropyl acetate^{[45](#page-202-1)} (2.04 g, 10.00 mmol), PCl₅ (2.28) g, 23.4 mmol), and 1,2-dichloroethane (6.0 mL, 72 mmol). The product was purified by flash column chromatography on silica gel (hexanes/EtOAc = $30/1 \rightarrow 20/1$). 721 mg (3.03 mmol, 30% yield). Pale-yellow oil.

¹H NMR (500 MHz, CDCl3) δ 7.65 (d, *J =* 7.1 Hz, 2H), 7.45 (q, *J =* 8.5, 7.5 Hz, 3H), 4.31 (dt, *J =* 12.5, 6.4 Hz, 1H), 4.19 (dt, *J =* 12.0, 6.3 Hz, 1H), 2.80 (dt, *J =* 14.5, 6.2 Hz, 1H), 2.73 (dt, *J =* 14.7, 6.4 Hz, 1H), 2.00 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 170.8, 136.6, 130.1, 129.3, 125.9, 117.9, 60.1, 59.5, 43.7, 20.7.

FT-IR (film): 3066, 2961, 2903, 2245, 1745, 1493, 1451, 1367, 1230, 1052, 830, 752, 696 cm⁻¹.

HRMS (FI+) m/z [M]⁺ calcd for C₁₂H₁₂ClNO₂⁺: 237.0551, found: 237.0539.

2-Chloro-2-phenylpent-4-enenitrile. The title compound was synthesized according to **GP-1** from 2-phenylacetonitrile (5.86 g, 50.0 mmol), diisopropylamine (7.1 mL, 50 mmol), *n*-butyllithium (20.0 mL of 2.5 M solution, 50.0 mmol), allyl bromide (4.8 mL, 55 mmol), and THF (120 mL) for the first step, and from the intermediate (19.1 mmol), PCl₅ (5.16 g, 24.8 mmol), and 1,2-dichloroethane (10 mL, 120 mmol) for the second step. The product was purified by flash column chromatography on silica gel. 1.00 g (5.22 mmol, 27% yield over 2 steps). Colorless oil.

¹H NMR (500 MHz, CDCl₃) δ 7.69 – 7.60 (m, 2H), 7.51 – 7.37 (m, 3H), 5.74 (ddt, *J* = 17.1, 10.3, 7.1 Hz, 1H), 5.35 – 5.23 (m, 2H), 3.15 (dd, *J =* 14.2, 7.3 Hz, 1H), 3.08 (dd, *J =* 14.2, 6.9 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 136.9, 129.94, 129.91, 129.1, 126.0, 122.3, 118.0, 61.5, 49.4.

FT-IR (film): 3085, 3067, 2985, 2918, 2244, 1491, 1450, 1290, 1265, 1191, 993, 933, 820, 758, 715, 694 cm⁻¹.

HRMS (FI+) m/z [M]⁺ calcd for C₁₁H₁₀ClN⁺: 191.0496, found: 191.0495.

2,4-Dichloro-2-phenylbutanenitrile. The title compound was synthesized according to a modified **GP-1** from 4-hydroxy-2-phenylbutanenitrile⁴ $(1.00 \text{ g}, 6.20 \text{ mmol})$, PCl₅ $(2.71 \text{ g},$ 13.0 mmol), and 1,2-dichloroethane (4.0 mL, 76 mmol). An oven-dried 100 mL roundbottom flask (equipped with a reflux condenser with a CaCl₂-filled drying tube) was charged with 4-hydroxy-2-phenylbutanenitrile, PCl5, anhydrous 1,2-dichloroethane, and a magnetic stir bar. The reaction mixture was stirred at room temperature for 30 min, and then it was heated to 80 °C. After full consumption of the starting material (monitored by TLC), the reaction mixture was cooled to room temperature and poured into ice (10 g). Dichloromethane (30 mL) was added, and then the organic layer was separated, dried over Na2SO4, filtered, and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (hexane/dichloromethane = $10/1 \rightarrow 5/1$). 821 mg (5.52 mmol, 62% yield). Colorless oil.

¹H NMR (500 MHz, CDCl3) δ 7.65 (d, *J =* 7.3 Hz, 2H), 7.51 – 7.42 (m, 3H), 3.72 (td, *J =* 10.9, 5.2 Hz, 1H), 3.60 (td, *J =* 10.9, 5.1 Hz, 1H), 2.90 (ddd, *J =* 14.3, 10.8, 5.2 Hz, 1H), 2.79 (ddd, *J =* 14.4, 10.7, 5.1 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 136.2, 130.4, 129.5, 125.8, 117.4, 59.8, 47.4, 38.8.

FT-IR (film): 3065, 3040, 2967, 2245, 1491, 1450, 1339, 1261, 1182, 837, 758, 694, 672, 652 cm⁻¹.

HRMS (FI+) m/z [M]⁺ calcd for C₁₀H₉Cl₂N⁺: 213.0107, found: 213.0104.

2-Chloro-2-(3-fluorophenyl)butanenitrile. The title compound was synthesized according to **GP-1** from 2-(3-fluorophenyl)acetonitrile (4.73 g, 35.0 mmol),

diisopropylamine (5.3 mL, 38 mmol), *n*-butyllithium (14.4 mL of 2.5 M solution, 36.0 mmol), iodoethane (3.1 mL, 38 mmol), and THF (70 mL) for the first step, and from the intermediate (9.20 mmol), PCl₅ (2.10 g, 10.1 mmol), and 1,2-dichloroethane (2.2 mL, 28) mmol) for the second step. The product was purified by flash column chromatography on silica gel. 842 mg (4.26 mmol, 25% yield over 2 steps). Colorless oil.

¹H NMR (500 MHz, CDCl3) δ 7.47 – 7.38 (m, 2H), 7.35 (d, *J =* 9.8 Hz, 1H), 7.16 – 7.08 (m, 1H), 2.45 (dq, *J =* 14.5, 7.3 Hz, 1H), 2.36 (dq, *J =* 14.4, 7.2 Hz, 1H), 1.16 (t, *J =* 7.3 Hz, 3H).

¹³C NMR (126 MHz, CDCl3) δ 162.8 (d, *J =* 248.4 Hz), 139.6 (d, *J =* 7.3 Hz), 130.7 (d, *J =* 8.2 Hz), 121.6 (d, *J =* 3.1 Hz), 117.7, 116.8 (d, *J =* 21.1 Hz), 113.4 (d, *J =* 24.4 Hz), 62.4, 38.9, 10.2.

¹⁹F NMR (376 MHz, CDCl₃) δ –110.7.

FT-IR (film): 3072, 2983, 2942, 2883, 2245, 1614, 1594, 1490, 1446, 1386, 1254, 1164, 859, 809, 785, 689 cm⁻¹.

HRMS (FI+) m/z [M]⁺ calcd for C₁₀H₉ClFN⁺: 197.0402, found: 197.0400.

2-Chloro-2-(4-chlorophenyl)butanenitrile. The title compound was synthesized according to **GP-1** from 2-(4-chlorophenyl)acetonitrile (7.58 g, 50.0 mmol), diisopropylamine (7.1 mL, 50 mmol), *n*-butyllithium (31.3 mL of 1.6 M solution, 50.1 mmol), iodoethane (4.8 mL, 60 mmol), and THF (120 mL) for the first step, and from the intermediate (14.0 mmol), $PCl₅$ (4.35 g, 20.9 mmol), and 1,2-dichloroethane (2.4 mL, 30 mmol) for the second step. The product was purified by flash column chromatography on silica gel. 1.59 g (7.42 mmol, 37% yield over 2 steps). Colorless oil.

¹H NMR (500 MHz, CDCl3) δ 7.57 (d, *J =* 8.6 Hz, 2H), 7.42 (d, *J =* 8.6 Hz, 2H), 2.45 (dq, *J =* 14.5, 7.3 Hz, 1H), 2.35 (dq, *J =* 14.4, 7.2 Hz, 1H), 1.15 (t, *J =* 7.3 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 135.95, 135.88, 129.4, 127.4, 117.9, 62.6, 39.0, 10.3.

FT-IR (film): 2982, 2941, 2882, 2242, 1596, 1493, 1457, 1403, 1208, 1097, 1015, 917, 848, 815, 764, 722 cm⁻¹.

HRMS (FI+) m/z [M]⁺ calcd for C₁₀H₉Cl₂N⁺: 213.0107, found: 213.0104.

2-Chloro-2-(4-bromophenyl)butanenitrile. The title compound was synthesized according to **GP-1** from 2-(4-bromophenyl)acetonitrile (5.90 g, 30.0 mmol), diisopropylamine (5.1 mL, 36 mmol), *n*-butyllithium (13.2 mL of 2.5 M solution, 33.0 mmol), iodoethane (2.9 mL, 36 mmol), and THF (80 mL) for the first step, and from the intermediate (5.2 mmol), PCl₅ (4.35 g, 5.77 mmol), and 1,2-dichloroethane (2.5 mL, 30 mmol) for the second step. The product was purified by flash column chromatography on silica gel. 843 mg (3.26 mmol, 32% yield over 2 steps). Colorless oil.

¹H NMR (500 MHz, CDCl3) δ 7.58 (d, *J =* 8.5 Hz, 2H), 7.50 (d, *J =* 8.6 Hz, 2H), 2.44 (dq, *J =* 14.5, 7.3 Hz, 1H), 2.34 (dq, *J =* 14.4, 7.2 Hz, 1H), 1.15 (t, *J =* 7.3 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 136.4, 132.3, 127.7, 124.1, 117.8, 62.7, 38.9, 10.3.

FT-IR (film): 2981, 2941, 2881, 2243, 1589, 1490, 1457, 1399, 1208, 1076, 1011, 916, 845, 813 cm⁻¹.

HRMS (FI+) m/z [M]⁺ calcd for C₁₀H₉BrClN⁺: 256.9601, found: 256.9591.

*N-***Benzyl-2-bromo-2-cyano-***N***-phenylbutanamide.** A 250 mL round-bottom flask charged with a magnetic stir bar, *N*-benzyl-2-cyano-*N*-phenylacetamide^{[46](#page-202-2)} (3.30 g, 13.2) mmol), K_2CO_3 (1.83 g, 13.2 mmol), tetrabutylammonium iodide (851 mg, 2.64 mmol), and acetonitrile (37 mL) was capped with a rubber septum. Iodoethane (1.2 mL, 15 mmol) was added, and a nitrogen-filled balloon was attached. The reaction mixture was heated to 70 °C and stirred for 24 h. Next, the solid was removed by filtration, and the filtrate was concentrated in vacuo. *N*-benzyl-2-cyano-*N*-phenylbutanamide was purified by flash

column chromatography on silica gel (10% EtOAc/hexane). 2.65 g (9.52 mmol, 72% yield). White solid.

Next, an oven-dried 100 mL round-bottom flask charged with *N*-benzyl-2-cyano-*N*phenylbutanamide (1.05 g, 3.77 mmol) was capped with a rubber septum and placed under a nitrogen atmosphere by evacuating and backfilling the flask (three cycles). Anhydrous THF (18 mL) was added, and the resulting solution was cooled to 0 °C. Sodium hydride (158 mg, 3.96 mmol, 60% dispersion in mineral oil) was added under a gentle flow of nitrogen, and the reaction mixture was stirred for 30 min. *N*-Bromosuccinimide (805 mg, 4.15 mmol) was added, and the reaction mixture was stirred at 0 \degree C for 2 h. Then, the reaction was quenched by the addition of water (20 mL) . Et₂O (20 mL) was added, and the organic layer was separated. The aqueous layer was extracted with $Et₂O$ (20 mL), and the combined organic layer was dried over $Na₂SO₄$, filtered, and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (hexanes/EtOAc = $20/1 \rightarrow 10/1$). 1.08 g (3.02 mmol, 58% yield over 2 steps). White solid.

¹H NMR (500 MHz, CDCl3) δ 7.44 (t, *J =* 7.0 Hz, 1H), 7.38 (s, 2H), 7.32 – 7.26 (m, 4H), 7.25 – 7.12 (m, 3H), 5.11 (d, *J =* 14.0 Hz, 1H), 4.75 (d, *J =* 14.0 Hz, 1H), 2.62 (dq, *J =* 14.2, 7.1 Hz, 1H), 2.39 (dq, *J =* 14.4, 7.3 Hz, 1H), 1.14 (t, *J =* 7.2 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 162.3, 139.6, 136.0, 130.1, 129.8, 129.4, 129.2, 128.7, 128.1, 115.5, 56.8, 43.6, 36.6, 11.0.

FT-IR (film): 3064, 3031, 2977, 2939, 2230, 1663, 1595, 1496, 1456, 1394, 1260, 1157, 1080, 1014, 690 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for $C_{18}H_{18}BrN_2O^+$: 357.0597, found: 357.0623.

Diisopropyl (1-bromo-1-cyanopropyl)phosphonate. The title compound was synthesized according to a modified literature procedure.^{[47](#page-202-3)} An oven-dried 250 mL roundbottom flask charged with a magnetic stir bar and capped with a rubber septum was placed under a nitrogen atmosphere by evacuating and backfilling the flask (three cycles). Anhydrous THF (40 mL) and anhydrous diisopropylamine (6.7 mL, 48 mmol) were added, and the mixture was cooled to -78 °C. A solution of *n*-butyllithium (18.3 mL, 45.8 mmol, 2.5 M solution in hexane) was slowly added into the flask, and then the reaction mixture was stirred at –78 °C for 15 min. Next, butyronitrile (1.44 g, 20.8 mmol) was added dropwise into the flask over 2 min, and the resulting mixture was stirred at -78 °C for 40 min. Then, a solution of diisopropyl chlorophosphate (4.81 g, 24.0 mmol) in 5 mL of THF was added over 2 min to the reaction mixture. The reaction mixture was stirred at –78 °C for 30 min, and then it was warmed to 0 °C. *N*-Bromosuccinimide (4.27 g, 24.0 mmol) was added, and the reaction mixture was stirred for 2 h. Next, the reaction was quenched with 20 mL of water . Et₂O (20 mL) was added to the mixture, and the organic layer was separated. The aqueous layer was extracted with $Et₂O$ (20 mL), and the combined organic layer was dried over Na2SO4, filtered, and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (hexanes/EtOAc = $10/1$). 4.37 g (14.0 mmol, 70% yield). Pale-brown oil.

¹H NMR (500 MHz, CDCl₃) δ 5.01 – 4.83 (m, 2H), 2.30 (dp, $J = 14.2, 7.1$ Hz, 1H), 2.17 (dp, *J =* 14.8, 7.4 Hz, 1H), 1.48 – 1.37 (m, 12H), 1.30 (t, *J =* 7.2 Hz, 3H).

¹³C NMR (126 MHz, CDCl3) δ 116.0 (d, *J =* 3.9 Hz), 75.6 (d, *J =* 7.4 Hz), 74.9 (d, *J =* 7.6 Hz), 43.3 (d, *J =* 159.7 Hz), 31.2 (d, *J =* 2.0 Hz), 24.4 (d, *J =* 2.8 Hz), 24.1 (d, *J =* 3.6 Hz), 23.8 (d, *J =* 5.5 Hz), 23.6 (d, *J =* 6.5 Hz), 10.7 (d, *J =* 9.1 Hz).

³¹P NMR (162 MHz, CDCl₃) δ 9.31.

FT-IR (film): 2983, 2940, 2882, 2235, 1725, 1457, 1388, 1377, 1268, 1180, 1144, 1102, 988, 826, 766, 730 cm⁻¹.

2-Fluoro-2-phenylbutanenitrile. An oven-dried 250 mL round-bottom flask charged with a magnetic stir bar and capped with a rubber septum was placed under a nitrogen atmosphere by evacuating and backfilling the flask (three cycles). ZnI_2 (3.86 g, 11.0 mmol) and anhydrous dichloromethane (50 mL) were added, and a nitrogen-filled balloon was attached. The reaction mixture was cooled to 0 $^{\circ}$ C, and propiophenone (1.46 mL, 11.0) mmol) was added. TMSCN (3.15 mL, 22.0 mmol) was carefully added dropwise, and the

reaction mixture was warmed to room temperature and stirred overnight. A saturated aqueous solution of NaHCO₃ (50 mL) was added, and the mixture was filtered through a short plug of CeliteTM, washing with dichloromethane (20 mL). The water layer was extracted with dichloromethane (2 x 25 mL), and the combined organic layer was dried over Na2SO4, filtered, and concentrated in vacuo. To the residue was added 30 mL of EtOH and 30 mL of 2 N HCl (aq), and the resulting mixture was stirred at room temperature for 3 h. Deionized water (50 mL) and dichloromethane (50 mL) were added, and then the aqueous layer was extracted with dichloromethane (2 x 30 mL). The combined organic layer was dried over $Na₂SO₄$, filtered, and concentrated in vacuo to afford the cyanohydrin, which was used without further purification. 1.57 g (9.74 mmol, 89% yield).

An oven-dried 50 mL round-bottom flask charged with a magnetic stir bar and capped with a rubber septum was placed under a nitrogen atmosphere by evacuating and backfilling the flask (three cycles). The cyanohydrin (967 mg, 6.00 mmol) and anhydrous dichloromethane (8 mL) were added, and a nitrogen-filled balloon was attached. The reaction mixture was cooled to 0 °C, and then DAST (910 μ L, 6.9 mmol) was added dropwise over 2 min. The reaction mixture was stirred for 90 min, and then a saturated aqueous solution of NaHCO₃ (10 mL) was carefully added at 0 °C. The reaction mixture was diluted with water (10 mL), and the water layer was extracted with dichloromethane (2 x 10 mL). The combined organic layer was dried over Na₂SO₄, filtered, and concentrated in vacuo. The product was purified by column chromatography (hexanes/dichloromethane = 8/1). 825 mg (5.06 mmol, 84% yield). Colorless oil.

¹H NMR (500 MHz, CDCl₃) δ 7.54 – 7.48 (m, 2H), 7.48 – 7.41 (m, 3H), 2.36 – 2.14 (m, 2H), 1.14 (t, *J =* 7.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl3) δ 136.2 (d, *J =* 22.9 Hz), 130.0 (d, *J =* 1.8 Hz), 129.0, 124.8 (d, *J =* 5.9 Hz), 117.4 (d, *J =* 34.1 Hz), 92.8 (d, *J =* 183.9 Hz), 35.2 (d, *J =* 25.9 Hz), 8.3 (d, $J = 4.7$ Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ –148.2.

FT-IR (film): 3067, 3037, 2982, 2944, 2886, 2247, 1494, 1452, 1300, 1229, 1068, 1008, 973, 760, 697 cm⁻¹.

HRMS (FI+) m/z [M]⁺ calcd for C₁₀H₁₀FN⁺: 163.0792, found: 163.0792.

4-Chlorophenyl (1-cyano-1-phenylpropyl) carbonate. An oven-dried 50 mL roundbottom flask charged with a magnetic stir bar and capped with a rubber septum was placed under a nitrogen atmosphere by evacuating and backfilling the flask (three cycles). The cyanohydrin (1.61 g, 10.0 mmol) and dichloromethane 10 mL were added, and a nitrogenfilled balloon was attached. The reaction mixture was cooled to 0° C, and 4-chlorophenyl chloroformate (2.87 g, 15.0 mmol) and pyridine (2.4 mL, 30 mmol) were sequentially added. The reaction mixture was warmed to room temperature and stirred overnight. Next, water was added (5 mL), and then the water layer was extracted with dichloromethane (5 mL x 2). The combined organic layer was dried over $Na₂SO₄$, filtered, and concentrated in vacuo. The product was purified by column chromatography (hexanes/EtOAc $=$ 40/1). 2.90 g (9.18 mmol, 92% yield). Colorless oil.

¹H NMR (500 MHz, CDCl3) δ 7.57 (d, *J =* 7.3 Hz, 2H), 7.49 – 7.38 (m, 3H), 7.30 (d, *J =* 8.8 Hz, 2H), 7.07 (d, *J =* 8.8 Hz, 2H), 2.43 (dq, *J =* 14.5, 7.3 Hz, 1H), 2.20 (dq, *J =* 14.7, 7.4 Hz, 1H), 1.12 (t, *J =* 7.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 150.6, 149.2, 136.1, 131.9, 129.8, 129.7, 129.2, 125.0, 122.2, 116.7, 81.5, 36.0, 8.7.

FT-IR (film): 3066, 2982, 2942, 2884, 2272, 1777, 1488, 1432, 1247, 1090, 1014, 829, 761, 698 cm⁻¹.

General Procedure 2 (GP-2): Preparation of α-chloroamides. A 100 mL three-neck round-bottom flask was charged with a magnetic stir bar, the carboxylic acid, and SOCl₂ $(5-7$ equiv), and then it was fitted with a reflux condenser to which a CaCl₂ drying tube was attached. The resulting solution was heated to reflux for 30 min with vigorous stirring, and then it was cooled to room temperature. *N*-Chlorosuccinimide (2.5 equiv) , $S OCl₂$ (1 equiv), and HCl $(35\%$ in H₂O; 4 drops) were carefully added. The resulting mixture was heated to

reflux for 2.5 h, and then it was cooled to room temperature. Next, with vigorous stirring of the reaction mixture, 20 mL of anhydrous hexane/ $Et_2O(1:2)$ was slowly added, resulting in a black precipitate. The precipitate was removed by filtration, and the volatiles were removed by evaporation. The resulting brown liquid was purified by distillation or directly used without further purification.

Next, an oven-dried 100 mL round-bottom flask capped with a rubber septum was placed under a nitrogen atmosphere by evacuating and backfilling the flask (three cycles). The flask was charged with a magnetic stir bar, anhydrous dichloromethane (0.13-0.16 M), and the 2-alkyl-2-chloroalkanoyl chloride, and then it was cooled to 0 °C. The amine (1.05 equiv) and triethylamine (1.2 equiv) were sequentially added dropwise over 5 min. The reaction mixture was stirred at 0° C for 2 h, and then it was warmed to room temperature. The reaction mixture was acidified with 50 mL of 0.2 N HCl, and the organic layer was separated. The aqueous layer was extracted with 20 mL of dichloromethane, and the combined organic layer was dried over Na2SO4, filtered, and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (hexanes/ $EtOAc =$ 40/1).

*N***-(***tert***-butyl)-2-chloro-2-phenylbutanamide.** The title compound was synthesized according to GP-2 from 2-phenylbutanoic acid (16.4 g, 100 mmol), $\text{SOC}_2(29 \text{ mL}, 500$ mmol), and NCS (33.4 g, 250 mmol) for the first step (18.2 g, 84% yield), and from the intermediate (1.09 g, 5.00 mmol), *t*-butyl amine (1.1 mL, 10 mmol), triethylamine (830 μL, 6.0 mmol), and dichloromethane (40 mL) for the second step. The product was purified by flash column chromatography on silica gel. 1.17 g (3.89 mmol, 57% yield over 2 steps). White solid.

¹H NMR (500 MHz, CDCl3) δ 7.52 (d, *J =* 7.8 Hz, 2H), 7.35 (t, *J =* 7.5 Hz, 2H), 7.29 (t, *J =* 7.2 Hz, 1H), 6.74 (s, 1H), 2.59 (dq, *J =* 14.2, 7.1 Hz, 1H), 2.30 (dq, *J =* 14.3, 7.2 Hz, 1H), 1.39 (s, 9H), 1.06 (t, *J =* 7.1 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 168.9, 141.8, 128.5, 128.3, 126.4, 79.3, 52.0, 34.7, 28.5, 9.6.

FT-IR (film): 3423, 3061, 2972, 2938, 2882, 1678, 1514, 1455, 1393, 1366, 1269, 1226, 1085, 1034, 854, 826, 756, 736, 696 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₁₄H₂₁ClNO⁺: 254.1306, found: 254.1328.

2-Chloro-*N***,2-diphenylbutanamide.** The title compound was synthesized according to **GP-2** from 2-phenylbutanoic acid (16.4 g, 100 mmol), $S OCl₂ (29 mL, 500 mmol)$, and NCS (33.4 g, 250 mmol) for the first step, and from the intermediate (5.40 g, 25 mmol), aniline (2.4 mL, 26 mmol), triethylamine (4.2 mL, 30 mmol), and dichloromethane (150 mL) for the second step. The product was purified by flash column chromatography on silica gel and further purified by recrystallization (hexanes/dichloromethane). 4.17 g (15.2) mmol, 51% yield over 2 steps). White solid.

¹H NMR (500 MHz, CDCl3) δ 8.42 (s, 1H), 7.60 (d, *J =* 7.9 Hz, 2H), 7.54 (d, *J =* 8.1 Hz, 2H), 7.41 – 7.30 (m, 5H), 7.14 (t, *J =* 7.4 Hz, 1H), 2.67 (dq, *J =* 14.3, 7.1 Hz, 1H), 2.43 (dq, *J =* 14.4, 7.2 Hz, 1H), 1.08 (t, *J =* 7.1 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 168.1, 140.5, 137.3, 129.2, 128.7, 128.6, 126.5, 125.1, 120.1, 79.6, 35.1, 9.6.

FT-IR (film): 3399, 3341, 3060, 2978, 2938, 1685, 1600, 1526, 1442, 1314, 1240, 1078, 753, 691 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₁₆H₁₇ClNO⁺: 274.0993, found: 274.1016.

*N***-(***tert-***butyl)-2-chloro-2,3-diphenylpropanamide.** The title compound was synthesized according to a modified **GP-2**.^{[16](#page-27-0)} A 100 mL three-neck round-bottom flask was charged with a magnetic stir bar, 2,3-diphenylpropanoic acid (2.16 g, 9.54 mmol), and $S OCl₂ (4.0 mL, 69 mmol)$, and then a reflux condenser with a CaCl₂ drying tube was

attached. The reaction mixture was heated to 76 °C for 30 min, and then it was cooled to room temperature. SO_2Cl_2 (7.7 mL, 95.4 mmol) was added, and the reaction mixture was heated to reflux for 24 h. The volatiles were distilled off, and the residue was transferred to an oven-dried 250 mL round-bottom flask charged with a magnetic stir bar and anhydrous dichloromethane (100 mL) under a nitrogen atmosphere. The mixture was cooled to 0 °C, and then *tert*-butyl amine (3.0 mL, 29 mmol) was added dropwise over 2 min. Next, the reaction mixture was stirred at 0° C for 2 h. Then, the reaction mixture was acidified with 20 mL of 0.2 N HCl, and the organic layer was separated. The aqueous layer was extracted with 40 mL of dichloromethane, and the combined organic layer was dried over $Na₂SO₄$, filtered, and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (hexanes/dichloromethane $= 3/1$). The product was further purified by recrystallization (hexanes, 40 °C to 0 °C). 1.02 g (3.22 mmol, 34% yield over 2 steps). White solid.

¹H NMR (500 MHz, CDCl3) δ 7.55 (d, *J =* 7.7 Hz, 2H), 7.36 (t, *J =* 7.3 Hz, 2H), 7.34 – 7.29 (m, 1H), 7.26 – 7.20 (m, 5H), 6.54 (s, 1H), 3.97 (d, *J =* 13.7 Hz, 1H), 3.47 (d, *J =* 13.7 Hz, 1H), 1.31 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 168.3, 141.6, 135.7, 131.5, 128.5, 128.4, 127.8, 127.1, 126.7, 77.8, 52.0, 46.7, 28.4.

FT-IR (film): 3421, 3064, 3032, 2967, 2932, 1670, 1515, 1496, 1455, 1393, 1366, 1272, 1224, 700 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₁₉H₂₃ClNO⁺: 316.1463, found: 316.1485.

General Procedure 3 (GP-3): Preparation of α-bromoamides. The title compounds were synthesized according to a modified literature procedure.^{[48](#page-202-4)} A 100 mL two-neck round-bottom flask was charged with a magnetic stir bar, the carboxylic acid, and PBr₃ (0.4) equiv), and then one neck was capped with a rubber stopper and a reflux condenser with a CaCl² drying tube (connected to a Tygon® tube submerged in a saturated aqueous solution of sodium thiosulfate hydrate) was attached to the other neck. The reaction mixture was

warmed to 60 °C and stirred for 3 h or until all of the acid was consumed. The reaction mixture was cooled to room temperature, and bromine (1.4 equiv) was carefully added dropwise (vigorous gas evolution was observed). The heating bath was warmed to 100 °C for 3 h. Next, additional bromine (1.4 equiv) was added, and the reaction mixture was stirred for 2 h to complete the reaction. The reaction mixture was cooled to room temperature, and cyclohexene (2.5 equiv) was slowly added to quench the residual bromine. The volatiles were distilled off, and the residue was transferred to an oven-dried round-bottom flask charged with a magnetic stir bar and anhydrous dichloromethane (0.16 M, based on the carboxylic acid) under a nitrogen atmosphere. The mixture was cooled to 0 °C, and aniline (3 equiv) was added dropwise over 2 min. The reaction mixture was stirred at 0° C for 2 h, and then it was acidified with 20 mL of 0.2 N HCl. The organic layer was separated, and the aqueous layer was extracted with 40 mL of dichloromethane. The combined organic layer was dried over Na2SO4, filtered, and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (hexanes/ $EtOAc =$ $100/1 \rightarrow 50/1$) or recrystallized from pentane/EtOAc.

2-Bromo-2-cyclopentyl-*N***-phenylpropanamide.** The title compound was synthesized according to GP-3 from 2-cyclopentylpropanoic acid (960 mg, 6.80 mmol), PBr_3 (320 μL , 3.4 mmol), bromine (990 μL, 19 mmol), cyclohexene (1.7 mL, 17 mmol), aniline (1.9 mL, 20 mmol), and dichloromethane (45 mL). 1.13 g (3.81 mmol, 56% yield over 2 steps). Pale-brown solid.

¹H NMR (500 MHz, CDCl3) δ 8.68 (s, 1H), 7.54 (d, *J =* 7.9 Hz, 2H), 7.35 (t, *J =* 7.9 Hz, 2H), 7.15 (t, *J =* 7.4 Hz, 1H), 2.51 (p, *J =* 8.4 Hz, 1H), 2.02 (s, 3H), 1.91 – 1.75 (m, 2H), 1.75 – 1.45 (m, 6H).

¹³C NMR (126 MHz, CDCl₃) δ 169.8, 137.5, 129.2, 125.0, 120.2, 77.5, 50.4, 30.04, 30.01, 29.1, 25.7, 25.7.

FT-IR (film): 3386, 3349, 2956, 2868, 1673, 1598, 1529, 1499, 1442, 1314, 1242, 1163, 1074, 1029, 755, 690 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₁₄H₁₉BrNO⁺: 296.0645/298.0625, found: 296.0670/298.0651.

2-Bromo-2,4-dimethyl-*N***-phenylpentanamide.** The title compound was synthesized according to GP-3 from 2,4-dimethylpentanoic acid $(5.20 \text{ g}, 40.0 \text{ mmol})$, $\text{PBr}_3 (1.5 \text{ mL}, 16$ mmol) bromine (2 x 2.9 mL, 112 mmol), cyclohexene (10.1 mL, 100 mmol), aniline (11.0 mL, 120 mmol), and dichloromethane (250 mL). The product was further purified by recrystallization (pentane/EtOAc). 7.56 g (26.6 mmol, 67% yield over 2 steps). White solid.

¹H NMR (500 MHz, CDCl3) δ 8.64 (s, 1H), 7.54 (d, *J =* 8.0 Hz, 2H), 7.36 (t, *J =* 7.8 Hz, 2H), 7.16 (t, *J =* 7.4 Hz, 1H), 2.28 (dd, *J =* 14.3, 6.8 Hz, 1H), 2.05 (s, 3H), 2.00 (dd, *J =* 14.4, 4.5 Hz, 1H), 1.97 – 1.91 (m, 1H), 1.00 (d, *J =* 6.5 Hz, 3H), 0.94 (d, *J =* 6.5 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 169.6, 137.5, 129.2, 125.0, 120.1, 70.2, 52.3, 32.5, 27.3, 24.2, 22.9.

FT-IR (film): 3386, 3343, 2959, 2931, 2871, 1669, 1598, 1529, 1442, 1316, 1241, 1153, 854, 691 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₁₃H₁₉BrNO⁺: 264.0958/266.0938, found: 264.0979/266.0959.

2.4.3. Enantioconvergent coupling reactions

General Procedure 4 (GP-4). In the air, an oven-dried 20 mL vial equipped with a PTFE magnetic stir bar was charged with CuCl (8.0 mg, 0.080 mmol, white to off-white solid) and (R) - $P(113 \text{ mg}, 0.0960 \text{ mmol})$. The vial was capped with a PTFE-lined open top cap, and the joint was wrapped with electrical tape. The vial was evacuated/backfilled with nitrogen three times, using a 21G needle attached to a Schlenk line. Then, a nitrogen-filled balloon was attached, toluene (5 mL) and BTPP (290 μ L, 0.960 mmol) were added using standard air-free technique by syringe, and the reaction mixture was stirred for 4 h (see **Note 1** below).

The aniline nucleophile (0.960 mmol) was weighed into an oven-dried 40 mL vial equipped with a PTFE magnetic stir bar. The vial was capped with a PTFE-lined open top cap, and the joint was wrapped with electrical tape. The vial was attached to a Schlenk line and evacuated/backfilled with nitrogen three times. A nitrogen-filled balloon was attached, and 5 mL of toluene was added. The catalyst solution was transferred to the vial containing the nucleophile. An additional 3 mL of toluene was used to wash the vial that had contained the catalyst, and the wash was added to the reaction vial. The reaction vial was submerged in a low-form hemispherical Dewar flask filled with pre-cooled methanol (– 78 °C) and stirred for 10 min.

The electrophile (0.800 mmol) was weighed into a 4 mL vial, which was capped with a PTFE-lined open top cap. The vial was attached to a Schlenk line and evacuated/backfilled with nitrogen three times. A nitrogen-filled balloon was attached, and 1.5 mL of toluene was added. The resulting solution was transferred to the pre-cooled reaction vial. An

additional 1.5 mL of toluene was used to wash the vial that had contained the electrophile, and the wash was added to the reaction mixture. The reaction mixture was stirred for 5 min, the balloon was removed, and vacuum grease was applied to the PTFE cap to seal the needle puncture. Kessil® PR160-440 nm lamps (one lamp per reaction, 3-4 lamps per cooling bath) were installed 5 cm above the reaction vial, and the vial was irradiated for 12- 48 h.

Work-up. The reaction was quenched by turning off the lamps. The reaction mixture was warmed to room temperature and opened to air. 2 mL of 30% NH4OH (aq) and 2 mL of EtOAc were added, and the reaction mixture was vigorously stirred for 5 min, at which point the aqueous layer was blue (see **Note 2** below). The aqueous layer was extracted with EtOAc (2 x 3 mL), and the combined organic layer was passed through a short plug of silica gel with EtOAc washings (2 x 4 mL). The resulting filtrate was concentrated and purified by flash column chromatography.

Left: immediately after irradiation and warming to room temperature. Right: after the addition of NH4OH and EtOAc and 5 min of stirring.

Note 1. In the presence of BTPP, we observe fast solubilization of CuCl in toluene, resulting in an apparently homogenous reaction mixture within 10 min. However, 4 h of stirring was necessary to obtain reproducible ee values with **GP-4**. If pre-made catalyst (**CuA**) is used (**GP-5** below), the 4 h of stirring can be omitted.

Note 2. The coupling products and **Cu^A** often have similar Rf values. Workup with NH4OH results in the consumption of **Cu^A** and allows recovery of the ligand (if desired) and facile purification of the coupling product. This step is not required for the purification of substrates that have an Rf value significantly different from that of **CuA**.

General Procedure 5 (GP-5): Use of Cu^A as a discrete catalyst. In the air, a 40 mL vial equipped with a PTFE stir bar was charged with **Cu^A** (synthesis: Section 2.4.10; 103 mg, 0.0800 mmol) and the nucleophile (0.960 mmol). The vial was capped with a PTFElined open top cap, and the joint was wrapped with electrical tape. The vial was evacuated/backfilled with nitrogen three times, using a 21G needle attached to a Schlenk line, and then a nitrogen-filled balloon was attached. Toluene (13 mL) and BTPP (290 μL, 0.960 mmol) were added, and the reaction mixture was stirred for 5 min. The remainder of the procedure is the same as **GP-4**.

2-Phenyl-2-(*p***-tolylamino)propanenitrile.** The title compound was prepared according to **GP-4** from 2-chloro-2-phenylpropanenitrile and *p*-toluidine with 12 h of reaction time. The product was purified by column chromatography on silica gel $(hexanes/EtOAc/dichloromethane = 25/1/1)$. Pale-yellow solid.

(R)-**P**: 112 mg, 59% yield, 89% ee; *(S)*-**P**: 113 mg, 60% yield, 88% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (5% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 9.9 min (major), 14.6 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.64 (d, *J =* 7.5 Hz, 2H), 7.41 (t, *J =* 7.4 Hz, 2H), 7.36 (t, *J =* 7.3 Hz, 1H), 6.94 (d, *J =* 8.3 Hz, 2H), 6.48 (d, *J =* 8.4 Hz, 2H), 4.19 (s, 1H), 2.22 (s, 3H), 1.94 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 141.2, 140.2, 129.7, 129.5, 129.3, 128.7, 125.1, 116.2, 57.5, 33.4, 20.6.

FT-IR (film): 3383, 3026, 2989, 2917, 2865, 2230, 1618, 1518, 1448, 1303, 1258, 1219, 807, 759, 698 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for $C_{16}H_{17}N_2$ ⁺: 237.1386, found: 237.1384.

 $[\alpha]^{23}$ _D = –240 (*c* 1.0, CHCl₃); 89% ee, from *(R)*-**P**.

2-Phenyl-2-(*p***-tolylamino)butanenitrile.** The title compound was prepared according to **GP-4** and **GP-5** from 2-chloro-2-phenylbutanenitrile and *p*-toluidine with 12 h of reaction time. The product was purified by column chromatography on silica gel $(hexanes/EtOAc/dichloromethane = 30/1/1)$. White solid.

From **GP-4**, *(R)*-**P**: 143 mg, 72% yield, 93% ee; *(S)*-**P**: 142 mg, 71% yield, 92% ee.

From **GP-5**, *(R)*-**P**: 143 mg, 72% yield, 93% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (5% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 7.7 min (major), 11.3 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.61 (d, *J =* 7.5 Hz, 2H), 7.41 (t, *J =* 7.4 Hz, 2H), 7.36 (t, *J =* 7.2 Hz, 1H), 6.92 (d, *J =* 8.4 Hz, 2H), 6.47 (d, *J =* 8.4 Hz, 2H), 4.22 (s, 1H), 2.24 – 2.16 (m, 4H), 2.09 (dq, *J =* 14.8, 7.4 Hz, 1H), 1.07 (t, *J =* 7.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 141.3, 138.6, 129.6, 129.3, 129.1, 128.6, 125.8, 120.2, 116.1, 62.5, 38.4, 20.5, 8.9.

FT-IR (film): 3387, 3026, 2975, 2924, 2231, 1620, 1522, 1449, 1318, 1305, 1254, 805, 757, 697 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₁₇H₁₉N₂⁺: 251.1543, found: 251.1547.

 $[\alpha]^{23}$ _D = –211 (*c* 1.0, CHCl₃); 93% ee, from *(R)*-**P**.

4-Methyl-2-phenyl-2-(*p***-tolylamino)pentanenitrile.** The title compound was prepared according to **GP-4** from 2-chloro-4-methyl-2-phenylpentanenitrile and *p*-toluidine with 24 h of reaction time. The product was purified by column chromatography on silica gel $(hexanes/EtOAC = 50/1)$. Pale-yellow solid.

(R)-**P**: 93 mg, 42% yield, 88% ee; *(S)*-**P**: 94 mg, 42% yield, 88% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (5% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 5.8 min (major), 9.0 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.62 (d, *J =* 7.9 Hz, 2H), 7.40 (t, *J =* 7.5 Hz, 2H), 7.35 (t, *J =* 7.2 Hz, 1H), 6.91 (d, *J =* 8.3 Hz, 2H), 6.44 (d, *J =* 8.3 Hz, 2H), 4.19 (s, 1H), 2.20 (s, 3H), 2.07 (dd, *J =* 13.9, 6.2 Hz, 1H), 2.00 (dd, *J =* 13.9, 5.8 Hz, 1H), 1.84 (dh, *J =* 12.9, 6.5 Hz, 1H), 1.07 (d, *J =* 6.7 Hz, 3H), 0.85 (d, *J =* 6.7 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 141.2, 139.3, 129.6, 129.4, 129.2, 128.6, 120.6, 116.3, 61.2, 53.5, 24.9, 23.9, 20.6.

FT-IR (film): 3373, 3022, 2956, 2927, 2873, 2244, 1617, 1521, 1448, 1304, 1252, 1189, 800, 754, 697 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₁₉H₂₃N₂⁺: 279.1856, found: 279.1854.

 $[\alpha]^{23}$ _D = –186 (*c* 1.0, CHCl₃); 88% ee, from *(R)*-**P**.

2,3-Diphenyl-2-(*p***-tolylamino)propanenitrile.** The title compound was prepared according to **GP-4** from 2-chloro-2,3-diphenylpropanenitrile and *p*-toluidine with 14 h of reaction time. The product was purified by column chromatography on silica gel: column #1 (hexanes/EtOAc/dichloromethane = $35/1/3$); column #2 (hexanes/toluene = $1/1$). Paleyellow solid.

(R)-**P**: 173 mg, 69% yield, 92% ee; *(S)*-**P**: 171 mg, 68% yield, 91% ee.
HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (5% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 9.7 min (major), 14.0 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.58 (d, *J =* 7.4 Hz, 2H), 7.41 – 7.33 (m, 6H), 7.29 – 7.22 (m, 2H), 6.38 (d, *J =* 8.3 Hz, 2H), 4.28 (s, 1H), 3.35 (d, *J =* 13.5 Hz, 1H), 3.23 (d, *J =* 13.5 Hz, 1H), 2.18 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 141.1, 138.9, 133.1, 130.7, 129.63, 129.57, 129.2, 129.0, 128.8, 128.4, 125.8, 119.9, 116.5, 62.1, 51.2, 20.6.

FT-IR (film): 3380, 3031, 2920, 2858, 2234, 1620, 1522, 1449, 1303, 1256, 805, 758, 702 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₂₂H₂₁N₂⁺: 313.1699, found: 313.1708.

 $[\alpha]^{23}$ _D = +224 (*c* 1.0, CHCl₃); 91% ee, from *(S)*-**P**.

3-Cyano-3-phenyl-3-(*p***-tolylamino)propyl acetate.** The title compound was prepared according to **GP-4** from 3-chloro-3-cyano-3-phenylpropyl acetate and *p*-toluidine with 24 h of reaction time. The product was purified by column chromatography on silica gel $(hexanes/EtOAc = 10/1 \rightarrow 5/1)$. Pale-yellow solid.

(R)-**P**: 167 mg, 67% yield, 91% ee; *(S)*-**P**: 163 mg, 66% yield, 91% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (10% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 10.1 min (major), 14.1 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.61 (d, *J =* 7.5 Hz, 2H), 7.41 (t, *J =* 7.3 Hz, 2H), 7.38 – 7.34 (m, 1H), 6.92 (d, *J =* 8.1 Hz, 2H), 6.44 (d, *J =* 8.4 Hz, 2H), 4.75 (s, 1H), 4.35 (dt, *J =* 11.4, 5.5 Hz, 1H), 4.25 (ddd, *J =* 12.4, 8.0, 5.0 Hz, 1H), 2.49 (ddd, *J =* 14.7, 8.0, 5.3 Hz, 1H), 2.37 (dt, *J =* 14.7, 5.4 Hz, 1H), 2.21 (s, 3H), 2.10 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 170.6, 141.0, 138.4, 129.71, 129.69, 129.4, 129.0, 125.6, 119.6, 116.3, 60.8, 60.6, 43.2, 21.0, 20.6.

FT-IR (film): 3366, 3027, 2962, 2919, 2865, 2229, 1740, 1617, 1521, 1448, 1366, 1232, 1049, 809, 769, 701 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₁₉H₂₁N₂O₂⁺: 309.1598, found: 309.1610. $[\alpha]^{23}$ _D = +202 (*c* 1.0, CHCl₃); 91% ee, from *(S)*-**P**.

2-Phenyl-2-(*p***-tolylamino)pent-4-enenitrile.** The title compound was prepared according to **GP-4** from 2-chloro-2-phenylpent-4-enenitrile and *p*-toluidine with 18 h of reaction time. The product was purified by column chromatography on silica gel $(hexanes/EtOAc = 45/1)$. Pale-yellow solid.

(R)-**P**: 149 mg, 71% yield, 91% ee; *(S)*-**P**: 147 mg, 70% yield, 92% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (5% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 6.4 min (major), 8.8 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.62 (d, *J =* 7.7 Hz, 2H), 7.41 (t, *J =* 7.4 Hz, 2H), 7.36 (t, *J =* 7.2 Hz, 1H), 6.91 (d, *J =* 8.2 Hz, 2H), 6.43 (d, *J =* 8.2 Hz, 2H), 5.95 (dtd, *J =* 19.4, 9.7, 5.3 Hz, 1H), 5.45 – 5.34 (m, 2H), 4.32 (s, 1H), 2.89 (dd, *J =* 13.7, 5.1 Hz, 1H), 2.66 (dd, *J =* 13.9, 9.4 Hz, 1H), 2.20 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 141.1, 139.0, 131.0, 129.7, 129.5, 129.3, 128.8, 125.6, 122.8, 119.9, 116.1, 60.3, 49.9, 20.6.

FT-IR (film): 3370, 3078, 3026, 2981, 2918, 2864, 2230, 1617, 1558, 1519, 1448, 1301, 995, 931, 808, 772, 700 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for $C_{18}H_{19}N_2^+$: 263.1543, found: 263.1545. $[\alpha]^{23}$ _D = –245 (*c* 1.0, CHCl₃); 91% ee, from *(R)*-**P**.

4-Chloro-2-phenyl-2-(*p***-tolylamino)butanenitrile.** The title compound was prepared according to **GP-4** from 2,4-dichloro-2-phenylbutanenitrile and *p*-toluidine with 20 h of reaction time. The product was purified by column chromatography on silica gel $(hexanes/EtOAc = 40/1)$. Pale-yellow solid.

(R)-**P**: 178 mg, 78% yield, 90% ee; *(S)*-**P**: 177 mg, 77% yield, 91% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (5% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 11.6 min (major), 18.2 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.60 (d, *J =* 7.5 Hz, 2H), 7.43 (t, *J =* 7.4 Hz, 2H), 7.39 (t, *J =* 7.1 Hz, 1H), 6.93 (d, *J =* 8.3 Hz, 2H), 6.46 (d, *J =* 8.3 Hz, 2H), 4.53 (s, 1H), 3.72 (ddd, *J =* 11.2, 8.7, 6.8 Hz, 1H), 3.50 (ddd, *J =* 11.3, 8.7, 5.4 Hz, 1H), 2.64 (ddd, *J =* 15.1, 8.5, 6.8 Hz, 1H), 2.55 (ddd, *J =* 14.1, 8.8, 5.5 Hz, 1H), 2.21 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 140.7, 137.7, 130.1, 129.7, 129.5, 129.3, 125.6, 119.4, 116.6, 61.4, 46.6, 39.1, 20.6.

FT-IR (film): 3362, 3061, 3026, 2962, 2919, 2864, 2231, 1617, 1518, 1490, 1448, 1301, 1249, 809, 770, 699 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₁₇H₁₈ClN₂⁺: 285.1154, found: 285.1150.

 $[\alpha]^{23}$ _D = –173 (*c* 1.0, CHCl₃); 89% ee, from *(R)*-**P**.

2-(3-Fluorophenyl)-2-(*p***-tolylamino)butanenitrile.** The title compound was prepared according to **GP-4** from 2-chloro-2-(3-fluorophenyl)butanenitrile and *p*-toluidine with 18 h of reaction time. The product was purified by column chromatography on silica gel $(hexanes/EtOAc/dichlorome thane = 25/1/1)$. Pale-yellow solid.

(R)-**P**: 141 mg, 66% yield, 90% ee; *(S)*-**P**: 143 mg, 67% yield, 90% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (5% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 8.4 min (major), 13.1 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.44 – 7.35 (m, 2H), 7.31 (dt, *J =* 9.9, 2.0 Hz, 1H), 7.08 – 7.02 (m, 1H), 6.93 (d, *J =* 8.4 Hz, 2H), 6.46 (d, *J =* 8.4 Hz, 2H), 4.20 (s, 1H), 2.24 – 2.13 (m, 4H), 2.08 (dq, *J =* 14.9, 7.4 Hz, 1H), 1.08 (t, *J =* 7.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl3) δ 163.3 (d, *J =* 247.2 Hz), 141.5 (d, *J =* 6.9 Hz), 141.0, 130.7 (d, *J =* 8.2 Hz), 129.71, 129.67, 121.6 (d, *J =* 2.9 Hz), 119.7, 116.1, 115.8 (d, *J =* 21.2 Hz), 113.0 (d, *J =* 23.6 Hz), 62.2 (d, *J =* 2.2 Hz), 38.3, 20.5, 8.9.

¹⁹F NMR (376 MHz, CDCl₃) δ –111.4.

FT-IR (film): 3370, 3027, 2976, 2939, 2922, 2882, 2865, 2229, 1617, 1592, 1521, 1484, 1457, 1443, 1302, 1175, 1159, 974, 848, 808, 789, 694 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₁₇H₁₈FN₂⁺: 269.1449, found: 269.1458.

 $[\alpha]^{23}$ _D = –217 (*c* 1.0, CHCl₃); 90% ee, from *(R)*-**P**.

2-(4-Chlorophenyl)-2-(*p***-tolylamino)butanenitrile**. The title compound was prepared according to **GP-4** from 2-chloro-2-(4-chlorophenyl)butanenitrile and *p*-toluidine with 14 h of reaction time. The product was purified by column chromatography on silica gel $(hexanes/EtOAc/dichlorome thane = 20/1/1)$. White solid.

(R)-**P**: 177 mg, 78% yield, 91% ee; *(S)*-**P**: 177 mg, 78% yield, 91% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (5% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 9.1 min (major), 13.4 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.54 (d, *J =* 8.6 Hz, 2H), 7.38 (d, *J =* 8.6 Hz, 2H), 6.93 (d, *J =* 8.3 Hz, 2H), 6.45 (d, *J =* 8.4 Hz, 2H), 4.20 (s, 1H), 2.26 – 2.13 (m, 4H), 2.06 (dq, *J =* 14.9, 7.4 Hz, 1H), 1.07 (t, *J =* 7.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 141.0, 137.3, 134.5, 129.70, 129.65, 129.3, 127.3, 119.8, 116.1, 62.0, 38.3, 20.5, 8.9.

FT-IR (film): 3372, 3028, 2976, 2921, 2880, 2230, 1614, 1518, 1488, 1401, 1303, 1251, 1094, 1013, 920, 808, 751 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₁₇H₁₇ClN₂: 285.1154, found: 285.1158. $[\alpha]^{23}$ _D = –187 (*c* 1.0, CHCl₃); 91% ee, from *(R)*-**P**.

2-(4-Bromophenyl)-2-(*p***-tolylamino)butanenitrile.** The title compound was prepared according to **GP-4** from 2-chloro-2-(4-bromophenyl)butanenitrile and *p*-toluidine with 16 h of reaction time. The product was purified by column chromatography on silica gel $(hexanes/EtOAc/dichlorome thane = 25/1/1)$. White solid.

(R)-**P**: 200 mg, 76% yield, 91% ee; *(S)*-**P**: 203 mg, 77% yield, 91% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (5% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 9.4 min (major), 13.5 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.53 (d, *J =* 8.6 Hz, 2H), 7.47 (d, *J =* 8.7 Hz, 2H), 6.93 $(d, J = 8.4 \text{ Hz}, 2H)$, 6.45 $(d, J = 8.4 \text{ Hz}, 2H)$, 4.20 $(s, 1H)$, 2.25 – 2.13 $(m, 4H)$, 2.05 (dq, J) *=* 14.8, 7.4 Hz, 1H), 1.06 (t, *J =* 7.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 141.0, 137.9, 132.3, 129.70, 129.67, 127.6, 122.7, 119.7, 116.1, 62.1, 38.3, 20.5, 8.9.

FT-IR (film): 3369, 3024, 2976, 2938, 2921, 2880, 2864, 2229, 1617, 1518, 1486, 1396, 1303, 1251, 1219, 1073, 1009, 920, 808, 772 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₁₇H₁₈BrN₂⁺: 329.0648, found: 329.0642.

 $[\alpha]^{23}$ _D = –154 (*c* 1.0, CHCl₃); 91% ee, from *(R)*-**P**.

*N***-Benzyl-2-cyano-***N***-phenyl-2-(***p***-tolylamino)butanamide.** The title compound was prepared according to **GP-4** from *N*-benzyl-2-bromo-2-cyano-*N*-phenylbutanamide and *p*toluidine with 18 h of reaction time. The product was purified by column chromatography on silica gel (hexanes/EtOAc = $10/1 \rightarrow 7/1$). Pale-brown solid.

(R)-**P**: 223 mg, 73% yield, 97% ee; *(S)*-**P**: 226 mg, 74% yield, 97% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK AD column (10% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 12.9 min (major), 17.1 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.32 – 7.17 (m, 5H), 7.11 (d, *J =* 6.8 Hz, 3H), 7.03 (d, *J =* 8.2 Hz, 2H), 6.77 (s, 1H), 6.50 (d, *J =* 8.0 Hz, 2H), 6.42 (s, 1H), 4.93 (d, *J =* 13.8 Hz, 1H), 4.66 (d, *J =* 13.8 Hz, 1H), 3.40 (s, 1H), 2.29 (s, 3H), 2.05 (q, *J =* 7.2 Hz, 2H), 1.22 (t, $J = 7.4$ Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 166.5, 140.5, 139.9, 136.4, 129.9, 129.4, 129.3, 129.0, 128.8, 128.7, 128.4, 128.2, 127.9, 118.1, 114.9, 66.2, 56.9, 32.7, 20.6, 9.0.

FT-IR (film): 3369, 3064, 3030, 2982, 2939, 2921, 2236, 1646, 1593, 1522, 1496, 1457, 1395, 1320, 1252, 809, 753, 701 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₂₅H₂₆N₃O⁺: 384.2070, found: 384.2078.

 $[\alpha]^{23}$ _D = +17.6 (*c* 1.0, CHCl₃); 97% ee, from *(R)*-**P**.

Diisopropyl (1-cyano-1-(*p***-tolylamino)propyl)phosphonate.** The title compound was prepared according to **GP-4** from diisopropyl (1-bromo-1-cyanopropyl)phosphonate and *p*toluidine with 24 h of reaction time. The product was purified by column chromatography on silica gel (hexanes/EtOAc = $3/1$). Pale-brown solid.

(R)-**P**: 144 mg, 53% yield, 95% ee; *(S)*-**P**: 145 mg, 53% yield, 96% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK AD column (10% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 7.7 min (major), 10.8 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.06 (d, *J =* 8.3 Hz, 2H), 6.98 (d, *J =* 8.4 Hz, 2H), 4.88 (hept, *J =* 6.2 Hz, 2H), 3.94 (d, *J =* 7.9 Hz, 1H), 2.28 (s, 3H), 2.28 – 2.14 (m, 1H), 2.15 – 2.01 (m, 1H), 1.42 – 1.36 (m, 12H), 1.13 (t, *J =* 7.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl3) δ 141.0 (d, *J =* 11.2 Hz), 131.8, 129.9, 120.0, 117.9 (d, *J =* 4.5 Hz), 74.3 (d, *J =* 7.3 Hz), 74.0 (d, *J =* 7.8 Hz), 57.6 (d, *J =* 157.8 Hz), 27.9 (d, *J =*

2.7 Hz), 24.4 (d, *J =* 3.0 Hz), 24.1 (d, *J =* 3.8 Hz), 24.0 (d, *J =* 5.2 Hz), 23.7 (d, *J =* 5.9 Hz), 20.7, 9.0 (d, *J =* 4.1 Hz).

³¹P NMR (162 MHz, CDCl₃) δ 15.0.

FT-IR (film): 3306, 3026, 2982, 2938, 2884, 2873, 2237, 1617, 1515, 1453, 1387, 1377, 1243, 1178, 1143, 1192, 995, 811, 768, 582 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₁₇H₂₈N₂O₃P⁺: 339.1833, found: 339.1835. $[\alpha]^{23}$ _D = +60.7 (*c* 1.0, CHCl₃); 95% ee, from *(R)*-**P**.

Ethyl 4-((1-cyano-1-phenylpropyl)amino)benzoate. The title compound was prepared according to **GP-4** from 2-chloro-2-phenylbutanenitrile and ethyl 4 aminobenzoate with 24 h of reaction time. The product was purified by column chromatography on silica gel (hexanes/EtOAc/dichloromethane = 8/1/1). Pale-yellow solid.

(R)-**P**: 186 mg, 75% yield, 93% ee; *(S)*-**P**: 181 mg, 73% yield, 93% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK AD column (10% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 9.4 min (major), 10.3 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.80 (d, *J =* 8.7 Hz, 2H), 7.54 (d, *J =* 7.5 Hz, 2H), 7.39 (t, *J =* 7.4 Hz, 2H), 7.35 (t, *J =* 7.2 Hz, 1H), 6.54 (d, *J =* 8.7 Hz, 2H), 4.86 (s, 1H), 4.28 (q, *J =* 7.1 Hz, 2H), 2.24 (dq, *J =* 14.5, 7.3 Hz, 1H), 2.10 (dq, *J =* 14.8, 7.4 Hz, 1H), 1.32 (t, *J =* 7.1 Hz, 3H), 1.08 (t, *J =* 7.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 166.6, 147.5, 137.7, 131.1, 129.3, 128.9, 125.6, 121.4, 119.4, 114.6, 61.7, 60.5, 38.3, 14.5, 8.9.

FT-IR (film): 3364, 3063, 3029, 2979, 2938, 2903, 2881, 2233, 1689, 1608, 1521, 1490, 1449, 1367, 1315, 1279, 1180, 1108, 1025, 925, 841, 771, 699 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₁₉H₂₁N₂O₂⁺: 309.1598, found: 309.1612.

 $[\alpha]^{23}$ _D = –238 (*c* 1.0, CHCl₃); 93% ee, from *(R)*-**P**.

4-((1-Cyano-1-phenylpropyl)amino)benzonitrile. The title compound was prepared according to **GP-4** from 2-chloro-2-phenylbutanenitrile and 4-aminobenzonitrile with 24 h of reaction time. The product was purified by column chromatography on silica gel $(hexanes/EtOAc = 15/1)$. Pale-yellow solid.

(R)-**P**: 132 mg, 63% yield, 92% ee; *(S)*-**P**: 128 mg, 61% yield, 92% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (10% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 13.2 min (major), 17.2 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.51 (d, *J =* 7.5 Hz, 2H), 7.44 – 7.33 (m, 5H), 6.56 (d, *J =* 8.7 Hz, 2H), 4.97 (s, 1H), 2.25 (dq, *J =* 14.5, 7.3 Hz, 1H), 2.11 (dq, *J =* 14.8, 7.4 Hz, 1H), 1.08 (t, *J =* 7.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 147.3, 137.1, 133.5, 129.4, 129.2, 125.5, 119.8, 119.1, 115.3, 101.9, 61.6, 38.2, 8.9.

FT-IR (film): 3351, 2977, 2939, 2881, 2218, 1608, 1521, 1449, 1328, 1265, 1177, 925, 827, 760, 700 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₁₇H₁₆N₃⁺: 262.1339, found: 262.1356. $[\alpha]^{23}$ _D = +267 (*c* 1.0, CHCl₃); 92% ee, from *(S*)-**P**.

2-Phenyl-2-((4-(trifluoromethyl)phenyl)amino)butanenitrile. The title compound was prepared according to **GP-4** from 2-chloro-2-phenylbutanenitrile and 4 trifluoromethylaniline with 24 h of reaction time. The product was purified by column chromatography on silica gel (hexanes/ $EtOAc = 20/1$). Pale-yellow solid.

(R)-**P**: 120 mg, 49% yield, 89% ee; *(S)*-**P**: 123 mg, 51% yield, 89% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (10% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 6.5 min (major), 7.6 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.55 (d, *J =* 7.6 Hz, 2H), 7.46 – 7.36 (m, 3H), 7.34 (d, *J =* 8.4 Hz, 2H), 6.57 (d, *J =* 8.4 Hz, 2H), 4.61 (s, 1H), 2.24 (dq, *J =* 14.7, 7.3 Hz, 1H), 2.12 (dq, *J =* 14.7, 7.4 Hz, 1H), 1.09 (t, *J =* 7.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl3) 146.3, 137.6, 129.4, 129.1, 126.5 (q, *J =* 3.8 Hz), 125.6, 124.6 (q, *J =* 270.5 Hz), 121.8, 121.5, 119.4, 61.8, 38.4, 8.9.

¹⁹F NMR (376 MHz, CDCl₃) δ –61.6.

FT-IR (film): 3350, 3027, 2982, 2947, 2925, 2248, 1617, 1536, 1449, 1327, 1162, 1117, 1072, 834, 815, 755, 700 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₁₇H₁₆F₃N₂⁺: 305.1260, found: 305.1286.

 $[\alpha]^{23}$ _D = –204 (*c* 1.0, CHCl₃); 89% ee, from *(R)*-**P**.

*tert***-Butyl (4-((1-cyano-1-phenylpropyl)amino)phenethyl)carbamate.** The title compound was prepared according to **GP-4** from 2-chloro-2-phenylbutanenitrile and *tert*butyl (4-aminophenethyl)carbamate with 24 h of reaction time. The product was purified by column chromatography on silica gel (hexanes/EtOAc = $8/1\rightarrow 5/1$). White solid.

(R)-**P**: 201 mg, 66% yield, 89% ee; *(S)*-**P**: 180 mg, 59% yield, 89% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (15% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 7.8 min (major), 10.7 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.58 (d, *J =* 7.7 Hz, 2H), 7.40 (t, *J =* 7.5 Hz, 2H), 7.35 (t, *J =* 7.2 Hz, 1H), 6.91 (d, *J =* 8.2 Hz, 2H), 6.48 (d, *J =* 8.3 Hz, 2H), 4.52 (s, 1H), 4.32 (s, 1H), 3.28 (t, *J =* 6.9 Hz, 2H), 2.63 (t, *J =* 6.5 Hz, 2H), 2.20 (dq, *J =* 14.5, 7.3 Hz, 1H), 2.08 (dq, *J =* 14.3, 7.4 Hz, 1H), 1.43 (s, 9H), 1.06 (t, *J =* 7.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 156.0, 142.2, 138.5, 130.3, 129.4, 129.1, 128.7, 125.7, 120.1, 116.0, 79.2, 62.3, 41.8, 38.4, 35.3, 28.5, 8.9.

FT-IR (film): 3366, 2977, 2930, 2229, 1696, 1616, 1521, 1456, 1251, 1168, 1053, 924, 824, 759, 700 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₂₃H₃₀N₃O₂⁺: 380.2333, found: 380.2348.

 $[\alpha]^{23}$ _D = +133 (*c* 1.0, CHCl₃); 89% ee, from (*S*)-**P**.

2-((4-Methoxyphenyl)amino)-2-phenylbutanenitrile. The title compound was prepared according to **GP-4** from 2-chloro-2-phenylbutanenitrile and 4-methoxyaniline with 24 h of reaction time. The product was purified by column chromatography on silica gel (hexanes/EtOAc/dichloromethane = 15/1/1). Pale-brown solid.

(R)-**P**: 90 mg, 42% yield, 88% ee; *(S)*-**P**: 90 mg, 42% yield, 88% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (10% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 8.2 min (major), 15.1 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.60 (d, *J =* 7.5 Hz, 2H), 7.40 (t, *J =* 7.5 Hz, 2H), 7.35 (t, *J =* 7.2 Hz, 1H), 6.68 (d, *J =* 8.9 Hz, 2H), 6.53 (d, *J =* 8.9 Hz, 2H), 4.04 (s, 1H), 3.70 (s, 3H), 2.18 (dq, *J =* 14.7, 7.3 Hz, 1H), 2.09 (dq, *J =* 13.6, 7.3 Hz, 1H), 1.04 (t, *J =* 7.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 154.0, 138.6, 137.4, 129.1, 128.7, 126.0, 120.3, 118.2, 114.6, 63.3, 55.6, 38.2, 9.0.

FT-IR (film): 3370, 3031, 2975, 2937, 2839, 2229, 1617, 1601, 1457, 1205, 1153, 1069, 925, 816, 761, 700 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₁₇H₁₉N₂O: 267.1492, found: 267.1507.

 $[\alpha]^{23}$ _D = –161 (*c* 1.0, CHCl₃); 88% ee, from (*R*)-**P**.

2-(Dibenzo[*b,d***]furan-3-ylamino)-2-phenylbutanenitrile.** The title compound was prepared according to **GP-4** from 2-chloro-2-phenylbutanenitrile and 3-(4,4,5-trimethyl-1,3,2-dioxaborolan-2-yl)aniline with 24 h of reaction time. The product was purified by column chromatography on silica gel: column #1 (hexanes/EtOAc/dichloromethane =

10/1/1→4/1/1); column #2 (toluene). The reaction mixture was dry-loaded on the silica due to the poor solubility of the title compound. Pale-brown solid.

(R)-**P**: 175 mg, 67% yield, 94% ee; *(S)*-**P**: 161 mg, 62% yield, 94% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (5% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 22.0 min (major), 24.9 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.76 (d, *J =* 7.6 Hz, 1H), 7.66 (d, *J =* 9.0 Hz, 1H), 7.63 (d, *J =* 7.5 Hz, 2H), 7.42 (t, *J =* 7.3 Hz, 3H), 7.37 (t, *J =* 7.3 Hz, 1H), 7.30 (t, *J =* 7.7 Hz, 1H), 7.28 – 7.21 (m, 1H), 6.67 – 6.59 (m, 2H), 4.55 (s, 1H), 2.26 (dq, *J =* 14.6, 7.3 Hz, 1H), 2.14 (dq, *J =* 14.8, 7.4 Hz, 1H), 1.10 (t, *J =* 7.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 157.4, 156.1, 143.8, 138.1, 129.3, 128.9, 125.8, 125.7, 124.6, 122.7, 121.1, 119.8, 119.7, 116.8, 112.2, 111.3, 98.6, 62.5, 38.6, 9.0.

FT-IR (film): 3381, 3062, 2976, 2925, 2230, 1636, 1606, 1506, 1457, 1260, 1165, 1130, 747, 721, 699 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₂₂H₁₉N₂O⁺: 327.1492, found: 327.1509.

 $[\alpha]^{23}$ _D = +199 (*c* 1.0, CHCl₃); 94% ee, from (*S*)-**P**.

$$
\textrm{NC} \xrightarrow{\textrm{H}} \textrm{SiMe}_3
$$

2-Phenyl-2-((3-(trimethylsilyl)phenyl)amino)butanenitrile. The title compound was prepared according to **GP-4** from 2-chloro-2-phenylbutanenitrile and 3- $(t$ rimethylsilyl)aniline^{[49](#page-202-0)} with 24 h of reaction time. The product was purified by column chromatography on silica gel (hexanes/ $EtOAc = 45/1$). Pale-yellow solid.

(R)-**P**: 167 mg, 68% yield, 91% ee; *(S)*-**P**: 158 mg, 64% yield, 92% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (5% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 5.7 min (major), 6.2 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.61 (d, *J =* 7.6 Hz, 2H), 7.41 (t, *J =* 7.5 Hz, 2H), 7.35 (t, *J =* 7.3 Hz, 1H), 7.09 (t, *J =* 7.6 Hz, 1H), 6.94 (d, *J =* 7.2 Hz, 1H), 6.70 (d, *J =* 2.3 Hz, 1H), 6.51 (ddd, *J =* 8.1, 2.7, 1.0 Hz, 1H), 4.33 (s, 1H), 2.23 (dq, *J =* 14.7, 7.3 Hz, 1H), 2.12 (dq, *J =* 14.9, 7.4 Hz, 1H), 1.09 (t, *J =* 7.4 Hz, 3H), 0.16 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 142.9, 141.3, 138.6, 129.1, 128.6, 128.5, 125.8, 124.8, 120.9, 120.0, 116.1, 62.3, 38.4, 9.0, –1.2.

FT-IR (film): 3370, 3033, 2953, 2230, 1594, 1576, 1399, 1304, 1248, 1123, 936, 754, 699 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₁₉H₂₅N₂Si⁺: 309.1782, found: 309.1797.

 $[\alpha]^{23}$ _D = +199 (*c* 1.0, CHCl₃); 92% ee, from (*S*)-**P**.

$$
\mathsf{NC}\underset{\mathsf{Ph}}{\underset{\mathsf{E}}{\overset{\mathsf{H}}{\underset{\mathsf{E}}{\bigwedge}}\longrightarrow}}\mathsf{Bpin}
$$

2-Phenyl-2-((3-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-

yl)phenyl)amino)butanenitrile. The title compound was prepared according to **GP-4** from 2-chloro-2-phenylbutanenitrile and 3-(4,4,5-trimethyl-1,3,2-dioxaborolan-2-yl)aniline with 24 h of reaction time. The product was purified by column chromatography on silica gel $(hexanes/EtOAc = 8/1 \rightarrow 5/1)$. Pale-yellow solid.

(R)-**P**: 110 mg, 38% yield, 93% ee; *(S)*-**P**: 116 mg, 40% yield, 93% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (2% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 7.9 min (major), 9.8 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.57 (d, *J =* 7.5 Hz, 2H), 7.38 (t, *J =* 7.3 Hz, 2H), 7.34 (t, *J =* 7.2 Hz, 1H), 7.22 (d, *J =* 7.3 Hz, 1H), 7.19 (d, *J =* 2.1 Hz, 1H), 7.04 (t, *J =* 7.7 Hz, 1H), 6.44 (dd, *J =* 8.1, 1.9 Hz, 1H), 4.32 (s, 1H), 2.17 (dq, *J =* 14.5, 7.3 Hz, 1H), 2.08 (dq, *J =* 14.4, 7.4 Hz, 1H), 1.33 (d, *J =* 3.1 Hz, 12H), 1.07 (t, *J =* 7.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 143.1, 138.5, 129.2, 128.7, 128.6, 126.1, 125.8, 123.2, 119.9, 117.5, 83.9, 62.1, 38.5, 25.02, 24.98, 9.0.

¹¹B NMR (128 MHz, CDCl₃) δ 31.2.

FT-IR (film): 3384, 2978, 2929, 2229, 1576, 1361, 1319, 1144, 995, 854, 760, 702 cm-1 .

HRMS (ESI+) m/z [M+H]⁺ calcd for $C_{22}H_{28}BN_2O_2^{\text{+}}$: 363.2238, found: 363.2261.

 $[\alpha]^{23}$ _D = +174 (*c* 1.0, CHCl₃); 93% ee, from (*S*)-**P**.

2-((3,5-Dimethoxyphenyl)amino)-2-phenylbutanenitrile. The title compound was prepared according to **GP-4** from 2-chloro-2-phenylbutanenitrile and 3,5-dimethoxyaniline with 24 h of reaction time. The product was purified by column chromatography on silica gel (hexanes/EtOAc/dichloromethane $= 10/1/2$). Yellow solid.

(R)-**P**: 127 mg, 54% yield, 90% ee; *(S)*-**P**: 116 mg, 49% yield, 89% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (10% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 12.3 min (major), 19.4 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.58 (d, *J =* 7.6 Hz, 2H), 7.40 (t, *J =* 7.5 Hz, 2H), 7.34 (t, *J =* 7.3 Hz, 1H), 5.93 (t, *J =* 2.0 Hz, 1H), 5.71 (d, *J =* 2.0 Hz, 2H), 4.33 (s, 1H), 3.61 (s, 6H), 2.19 (dq, *J =* 14.5, 7.3 Hz, 1H), 2.08 (dq, *J =* 14.8, 7.5 Hz, 1H), 1.06 (t, *J =* 7.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 161.3, 145.6, 138.6, 129.2, 128.7, 125.7, 119.9, 94.7, 92.4, 62.3, 55.2, 38.4, 8.9.

FT-IR (film): 3370, 2972, 2937, 2839, 2229, 1617, 1601, 1457, 1205, 1153, 1069, 925, 816, 761, 700 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₁₈H₂₁N₂O₂⁺: 297.1598, found: 297.1615.

 $[\alpha]^{23}$ _D = –176 (*c* 1.0, CHCl₃); 90% ee, from *(R)*-**P**.

*N***-(***tert***-butyl)-2-phenyl-2-(***p***-tolylamino)butanamide.** The title compound was prepared according to **GP-4** from *N*-(*tert*-butyl)-2-chloro-2-phenylbutanamide and *p*toluidine with 48 h of reaction time. The product was purified by column chromatography on silica gel (hexanes/EtOAc/dichloromethane $= 30/1/1$). Pale-yellow solid.

(R)-**P**: 128 mg, 49% yield, 88% ee; *(S)*-**P**: 146 mg, 59% yield, 88% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (3% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 4.4 min (major), 4.8 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.60 (d, *J =* 7.9 Hz, 2H), 7.38 (t, *J =* 7.7 Hz, 2H), 7.29 (t, *J =* 7.3 Hz, 1H), 6.87 (d, *J =* 8.4 Hz, 2H), 6.41 (d, *J =* 8.4 Hz, 2H), 5.87 (s, 1H), 5.08 (s, 1H), 2.54 (dq, *J =* 14.6, 7.4 Hz, 1H), 2.28 (dq, *J =* 14.3, 7.2 Hz, 1H), 2.20 (s, 3H), 1.23 (s, 9H), 0.77 (t, $J = 7.3$ Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 172.5, 142.7, 142.2, 129.5, 129.0, 127.5, 127.2, 126.3, 115.7, 66.5, 51.2, 28.5, 26.0, 20.5, 7.9.

FT-IR (film): 3379, 3055, 3021, 2967, 2934, 2877, 1669, 1617, 1517, 1394, 1364, 1296, 1227, 1034, 812, 757, 700 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₂₁H₂₉N₂O⁺: 325.2274, found: 325.2296.

 $[\alpha]^{23}$ _D = +24.3 (*c* 1.0, CHCl₃); 88% ee, from *(R)*-**P**.

*N***-(***tert***-Butyl)-2,3-diphenyl-2-(***p***-tolylamino)propenamide.** The title compound was prepared according to **GP-4** from *N*-(*tert-*butyl)-2-chloro-2,3-diphenylpropanamide and *p*toluidine with 48 h of reaction time. The product was purified by column chromatography on silica gel (hexanes/EtOAc/dichloromethane $= 50/1/1$). Pale-yellow solid.

(R)-**P**: 165 mg, 53% yield, 85% ee; *(S)*-**P**: 162 mg, 52% yield, 87% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK IC column (5% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 6.5 min (major), 5.5 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.51 (d, *J =* 7.6 Hz, 2H), 7.33 (t, *J =* 7.5 Hz, 2H), 7.28 (d, *J =* 7.2 Hz, 1H), 7.13 (t, *J =* 7.2 Hz, 1H), 7.08 (t, *J =* 7.4 Hz, 2H), 6.94 (d, *J =* 8.0 Hz, 2H), 6.73 (d, *J =* 7.5 Hz, 2H), 6.44 (d, *J =* 8.1 Hz, 2H), 6.20 (s, 1H), 4.57 (s, 1H), 3.72 (d, *J =* 13.7 Hz, 1H), 3.60 (d, *J =* 13.6 Hz, 1H), 2.24 (s, 3H), 1.22 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 171.9, 142.0, 141.6, 136.3, 130.8, 129.7, 128.7, 127.8, 127.7, 127.5, 126.7, 126.2, 116.1, 67.1, 51.2, 40.2, 28.4, 20.6.

FT-IR (film): 3386, 3062, 3029, 2965, 2919, 2864, 1675, 1617, 1517, 1453, 1365, 1226, 1033, 809, 757, 701 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₂₆H₃₁N₂O⁺: 387.2431, found: 387.2468.

 $[\alpha]^{23}$ _D = -28.5 (*c* 1.0, CHCl₃); 85% ee, from *(R)*-**P**.

*N***,2-Diphenyl-2-(***p***-tolylamino)butanamide.** The title compound was prepared according to **GP-4** from 2-chloro-*N*,2-diphenylbutanamide and *p*-toluidine with 48 h of reaction time. The product was purified by column chromatography on silica gel $(hexanes/EtOAc = 30/1)$. Pale-yellow solid.

(R)-**P**: 168 mg, 61% yield, 84% ee; *(S)*-**P**: 168 mg, 61% yield, 84% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK OD column (3% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 10.1 min (major), 14.3 min (minor).

¹H NMR (500 MHz, CDCl3) δ 8.33 (s, 1H), 7.70 (d, *J =* 8.0 Hz, 2H), 7.41 (t, *J =* 8.0 Hz, 4H), 7.31 (t, *J =* 7.3 Hz, 1H), 7.26 (t, *J =* 7.7 Hz, 2H), 7.06 (t, *J =* 7.4 Hz, 1H), 6.94 (d, *J =* 8.3 Hz, 2H), 6.55 (d, *J =* 8.4 Hz, 2H), 4.82 (s, 1H), 2.64 (dq, *J =* 14.7, 7.4 Hz, 1H), 2.44 (dq, *J =* 14.5, 7.3 Hz, 1H), 2.22 (s, 3H), 0.74 (t, *J =* 7.3 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 171.8, 141.8, 141.1, 137.8, 129.8, 129.2, 129.0, 128.6, 127.9, 126.2, 124.4, 119.9, 116.4, 67.5, 25.7, 20.5, 7.8.

FT-IR (film): 3351, 3055, 3023, 2975, 2934, 2876, 1675, 1617, 1599, 1517, 1439, 1312, 1245, 1185, 812, 753, 693 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₂₃H₂₅N₂O⁺: 345.1961, found: 345.1978. $[\alpha]^{23}$ _D = +4.5 (*c* 1.0, CHCl₃); 84% ee, from *(S*)-**P**.

2-Cyclopentyl-*N***-phenyl-2-(***p***-tolylamino)propenamide.** The title compound was prepared according to **GP-4** from 2-bromo-2-cyclopentyl-*N*-phenylpropanamide and *p*toluidine with 48 h of reaction time. The product was purified by column chromatography on silica gel (hexanes/EtOAc = $30/1$). Pale-yellow solid.

(R)-**P**: 118 mg, 46% yield, 60% ee; *(S)*-**P**: 117 mg, 45% yield, 60% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK IC column (10% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 6.5 min (minor), 7.0 min (major).

¹H NMR (500 MHz, CDCl3) δ 9.04 (s, 1H), 7.52 (d, *J =* 8.1 Hz, 2H), 7.31 (t, *J =* 7.9 Hz, 2H), 7.09 (t, *J =* 7.4 Hz, 1H), 6.99 (d, *J =* 8.2 Hz, 2H), 6.59 (d, *J =* 8.3 Hz, 2H), 3.77 (s, 1H), 2.36 (p, *J =* 9.3 Hz, 1H), 2.25 (s, 3H), 1.91 – 1.73 (m, 2H), 1.73 – 1.50 (m, 6H), 1.45 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 173.8, 142.3, 138.0, 129.9, 129.3, 129.1, 124.2, 119.9, 117.0, 63.7, 50.3, 27.4, 27.0, 25.54, 25.49, 20.6, 17.8.

FT-IR (film): 3386, 3349, 2956, 2868, 1673, 1598, 1529, 1499, 1442, 1314, 1242, 1163, 1074, 1029, 755, 690 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for C₂₆H₃₁N₂O⁺: 387.2431, found: 387.2468.

 $[\alpha]^{23}$ _D = +44.1 (*c* 1.0, CHCl₃); 60% ee, from *(S*)-**P**.

2,4-Dimethyl-*N***-phenyl-2-(***p***-tolylamino)pentanamide.** The title compound was prepared according to **GP-4** from 2-bromo-2,4-dimethyl-*N*-phenylpentanamide and *p*toluidine with 48 h of reaction time. The product was purified by column chromatography on silica gel (hexanes/EtOAc = $40/1$). Pale-orange solid.

(R)-**P**: 182 mg, 73% yield, 61% ee; *(S)*-**P**: 170 mg, 68% yield, 59% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK IC column (2% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 9.3 min (major), 8.7 min (minor).

¹H NMR (500 MHz, CDCl3) δ 9.04 (s, 1H), 7.50 (d, *J =* 7.9 Hz, 2H), 7.30 (t, *J =* 7.9 Hz, 2H), 7.09 (t, *J =* 7.4 Hz, 1H), 6.97 (d, *J =* 8.0 Hz, 2H), 6.55 (d, *J =* 8.2 Hz, 2H), 3.76 (s, 1H), 2.24 (s, 3H), 1.94 – 1.85 (m, 1H), 1.85 – 1.79 (m, 2H), 1.50 (s, 3H), 0.99 (d, *J =* 6.4 Hz, 3H), 0.92 (d, *J =* 6.5 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 174.0, 142.0, 138.0, 129.9, 129.2, 129.1, 124.3, 119.9, 116.9, 62.2, 48.9, 25.0, 24.5, 24.1, 21.7, 20.6.

FT-IR (film): 3344, 2956, 2927, 2868, 1675, 1617, 1601, 1512, 1439, 1311, 1250, 812, 754, 693 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for $C_{20}H_{27}N_2O^+$: 311.2118, found: 311.2145. $[\alpha]^{21}$ _D = +21.2 (*c* 1.0, CHCl₃); 61% ee, from *(S)*-**P**.

2.4.4. Effect of reaction parameters

General Procedure 6 (GP-6). In a nitrogen-filled glovebox, a 4 mL vial was charged with a PTFE stir bar, CuCl (1.0 mg, 0.010 mmol), **P** (14 mg, 0.012 mmol), toluene (1 mL), and BTPP (38 μL, 0.12 mmol). The resulting mixture was stirred for 4 h. A separate 4 mL vial was charged with a PTFE stir bar, *p*-toluidine (13 mg, 0.12 mmol), 2-chloro-2 phenylbutanenitrile (18 mg, 0.10 mmol), and toluene (0.5 mL). This mixture was stirred for 1 min, and then the solution of the catalyst was added; additional toluene (0.5 mL) was used to wash the vial that had contained the catalyst, and the washing was added to the reaction mixture. The reaction vial was capped with a PTFE-lined cap, removed from the glovebox, submerged in a low-form hemispherical Dewar flask filled with pre-cooled methanol (–78 °C), and stirred for 5 min. Two Kessil[®] PR160-440nm lamps were installed 5 cm above the vial, and the vial was irradiated for 4 h.

Work-up. The reaction was quenched by turning off the lamps. The reaction mixture was opened to air and passed through a 2 cm plug of silica gel loaded in a pipet, with EtOAc washings (3 x 4 mL). The filtrate was concentrated in vacuo, 1,3,5 trimethoxybenzene (internal standard; 17 mg, 0.10 mmol) was added, and the mixture was dissolved in CDCl₃ (0.6 mL). The yield was determined via ¹H NMR spectroscopy. Part of the solution in CDCl₃ was purified by preparative TLC (hexane/EtOAc 5:1 eluent) and analyzed via chiral HPLC analysis to determine the enantiomeric excess.

Note. The model electrophile, 2-chloro-2-phenylbutanenitrile, was stable for at least 10 min under the reaction conditions in the absence of irradiation.

Table 2.2. Effect of the Reaction Parameters.

| entry | variation from the "standard conditions" | yield (%) | ee (%) | entry | variation from the "standard conditions" | yield (%) | ee (%) |
|-----------------|-------------------------------------------------------|-----------|----------------|-----------------|------------------------------------------|------------|------------|
| | none | 77 | 92 | 21 ^b | BTMG, instead of BTPP | 75 | 92 |
| 2^a | no CuCl | 2 | | 22^b | DBU, instead of BTPP | 14 | 52 |
| 3 | no P | 2 | \blacksquare | 23 | CsOPh-H ₂ O, instead of BTPP | 16 | 86 |
| 4 | no BTPP | 2 | \blacksquare | 24 | Et ₂ O, instead of Toluene | 72 | 92 |
| 5 | no hv | 2 | | 25 | EtOAc, instead of Toluene | 68 | 94 |
| 6 | no hv, room temperature | 5 | | 26 | -60 °C, instead of -78 °C | 49 | 81 |
| | no hv, no P | 5 | | 27 | -10 °C, instead of -78 °C | 16 | 44 |
| 8 | no hv, 80 °C | 14 | 13 | 28 | 5 mol% P | 60 | 76 |
| 9 | no hv, no CuCl, no P, 80 °C | 5 | \sim | 29 | 20 mol% P | 79 | 92 |
| 10 ^b | CuBr instead of CuCl | 77 | 92 | 30 | 2 mol% CuCl, 2.4 mol% P | $38(56^c)$ | $90(90^c)$ |
| 11 ^b | CuCl ₂ instead of CuCl | 68 | 87 | 31 | 5 mol% CuCl, 6 mol% P | $70(73^c)$ | $92(92^c)$ |
| 12^b | (MeCN) ₄ CuBF ₄ instead of CuCl | 71 | 92 | 32 | 1.1 equiv p-Toluidine | 73 | 92 |
| 13 ^c | L1, instead of P | 33 | 41 | 33 | 1.5 equiv p-Toluidine | 77 | 92 |
| 14 ^c | L2, instead of P | 5 | 7 | 34 | 0.1 M, instead of 0.05 M | 75 | 92 |
| 15 ^c | L3, instead of P | <2 | | 35 | 1 lamp | 78 | 92 |
| 16 ^c | L4. instead of P | 21 | 20 | 36 | 3 lamps | 78 | 93 |
| 17 ^c | L5, instead of P | 8 | 4 | 37 | 2 h, instead of 4 h | 72 | 92 |
| 18 ^c | L6, instead of P | 2 | \blacksquare | 38 | 5 equiv H_2O | 78 | 92 |
| 19 ^c | L7, instead of P | 12 | 2 | 39 | air (5 mL) purged | 68 | 92 |
| 20 ^c | L8, instead of P | 6 | rac | 40 ^d | HPLC grade toluene without degassing | 76 | 92 |
| | | | | 41 ^d | 10 mol% CuA | 76 | 92 |

All data are the average of two or more runs. ^{*a*} Conversion of the electrophile: <1%. ^{*b*} To ensure reproducibility, stirring time during catalyst preparation: 6 h. *^c* Reaction time: 24 h. *^d* The reaction was set up outside of the glovebox, using standard air-free techniques.

2.4.5. Functional-group compatibility

2-Chloro-2-phenylbutanenitrile was reacted with *p*-toluidine according to modified **GP-6** in the presence of 1.0 equiv of each of the additives shown below. The additive was added before the addition of the catalyst solution. The yield and the recovery of the additive was determined by GC analysis using dodecane as an internal standard, and the ee was determined by chiral HPLC.

All data are the average of two or more runs. *^a* Reaction time: 24 h.

2.4.6. Gram-scale coupling

In the air, an oven-dried 250 mL round-bottom flask equipped with a PTFE stir bar was charged with CuCl (80 mg, 0.80 mmol) and (R) - P (1.13 g, 0.960 mmol). The flask was sealed with a rubber stopper, and the joint was wrapped with electrical tape. The flask was evacuated/backfilled with nitrogen three times, using a 20G needle attached to a Schlenk line. Then, a nitrogen-filled balloon was attached, and toluene (50 mL) and BTPP (2.90 mL, 9.60 mmol) were added via syringe. The mixture was stirred for 4 h, and then additional toluene (80 mL) was added. *p*-Toluidine (1.03 g, 9.60 mmol) was weighed into an oven-dried 40 mL vial equipped with a PTFE stir bar. The vial was capped with a PTFE-lined open top cap, and the joint was wrapped with electrical tape. The vial was attached to a Schlenk line and evacuated/backfilled with nitrogen three times. A nitrogenfilled balloon was attached, and 15 mL of toluene was added. This solution of *p*-toluidine was transferred to the round-bottom flask containing the catalyst solution. An additional 15 mL of toluene was used to wash the vial, and the washing was added to the flask. The flask was submerged in a low-form hemispherical Dewar flask filled with pre-cooled methanol $(-78 \degree C)$, and the reaction mixture was stirred for 10 min. 2-Chloro-2-phenylbutanenitrile (1.43 g, 8.00 mmol) was weighed into a syringe and transferred to the pre-cooled reaction mixture. The reaction mixture was stirred for 5 min, and then four Kessil® PR160-440 nm lamps were installed 5 cm above the reaction mixture, and the mixture was irradiated for 24 h under a positive pressure of nitrogen from the nitrogen-filled balloon.

Work-up. The reaction was quenched by turning off the lamps. The reaction mixture was warmed to room temperature, opened to air, and concentrated in vacuo. 30% aq NH4OH (20 mL) and EtOAc (30 mL) were added, and the reaction mixture was vigorously stirred for 15 min, at which point the aqueous layer was blue (see **Note 2** in **GP-4**). The aqueous layer was extracted with EtOAc (3 x 10 mL), and the combined organic layer was dried over $Na₂SO₄$ and then passed through a plug of silica gel (2 cm) with an EtOAc washing (50 mL). The organic layer was concentrated in vacuo and purified by flash

column chromatography on silica gel (hexanes/EtOAc/dichloromethane $= 40/1/1$) to afford a pale-yellow solid (1.28 g, 64% yield). The ee was determined via HPLC analysis to be 92%.

2.4.7. Ligand recovery

2-Chloro-2-phenylbutanenitrile was reacted with *p*-toluidine (Run 1) and 4 methoxyaniline (Run 2) according to **GP-4**. Fractions containing the ligand were collected via column chromatography during the purification of the coupling product. The ligand was further purified by column chromatography on silica gel: column #1 (hexanes/EtOAc = $100/1$; column #2 (hexanes/toluene = $1/1$).

Run 1: recovered *(S)*-**P**: 92 mg (82% of the initial loading). Run 2: recovered *(R)*-**P**: 91 mg (81% of the initial loading).

Recovered *(R)*-**P** and *(S)*-**P** were used in a copper-catalyzed coupling between 2-chloro-2-phenylbutanenitrile and *p*-toluidine, following **GP-6**. Recovered *(R)*-**P**: 75% yield and 92% ee. Recovered *(S)*-**P**, 75% yield and 92% ee.

2.4.8. Couplings of an alkyl carbonate and an alkyl fluoride

Procedure. In a nitrogen-filled glovebox, a 40 mL vial equipped with a PTFE magnetic stir bar was charged with CuCl (8.0 mg, 0.080 mmol), **P** (113 mg, 0.096 mmol), toluene (5 mL), and BTPP (290 μL, 0.960 mmol). The reaction mixture was stirred for 4 h. Next, *p*toluidine (103 mg, 0.960 mmol), tetrabutylammonium chloride (289 mg, 1.04 mmol), and toluene (8 mL) were added, and the mixture was stirred for 5 min. The vial was capped with a PTFE-lined open top cap, and the joint was wrapped with electrical tape. The vial was removed from the glovebox, and a nitrogen-filled balloon was attached. The vial was

submerged in a low-form hemispherical Dewar flask filled with pre-cooled methanol (– 78 °C), and the reaction mixture was stirred for 5 min. The electrophile (0.800 mmol) was weighed into a 4 mL vial, and the vial was capped with a PTFE-lined open top cap. The vial was attached to a Schlenk line and evacuated/backfilled with nitrogen three times. A nitrogen-filled balloon was attached, and 1.5 mL of toluene was added. The resulting solution was transferred to the pre-cooled reaction mixture; additional toluene (1.5 mL) was used to wash the vial, and the washing was added to the reaction mixture. $B(OMe)_{3}$ (180 μL, 1.6 mmol) was added via syringe, and the resulting mixture was stirred for 5 min. The balloon was removed, and vacuum grease was applied to the PTFE cap to seal the needle puncture. Kessil® PR160-440 nm lamps (one per reaction) were installed 5 cm above the reaction, and the vial was irradiated for 48 h.

Work-up. Same as **GP-4.**

2-Phenyl-2-(*p***-tolylamino)butanenitrile**. The title compound was prepared according to the above procedure from *p*-toluidine and the two electrophiles indicated below, with 48 h of reaction time. The product was purified by column chromatography on silica gel $(hexanes/EtOAc/dichlorome thane = 30/1/1)$. Pale-yellow solid.

From 2-fluoro-2-phenylbutanenitrile: *(R)*-**P**: 152 mg, 76% yield, 88% ee; *(S)*-**P**: 147 mg, 73% yield, 88% ee.

From 4-chlorophenyl (1-cyano-1-phenylpropyl) carbonate: *(R)*-**P**: 113 mg, 57% yield, 87% ee; *(S)*-**P**: 121 mg, 60% yield, 86% ee.

Control Experiments. 2-Fluoro-2-phenylbutanenitrile was reacted with *p*-toluidine according to modified **GP-6** in the presence of 1.0 equiv the additives. The yield of the product and the conversion of the electrophile were determined by ${}^{1}H$ NMR analysis using 1,3,5-trimethoxybenzene as an internal standard, and the ee was determined by chiral HPLC.

Overview of the X-ray crystal structures of the coupling products, with thermal ellipsoids at 50% probability level.

All other absolute configurations are assigned by analogy.

Thermal ellipsoid plot at 50% probability level. For clarity, the hydrogen atoms are not shown. The N–H hydrogen atom was found in the residual density map and refined isotropically.

A colorless needle-like specimen of $C_{17}H_{18}N_2$ was used for the X-ray crystallographic analysis. The X-ray intensity data were measured ($\lambda = 1.54178$ Å). The total exposure time was 21.48 h. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using an orthorhombic unit cell yielded a total of 16065 reflections to a maximum θ angle of 70.05 \degree (0.82 Å resolution), of which 2665 were independent (average redundancy 6.028, completeness = 100.0% , R_{int} = 7.28%, $R_{sig} = 4.40\%$) and 2477 (92.95%) were greater than $2\sigma(F^2)$. The final cell constants of <u>a</u> = 5.8710(6) Å, <u>b</u> = 9.1726(9) Å, <u>c</u> = 25.984(4) Å, volume = 1399.3(3) Å³, are based upon the refinement of the XYZ-centroids of 8448 reflections above 20 σ(I) with $6.803^\circ < 2θ <$ 144.9°. Data were corrected for absorption effects using the Multi-Scan method (SADABS). The ratio of minimum to maximum apparent transmission was 0.765.

The structure was solved and refined using the Bruker SHELXTL Software Package, using the space group P 21 21 21, with $Z = 4$ for the formula unit, $C_{17}H_{18}N_2$. The final anisotropic full-matrix least-squares refinement on F^2 with 178 variables converged at R1 = 3.81%, for the observed data and $wR2 = 8.87\%$ for all data. The goodness-of-fit was 1.101. The largest peak in the final difference electron density synthesis was 0.191 e/\hat{A}^3 , and the largest hole was -0.156 e⁻/ \AA ³ with an RMS deviation of 0.038 e^{-/ \AA 3. On the basis of the} final model, the calculated density was 1.188 g/cm^3 and F(000), 536 e.

Sample and crystal data

Data collection and structural refinement

Thermal ellipsoid plot at 50% probability level. For clarity, the hydrogen atoms are not shown. The N–H hydrogen atom was found in the residual density map and refined isotropically.

A colorless needle-like specimen of $C_{25}H_{25}N_3O$, approximate dimensions 0.050 mm x 0.050 mm x 0.200 mm, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured ($\lambda = 1.54178$ Å). The integration of the data using an orthorhombic unit cell yielded a total of 24918 reflections to a maximum θ angle of 70.07° (0.82 Å resolution), of which 4074 were independent (average redundancy 6.116, completeness = 100.0%, R_{int} = 4.91%, R_{sig} = 2.65%) and 3848 (94.45%) were greater than $2\sigma(F^2)$. The final cell constants of <u>a</u> = 10.1183(4) Å, <u>b</u> = 13.4557(5) Å, <u>c</u> = 15.7406(6) Å, volume = 2143.06(14) \AA^3 , are based upon the refinement of the XYZ-centroids of 9977 reflections above 20 σ(I) with 8.645° < 2θ < 144.7°. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.8930 and 0.9720.

The structure was solved and refined using the Bruker SHELXTL Software Package, using the space group P 21 21 21, with $Z = 4$ for the formula unit, $C_{25}H_{25}N_3O$. The final anisotropic full-matrix least-squares refinement on $F²$ with 268 variables converged at R1 $= 3.06\%$, for the observed data and wR2 = 7.66% for all data. The goodness-of-fit was 1.085. The largest peak in the final difference electron density synthesis was 0.138 e⁻ $/\AA$ ³, and the largest hole was -0.186 e^{$/\AA$ 3} with an RMS deviation of 0.035 e $/\AA$ ³. On the basis of the final model, the calculated density was 1.189 g/cm³ and $F(000)$, 816 e.

Sample and crystal data

Data collection and structural refinement

Thermal ellipsoid plot at 50% probability level. For clarity, the hydrogen atoms are not shown. The N–H hydrogen atom was found in the residual density map and refined isotropically.

A clear colorless block-like specimen of $C_{17}H_{27}N_2O_3P$, approximate dimensions 0.100 mm x 0.150 mm x 0.300 mm, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured ($\lambda = 1.54178$ Å). The total exposure time was 2.67 h. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using a monoclinic unit cell yielded a total of 18004 reflections to a maximum θ angle of 70.04 \degree (0.82 Å resolution), of which 3476 were independent (average redundancy 5.180, completeness = 100.0% , $R_{int} = 3.31\%$, $R_{sig} = 2.68\%$) and 3405 (97.96%) were greater than $2\sigma(F^2)$. The final cell constants of <u>a</u> = 9.7328(3) Å, <u>b</u> = 7.5268(2) Å, <u>c</u> = 12.9041(4) Å, β = 100.3380(10)°, volume = 929.97(5) \AA^3 , are based upon the refinement of the XYZ-centroids of 9861 reflections above 20 σ(I) with 6.963° < 2θ < 144.8°. Data were corrected for absorption effects using the Multi-Scan method (SADABS). The ratio of minimum to maximum apparent transmission was 0.871.

The structure was solved and refined using the Bruker SHELXTL Software Package, using the space group P 1 21 1, with $Z = 2$ for the formula unit, $C_{17}H_{27}N_2O_3P$. The final anisotropic full-matrix least-squares refinement on F^2 with 218 variables converged at R1 $= 2.41\%$, for the observed data and wR2 = 6.31% for all data. The goodness-of-fit was 1.056. The largest peak in the final difference electron density synthesis was 0.230 e $/\AA$ ³, and the largest hole was -0.235 e $/\AA$ ³ with an RMS deviation of 0.037 e $/\AA$ ³. On the basis of the final model, the calculated density was 1.208 g/cm³ and F(000), 364 e.

Sample and crystal data

Data collection and structural refinement

Thermal ellipsoid plot at 50% probability level. For clarity, the hydrogen atoms are not shown. The N–H hydrogen atom was found in the residual density map and refined isotropically.

A specimen of $C_{21}H_{28}N_2O$ was used for the X-ray crystallographic analysis. The X-ray intensity data were measured ($\lambda = 1.54178$ Å). The total exposure time was 10.53 h. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using an orthorhombic unit cell yielded a total of 22825 reflections to a maximum θ angle of 70.10° (0.82 Å resolution), of which 3758 were independent (average redundancy 6.074, completeness = 100.0%, $R_{int} = 6.46\%, R_{sig} = 3.78\%$) and 3540 (94.20%) were greater than $2\sigma(F^2)$. The final cell constants of <u>a</u> = 6.3131(14) Å, <u>b</u> = 16.797(4) Å, <u>c</u> = 18.605(3) Å, volume = 1972.9(7) Å³, are based upon the refinement of the XYZ-centroids of 9931 reflections above 20 $\sigma(I)$ with $7.088^{\circ} < 2\theta < 144.7^{\circ}$. Data were corrected for absorption effects using the Multi-Scan method (SADABS). The ratio of minimum to maximum apparent transmission was 0.883.

The structure was solved and refined using the Bruker SHELXTL Software Package, using the space group P 21 21 21, with $Z = 4$ for the formula unit, $C_{21}H_{28}N_2O$. The final anisotropic full-matrix least-squares refinement on F^2 with 230 variables converged at R1 $= 3.22\%$, for the observed data and wR2 = 7.79% for all data. The goodness-of-fit was 1.039. The largest peak in the final difference electron density synthesis was 0.165 e $/\AA$ ³, and the largest hole was -0.154 e^{$/\AA$ 3} with an RMS deviation of 0.032 e $/\AA$ ³. On the basis of the final model, the calculated density was 1.092 g/cm^3 and F(000), 704 e .

Sample and crystal data

Data collection and structural refinement

Thermal ellipsoid plot at 50% probability level. For clarity, the hydrogen atoms are not shown. The N–H hydrogen atom was found in the residual density map and refined isotropically.

A clear white needle-like specimen of $C_{21}H_{26}N_2O$ was used for the X-ray crystallographic analysis. The X-ray intensity data were measured ($\lambda = 1.54178$ Å). The total exposure time was 4.67 hours. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using a monoclinic unit cell yielded a total of 15223 reflections to a maximum θ angle of 74.68° (0.80 Å resolution), of which 3600 were independent (average redundancy 4.229, completeness = 99.4%, R_{int} = 3.42%, R_{sig} = 2.54%) and 3506 (97.39%) were greater than $2\sigma(F^2)$. The final cell constants of <u>a</u> = 11.3104(15) Å, <u>b</u> = 6.3399(4) Å, <u>c</u> = 13.0915(13) Å, $β = 108.923(9)°$, volume = 888.02(16) Å³, are based upon the refinement of the XYZcentroids of 9981 reflections above 20 $\sigma(I)$ with 7.138° < 2 θ < 144.4°. Data were corrected for absorption effects using the Multi-Scan method (SADABS). The ratio of minimum to maximum apparent transmission was 0.895.

The structure was solved and refined using the Bruker SHELXTL Software Package, using the space group P2(1), with $Z = 2$ for the formula unit, $C_{21}H_{26}N_2O$. The final anisotropic full-matrix least-squares refinement on F^2 with 231 variables converged at R1 $= 2.99\%$, for the observed data and wR2 = 7.80% for all data. The goodness-of-fit was 1.040. The largest peak in the final difference electron density synthesis was 0.186 e $/\AA$ ³, and the largest hole was -0.156 e^{$/\AA$ 3} with an RMS deviation of 0.031 e $/\AA$ ³. On the basis of the final model, the calculated density was 1.206 g/cm³ and F(000), 348 e.

Sample and crystal data

Data collection and structure refinement

2.4.10. Mechanistic studies

2.4.10.1.Preparation of *(S)***-DTBM-SEGPHOS-CuCl (CuA)**

*(S)***-DTBM-SEGPHOS-CuCl (CuA).** The title compound was synthesized via a slight modification of a literature procedure.^{10,11} An oven-dried 20 mL vial was charged with CuCl (104 mg, 1.05 mmol), (*S*)-**P** (1.18 g, 1.00 mmol), and a magnetic stir bar. THF (4 mL) was added, and the vial was capped and covered with aluminum foil. The reaction mixture was stirred for 6 h, and then it was filtered through Grade GF/D Glass Microfiber (Whatman products, Cytiva) tightly fixed in an oven-dried glass pipette. The solvent was removed in vacuo, and the resulting solid was triturated with pentane (2 mL) three times to afford a white solid (1.27 g, 99% yield). Spectroscopic data match those reported in the literature.^{[34](#page-30-0)[,50](#page-202-0)} The title compound was stored in a glovebox (-35 °C) .

¹H NMR (400 MHz, toluene- d_8) δ 8.51 (br, 4H), 7.80 (t, $J = 5.8$ Hz, 4H), 6.77 (dt, J *=* 9.1, 4.8 Hz, 2H), 6.19 (d, *J =* 8.0 Hz, 2H), 5.28 (s, 2H), 5.13 (s, 2H), 3.32 (s, 6H), 3.22 (s, 6H), 1.58 (s, 36H), 1.31 (s, 36H).

³¹P NMR (162 MHz, toluene- d_8) δ -4.8.

2.4.10.2.Representative side products from a coupling reaction

2-Chloro-2-phenylbutanenitrile was reacted with *p*-toluidine according to **GP-6**. The yields of 2-phenyl-2-(*p*-tolylamino)butanenitrile, 2,3-diethyl-2,3-diphenylsuccinonitrile, and (E) -1,2-di-*p*-tolyldiazene were analyzed via ¹H NMR spectroscopy with 1,3,5trimethoxybenzene (17 mg, 0.10 mmol) as an internal standard.

Figure 2.18. Representative ¹H NMR spectrum (CDCl₃, 500 MHz) of an unpurified reaction mixture showing a 78% yield of the C–N coupling product (calibrated using 1,3,5 trimethoxybenzene).

2,3-Diethyl-2,3-diphenylsuccinonitrile. The title compound was isolated via preparative TLC (hexanes/EtOAc = $10/1$) as a mixture of two diastereomers. Spectroscopic data match those reported in the literature.^{[51](#page-202-1)}

Meso: ¹H NMR (500 MHz, CDCl₃) δ 7.57 (d, *J* = 7.1 Hz, 4H), 7.44 (d, *J* = 7.0 Hz, 6H), 2.43 (dq, *J =* 14.3, 7.3 Hz, 2H), 1.84 (dq, *J =* 14.4, 7.3 Hz, 2H), 0.81 (t, *J =* 7.2 Hz, 6H). (S, S) and (R, R) : ¹H NMR (500 MHz, CDCl₃) δ 7.24 (d, *J* = 7.2 Hz, 2H), 7.18 (t, *J* = 7.5 Hz, 4H), 7.06 (d, *J =* 7.7 Hz, 4H), 2.52 (qd, *J =* 7.1, 2.4 Hz, 4H), 0.97 (t, *J =* 7.2 Hz, 6H).

$$
p\text{-}\mathrm{Hol} \setminus \bigwedge_{N} N \setminus p\text{-}\mathrm{Hol}
$$

*(E)***-1,2-di-***p***-tolyldiazene.** The title compound was purified via preparative TLC (hexanes/EtOAc = $10/1$). Spectroscopic data match those reported in the literature.^{[52](#page-202-2)}

¹H NMR (500 MHz, CDCl3) δ 7.81 (d, *J =* 8.1 Hz, 4H), 7.31 (d, *J =* 8.0 Hz, 4H), 2.44 (s, 6H).

2.4.10.3. Coupling with aniline (PhNH2) as the nucleophile

2-Chloro-2-phenylbutanenitrile was reacted with aniline according to **GP-6**. The yields of 2-phenyl-2-(phenylamino)butanenitrile and 2-(4-aminophenyl)-2-phenylbutanenitrile were analyzed via 1 H NMR spectroscopy with 1,3,5-trimethoxybenzene (17 mg, 0.10) mmol) as an internal standard. Samples of pure products were obtained via preparative TLC (hexanes/EtOAc $= 5/1$) for chiral HPLC analysis.

2-Phenyl-2-(phenylamino)butanenitrile. Yellow solid. *(R)*-**P**: 17% yield, 83% ee; *(S)*-**P**: 18% yield, 84% ee

HPLC analysis: The ee was determined via HPLC on a CHIRALCEL OD-H column (10% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 6.6 min (major), 8.2 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.59 (d, *J =* 7.7 Hz, 2H), 7.40 (t, *J =* 7.4 Hz, 2H), 7.35 (t, *J =* 7.2 Hz, 1H), 7.10 (t, *J =* 7.8 Hz, 2H), 6.78 (t, *J =* 7.3 Hz, 1H), 6.54 (d, *J =* 8.2 Hz, 2H), 4.30 (s, 1H), 2.21 (dq, *J =* 14.5, 7.3 Hz, 1H), 2.10 (dq, *J =* 14.5, 7.5 Hz, 1H), 1.07 (t, *J =* 7.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 143.7, 138.5, 129.2, 129.1, 128.7, 125.8, 120.0, 119.9, 115.8, 62.2, 38.5, 9.0.

FT-IR (film): 3382, 3057, 3027, 2973, 2851, 2232, 1604, 1518, 1499, 1449, 1318, 1258, 1204, 1181, 1078, 1038, 919, 872, 747, 697, 691 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for $C_{16}H_{17}N_2^+$: 237.1386, found: 237.1398.

 $[\alpha]^{23}$ _D = –154 (*c* 1.0, CHCl₃); 83% ee, from (*R*)-**P**.

2-(4-Aminophenyl)-2-phenylbutanenitrile. Brown solid.

(R)-**P**: 22% yield, 23% ee; *(S)*-**P**: 26% yield, 24% ee.

HPLC analysis: The ee was determined via HPLC on a CHIRALPAK IC column (30% *i*-PrOH in hexane, 1 mL/min); retention times for compound obtained using *(R)*-**P**: 9.2 min (major), 7.7 min (minor).

¹H NMR (500 MHz, CDCl3) δ 7.40 – 7.29 (m, 4H), 7.27 (t, *J =* 7.3 Hz, 1H), 7.14 (d, *J =* 8.5 Hz, 2H), 6.64 (d, *J =* 8.5 Hz, 2H), 3.72 (s, 2H), 2.47 – 2.27 (m, *J =* 7.0 Hz, 2H), 1.05 (t, $J = 7.3$ Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 146.0, 140.9, 130.0, 128.8, 128.2, 127.7, 127.0, 122.8, 115.2, 51.9, 33.0, 10.3.

FT-IR (film): 3370, 3031, 2973, 2937, 2879, 2232, 1624, 1515, 1448, 1291, 1187, 1139, 824, 756, 699 cm⁻¹.

HRMS (ESI+) m/z [M+H]⁺ calcd for $C_{16}H_{17}N_2^+$: 237.1386, found: 237.1400.

 $[\alpha]^{23}$ _D = +13.5 (*c* 1.0, CHCl₃); 23% ee, from (R)-**P**.

2.4.10.4.Time-course profile of the standard coupling reaction

2-Chloro-2-phenylbutanenitrile and *p*-toluidine were reacted and analyzed (**GP-6**).

Figure 2.19. Time-course profile of the standard coupling reaction, analyzed via ¹H NMR spectroscopy to determine the conversion of the electrophile and the yield of the desired product and byproducts, and analyzed via chiral HPLC to determine the ee of the desired product. "time" = time of irradiation. Yields of the byproducts are based on the electrophile and yields of the "El dimer" are the sum of the two diastereomers.

2.4.10.5. Cu^A as the dominant Cu(I) species during catalysis

UV-vis analysis of the initial stage of the reaction at -78 **°C.** A solution of Cu_A (0.5) mM) was prepared by dissolving 6.4 mg (5.0 μmol) of **Cu^A** in 10 mL of toluene. A solution of **CuA** (0.5 mM), BTPP (6.0 mM), *p*-toluidine (6.0 mM), and 2-chloro-2 phenylbutanenitrile (5.0 mM) was prepared by dissolving 6.4 mg (5.0 μmol) of **CuA,** 18.8 mg (60 μmol) of BTPP, 6.4 mg of *p*-toluidine (60 μmol), and 9.0 mg of 2-chloro-2 phenylbutanenitrile (50 μmol) in 10 mL of toluene. Each solution was pipetted into a 1 cm path length cuvette, which was sealed with a high-vacuum Teflon valve (Kontes). The absorption spectra were acquired at -78 °C (**Figure 2.7a**). The absorption of **Cu**A has λ_{max} $=$ 376 nm with ε = 3890 M⁻¹ cm⁻¹.

The UV-vis absorption of each reagent in the catalytic reaction was measured to ensure that there is no significant overlap between **Cu^A** and the other reagents (**Figure 2.20**). Time-course UV-vis monitoring of **Cu^A** during catalysis could not be achieved due to overlap of the absorption peak of **Cu**A with peaks that emerge upon irradiation.

The emission spectrum of a Kessil® PR160-440 nm lamp was measured at the Beckman Institute Molecular Materials Resource Center (MMRC) using a StellarNet BlackComet Spectroradiometer.

Figure 2.20. UV-vis absorption of **Cu^A** (0.5 mM, black), 2-chloro-2-phenylbutanenitrile (5 mM, red), *p*-toluidine (6 mM, green), and BTPP (6 mM, blue) in toluene at -78 °C.

NMR analysis at room temperature. A solution of **Cu^A** (12.8 mg, 5.0 mM), BTPP (37.5 mg, 60 mM), and *p*-toluidine(12.9 mg, 60 mM) in toluene- d_8 (2 mL), as well as a solution of CuA (12.8 mg, 5.0 mM) in toluene- d_8 (2 mL), were prepared, and 500 μ L of each solution was loaded into a J. Young NMR tube. The samples were analyzed via ¹H NMR (400 MHz) and ³¹P NMR (162 MHz) spectroscopy at room temperature (**Figure 2.21**). ~3% free **P** is observed in the three-component mixture.

Figure 2.21. Blue: Cu_A only in toluene-d₈. Red: Cu_A, *p*-toluidine, and BTPP in toluene-d₈.

Temperature-dependent ¹H NMR spectra NMR spectra of CuA. A solution of **Cu^A** (3.2 mg, 5.0 mM) in toluene- d_8 (500 μ L) was analyzed via ¹H NMR (400 MHz) and ³¹P NMR (162 MHz) spectroscopy at different temperatures (**Figures 2.22** and **2.23**).

Figure 2.22. Temperature-dependent ¹H NMR spectra (400 MHz) of **CuA**.

Figure 2.23. Temperature-dependent ¹H NMR (400 MHz) and ³¹P NMR (162 MHz) spectra of **CuA**.

The complexity of the ³¹P NMR spectra at lower temperature is presumably due to hindered rotation of the 4-methoxy groups of DTBM-SEGPHOS (P) at low temperature.^{[34](#page-30-0)}

¹H NMR (–78 °C, 400 MHz, toluene-*d8*) δ 9.72 – 9.13 (m, 2H), 8.30 – 7.84 (m, 4H), 7.84 -7.48 (m, 2H), $6.97 - 6.82$ (m, 2H), $6.33 - 5.66$ (m, 2H), $5.24 - 4.55$ (m, 4H), $3.50 - 2.71$ (m, 12H), 1.82 (s, 18H), 1.55 (s, 18H), 1.47 (s, 18H), 1.18 (s, 18H).

³¹P NMR (–78 °C, 162 MHz, toluene-*d8*) δ –2.16, –2.81, –3.42, –3.87, –4.11, –4.50, – 5.11, –6.00, –6.38, –7.28.

Low-temperature monitoring of a catalyzed reaction via NMR spectroscopy.

Control reaction: 2-Chloro-2-phenylbutanenitrile was reacted with *p*-toluidine according to modified **GP-6** (in the presence of 1.0 equiv (17 mg, 0.10 mmol) of 1,3,5 trimethoxybenzene). The yield (80%, analyzed by GC using dodecane as an internal standard; average of two runs) and the ee (92%) of the coupling product were essentially identical to a reaction run in the absence of 1,3,5-trimethoxybenzene (77% yield and 92% ee), and 1,3,5-trimethoxybenzene was fully recovered from the reaction mixture (analyzed by GC). On the basis of these data, we conclude that the presence of 1,3,5 trimethoxybenzene does not interfere with the reaction of interest.

In a nitrogen-filled glovebox, a 4 mL vial equipped with a PTFE stir bar was sequentially charged with Cu_A (13 mg, 0.010 mmol), toluene- d_8 (2 mL), *p*-toluidine (13 mg, 0.12 mmol), BTPP (38 μL, 0.12 mmol), 1,3,5-trimethoxybenzene (17 mg, 0.10 mmol), and 2-chloro-2-phenylbutanenitrile (18 mg, 0.10 mmol). The mixture was stirred for 3 min, and then the resulting homogenous solution was transferred to three J. Young NMR tubes (500 μL each). The tubes were sealed, removed from the glovebox, and cooled for 5 min in a low-form hemispherical Dewar flask filled with pre-cooled methanol (–78 °C). Two Kessil® PR160-440 nm lamps were set at 50% intensity and placed 5 cm above the reaction mixtures. The mixtures were irradiated for the indicated times and then immediately frozen in a –116 °C bath (frozen ethanol). The tubes were quickly placed in a pre-cooled (–78 °C) Bruker 400 MHz NMR spectrometer and analyzed via ¹H NMR spectroscopy (**Figures 2.24** and **2.25**). The concentration of **Cu^A** was determined based on calibration of the resonance of Cu_A (δ 9.72 – 9.13 ppm, 2H) and the resonance of the internal standard (**Table 2.4**). The NMR tubes were warmed to room temperature and again analyzed via ¹H NMR spectroscopy to determine the yield and the conversion based on the internal standard. Overlap of the phosphorus chemical shifts of BTPP and Cu_A at –78°C prevented the reaction from being analyzed by ${}^{31}P$ NMR spectroscopy at $-78^{\circ}C$.

Figure 2.24. Monitoring Cu_A at low temperature during a coupling reaction.

Figure 2.25. Characteristic resonances of **Cu^A** are maintained during a coupling reaction monitored at low temperature.

| Reaction time | [CuA]/[CuA] | | Conversion $(\%)$ Product Yield $(\%)$ |
|----------------------|-------------|----|------------------------------------------|
| 10 min | 93% | 19 | 16 |
| 20 min | 79% | 42 | 36 |
| 30 min | 54% | 64 | 55 |

Table 2.4. Concentration of **CuA**, conversion of the electrophile, and yield of the coupling product according to NMR spectroscopy.

Note: The rate of these NMR-tube reactions was faster than that of the standard 0.10 mmol reaction (**GP-6**). The intensity of the light for the NMR-tube reactions was accordingly decreased so that the reactions have a similar rate (see the **Section 2.4.10.4**).

2.4.10.6. Steady-state and time-resolved emission spectroscopy of Cu^A

Steady-state emission spectra of CuA. Steady-state emission measurements at room temperature were performed with a 1 cm fluorescence cuvette containing a solution of **Cu^A** in toluene (0.50 mM). Steady-state emission measurements at 195 K and 77 K were performed by immersing EPR tubes containing solutions or glassed samples of **Cu^A** in toluene (0.5 mM) in a glass Dewar filled with dry ice/acetone or liquid nitrogen (**Figure 2.26**).

Figure 2.26. Steady-state emission of **Cu***A* in toluene at different temperatures (λexcitation = 355 nm).

Luminescence decay of Cu_A (298 K). Nanosecond \sim microsecond time scale luminescence decay measurements were performed with a 1 cm fluorescence cuvette containing a solution of **Cu**A in toluene (5 mM, $\lambda_{\text{pump}} = 355$ nm, $\lambda_{\text{probe}} = 480$ nm). Two sets of decay were observed (long-lived, microsecond scale and short-lived, nanosecond scale). The lifetime of the long-lived luminescence of **Cu^A** was determined by the default exponential fitting tool provided in OriginPro 8 (**Figure 2.27**).

The lifetime of the short-lived luminescence of **Cu^A** was independently measured with a 1 cm fluorescence cuvette containing a solution of **Cu**A in toluene (5 mM, $\lambda_{\text{pump}} = 255$ nm, $\lambda_{\text{probe}} = 480$ nm). The lifetime of the short-lived luminescence of Cu_A was determined by the default exponential fitting tool provided in OriginPro 8 (**Figure 2.28**).

Figure 2.27. Long-lived luminescence decay for **Cu^A** measured at 298 K.

Figure 2.28. Short-lived luminescence decay for **Cu^A** measured at 298 K.

Excitation nature of CuA. Two sets of luminescence decay at the same observation wavelength are frequently observed for thermally activated delayed fluorescence (TADF) materials.^{[53](#page-202-3)} While these materials typically have a red shift of the emission at lower temperature due to the diminished delayed fluorescence (larger contribution of the phosphorescence),^{[54](#page-202-4)} emission of Cu_A at 77 K shows a blue shift compared to that measured at 195 K. The luminescence decay measured at this temperature performed by immersing EPR tubes containing solutions or glassed samples of **Cu^A** in toluene (5 mM) in a glass Dewar filled with dry ice/acetone (195 K, **Figure 2.29**, left) or liquid nitrogen (77 K, **Figure 2.29**, right) shows that the observed slow-decay process still contributes significantly. We therefore attribute the two sets of the decay directly to the fluorescence and phosphorescence of **CuA**. Based on their lifetimes, we propose the phosphorescence to be the dominant process of **Cu^A** for the photoinduced reduction of the electrophiles. MO visualization of the ground and triplet excited states of **Cu^A** suggests ³MLCT from the copper *d* orbital to the ligand backbone (see the **Section 2.4.11**; DFT calculations).

Figure 2.29. Long-lived luminescence decay for **Cu^A** measured at 195 K (left) and 77 K (right).

2.4.10.7. Stern-Volmer quenching of Cu^A

In a glovebox, **Cu^A** was dissolved in toluene to provide a 0.010 M solution. In a 4 mL vial with a PTFE-lined open-top screw cap, 2-chloro-2-phenylbutanenitrile was dissolved in toluene to make 0.0050, 0.010, 0.020, 0.044, 0.054, 0.072 M solutions. 1.5 mL of the solutions containing **Cu^A** was charged into 1 cm fluorescence cuvettes with a magnetic stir bar and PTFE-lined open-top screw cap (Starna Cells), and the cuvettes were sealed. The sealed cuvettes and vials were removed from the glovebox. Before the measurement, 1.5 mL of the solutions containing the electrophile were added to the cuvettes by syringe using air-free technique, and the solutions were vigorously stirred for 1 min and immediately subjected to the measurement. The lifetime of a non-emissive excited state of **CuA** as a function of electrophile concentration was measured at room temperature by transient absorption spectroscopy ($\lambda_{\text{pump}} = 355$ nm, $\lambda_{\text{probe}} = 580$ nm) (**Figure 2.30**). Data were analyzed using OriginPro 8 with the default exponential curve fitting function. The rate of quenching was determined to be $k_q = 3.7 \times 10^8$ M⁻¹ s⁻¹ (**Figure 2.9**).

Figure 2.30. Transient absorbance decays for **Cu^A** with varying electrophile concentrations.

2.4.10.8. Electrochemistry

Electrochemical measurements were carried out using a CD instrument 600B electrochemical analyzer (**Figure 2.31** and **2.32**). A freshly polished glassy carbon electrode was used as the working electrode, and a graphite rod was used as the auxiliary electrode. An Ag/AgOTf reference electrode was employed, and solutions (THF) of electrolyte $(0.1 \text{ M} \text{ TBAPF}_6)$ and analyte (1 mM) were used for each measurement. All redox potentials in the present work are reported versus the Fc/Fc^+ couple $(+0.115 \text{ V}$ versus our Ag/AgOTf reference electrode).

Figure 2.31. Cyclic voltammogram of Cu_A with scan rates ranging from 10 mV/s to 180 mV/s (left) and plot of the peak potentials vs the log of the scan rate (right), showing quasireversible features at $E_{1/2} = \sim 0.4$ V (versus Fc/Fc⁺).

Figure 2.32. Cyclic voltammograms of representative electrophiles, showing irreversible features at $E_P = -2.17$ V and -2.75 V (versus Fc/Fc⁺), respectively. Scan rate = 50 mV/s.

2.4.10.9. Estimated excited-state oxidation potential of Cu^A in THF

Steady-state emission spectra of CuA. Steady-state emission spectra were collected with a 1 cm fluorescence cuvette containing a solution of Cu_A in THF (0.5 mM). When light with λ < 360 nm was used, decomposition of **Cu**A in THF generated a new emission band at ~400 nm and prevented the emission spectrum from being accurately measured. Therefore, the emission spectrum measured at $\lambda_{\text{excitation}} = 370$ nm was extrapolated using the default GaussAmp fitting function in OriginPro 8. The intensity plot of the emission of **Cu**A at different excitation wavelengths was measured using $\lambda_{\text{probe}} = 480$ nm (**Figure 2.33**). E^{00} is estimated from the crossing point of the two plots (410 nm, \sim 3.0 eV). From the cyclic voltammogram of **Cu^A** (**Figure 2.31**), the excited-state oxidation potential of **Cu^A** is estimated to be $E_{1/2}$ (Cu^{II/I*}) ~ -2.6 V (versus Fc/Fc⁺).

Figure 2.33. Steady-state emission of Cu_A in THF at room temperature ($\lambda_{\text{excitation}} = 370$ nm) and excitation scan of Cu_A in THF ($\lambda_{\text{probe}} = 480$ nm).

2.4.10.10. Reaction of Cu^A to form CuB; independent preparation of Cu^B

Photoinduced reduction of the electrophile by CuA: Quantification of the formation of the C–C coupled dimer of the electrophile. In a glovebox, a 4 mL vial equipped with a PTFE stir bar was sequentially charged with **Cu^A** (13 mg, 0.010 mmol), toluene (2 mL), and 2-chloro-2-phenylbutanenitrile (18 mg, 0.10 mmol). The vial was capped with a PTFE-lined cap and removed from the glovebox. The vial was cooled for 5 min in a low-form hemispherical Dewar flask filled with pre-cooled methanol (–78 °C). Two Kessil[®] PR160-440 nm lamps were installed 5 cm above the vials, and the vials were irradiated for 30 min, during which time the solution turned dark purple. The reaction mixture was warmed to room temperature, and 30% aqueous NH4OH (0.5 mL) and EtOAc (0.5 mL) were added. The mixture was stirred for 1 min, and then the aqueous layer was extracted with 2 mL of EtOAc. The organic layer was dried over $Na₂SO₄$ and passed through a 2 cm plug of silica gel loaded in a pipet (the $Na₂SO₄$ was washed with EtOAc (2) x 2 mL), and the washings were passed through the silica gel). The combined organic filtrate was concentrated in vacuo, and an internal standard (1,3,5-trimethoxybenzene, 17 mg, 0.10 mmol) was added. The residue was dissolved in CDCl₃ and analyzed by ¹H NMR spectroscopy. The yield of the dimer was 43% based on **Cu^A** (average of two runs).

For control reactions, an analogous procedure was followed, using a vial covered with aluminum foil to block the light. The reaction mixture remained colorless, and the electrophile remained intact according to 1 H NMR spectroscopy.

Stoichiometric reactions with irradiation (left two) and control reactions (right two).

Photoinduced reduction of the electrophile by Cu^A to form Cu^B (X-band EPR, 77

K). In a glovebox, a 4 mL vial equipped with a PTFE stir bar was sequentially charged with **Cu**_A (13 mg, 0.010 mmol), toluene (2 mL), and 2-chloro-2-phenylbutanenitrile (18 mg, 0.10 mmol). The resulting solution $(300 \mu L)$ was loaded into an X-band EPR tube. The tube was sealed with a rubber stopper and removed from the glovebox. A nitrogen-filled balloon was quickly attached, and the reaction mixture was cooled for 5 min in a low-form hemispherical Dewar flask filled with pre-cooled methanol (–78 °C). Two Kessil[®] PR160-440 nm lamps were installed 5 cm above the tube, and the tube was irradiated for 40 min. The solution was frozen at 77 K and analyzed by X-band EPR spectroscopy (**Figure 2.34**).

Figure 2.34. Normalized X-band EPR spectra of Cu_B (red trace) and a photoinduced stoichiometric reaction between **Cu^A** and 2-chloro-2-phenylbutanenitrile (black trace). Acquisition parameters: MW frequency = $9.372 - 9.374$ GHz; temperature = 77 K; MW power = 140 μ W; modulation amplitude = 0.4 mT; conversion time = 5.0 ms.

Independent Synthesis of DTBM-SEGPHOS-CuCl² (CuB).

In-situ preparation (GP-8). In a glovebox, a 4 mL vial equipped with a PTFE stir bar was sequentially charged with $CuCl₂ (1.6 mg, 0.012mmol)$, (R) -**P** (12 mg, 0.010) mmol), and toluene (1 mL). The reaction mixture was vigorously stirred for 30 min. The resulting dark-purple solution was filtered through a PTFE syringe filter (0.45 μm, InnoSepTM). The yield of Cu **B** was determined to be >95% by quantitative X- band EPR spectroscopy, using a solution of $CuSO_4\bullet 5H_2O$ in deionized water/glycerol (4:1, v/v) as an external standard. The dark-purple solution lost its color over 24 h at room temperature. Due to the instability of **CuB**, this solution was used immediately for further transformations. The identity of **Cu^B** was determined by its isolation (below).

Isolation*.* In a glovebox, a 20 mL vial equipped with a PTFE stir bar was sequentially charged with CuCl₂ (8.1 mg, 0.060 mmol), (R) -**P** (59 mg, 0.05 mmol), and 5 mL of toluene. The reaction mixture was vigorously stirred for 40 min. The resulting dark-purple solution was filtered through a PTFE syringe filter $(0.45 \mu m, \text{InnoSep}^{\text{TM}})$. The solution was cooled to –78 °C (glovebox cold well) for 5 min, and pre-chilled pentane (–78 °C, 12 mL) was layered onto the solution. The vial was capped and allowed to stand at -78 °C for 48 h. The resulting dark-purple solids were washed with pre-chilled pentane $(-78 \degree C, 2 \text{ mL})$ two times and placed under vacuum to remove residual solvent. The purified material was obtained as DTBMSEGPHOS-CuCl₂(toluene)₂(pentane), as determined by X-ray crystallography and elemental analysis (40.9 mg, 54% yield). The isolated compound dissolved in toluene showed an identical X-band EPR spectrum to the solution prepared in situ (**Figure 2.34**).

Elemental analysis. Calcd for C₈₈H₁₁₆O₈P₂Cl₂Cu: C, 70.55; H, 7.80; Cl, 4.73; Cu, 4.24; O, 8.54; P, 4.13. Found: C, 70.11; H, 7.52.

CCDC 2098022

Thermal ellipsoid plot of **Cu^B** at 50% probability level. For clarity, the hydrogen atoms are not shown. The asymmetric unit contains two molecules of toluene and one molecule of pentane (left), which are omitted on the right.

X-ray analysis. A brown needle-like specimen of $C_{93}H_{128}Cl_2CuO_8P_2$, approximate dimensions 0.100 mm x 0.100 mm x 0.250 mm, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured ($\lambda = 0.71073$ Å). The total exposure time was 12.22 h. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using an orthorhombic unit cell yielded a total of 61680 reflections to a maximum θ angle of 26.48° (0.80 Å resolution), of which 18034 were independent (average redundancy 3.420, completeness = 99.6%, R_{int} = 4.42%, R_{sig} = 5.08%) and 15750 (87.34%) were greater than $2\sigma(F^2)$. The final cell constants of <u>a</u> = 15.959(7) Å, <u>b</u> = 21.778(6) Å, <u>c</u> = 25.211(13) Å, volume $= 8762(6)$ Å³, are based upon the refinement of the XYZ-centroids of 9095 reflections above 20 $\sigma(I)$ with $4.519^{\circ} < 2\theta < 52.82^{\circ}$. Data were corrected for absorption effects using the Multi-Scan method (SADABS). The ratio of minimum to maximum apparent transmission was 0.880. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.9070 and 0.9610.

The structure was solved and refined using the Bruker SHELXTL Software Package, using the space group P 21 21 21, with $Z = 4$ for the formula unit, $C_{93}H_{128}Cl_2CuO_8P_2$. The final anisotropic full-matrix least-squares refinement on

 F^2 with 987 variables converged at R1 = 3.95%, for the observed data and wR2 = 8.71% for all data. The goodness-of-fit was 1.020. The largest peak in the final difference electron density synthesis was 0.568 e⁻/ \AA ³, and the largest hole was -0.390 e⁻/ \AA ³ with an RMS deviation of 0.056 $e^{\frac{1}{A^3}}$. On the basis of the final model, the calculated density was 1.190 g/cm^3 and F(000), 3372 e.

Sample and crystal data of CuB(toluene)2(pentane)

Data collection and structural refinement for CuB(toluene)2(pentane)

2.4.10.11. Monitoring the standard coupling reaction by EPR spectroscopy

Under a nitrogen atmosphere, a 4 mL vial equipped with a PTFE stir bar was sequentially charged with **Cu^A** (13 mg, 0.010 mmol), toluene (2 mL), *p*-toluidine (13 mg, 0.12 mmol), BTPP (38 μ L, 0.12 mmol), and 2-chloro-2-phenylbutanenitrile (18 mg, 0.10 mmol) and then stirred for 3 min. The resulting homogenous solution was transferred to EPR tubes (300 μL per tube). The tubes were sealed with a rubber stopper and removed from the glovebox, and a nitrogen-filled balloon was attached. The tubes were cooled for 5 min in a low-form hemispherical Dewar flask filled with pre-cooled methanol $(-78 \degree C)$. Two Kessil® PR160-440 nm lamps were set to have 50% intensity by adjusting the dial and placed 5 cm above the reactions. The reaction mixtures were irradiated for the given times and then immediately frozen at 77 K. The stoppers and the balloons were removed, and the samples were analyzed by X-band EPR spectroscopy (**Figure 2.35**). The concentrations of the paramagnet were determined using a solution of $CuSO_4\bullet 5H_2O$ in deionized water/glycerol (4:1, v/v) as an external standard. Selected samples were carefully warmed, and an aliquot was recovered with the aid of dichloromethane. The volatiles were removed, and the residue was re-dissolved in CDCl3. The reactions were analyzed via ${}^{1}H$ NMR spectroscopy to determine the conversion and the yield, assuming that the sum of the remaining electrophile, the C–N coupling product, and the electrophile dimers gives 100% of the initially loaded electrophile (see section **B** of the mechanistic section: the sum of the C–N coupling product and the electrophile dimers gave 99% of the initially loaded electrophile) (**Table 2.5**).

Note. The rate of these EPR-tube reactions was faster than that of the standard 0.10 mmol reaction (**GP-6)**. The intensity of the light for the EPR-tube reactions was accordingly decreased so that the reactions have a similar rate of conversion.

Figure 2.35.Time-course X-band EPR spectra of the catalytic reaction. Acquisition parameters: MW frequency = 9.372 -9.374 GHz; temperature = 77 K; MW power = 140 μ W; modulation amplitude = 0.4 mT; conversion time = 5.0 ms.

| Reaction time | $[Cu(II)]/[CuA]$ ₀ (%) | Conversion $(\%)$ | Product Yield (%) | Product ee $(\%)$ | |
|----------------------|-----------------------------------|--------------------|--------------------------|--------------------|--|
| 10 min | 10 | 32 | | | |
| 20 min | 12 | 53 | 40 | | |
| 30 min | | 82 | 66 | 90 | |

Table 2.5. Concentration of the paramagnet, conversion of the electrophile, and yield of the coupling product from the EPR study of the catalytic reaction.

2.4.10.12. In Situ preparation and decomposition of Cu^C

In-situ preparation (GP-9). Due to the temperature sensitivity of Cu_c, the synthesis of **Cu^C** was performed on an EPR-tube scale for immediate freeze-quenching. In a glovebox, a solution was prepared in a 4 mL vial of *p*-toluidine (13 mg, 0.12 mmol), BTPP (9.5 μL, 0.030 mmol or 38 μ L, 0.12 mmol), and 1 mL of toluene. The vial was capped with a PTFElined open-top cap. An \sim 10 mM solution of $\mathbf{C}\mathbf{u}_\mathbf{B}$ in toluene was separately prepared according to **GP-8**. An EPR tube was charged with a magnetic stir bar and 150 μL of the **Cu^B** solution. The tube was sealed with a rubber stopper and removed from the glovebox. A nitrogen-filled balloon was attached to the top of the tube, and the solution was cooled for 2 min in a low-form hemispherical Dewar flask filled with pre-cooled methanol (– 90 °C). The solution of *p*-toluidine/BTPP (150 μL) was carefully layered over the solution of **CuB**, and the tube was incubated for an additional 1 min so that both layers reached the bath temperature. Mixing of the two layers was performed by vigorously moving the internal magnetic stir bar up and down using an external magnet for 90 seconds, at which time the dark-purple color of the solution had changed to dark-green. The magnetic stir bar inside the EPR tube was held above the solution using the external magnet, and the solution was freeze-quenched at 77 K. The magnetic stir bar was removed, and the sample was analyzed by X-band EPR spectroscopy (**Figure 2.11**). The yield of Cu_c was determined to be \sim 60% (with 3 equiv of BTPP) or \sim 30% (with 12 equiv of BTPP) by quantitative X-band EPR spectroscopy, using a solution of $CuSO_4\bullet 5H_2O$ in deionized water/glycerol (4:1, v/v) as an external standard.

Decomposition. Following the same procedure, the dark-green solution after 90 seconds was further incubated at -78 °C for 30 min, at which point the solution became bright-yellow. The solution was subjected to X-band EPR spectroscopy at 77 K, following the above procedure. No significant amount of a paramagnet was detected. The sample was carefully warmed to room temperature, concentrated in vacuo, and analyzed by preparative TLC and ¹H NMR.

2.4.10.13. Synthesis of Cu^C from different CuX² and bases

A solution of "Cu_B" prepared in situ according to GP-8 from CuCl₂ and CuBr₂ (**Figure 2.36**) showed distinctly different EPR spectra, due to the different halide ligands. **GP-9** was followed with these solutions, to synthesize **CuC**. Analysis of the first and second derivatives (**Figures 2.37** and **2.38**) of the X-band EPR spectra indicates that an identical species was formed from the different "**CuB**" intermediates.

Figure 2.36. X-band EPR spectra of "Cu_B" (PCuX₂) prepared from CuCl₂ (red trace) and CuBr₂ (blue trace). Acquisition parameters: MW frequency = 9.37 GHz; temperature = 77 K; MW power = 140 μ W; modulation amplitude = 0.4 mT; conversion time = 5.0 ms.

Figure 2.37. X-band EPR spectra of **Cu**c prepared from CuCl₂ (red trace) and CuBr₂ (blue trace). Acquisition parameters: MW frequency = 9.37 GHz; temperature = 77 K; MW power = 140 μ W; modulation amplitude = 0.4 mT; conversion time = 5.0 ms.

Figure 2.38. 2nd Derivative X-band EPR spectra of **Cu**c prepared from CuCl₂ (red trace) and CuBr² (blue trace). Overlapping spectra from two independent sets of syntheses are provided. Acquisition parameters: MW frequency = 9.37 GHz; temperature = 77 K; MW power = 140 μ W; modulation amplitude = 0.1 mT; conversion time = 5.0 ms.

A solution of **Cu^B** prepared in situ according to **GP-8** was followed with **GP-9**, using BTMG instead of BTPP to synthesize Cu_C. The mixing time of the two layers was shortened to 20 seconds, as the consumption of Cu **B** and the decomposition of Cu ^C were more rapid. Analysis of the first and second derivatives (**Figure 2.39**) of the X-band EPR spectra indicated that an identical species was formed from the two reactions.

Figure 2.39. 1st and 2nd derivative X-band EPR spectra of Cu_c prepared from BTPP (red trace) and BTMG (blue trace). Acquisition parameters: MW frequency $= 9.37$ GHz; temperature = 77 K; MW power = 140 μ W; modulation amplitude = 0.1-0.4 mT; conversion time $= 5.0$ ms.

2.4.10.14. Isotopologues of Cu^C and Q-band pulse EPR experiments

Preparation of *d***₂-labeled** *p***-toluidine.** *p*-Toluidine-*ND*₂ was synthesized by hydrogendeuterium exchange in MeOD (three cycles). An oven-dried 20 mL vial was charged with *p-*toluidine (500 mg, 4.67 mmol) and a magnetic stir bar. MeOD (5 mL) was added, and the vial was purged with nitrogen. The solution was stirred for 12 h, and then the solvents were evaporated. This process was repeated three times, providing a white solid (503 mg, 99% yield). The deuterium incorporation was determined to be 98% via ${}^{1}H$ NMR spectroscopic analysis. The product was further purified by sublimation before use.

¹H NMR (400 MHz, C_6D_6) δ 6.94 – 6.85 (m, 2H), 6.40 – 6.27 (m, 2H), 2.15 (s, 3H). ²H NMR (61 MHz, C₆D₆) δ 2.68.

¹³C NMR (101 MHz, C₆D₆) δ 144.7, 130.0, 127.1, 115.2, 20.6.

HRMS (ESI+) m/z [M(D₂)+H]⁺ calcd for C₇H₈D₂N⁺: 110.0933, found: 110.0880.

Preparation of ¹⁵N-labeled *p***-toluidine.** 4-Methylbenzoyl chloride (1.35 g, 8.74 mmol), a magnetic stir bar, dichloromethane (80 mL) , and 15 NH₄Cl $(500 \text{ mg}, 9.2 \text{ mmol})$ were added sequentially to an oven-dried, nitrogen-filled 250 mL round-bottom flask. The flask was sealed with a rubber stopper, and a nitrogen-filled balloon was attached. The reaction mixture was cooled to 0° C, and then triethylamine (3.65 mL, 26.2 mmol) was added dropwise. The reaction mixture was warmed to room temperature and stirred for 2 h. The reaction mixture was then acidified with 10 mL of 0.2 N HCl, and the organic layer was separated. The aqueous layer was extracted with 40 mL of dichloromethane, and the combined organic layer was dried over $Na₂SO₄$, filtered, and concentrated in vacuo. 4-Methylbenzamide-¹⁵*N* was purified by flash column chromatography on silica gel $(hexanes/EtOAC = 3/1 \rightarrow 1/1)$. White solid (993 mg, 84% yield).

To a 50 mL round-bottom flask was added a magnetic stir bar, KOH (750 mg, 13.3 mmol), and deionized water (5 mL). The solution was cooled to 0 $^{\circ}$ C, and Br₂ (3.23 mmol, 170 μL) was added dropwise. 4-Methylbenzamide-¹⁵*N* was slowly added to the resulting solution, and the reaction mixture was stirred for 2 h. The flask was fitted with a reflux condenser, heated to 90 °C, and stirred for 2 h. The solution was then cooled to room temperature, diluted with 5 mL of water, and extracted with $Et₂O$ (25 mL). The solvent was removed in vacuo, and the product was purified by column chromatography $(hexanes/EtOAc = 10/1)$. Pale-orange solid (129 mg, 41%). The product was further purified by sublimation to obtain a white crystalline solid.

¹H NMR (400MHz, C_6D_6) δ 6.90 (dd, *J* = 7.8, 0.7 Hz, 1H), 6.37 – 6.30 (m, 2H), 2.74 (d, *J =* 77.4 Hz, 2H), 2.15 (s, 3H).

¹³C NMR (101 MHz, C₆D₆) δ 144.8 (d, *J* = 11.0 Hz), 130.0 (d, *J* = 1.2 Hz), 127.1, 115.3 (d, $J = 2.8$ Hz), 20.6.

¹⁵N NMR (41MHz, C₆D₆) δ 52.9 (t, *J* = 77.7 Hz).

HRMS (ESI+) m/z [M+H]⁺ calcd for C₇H₁₀¹⁵N⁺: 109.0778, found: 109.0785.

Preparation of Q-band EPR samples of isotopologues of Cu_c. In a glovebox under a nitrogen atmosphere, a 4 mL vial equipped with a PTFE stir bar was sequentially charged with **Cu**_A (13 mg, 0.010 mmol), toluene- d_8 (2 mL), *p*-toluidine (13 mg, 0.12 mmol), BTPP (38 μL, 0.12 mmol), 1,3,5-trimethoxybenzene (17 mg, 0.10 mmol), and 2-chloro-2 phenylbutanenitrile (18 mg, 0.10 mmol), and the resulting mixture was stirred for 3 min. This mixture was transferred using a microsyringe (30 μL per tube) to Q-band EPR tubes. The top of each tube was sealed with vacuum grease. Meanwhile, \sim 20 mL of methanol was charged into a 40 mL vial, and the vial was placed in a low-form hemispherical Dewar flask filled with pre-cooled methanol so that the temperature of the methanol inside the vial was maintained at –78 °C. The sealed EPR tubes were placed inside the 40 mL vial. Vacuum grease was maintained on the top of the tubes while the tubes were cooled, to ensure sealing of the tubes. Two Kessil[®] PR160-440 nm lamps were set at 50% intensity and placed 5 cm above the reactions. The reaction mixtures were irradiated for the given

times and then immediately frozen at 77 K. X-band EPR spectra were taken to determine the sample that contained the strongest and cleanest EPR signal. The sample that was irradiated for 5 min (**Figures 2.40** and **2.41**) was chosen for pulse EPR analysis (**Figures 2.42** – **2.55**).

Figure 2.40. X-band EPR spectra of catalytic reactions charged in Q-band EPR tubes and irradiated for the given times. Acquisition parameters: MW frequency = 9.39 GHz; temperature = 77 K; MW power = 2180 μ W; modulation amplitude = 0.4 mT; conversion time = 5.0 ms. The baseline was not corrected.

Figure 2.41. X-band EPR spectra of Cu_C generated in an X-band EPR tube from CuCl₂ (red trace, section **L** of this mechanism section) and from the catalytic reaction charged in a Q-band EPR tube after 5 min of light irradiation (blue trace, **Figure 2.40**). Acquisition parameters: MW frequency = $9.36 - 9.39$ GHz; temperature = 77 K.

EPR Simulation of Cu_C (Figure 2.12, 2.13, 2.14 and Table 2.6).

Simulation parameters of the *g*-values and the hyperfine tensors of ${}^{31}P$ and ${}^{63/65}Cu$ nuclei were obtained by a global fit of the Q-band (~34 GHz) ENDOR (which primarily provided constraints on the ${}^{31}P$ couplings) and X- and O-band field-swept EPR spectra (the higher fields associated with the Q-band spectra provided more robust constraints on simulated *g*-values) (**Figures 2.42** and **2.43**).

A single set of elongated correlation ridges in the $(-,+)$ quadrant in the field dependent Q-band HYSCORE of ¹⁵N labeled **Cu^C** (**Figures 2.44** – **2.47**) were simulated by a single class of highly anisotropic ¹⁵N hyperfine tensor, with $A(^{15}N) = \pm [6, 90, 6]$ MHz $(a_{iso}(^{15}N) =$ \pm 24.3 MHz), with a Euler rotation of the hyperfine tensor relative to the g tensor of (a, β, γ) $= (-11, 84, 0)$ °.

The ¹⁴N signals from the Q-band HYSCORE of natural abundance **Cu**_c (**Figures 2.44**, **2.48** – **2.50**) also fall into the strong coupling regime but are additionally complicated by

the influence of $14N$ nuclear quadrupole interaction which further splits the nuclear spin sublevels due to the electric interaction of the ¹⁴N nucleus ($I = 1$) with the inhomogeneous electric field induced by electron density in p orbitals at the nucleus. These spectra were simulated by scaling the $15N$ hyperfine tensor determined from $15N$ HYSCORE by the proportion of ¹⁵N/¹⁴N gyromagnetic ratios ($|\gamma^{15}N|/|\gamma^{14}N| = 1.403$), with $A(^{14}N) = \pm [4.3, 1.403]$ 64.2, 4.3] MHz. ¹⁴N nuclear quadrupole parameters were then varied independently, with the best match between data and simulations achieved with ¹⁴N quadrupole parameters of $e^2 qQ/h = 1.7$ MHz and $\eta = 1$. These parameters provide a point-specific measurement of the magnitude of the electric field gradient (EFG, parametrized by e^2qQ/h) and its symmetry, with the asymmetry parameter η ranging from $\eta = 0$ for pure axial symmetry to $\eta = 1$ for fully rhombic symmetry.

Intense ²H cross-peaks in the (+,+) quadrant of Q-band HYSCORE of ²H-labeled **Cu^C (Figures 2.44, 2.51 – 2.53)** were simulated by a single class of relatively anisotropic ²H hyperfine tensor with $A(^{2}H) = \pm [8.9, 4.3, 1.5]$ MHz $(a_{iso}(^{2}H) = \pm 4.9$ MHz).

The presence of multiple equivalent -NHAr ligands could be ruled out by simulations that showed that the equivalent $14/15$ N nuclei would generate combination peaks that are completely absent in the observed spectra (**Figure 2.54**).

Overall, the simulated EPR spectra from the parameters described in **Table 2.6** match with the observed CW-EPR spectra of **Cu^C** and its isotopologues (**Figure 2.55**).

| g-value | 2.012 | 2.014 | 2.023 | | | | |
|-----------------------|---------|---------|---------|-----------------|---------------------|------------|---------------|
| Nucleus | $ A_1 $ | $ A_2 $ | $ A_3 $ | $ a_{\rm iso} $ | T | e^2qQ/h | η |
| ${}^{1}H$ (amide) | 58 | 28 | 10 | 32 | $[26, -4, -22]$ | | |
| ${}^{2}H$ (amide) | 8.9 | 4.3 | 1.5 | 4.9 | $[4.0, -0.6, -3.4]$ | ${}_{0.4}$ | ≈ 0 |
| 15 N(amide) | 6 | 90 | 6 | 34 | $[-28, 56, -28]$ | | |
| ^{14}N (amide) | 4.3 | 64.2 | 4.3 | 24.3 | $[-20, 40, -20]$ | 1.7 | ≈ 1.0 |
| $\overline{63/65}$ Cu | 18 | 26 | 146 | 63 | | | |
| $^{31}P_a$ | 130 | 100 | 100 | 110 | | | |
| $^{31}P_b$ | 110 | 100 | 130 | 113 | | | |

Table 2.6. EPR simulation parameters for **Cu**_C. All hyperfine and nuclear quadrupole coupling values are reported in MHz. α

 α A(¹H_(amide)) is rotated relative to g-tensor by $(\alpha, \beta, \gamma) = (-10, 54, 50)$ ^o, while A(^{15/14}N_(amide)) and the ¹⁴N_(amide) nuclear quadrupole tensors are both rotated relative to the g-tensor by (α , $β, γ$ = (-11, 84, 0)^o. All other hyperfine tensors are collinear with g-tensor. ^{1/2}H and ^{14/15}N parameters primarily determined from simulations of Q-band HYSCORE, while ${}^{31}P_{a,b}$ primarily determined from fixing all other parameters and optimizing simulations of Qband ENDOR and field-swept EPR spectra.

Cu_c intermediate Pseudomodulated Q-band ESE-EPR

Figure 2.42. Q-band pseudomodulated ESE-EPR spectra of Cu_c in toluene formed using isotopologues of *p*-toluidine (black traces) with simulations using parameters in **Table 2.6** overlaid (red traces). Acquisition parameters: MW frequency = 34.005 GHz; temperature = 20 K; MW power = 140 μ W; modulation amplitude = 0.1 mT; conversion time = 5.3 ms. Acquisition parameters: MW frequency = 34.011 GHz (Nat. Abund.), 34.072 GHz (¹⁵N) labeled), 34.048 GHz (²H labeled); temperature = 30 K; π pulse length = 160 ns; τ = 300 ns; shot rep time (srt) = 1 ms.

Figure 2.43. Field-dependent Q-band Davies ENDOR of Cu_c in toluene formed using ²H p -toluidine (black) with simulations of total ENDOR contributions from the two $31P$ nuclei simulated in red, and individual contributions from ${}^{31}P_1$ and ${}^{31}P_2$ in green and blue, respectively, using values in **Table 2.6**. Intense ENDOR signals from ligand ¹H nuclei in the range from 40 to 60 MHz have been truncated for ease of evaluation of the ^{31}P signals. Experimental conditions: microwave frequency = 34.050 GHz; temperature = 20 K; MW π pulse length = 80 ns; interpulse delay $\tau = 200$ ns; π_{RF} pulse length = 15 µs; T_{RF} delay = 2 μs; shot repetition time (srt) = 5 ms.

Figure 2.44. Q-band HYSCORE of the **Cu**c intermediate in toluene formed using isotopologues of *p*-toluidine with natural abundance (top), ${}^{2}H$ (middle), and ${}^{15}N$ (bottom) measured at magnetic fields corresponding to $g = 2.032$, $g = 2.015$, and $g = 2.004$ from left to right. Spectra of ²H isotopologues have been plotted to 2 orders of magnitude further from the intensity maximum compared to Figures in which comparison to the ${}^{2}H$ simulations are made in order to show that the $14N$ signals in the $(-,+)$ quadrant observed the natural abundance HYSCORE are still present. Experimental conditions: MW frequency = 34.005 GHz (Nat. Abund), 34.072 (²H and ¹⁵N); temperature = 30 K; τ = 128 ns, $t_1 = t_2 = 100$ ns; $\Delta t_1 = \Delta t_2 = 12$ ns; shot repetition time (srt) = 1.5 ms.

Figure 2.45. Q-band HYSCORE of the **Cu**_c intermediate in toluene formed using ¹⁵N labeled *p*-toluidine (left) measured at 1198.2 mT ($g = 2.032$) with overlay of ¹⁵N simulation contours (right, red) with experimental contours (right, gray). Experimental conditions: MW frequency = 34.072 GHz; temperature = 30 K; τ = 128 ns, $t_1 = t_2 = 100$ ns; $\Delta t_1 = \Delta t_2$ = 12 ns; shot repetition time (srt) = 1.5 ms.

Figure 2.46. Q-band HYSCORE of the **Cu**_c intermediate in toluene formed using ¹⁵N labeled *p*-toluidine (left) measured at 1208.1 mT ($g = 2.015$) with overlay of ¹⁵N simulation contours (right, red) with experimental contours (right, gray). Experimental conditions: MW frequency = 34.072 GHz; temperature = 30 K; τ = 128 ns, $t_1 = t_2 = 100$ ns; $\Delta t_1 = \Delta t_2$ = 12 ns; shot repetition time (srt) = 1.5 ms.

Figure 2.47. Q-band HYSCORE of the Cu_C intermediate in toluene formed using ¹⁵N labeled *p*-toluidine (left) measured at 1214.7 mT ($g = 2.004$) with overlay of ¹⁵N simulation contours (right, red) with experimental contours (right, gray). Experimental conditions: MW frequency = 34.072 GHz; temperature = 30 K; τ = 128 ns, $t_1 = t_2 = 100$ ns; $\Delta t_1 = \Delta t_2$ = 12 ns; shot repetition time (srt) = 1.5 ms.

Figure 2.48. Q-band HYSCORE of the **Cu**c intermediate in toluene formed using natural abundance *p*-toluidine (left) measured at 1196 mT ($g = 2.032$) with overlay of ¹⁴N simulation contours (right, red) with experimental contours (right, gray). Experimental conditions: MW frequency = 34.005 GHz; temperature = 30 K; τ = 128 ns, $t_1 = t_2 = 100$ ns; $\Delta t_1 = \Delta t_2 = 12$ ns; shot repetition time (srt) = 1.5 ms.

Figure 2.49. Q-band HYSCORE of the **Cu**c intermediate in toluene formed using natural abundance *p*-toluidine (left) measured at 1206 mT ($g = 2.015$) with overlay of ¹⁴N simulation contours (right, red) with experimental contours (right, gray). Experimental conditions: MW frequency = 34.005 GHz; temperature = 30 K; τ = 128 ns, $t_1 = t_2 = 100$ ns; $\Delta t_1 = \Delta t_2 = 12$ ns; shot repetition time (srt) = 1.5 ms.

Figure 2.50. Q-band HYSCORE of the Cu_c intermediate in toluene formed using natural abundance *p*-toluidine (left) measured at 1212.5 mT ($g = 2.004$) with overlay of ¹⁴N simulation contours (right, red) with experimental contours (right, gray). Experimental conditions: MW frequency = 34.005 GHz; temperature = 30 K; τ = 128 ns, $t_1 = t_2 = 100$ ns; $\Delta t_1 = \Delta t_2 = 12$ ns; shot repetition time (srt) = 1.5 ms.

Figure 2.51. Q-band HYSCORE of the **Cu**_C intermediate in toluene formed using ²H labeled *p*-toluidine (left) measured at 1198.2 mT ($g = 2.032$) with overlay of ²H simulation contours (right, red) with experimental contours (right, gray). Experimental conditions: MW frequency = 34.072 GHz; temperature = 30 K; τ = 128 ns, $t_1 = t_2 = 100$ ns; $\Delta t_1 = \Delta t_2$ = 12 ns; shot repetition time (srt) = 1.5 ms.

Figure 2.52. Q-band HYSCORE of the Cu_c intermediate in toluene formed using ²H labeled *p*-toluidine (left) measured at 1208.1 mT ($g = 2.015$) with overlay of ²H simulation contours (right, red) with experimental contours (right, gray). Experimental conditions: MW frequency = 34.072 GHz; temperature = 30 K; τ = 128 ns, $t_1 = t_2 = 100$ ns; $\Delta t_1 = \Delta t_2$ = 12 ns; shot repetition time (srt) = 1.5 ms.

Figure 2.53. Q-band HYSCORE of the Cu_c intermediate in toluene formed using ²H labeled *p*-toluidine (top) measured at 1214.7 mT ($g = 2.004$) with overlay of ²H simulation contours (bottom, red) with experimental contours (bottom, gray). Experimental conditions: MW frequency = 34.072 GHz; temperature = 30 K; τ = 128 ns, $t_1 = t_2 = 100$ ns; $\Delta t_1 = \Delta t_2 = 12$ ns; shot repetition time (srt) = 1.5 ms.

Figure 2.54. Q-band HYSCORE of the Cu_C intermediate in toluene formed using ¹⁵N labeled *p*-toluidine (top) measured at 1208.1 mT ($g = 2.015$) with comparison of ¹⁵N simulations using a single nitrogen (left) and two equivalent nitrogen couplings (right), the latter of which produces combination frequency cross-peaks at (-18, 4) and (-4, 18) MHz that are completely absent in the experimental data. Experimental conditions: MW frequency = 34.072 GHz; tem-perature = 30 K; τ = 128 ns, $t_1 = t_2 = 100$ ns; $\Delta t_1 = \Delta t_2 = 12$ ns; shot repetition time (srt) = 1.5 ms.

Figure 2.55. X-band CW-EPR spectra (top panel) and 2nd derivative (bottom panel) of Cu_C in toluene and isotopologues of *p*-toluidine (black traces) with simulations overlaid (red traces); acquisition parameters: MW frequency = $9.372-9.374$ GHz, MW power = 140 μ W, modulation amplitude = 0.1 mT, conversion time = 5.3 ms, and temperature = 77 K.

2.4.10.15. Reaction of Cuc with an organic radical

$$
H_2N \longrightarrow \longrightarrow \text{Me} \quad \xrightarrow{Ar^1_3CBr} Ar^1_{3}C-NHAr
$$
\n
$$
Ar^1 = p-t \cdot BUC_6H_4 \quad 98\% \text{ yield}
$$

Synthesis of authentic coupling product. In a glovebox, an oven-dried 4 mL vial was charged with a magnetic stir bar, tris(4-(*tert*-butyl)phenyl)methyl bromide (98 mg, 0.20 mmol), and toluene (2 mL). *p*-Toluidine (43 mg, 0.40 mmol) was added, and the reaction mixture was stirred for 24 h, at which time a yellow precipitate was observed. The precipitate was removed via filtration with a PTFE syringe filter, and the solution was concentrated in vacuo. Pentane (2 mL) was added to the residue, and the resulting solution was filtered through a PTFE syringe filter. The solution was concentrated in vacuo to afford the desired product. White solid, 101 mg (0.20 mmol, 98% yield).

¹H NMR (500 MHz, CDCl₃) δ 7.28 – 7.17 (m, 12H), 6.71 (d, $J = 8.2$ Hz, 2H), 6.23 (d, *J =* 8.3 Hz, 2H), 4.88 (s, 1H), 2.13 (s, 3H), 1.29 (s, 27H).

¹³C NMR (126 MHz, CDCl₃) δ 149.3, 144.4, 142.9, 129.0, 128.8, 125.6, 124.7, 115.9, 70.4, 34.5, 31.5, 20.5.

HRMS (ESI+) m/z [M – (NHp-tol)]⁺ calcd for $C_{31}H_{39}$ ⁺: 411.3046, found: 411.3064.

Reaction of Cu_C with an Organic Radical. A solution of tris-(4-*tert*-butyl phenyl)methyl radical (15 mM in toluene) was prepared according to the literature,^{[55](#page-203-0)} from tris(4-(tert-butyl)phenyl)methyl bromide (46.4 mg, 0.094 mmol), activated copper powder (90 mg, 1.41 mmol), and toluene (6.3 mL). A solution of *p*-toluidine/BTPP was prepared in a 4 mL vial with *p*-toluidine (13 mg, 0.12 mmol), BTPP (10 μL, 0.030 mmol or 38 μL, 0.12 mmol) and 1.0 mL of toluene.

In a glovebox, an NMR tube was charged with a magnetic stir bar and a solution of **Cu^B** in toluene (10 mM, 200 μL), prepared in situ according to **GP-8**. The tube was sealed with a rubber stopper and removed from the glovebox. A nitrogen-filled balloon was

attached to the top of the tube, and the solution was cooled for 2 min in a low-form hemispherical Dewar flask filled with pre-cooled methanol (–90 °C). The solution of *p*toluidine/BTPP (200 μL) was carefully layered over the solution of C **u***B*, and the tube was incubated for an additional 1 min so that both layers reached the bath temperature. Mixing of the two layers was performed by moving the internal magnetic stir bar up and down using an external magnet for 90 seconds (at which time the dark-purple color of the solution changed to dark-green), at which point the solution of tris-(4-*tert*-butyl phenyl)methyl radical (200 μL) was carefully layered above the solution of **Cu**_C. Mixing of the two layers was performed by moving an external magnet for 5 min. The tube was quickly moved to another hemispherical Dewar flask filled with a dry ice/acetone bath (– 78 °C) and incubated for 30 min. The dark color of the solution turned bright yellow. The solution was warmed and transferred to a 20 mL vial using 3 mL of dichloromethane. The solvent was then concentrated, and the residue was dissolved in a solution of internal standard $(1,3,5$ -trimethoxybenzene in CDCl₃) and analyzed via ¹H NMR spectroscopy (**Table 2.7**). Control experiments with decomposed **Cu**_c were performed by warming the dark-green solution of **Cu**_c to room temperature (at which point it turned yellow) and cooling it back to –90 °C before layering the solution of tris-(4-*tert*-butyl phenyl)methyl radical.

| Entry | Variations | Yield of the C–N coupling |
|-------|-----------------------------------|---------------------------|
| 1a | $\overline{}$ | 41% |
| | Decomposed Cuc | 7% |
| 3 | Cu_B was omitted | 9% |

Table 2.7. Stoichiometric reaction between **Cu^C** and tris-(4-*tert*-butyl phenyl)methyl radical, including control experiments.

a. The yields are an average of two runs.

2.4.11. DFT calculations

General information. All geometry optimizations were performed using ORCA 4.2.1 with the BP86 functional, the def2-TZVP basis set for all atoms, dispersion correction with Becke-Johnson damping (D3BJ), and the RIJCOSX approximation to speed up optimizations. A solvation model (CPCM (toluene)) was applied. Structures, molecular orbitals, and spin densities were visualized using Chemcraft 1.8 software.

MO visualization of the ground and the excited triplet state of CuA. Cartesian coordinates from the crystal structure of **Cu^A** (**CCDC 1579920**) was used as the input for the geometry optimization. Molecular orbitals of the ground and excited triplet structures of **Cu^A** propose ³MLCT from copper 3*d* orbital to the aryl rings of the SEGPHOS backbone (**Figure 2.56**).

Figure 2.56. Calculated plots of HOMO and LUMO of the ground state of **Cu^A** (top) and corresponding SOMOs of the triplet excited state of **CuA*** (bottom). Hydrogen atoms were omitted for clarity. Contour value $= 0.05$.

Geometry optimization of Cuc. Given the C₂-symmetric structure of the DTBM-SEGPHOS ligand, the structures shown below were chosen for calculation (**Figure 2.57**). Due to limited resources, a frequency calculation was only performed for Cu_{C1}⁺ (the proposed structure of Cu_C from the EPR analysis), to ensure that the structure is a local minimum. Although Cu_{C2A} has a single point energy that is 2.11 kcal/mol lower than **CuC2B**, and **CuC3A** has a single point energy that is 2.5 kcal/mol lower than **CuC3B**, all structures were considered for the DFT-EPR calculations.

Figure 2.57. Candidate structures considered for **CuC**.

DFT-EPR calculations. Calculations of EPR parameters were performed using ORCA 4.2.1 with the TPSS functional. For basis sets: core-property basis set CP(PPP) for Cu atom; def2-TZVP for -NH_p-Tol, -N(p-Tol)₂, phosphorus, chlorine, and bromine. For the remaining ligand fragments, essentially the same EPR parameters were predicted when def2-SVP and def2-TZVP were both tested for **CuC1**⁺ . Therefore, def2-TZVP was used for **CuC2,** and def2-SVP was used for **CuC3** (due to limited resources), assuming that the basis set of the remaining ligand fragments does not have a significant effect on the predicted EPR parameters. An expanded grid was used (IntAccX 4.34,4.34,4.67 and GridX 2,2,2) to improve the accuracy of the calculations; dispersion correction with Becke-Johnson damping (D3BJ), and the RIJCOSX approximation to speed up optimizations. A solvation model (CPCM(toluene)) was used. The choice of the TPSS functional was based on the best reproduction of the reported EPR parameters of the formal bisphosphine Cu(II) amido complex[40](#page-42-0) (**Figure 2.58** and **Table 2.8**) among other basis sets tested (BP86, TPSS, OLYP, revTPSS, BHandHLYP, B3LYP, B3PW91, PBE0, TPSSH).

Figure 2.58. Structure of the reference compound (left); reported EPR spectrum (right, black trace) and EasySpin simulation obtained from DFT-predicted EPR parameters (right, red trace). Acquisition parameters: MW frequency = 9.38 GHz; temperature = 77 K; modulation amplitude $= 0.2$ mT.

| Reported | $g_{\rm x}$ | g_{y} | $g_{\rm z}$ | giso | Calculated | $g_{\rm x}$ | g_{y} | $g_{\rm z}$ | giso |
|-----------------|-------------|-----------|---------------|---------------|------------|-------------|---------|-------------|------------------|
| g-value | 2.008 | 2.008 | 2.030 | 2.015 | g-value | 2.013 | 2.014 | 2.031 | 2.019 |
| | $ A_x $ | $ A_{y} $ | $ {\rm A_z} $ | $a_{\rm iso}$ | | A_x | A_v | A_{Z} | a _{iso} |
| Cu | 34 | 34 | 170 | 79.3 | Cu | -27.1 | 7.85 | -215 | 78.3 |
| P1 | 148 | 148 | 173 | 156.3 | P1 | 230 | 141 | 139 | 170 |
| P2 | 148 | 148 | 173 | 156.3 | P2 | 155 | 254 | 153 | 187 |
| N | 24 | 100 | 24 | 49 | N | -6.8 | 37 | -7.1 | |

Table 2.8. Reported and calculated EPR parameters of the reference compound. A in MHz.

Predicted EPR parameters of the optimized structures (**Figures 2.59** – **2.64**; A in MHz).

Figure 2.59. Predicted EPR parameters of Cu ⁻⁺ compared to the simulation (left) and overlay of the predicted EPR spectrum (right, red trace) and the observed spectrum (right, black trace). Simulation line width $= 3$.

Figure 2.60. Predicted EPR parameters of Cu_{C2A} compared to the simulation (left) and overlay of the predicted EPR spectrum (right, red trace) and the observed spectrum (right, black trace). Simulation line width $= 2.5$.

Figure 2.61. Predicted EPR parameters of Cu_{C2A}-Br compared to the simulation (left) and overlay of the predicted EPR spectrum (right, red trace) and the observed spectrum (right, black trace). Simulation line width $= 2.5$.

Figure 2.62. Predicted EPR parameters of Cuc_{2B} compared to the simulation (left) and overlay of the predicted EPR spectrum (right, red trace) and the observed spectrum (right, black trace). Simulation line width $= 3$.

Figure 2.63. Predicted EPR parameters of Cu_{C3A} compared to the simulation (left) and overlay of the predicted EPR spectrum (right, red trace) and the observed spectrum (right, black trace). Simulation line width $= 2$.

Figure 2.64. Predicted EPR parameters of Cuc_{3B} compared to the simulation (left) and overlay of the predicted EPR spectrum (right, red trace) and the observed spectrum (right, black trace). Simulation line width $= 2$.

Calculated spin densities of Cu_c. Spin densities of the proposed structure of Cu_c (three coordinate $[PCu(NHAr)]^+$; Cuc₁₊) were calculated with five different functionals with the def2-TZVP basis set for all atoms, dispersion correction with Becke-Johnson damping (D3BJ), the RIJCOSX approximation, and the solvation model (CPCM (toluene)). All functionals tested support significant spin density on the -NHAr group (**Table 2.9**).

| Functional | BP86 | B3LYP | B3PW91 | TPSS | PBE ₀ |
|---------------------|--------------|--------------|---------------|--------------|-----------------------|
| Spin density | | | | | |
| | | | | | |
| Cu | $0.15 e^{-}$ | $0.13 e^{-}$ | $0.13 e^{-}$ | $0.15 e^{-}$ | $0.12 e^{-}$ |
| P ₁ | $0.08 e^{-}$ | $0.06 e^{-}$ | $0.07 e^{-}$ | $0.08 e^{-}$ | $0.07 e^{-}$ |
| P ₂ | $0.08 e^{-}$ | $0.06 e^{-}$ | $0.07 e^{-}$ | $0.08 e^{-}$ | $0.06 e^{-}$ |
| NH | $0.33 e^-$ | $0.40 e^{-}$ | $0.41 e^{-}$ | $0.35 e^-$ | $0.43 e^{-}$ |
| Ar | $0.32 e^{-}$ | $0.32 e^{-}$ | $0.31 e^-$ | $0.31 e^-$ | 0.31 e ⁻ |

Table 2.9. Calculated spin densities of Cu_C with five different functionals.

Cartesian coordinates of the optimized structures.

Cu^A (ground)

 $Cu_A* (triplet)$

CuC2A

CuC2A – Br

CuC2B

Cu_{C3A}

 $CuC3B$

2.5. Notes and references.

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C h a p t e r 3

SYNTHESIS OF TERTIARY ALKYL AMINES VIA PHOTOINDUCED, COPPER-CATALYZED NUCLEOPHILIC SUBSTITUTION OF UNACTIVATED ALKYL HALIDES BY SECONDARY ALKYL AMINES

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