Supplementary Information for Chapter 5

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D.1 General procedures

General Considerations: All manipulations of inorganic complexes were carried out using standard Schlenk or glovebox techniques under an N2 or Ar atmosphere. Unless otherwise noted, solvents were deoxygenated and dried by thoroughly sparging with N₂ gas followed by passage through an activated alumina column in a solvent purification system (SG Water, USA LLC). For electrochemical measurements under an Ar atmosphere, solvents were further degassed and then stored under Ar. All solvents were stored over activated 3 or 4 Å molecular sieves prior to use. Anhydrous ammonia gas was dried by passage through a calcium oxide drying tube. All reagents were purchased from commercial vendors and used without further purification unless otherwise stated. Tris(2-pyridylmethyl)amine (TPA),¹ tris(2-pyridylmethylamine) iron(II) triflate bis-acetonitrile ([(TPA)Fe(MeCN)₂]OTf₂),² 6- $(BPM)^3$ (1,1-di(pyridin-2-yl)ethyl)-2,2'-bipyridine 6-(1,1-di(pyridin-2-yl)ethyl)-2,2'bipyridine iron(II) triflate bis-acetonitrile $([(BPM)Fe(MeCN)_2]OTf_2),^4$ bis(4-(dimethylamino)pyridin-2-yl)methanone.⁵ tris(4-(dimethylamino)-2-picolyl)amine (TPA^{NMe2}),⁶ tris(4-methoxy-2-picolyl)amine (TPA^{OMe}),⁶ 4-(trifluoromethyl)pyridine-2carboxaldehyde,⁷ 4-(trifluoromethyl)-2-ethylpyridine,⁸ bis(4-(dimethylamino)pyridin-2vl)methanone,⁹ and methylenetriphenylphosphorane¹⁰ were synthesized according to literature procedures.

Electrochemistry: Voltammetry experiments were carried out with a Biologic VSP-300 or CH Instruments 600B potentiostat using a one-compartment three-electrode cell, and coulometry experiments were carried out with a Biologic VSP-300 potentiostat using a one-compartment three-electrode cell with a septum capped 14/20 joint for headspace analysis.

For voltammetry, a boron-doped diamond (BDD) working electrode (3 mm diameter), a Pt wire counter electrode, and an Ag/AgOTf reference electrode (5 mM AgOTf and 0.1 M TBAPF₆ in MeCN) were employed. For controlled potential coulometry (CPC), the same reference electrode was used, but a BDD plate (geometric area: 4 cm²) and a Pt mesh were used as working and counter electrodes, respectively. All redox potentials in the present work are reported versus the Fc/Fc⁺ couple, measured before each experiment to be approximately +0.12 V versus the Ag/AgOTf reference electrode.

Cyclic voltammograms (CVs) were collected at 100 mV·s⁻¹ unless otherwise specified. $E_{1/2}$ values for the reversible waves were obtained from the half potential between the oxidative and reductive peaks. CV measurements were performed applying IR compensation, compensating 85% of the resistance measured at one high frequency value (100 kHz).

Gas Chromatography: Gas chromatography was performed in the Caltech Environmental Analysis Center using HP 5890 Series II instruments. Gas quantification was performed with a molecular sieve column attached to a thermal conductivity detector using argon as the carrier gas. Standard curves were generated by direct injection of hydrogen or nitrogen gas. Quantification of background nitrogen was determined using the background oxygen signal. Isotopic measurements were performed with a separate HP 5890 Series II instrument equipped with a GasPro column using helium as the carrier gas.

NMR: NMR spectroscopy was performed using Varian and Bruker 400 MHz NMR spectrometers equipped with broadband auto-tune probes. ¹H NMR chemical shifts are reported in ppm relative to tetramethylsilane, using residual solvent resonances as internal standards.

UV-Vis: Spectra were collected using a Cary 60 instrument with Cary WinUV software.

X-ray Crystallography: XRD studies were carried out at the Caltech Beckman Institute Crystallography Facility on a Bruker D8 Venture diffractometer (Cu K α radiation). The crystals were mounted on a glass fiber under Paratone N oil. Structures were solved using direct methods with SHELXS or SHELXT and refined against F² on all data by full-matrix least squares with SHELXL.¹¹ All of the solutions were performed in the Olex2 program.¹²

TPA derivatives

TPA^{CF3} (tris(4-(trifluoromethyl)-2-picolyl)amine)



Ammonium chloride (15 mg, 1 eq), 4-(trifluoromethyl)-pyridine-2-carbaldehyde (250 mg, 5 eq), and triethylamine (44 μ L, 1.1 eq) were combined in dichloromethane (5 mL). Sodium triacetoxyborohydride (270 mg, 4.5 eq) was added as a solid, and the mixture was stirred at room temperature for 48 h. A saturated aqueous solution of sodium carbonate was added, and the organic phase was separated. The aqueous phase was extracted twice with dichloromethane, dried over sodium sulfate, filtered, and concentrated under reduced pressure. The remaining oil was dissolved in ethyl acetate and added to a silica plug. The silica plug was eluted with ethyl acetate until the eluent was colorless, then the product was eluted with methanol, dried over sodium sulfate, and concentrated under reduced pressure to yield a yellow oil (48 mg, 34% yield).

¹H NMR (CDCl₃, 400 MHz): δ (ppm) = 8.71 (d, *J* = 5.1 Hz, 3H), 7.72 (d, *J* = 1.8 Hz, 3H), 7.36 (dd, *J* = 5.1, 1.7 Hz, 3H), 4.07 (s, 6H).

¹³C{¹H} NMR (CDCl₃, 101 MHz): δ (ppm) = 160.55 (s), 150.19 (s), 138.85 (q, J = 34.1 Hz), 122.74 (q, J = 273.3 Hz), 118.82 (q, J = 3.6 Hz), 117.87 (q, J = 3.8 Hz), 60.55 (s).

¹⁹F{¹H} NMR (CDCl₃, 376 MHz): δ (ppm) = -65.1 (s).

MS (ESI, UHPLC-MS (CH₃CO₂H), m/z): calculated for C₂₁H₁₅F₉N₄ + H, [M+H]⁺: 495.1, found: 495.1.

General note about the synthesis of iron compounds: For the preparation of the following iron complexes, only the crystalline yield for the first crop of crystals is reported. Higher yields can be obtained if the supernatant is concentrated and recrystallized.

[(TPA^{NMe2})Fe(MeCN)2]OTf2



White solids TPA^{NMe2} (tris(4-(dimethylamino)-2-picolyl)amine) (30 mg, 1 eq) and FeOTf₂ \cdot 2 MeCN (31 mg, 1 eq) were each dissolved in 0.4 mL acetonitrile. The solution of FeOTf₂ was added to the solution of TPA^{NMe2}, instantly producing a pale orange-brown solution. The solution was filtered through Celite, and diethyl ether (2 mL) was layered on top of the filtrate. This mixture was placed in a freezer (-30 °C) until a purple-gray (this compound changes color with varying temperature) precipitate appeared. The precipitate was isolated by decanting the supernatant and drying under vacuum (50.7 mg, 83% yield).

¹H NMR (25 °C, CD₃CN, 400.15 MHz): δ (ppm) = 94.92 (s), 93.33 (s), 43.73 (s), 35.32 (s), 15.88 (s).

¹⁹F{¹H} NMR (25 °C, CD₃CN, 376 MHz): δ (ppm) = -77.1.

UV-vis (MeCN): nm [cm⁻¹M⁻¹]): 225 [20000], 283 [40000], 342 [3900].

MS (ESI, direct injection in MeCN, m/z): calculated for C₂₅H₃₃N₇O₃F₃SFe, [M]⁺: 624.2, found: 624.3.

Electrochemistry: $E_{1/2} = 0.21$ V vs Fc/Fc⁺ (MeCN, 0.1 M TBAPF₆, BDD disk electrode).

[(TPA^{OMe})Fe(MeCN)2]OTf2



White solids TPA^{OMe} (tris(4-methoxy-2-picolyl)amine) (25 mg, 1 eq) and FeOTf₂ · 2 MeCN (29 mg, 1 eq) were each dissolved in 0.25 mL acetonitrile. The solution of FeOTf₂ was added to the solution of TPA^{OMe}, instantly producing a purple-red solution. The solution was filtered through Celite, and diethyl ether (2 mL) was layered on top of the filtrate. This mixture was placed in a freezer (-30 °C) until dark purple needle-shaped crystals appeared. The precipitate was isolated by decanting the supernatant and drying under vacuum (34.7 mg, 64% yield).

¹H NMR (25 °C, CD₃CN, 400.15 MHz): δ (ppm) = 38.51 (s), 30.31 (s), 22.62 (s), 20.02 (s), 4.63 (s).

¹⁹F{¹H} NMR (25 °C, CD₃CN, 376 MHz): δ (ppm) = -78.1.

UV-vis (MeCN): nm [cm⁻¹M⁻¹]): 235 [24000], 339 [5100], 375 [4700].

MS (ESI, direct injection in MeCN, m/z): calculated for C₂₂H₂₄N₄O₆F₃SFe, [M]⁺: 585.1, found: 585.2.

Electrochemistry: $E_{1/2} = 0.55$ V vs Fc/Fc⁺ (MeCN, 0.1 M TBAPF₆, BDD disk electrode).

[(TPACF3)Fe(MeCN)2]OTf2



Yellow oil TPA^{CF3} (tris(4-(trifluoromethyl)-2-picolyl)amine) (72 mg, 1 eq) and white solid FeOTf₂ · 2 MeCN (63 mg, 1 eq) were each dissolved in 0.5 mL acetonitrile. The solution of FeOTf₂ was added to the solution of TPA^{CF3}, instantly producing a purple-red solution. The solution was filtered through Celite, and diethyl ether (2 mL) was layered on top of the filtrate. This mixture was placed in a freezer (-30 °C) until dark red-brown crystals appeared. The precipitate was isolated by decanting the supernatant and drying under vacuum (47.5 mg, 35% yield).

¹H NMR (25 °C, CD₃CN, 400.15 MHz): δ (ppm) = 11.07 (s), 8.43 (s), 5.86 (s), 3.66 (s). ¹⁹F{¹H} NMR (25 °C, CD₃CN, 376 MHz): δ (ppm) = -65.9 (s), -78.9 (s). UV-vis (MeCN): nm [cm⁻¹M⁻¹]): 225 [9900], 262 [13000], 388 [6600], 428 [8900].

MS (ESI, direct injection in MeCN, m/z): calculated for C₂₂H₁₅N₄O₃F₁₂SFe, [M]⁺: 699.0, found: 699.0.

Electrochemistry: $E_{p/2} = 1.09 \text{ V}$ (irreversible) vs Fc/Fc⁺ (MeCN, 0.1 M TBAPF₆, BDD disk electrode).

<u>BPM – dipyridyl substitution</u>

2,2'-(ethene-1,1-diyl)bis(4-(dimethylamino)pyridine)



Note: This compound was prepared with modifications to the literature procedure for 2,2'- (ethene-1,1-diyl)dipyridine.¹³ Notably, poor performance was observed when methylenetriphenylphosphorane was generated *in situ*, presumably due to residual potassium salts (i.e., KBr).

Bis(4-dimethylaminopyridin-2-yl)methanone (406 mg, 1.5 mmol, 1 eq) was suspended in anhydrous tetrahydrofuran (15 mL, 0.1 M) in a glovebox and cooled to -30 °C, after which isolated methylenetriphenylphosphorane (435 mg, 1.58 mmol, 1.05 eq) was added in one portion. This mixture was warmed to room temperature and allowed to react for at least 4 h (reaction times up to overnight have no deleterious impact). Outside of the glovebox, the reaction was quenched with minimal water and concentrated to yield a viscous orange-red oil. The oil was purified by silica gel column chromatography by loading with dichloromethane, eluting the triphenylphosphine oxide with 3:1 ethyl acetate:acetone plus 1% triethylamine, then eluting the product with acetone plus 1% triethylamine to afford a white crystalline solid (346 mg, 86% yield).

 $R_f = 0.3$ (acetone plus 1% triethylamine; TLC plates treated with triethylamine).

¹H NMR (CDCl₃, 400 MHz): δ (ppm) = 8.29 (d, *J* = 5.9 Hz, 2H), 6.67 (d, *J* = 2.7 Hz, 2H), 6.46 (dd, *J* = 5.9, 2.6 Hz, 2H), 5.94 (s, 2H), 3.00 (s, 12H).

¹³C{¹H} NMR (CDCl₃, 101 MHz): δ (ppm) = 158.66 (s), 154.77 (s), 150.57 (s), 149.32 (s), 118.66 (s), 106.38 (s), 105.47 (s), 39.18 (s).

HRMS (ESI, TOF (CH₃CO₂H), m/z): calculated for C₁₆H₂₀N₄ + H, [M+H]⁺: 269.1761, found: 269.1763.

2,2'-(ethane-1,1-diyl)bis(4-(dimethylamino)pyridine)



2,2'-(ethene-1,1-diyl)bis(4-(dimethylamino)pyridine) (268 mg, 1 mmol, 1 eq), Pd/C (43 mg, 5% by mass Pd, 0.02 eq), and activated carbon (134 mg, 0.5 mass eq) were suspended in methanol (10 mL, 0.1 M). The headspace of this mixture was purged with nitrogen and then hydrogen. A hydrogen atmosphere was maintained using a balloon, and the mixture was allowed to react for 12 h. The suspension was allowed to settle, and the solution was filtered through Celite. The remaining solids were washed with methanol, and the supernatant was filtered through Celite. The combined solution was concentrated, dissolved in dichloromethane, dried over sodium sulfate, and concentrated again to afford an off-white

crystalline solid after gentle heating under vacuum. (247 mg, 91% yield). This material was used without further purification.

¹H NMR (CDCl₃, 400 MHz): δ (ppm) = 8.19 (d, *J* = 6.0 Hz, 2H), 6.58 (d, *J* = 2.6 Hz, 2H), 6.34 (dd, *J* = 6.0, 2.6 Hz, 2H), 4.21 (q, *J* = 7.2 Hz, 1H), 2.95 (s, 12H), 1.71 (d, *J* = 7.2 Hz, 3H).

¹³C{¹H} NMR (CDCl₃, 101 MHz): δ (ppm) = 164.30 (s), 154.84 (s), 149.03 (s), 105.02 (s), 104.81 (s), 50.39 (s), 39.13 (s), 19.94 (s).

MS (ESI, UHPLC-MS (CH₃CO₂H), m/z): calculated for C₁₆H₂₂N₄ + H, [M+H]⁺: 271.2, found: 271.2.

HRMS (ESI, TOF (CH₃CO₂H), m/z): calculated for C₁₆H₂₂N₄ + H, [M+H]⁺: 271.1917, found: 271.1919.

BPM^{NMe2} (6-(1,1-bis(4-(dimethylamino)pyridin-2-yl)ethyl)-2,2'-bipyridine)



This compound was prepared by analogy to the parent ligand.³ In a Schlenk tube under nitrogen, 2,2'-(ethane-1,1-diyl)bis(4-(dimethylamino)pyridine (135 mg, 0.5 mmol, 1 eq) was

dissolved in 2 mL dry tetrahydrofuran and cooled in a dry ice/acetone bath. A 1.6 M *n*-BuLi solution in hexanes (0.31 mL, 0.5 mmol, 1 eq) was added dropwise via syringe, and the solution turned yellow-orange. The mixture was stirred for 45 additional minutes, then 6-bromo-2,2'-bipyridine (118 mg, 0.5 mmol, 1 eq) was added as a solution in 0.5 mL tetrahydrofuran. The solution was warmed to room temperature and stirred for 16 h, after which time the reaction was quenched with water. The solution was concentrated, transferred to a separatory funnel, and extracted with dichloromethane. The combined organic fractions were concentrated and purified via column chromatography on neutral alumina using methanol:dichloromethane (1:49) to afford a waxy colorless solid. (163 mg, 77% yield).

 $R_f = 0.2$ (methanol; TLC plates treated with methanol).

¹H NMR (CDCl₃, 400 MHz): δ (ppm) = 8.56 (ddd, J = 4.8, 1.9, 0.9 Hz, 1H), 8.31 (dt, J = 8.1, 1.1 Hz, 1H), 8.19 (dd, J = 5.8, 0.6 Hz, 2H), 8.15 (dd, J = 7.8, 0.9 Hz, 1H), 7.65 (td, J = 7.7, 1.8 Hz, 1H), 7.60 (t, J = 7.8 Hz, 1H), 7.17 (ddd, J = 7.5, 4.8, 1.3 Hz, 1H), 7.11 (dd, J = 7.9, 0.9 Hz, 1H), 6.31 – 6.25 (m, 4H), 2.79 (s, 12H), 2.33 (s, 3H).

¹³C{¹H} NMR (CDCl₃, 101 MHz): δ (ppm) = 166.41 (s), 165.44 (s), 156.89 (s), 154.50 (s), 154.27 (s), 148.86 (s), 148.79 (s), 136.66 (s), 136.48 (s), 124.42 (s), 123.30 (s), 121.47 (s), 117.99 (s), 106.72 (s), 104.38 (s), 60.37 (s), 39.04 (s), 27.16 (s).

MS (ESI, UHPLC-MS (CH₃CO₂H), m/z): calculated for C₂₆H₂₈N₆ + H, [M+H]⁺: 425.3, found: 425.3.

HRMS (ESI, TOF (CH₃CO₂H), m/z): calculated for C₂₆H₂₈N₆ + H, [M+H]⁺: 425.2448, found: 425.2448.

bis(4-methoxypyridin-2-yl)methanone



This compound was prepared by analogy to the known analogue, bis(4-(dimethylamino)pyridin-2-yl)methanone.⁵ In a 100 mL flask under nitrogen, 2-bromo-4methoxypyridine (0.7 g, 3.7 mmol, 1 eq) was dissolved in 30 mL dry diethyl ether and cooled in a dry ice/acetone bath. A 1.7 M t-BuLi solution in pentane (4.4 mL, 7.4 mmol, 2 eq) was added dropwise via syringe, and the solution slowly turned red-orange. The mixture was stirred for 5 additional minutes before neat ethyl chloroformate (0.18 mL, 1.8 mmol, 0.5 eq) was added all at once via syringe, after which the solution darkened. The solution was stirred 30 additional minutes in the dry ice/acetone bath, warmed to approximately 0 $^{\circ}$ C, and then quenched with water. The mixture was transferred to a separatory funnel and extracted with additional diethyl ether. The combined organic fractions were dried over sodium sulfate and concentrated under reduced pressure. The crude material was eluted through a plug of silica using acetone and then purified by silica gel column chromatography (3:1 ethyl acetate: acetone plus 1% triethylamine; $R_f = 0.3$) to yield an off-white solid (0.267 g, 59%) yield). This material can be crystallized from cold 1:3 acetone:diethyl ether.

¹H NMR (CDCl₃, 400 MHz): δ (ppm) = 8.56 (dd, *J* = 5.7, 0.5 Hz, 2H), 7.61 (dd, *J* = 2.6, 0.5 Hz, 2H), 6.99 (dd, *J* = 5.7, 2.6 Hz, 2H), 3.93 (s, 6H).

¹³C{¹H} NMR (CDCl₃, 101 MHz): δ (ppm) = 193.11 (s), 166.28 (s), 156.11 (s), 150.42 (s), 112.73 (s), 110.93 (s), 55.53 (s).

MS (ESI, UHPLC-MS (CH₃CO₂H), m/z): calculated for C₁₃H₁₂N₂O₃ + H, [M+H]⁺: 245.1, found: 245.1.

HRMS (ESI, TOF (CH₃CO₂H), m/z): calculated for C₁₃H₁₂N₂O₃ + H, [M+H]⁺: 245.0921, found: 245.0927.

2,2'-(ethene-1,1-diyl)bis(4-(methoxypyridine)



Note: This compound was prepared with modifications to the literature procedure for 2,2'- (ethene-1,1-diyl)dipyridine.¹³ Notably, poor performance was observed when methylenetriphenylphosphorane was generated *in-situ*, presumably due to residual potassium salts (i.e., KBr).

Bis(4-methoxypyridin-2-yl)methanone (366 mg, 1.5 mmol, 1 eq) was dissolved in anhydrous tetrahydrofuran (15 mL, 0.1 M) in a glovebox and cooled to -30 °C, after which isolated methylenetriphenylphosphorane (435 mg, 1.58 mmol, 1.05 eq) was added in one portion.

This mixture was warmed to room temperature and allowed to react for at least 4 h (reaction times up to overnight have no deleterious impact). Outside of the glovebox, the reaction was quenched with minimal water and concentrated to yield a viscous orange-red oil. The oil was purified by silica gel column chromatography by loading with dichloromethane, eluting the triphenylphosphine oxide with 4:1 ethyl acetate:acetone plus 1% triethylamine, then eluting the product with 3:1 ethyl acetate:acetone plus 1% triethylamine to afford an off-white crystalline solid (269 mg, 74% yield).

 $R_{\rm f}=0.3$ (4:1 ethyl acetate:acetone plus 1% triethylamine; TLC plates treated with triethylamine).

¹H NMR (CDCl₃, 400 MHz): δ (ppm) = 8.49 (d, *J* = 5.7 Hz, 2H), 6.95 (d, *J* = 2.4 Hz, 2H), 6.79 (dd, *J* = 5.7, 2.5 Hz, 2H), 6.04 (s, 2H), 3.86 (s, 6H).

¹³C{¹H} NMR (CDCl₃, 101 MHz): δ (ppm) = 166.05 (s), 159.50 (s), 150.57 (s), 148.72 (s), 120.38 (s), 109.55 (s), 108.54 (s), 55.14 (s).

HRMS (ESI, TOF (CH₃CO₂H), m/z): calculated for C₁₄H₁₄N₂O₂ + H, [M+H]⁺: 243.1128, found: 243.1133.

2,2'-(ethane-1,1-diyl)bis(4-methoxypyridine)



2,2'-(ethene-1,1-diyl)bis(4-methoxypyridine) (242 mg, 1 mmol, 1 eq), Pd/C (43 mg, 5% by mass Pd, 0.02 eq), and activated carbon (121 mg, 0.5 mass eq) were suspended in methanol (10 mL, 0.1 M). The headspace of this mixture was purged with nitrogen and then hydrogen. A hydrogen atmosphere was maintained using a balloon, and the mixture was allowed to react for 12 h. The suspension was allowed to settle, and the solution was filtered through Celite. The remaining solids were washed with methanol, and the supernatant was filtered through Celite. The combined solution was concentrated and purified by column chromatography to afford a white solid (127 mg, 52% yield).

Chromatography: Silica was slurry-packed with 1:19 methanol:dichloromethane then equilibrated with 1:49 methanol:dichloromethane. A gradient of 1:49 to 1:19 methanol:dichloromethane was used to elute the product, as well as a dimeric product characterized below.

 $R_f = 0.3$ (1:19 methanol:dichloromethane).

¹H NMR (CDCl₃, 400 MHz): δ (ppm) = 8.36 (d, *J* = 5.8 Hz, 2H), 6.81 (d, *J* = 2.5 Hz, 2H), 6.64 (dd, *J* = 5.7, 2.5 Hz, 2H), 4.34 (q, *J* = 7.2 Hz, 1H), 3.79 (s, 6H), 1.71 (d, *J* = 7.2 Hz, 3H). ¹³C{¹H} NMR (CDCl₃, 101 MHz): δ (ppm) = 166.09 (s), 165.52 (s), 150.34 (s), 108.26 (s), 107.90 (s), 55.03 (s), 49.99 (s), 19.67 (s).

MS (ESI, UHPLC-MS (CH₃CO₂H), m/z): calculated for C₁₄H₁₆N₂O₂ + H, [M+H]⁺: 245.1, found: 245.1.

HRMS (ESI, TOF (CH₃CO₂H), m/z): calculated for C₁₄H₁₆N₂O₂ + H, [M+H]⁺: 245.1285, found: 245.1287.

Byproduct from synthesis of 2,2'-(ethane-1,1-diyl)bis(4-methoxypyridine):

2,2',2'',2'''- 2,2',2'''-(butane-1,1,3,3-tetrayl)tetrakis(4-methoxypyridine)



Isolated from the above reaction as a pale oil (75.6 mg, 31% yield).

 $R_f = 0.1$ (1:19 methanol:dichloromethane).

¹H NMR (CDCl₃, 400 MHz): δ (ppm) = 8.31 (dd, *J* = 5.6, 0.9 Hz, 2H), 8.27 (dd, *J* = 5.7, 0.9 Hz, 2H), 6.78 (d, *J* = 2.5 Hz, 2H), 6.70 (d, *J* = 2.2 Hz, 2H), 6.55 (dd, *J* = 2.5, 1.1 Hz, 2H), 6.53 (dd, *J* = 2.4, 1.1 Hz, 2H), 4.27 (t, *J* = 6.0 Hz, 1H), 3.77 (s, 6H), 3.74 (s, 6H), 3.40 (d, *J* = 5.8 Hz, 2H), 1.72 (s, 3H).

¹³C{¹H} NMR (CDCl₃, 101 MHz): δ (ppm) = 168.19 (s), 165.72 (s), 165.58 (s), 165.11 (s), 149.76 (s), 149.64 (s), 108.52 (s), 108.12 (s), 107.20 (s), 54.96 (s), 54.87 (s), 52.99 (s), 51.92 (s), 44.21 (s), 25.63 (s).

HRMS (ESI, TOF (CH₃CO₂H), m/z): calculated for C₂₈H₃₀N₄O₄ + H, [M+H]⁺: 487.2340, found: 487.2361.

BPM^{OMe} (6-(1,1-bis(4-methoxypyridin-2-yl)ethyl)-2,2'-bipyridine)



This compound was prepared by analogy to the parent ligand.³ In a Schlenk tube under nitrogen, 2,2'-(ethane-1,1-diyl)bis(4-methoxypyridine (122 mg, 0.5 mmol, 1 eq) was dissolved in 2 mL dry tetrahydrofuran and cooled in a dry ice/acetone bath. A 1.6 M *n*-BuLi solution in hexanes (0.31 mL, 0.5 mmol, 1 eq) was added dropwise via syringe, and the solution turned yellow-orange. The mixture was stirred for 45 additional minutes, then 6-bromo-2,2'-bipyridine (118 mg, 0.5 mmol, 1 eq) was added as a solution in 0.5 mL tetrahydrofuran. The solution was warmed to room temperature and stirred for 24 h, after which time the reaction was quenched with water. The solution was concentrated, transferred to a separatory funnel, and extracted with dichloromethane. The combined organic fractions were concentrated and purified via column chromatography to afford a white solid (166 mg, 83% yield).

Chromatography: Silica was slurry-packed with 1:49 methanol:dichloromethane then equilibrated with 1:99 methanol:dichloromethane. A gradient of 1:99 to 1:19 methanol:dichloromethane was used to elute the product.

 $R_{\rm f} = 0.3$ (1:9 methanol:dichloromethane).

¹H NMR (CDCl₃, 400 MHz): δ (ppm) = 8.56 (ddd, *J* = 4.9, 1.9, 1.0 Hz, 1H), 8.38 (dd, *J* = 5.6, 0.8 Hz, 2H), 8.20 (ddt, *J* = 8.8, 7.7, 1.1 Hz, 2H), 7.64 (td, *J* = 7.8, 0.9 Hz, 2H), 7.19 – 7.15 (m, 1H), 7.09 (dd, *J* = 7.8, 1.0 Hz, 1H), 6.60 (ddd, *J* = 5.6, 2.4, 0.9 Hz, 2H), 6.56 (dd, *J* = 2.4, 0.8 Hz, 2H), 3.66 (d, *J* = 0.9 Hz, 6H), 2.32 (d, *J* = 1.0 Hz, 3H).

¹³C{¹H} NMR (CDCl₃, 101 MHz): δ (ppm) = 167.66 (s), 165.72 (s), 164.37 (s), 156.55 (s), 154.55 (s), 150.12 (s), 148.88 (s), 136.87 (s), 136.71 (s), 123.94 (s), 123.46 (s), 121.30 (s), 118.37 (s), 110.37 (s), 106.96 (s), 60.23 (s), 54.96 (s), 27.10 (s).

MS (ESI, UHPLC-MS (CH₃CO₂H), m/z): calculated for C₂₄H₂₂N₄O₂ + H, [M+H]⁺: 399.2, found: 399.1.

HRMS (ESI, TOF (CH₃CO₂H), m/z): calculated for C₂₄H₂₂N₄O₂ + H, [M+H]⁺: 399.1816, found: 399.1821.

2,2'-(ethane-1,1-diyl)bis(4-(trifluoromethyl)pyridine)



This compound was prepared by analogy to the procedure reported for the parent dipyridylethane.¹⁴ In a Schlenk tube under nitrogen, 2-ethyl-4-(trifluoromethyl)pyridine (876 mg, 2 eq) was dissolved in THF (7 mL) and cooled to -78 °C while stirring. A 2.0 M solution of lithium diisopropylamide in THF/heptane/ethylbenzene (2.5 mL, 2 eq) was added dropwise via syringe, and the solution turned dark purple. The solution was warmed to room temperature for 5 minutes then cooled to -78 °C prior to dropwise addition of a cooled solution of 2-fluoro-4-(trifluoromethyl)pyridine (413 mg, 1 eq) in THF (3 mL). The reaction was warmed to room temperature after which it was stirred for 30 minutes then quenched with water. The mixture was concentrated, diluted in water, and extracted with dichloromethane. The organic extract was dried over sodium sulfate, concentrated, subjected to a silica plug with 1:4 ethyl acetate:hexane, and concentrated to a yellow oil that was used without further purification (0.15 g, 19% yield).

 $R_f = 0.3$ (1:9 ethyl acetate:hexane).

¹H NMR (CDCl₃, 400 MHz): δ (ppm) = 8.73 (dt, *J* = 5.1, 0.8 Hz, 2H), 7.57 – 7.54 (m, 2H), 7.37 (ddd, *J* = 5.1, 1.7, 0.8 Hz, 2H), 4.61 (q, *J* = 7.2 Hz, 1H), 1.80 (d, *J* = 7.2 Hz, 3H). ¹³C{¹H} NMR (CDCl₃, 101 MHz): 164.46 (s), 150.34 (s), 138.89 (q, *J* = 33.9 Hz), 122.80 (q, *J* = 273.3 Hz), 118.10 (q, *J* = 3.6 Hz), 117.49 (q, *J* = 3.5 Hz), 49.90 (s), 19.79 (s).

¹⁹F{¹H} NMR (CDCl₃, 376 MHz): δ (ppm) = -64.8 (s).

MS (ESI, UHPLC-MS (CH₃CO₂H), m/z): calculated for C₁₄H₁₀N₂F₆ + H, [M+H]⁺: 321.1, found: 321.1.





This compound was prepared by analogy to the parent ligand.³ In a Schlenk tube under nitrogen, 2,2'-(ethane-1,1-diyl)bis(4-trifluoromethyl)pyridine (0.1 g, 1 eq) was dissolved in 3 mL tetrahydrofuran and cooled in a dry ice/acetone bath. A 2.0 M lithium diisopropylamide solution in THF/heptane/ethylbenzene (0.16 mL, 1.05 eq) was added dropwise via syringe, and the solution turned red. The mixture was stirred for 5 additional minutes then warmed to room temperature. The solution was returned to the dry ice/acetone bath before 6-bromo-2,2'-bipyridine (0.073 g, 1 eq) was added as a solid. The solution was warmed to room temperature then heated to 70 °C for 1.5 d, after which time the reaction was cooled to room temperature and quenched with water. The solution was concentrated, transferred to a separatory funnel, and extracted with dichloromethane. The combined organic fractions were dried over sodium sulfate, concentrated, and purified via silica gel column chromatography using 1:4 ethyl acetate:hexane to afford a viscous yellow oil (0.075 g, 51% yield).

 $R_f = 0.2$ (1:4 ethyl acetate:hexane).

¹H NMR (CDCl₃, 400 MHz): δ (ppm) = 8.68 (dt, *J* = 5.1, 0.8 Hz, 2H), 8.57 (ddd, *J* = 4.8, 1.8, 0.9 Hz, 1H), 8.26 (dd, *J* = 7.9, 0.9 Hz, 1H), 7.98 (dt, *J* = 8.1, 1.1 Hz, 1H), 7.73 (t, *J* = 7.9)

Hz, 1H), 7.62 (td, *J* = 7.7, 1.8 Hz, 1H), 7.41 (dt, *J* = 1.6, 0.8 Hz, 2H), 7.32 (ddd, *J* = 5.1, 1.7, 0.8 Hz, 2H), 7.23 – 7.15 (m, 2H), 2.33 (s, 3H).

¹³C{¹H} NMR (CDCl₃, 101 MHz): δ (ppm) = 167.10 (s), 162.96 (s), 155.88 (s), 154.91 (s), 149.53 (s), 148.97 (s), 138.11 (q, *J* = 33.7 Hz), 137.60 (s), 136.83 (s), 123.78 (s), 123.09 (s), 122.92 (q, *J* = 273.6 Hz), 121.08 (s), 119.57 (q, *J* = 3.8 Hz), 118.96 (s), 117.13 (q, *J* = 3.6 Hz), 60.54 (s), 27.10 (s).

¹⁹F{¹H} NMR (CDCl₃, 376 MHz): δ (ppm) = -64.7 (s).

MS (ESI, UHPLC-MS (CH₃CO₂H), m/z): calculated for C₂₄H₁₆N₄F₆ + H, [M+H]⁺: 475.1, found: 475.1.

[(BPM^{NMe2})Fe(MeCN)₂]OTf₂



White solids BPM^{NMe2} (6-(1,1-bis(4-(dimethylamino)pyridin-2-yl)ethyl)-2,2'-bipyridine) (30 mg, 1 eq) and FeOTf₂ · 2 MeCN (31 mg, 1 eq) were each dissolved in 0.5 mL acetonitrile. The solution of FeOTf₂ was added to the solution of BPM^{NMe2}, instantly producing a purple solution. The solution was filtered through a glass microfilter, and diethyl ether (2 mL) was layered on top of the filtrate. This mixture was placed in a freezer (-30 °C) until a purple precipitate appeared. The precipitate was isolated by decanting the supernatant and drying under vacuum (50.7 mg, 83% yield).

¹H NMR (25 °C, CD₃CN, 400.15 MHz): δ (ppm) = 9.55 (d, *J* = 5.5 Hz, 1H), 8.49 (d, *J* = 6.8 Hz, 2H), 8.39 (d, *J* = 8.0 Hz, 1H), 8.27 (d, *J* = 7.9 Hz, 1H), 8.17 (td, *J* = 7.9, 0.9 Hz, 1H), 8.13 (t, *J* = 7.9 Hz, 1H), 7.94 (d, *J* = 8.0 Hz, 1H), 7.74 (t, *J* = 6.5 Hz, 1H), 7.04 (d, *J* = 2.7 Hz, 2H), 6.60 (dd, *J* = 6.9, 2.7 Hz, 2H), 3.07 (d, *J* = 1.0 Hz, 12H), 2.70 (s, 3H).

¹⁹F{¹H} NMR (25 °C, CD₃CN, 376 MHz): δ (ppm) = -79.3.

UV-vis (MeCN): nm [cm⁻¹M⁻¹]): 331 [12000], 393 [4400], 498 [2800], 528 [3200].

MS (ESI, direct injection in MeCN, m/z): calculated for C₂₇H₂₈N₆O₃F₃SFe, [M]⁺: 629.1, found: 629.2.

Electrochemistry: $E_{1/2} = 0.45$ V vs Fc/Fc⁺ (MeCN, 0.1 M TBAPF₆, BDD disk electrode).

[(BPM^{OMe})Fe(MeCN)₂]OTf₂

White solids BPM^{OMe} (6-(1,1-bis(4-methoxypyridin-2-yl)ethyl)-2,2'-bipyridine) (26 mg, 1 eq) and FeOTf₂ · 2 MeCN (28.5 mg, 1 eq) were each dissolved in 0.3 mL acetonitrile. The solution of FeOTf₂ was added to the solution of BPM^{OMe}, instantly producing a purple solution. The solution was filtered through a glass microfilter, and diethyl ether (1.5 mL) was layered on top of the filtrate. This mixture was placed in a freezer (-30 °C) until a purple precipitate appeared. The precipitate was isolated by decanting the supernatant and drying under vacuum (50.2 mg, 92% yield).

¹H NMR (25 °C, CD₃CN, 400.15 MHz): δ (ppm) = 9.57 (d, *J* = 5.5 Hz, 1H), 8.94 (d, *J* = 6.5 Hz, 2H), 8.40 (d, *J* = 8.0 Hz, 1H), 8.28 (d, *J* = 7.9 Hz, 1H), 8.18 (dt, *J* = 13.5, 7.9 Hz, 2H), 7.99 (d, *J* = 7.9 Hz, 1H), 7.77 (t, *J* = 6.6 Hz, 1H), 7.46 (d, *J* = 2.6 Hz, 2H), 7.00 (dd, *J* = 6.7, 2.6 Hz, 2H), 3.93 (s, 6H), 2.74 (s, 3H).

¹⁹F{¹H} NMR (25 °C, CD₃CN, 376 MHz): δ (ppm) = -79.3.

UV-vis (MeCN): nm [cm⁻¹M⁻¹]): 305 [24000], 338 [5500], 387 [3500], 496 [2900].

MS (ESI, direct injection in MeCN, m/z): calculated for C₂₅H₂₂N₄O₅F₃SFe, [M]⁺: 603.1, found: 603.2.

Electrochemistry: $E_{1/2} = 0.69$ V vs Fc/Fc⁺ (MeCN, 0.1 M TBAPF₆, BDD disk electrode).

[(BPM^{CF3})Fe(MeCN)₂]OTf₂

Yellow oil BPM^{CF3} (6-(1,1-bis(4-(trifluoromethyl)pyridin-2-yl)ethyl)-2,2'-bipyridine) (87.8 mg, 1 eq) and white solid FeOTf₂ · 2 MeCN (80.6 mg, 1 eq) were each dissolved in 0.3 mL acetonitrile. The solution of FeOTf₂ was added to the solution of BPM^{CF3}, instantly producing an orange solution. The solution was filtered through a glass microfilter, and diethyl ether (1.5 mL) was layered on top of the filtrate. This mixture was placed in a freezer (-30 °C) until an orange-red precipitate appeared. The precipitate was isolated by decanting the supernatant and drying under vacuum (163.8 mg, 97% yield).

¹H NMR (25 °C, CD₃CN, 400.15 MHz): δ (ppm) = 9.57 (d, *J* = 5.6 Hz, 1H), 9.53 (d, *J* = 6.0 Hz, 2H), 8.42 (d, *J* = 7.9 Hz, 1H), 8.26 (dt, *J* = 15.7, 7.9 Hz, 3H), 8.15 (s, 2H), 8.04 (d, *J* = 7.8 Hz, 1H), 7.83 (t, *J* = 6.6 Hz, 1H), 7.68 (d, *J* = 5.9 Hz, 2H), 2.91 (s, 3H).

¹⁹F{¹H} NMR (25 °C, CD₃CN, 376 MHz): δ (ppm) = -65.3, -79.3.

UV-vis (MeCN): nm [cm⁻¹M⁻¹]): 259 [16000], 301 [19000], 361 [4100], 425 [6000], 463 [5000].

MS (ESI, direct injection in MeCN, m/z): calculated for C₂₅H₁₆N₄O₃F₉SFe: 679.0, found: 679.2.

Electrochemistry: $E_{p/2} = 0.96$ V (irreversible) vs Fc/Fc⁺ (MeCN, 0.1 M TBAPF₆, BDD disk electrode).

D3. Additional voltammetry data related to E°

Figure D1. Correlation between half-wave $(E_{1/2})$ and half-peak $(E_{p/2})$ potentials for both $[(TPA^R)Fe(MeCN)_2]^{2+}$ and $[(BPM^R)Fe(MeCN)_2]^{2+}$ for $R = NMe_2$, OMe, H. Potentials are reported versus Fc/Fc⁺.

Figure D2. Electrochemical data for $[(TPA^R)Fe(MeCN)_2]^{2+}$ (left) and $[(BPM^R)Fe(MeCN)_2]^{2+}$ (right) analyzed using a Hammett parameter. E° represents $E_{1/2}$ or $E_{p/2}$ and is plotted as a function of Hammett σ_p . From left to right, R = NMe₂, OMe, H, CF₃.

D4. Additional voltammetry data for R = CF₃

Figure D3. Cyclic voltammetry (2 scans at 100 mV/s) of $[(TPA^{CF3})Fe(MeCN)_2]^{2+}$ in acetonitrile using 0.1 M tetrabutylammonium hexafluorophosphate as supporting electrolyte.

Figure D4. Randles–Ševčík plot from cyclic voltammograms of $[(TPA^{CF3})Fe(MeCN)_2]^{2+}$ in acetonitrile using 0.1 M tetrabutylammonium hexafluorophosphate as supporting electrolyte. The linear behavior is indicative of a freely diffusing, non-catalytic electroactive species, which, when combined with similar peak currents between scans, corroborates assignment of this redox feature as reversible.

Figure D5. Cyclic voltammetry (2 scans at 100 mV/s) of $[(BPM^{CF3})Fe(MeCN)_2]^{2+}$ in acetonitrile using 0.1 M tetrabutylammonium hexafluorophosphate as supporting electrolyte.

Figure D6. Randles–Ševčík plot from cyclic voltammograms of $[(BPM^{CF3})Fe(MeCN)_2]^{2+}$ in acetonitrile using 0.1 M tetrabutylammonium hexafluorophosphate as supporting electrolyte. The linear behavior is indicative of a freely diffusing, non-catalytic electroactive species, which, when combined with similar peak currents between scans, corroborates assignment of this redox feature as reversible.

D5. *E*¹ voltammetry

Figure D7. Differential pulse voltammetry of $[(TPA^R)Fe(MeCN)_2]^{2+}$ in acetonitrile with 50 equiv. NH₃ using 0.05 M ammonium triflate as supporting electrolyte with BDD WE. Peak locations (indicated by vertical bars in plot) were picked using the peak picking function in EC-Lab.

Figure D8. Differential pulse voltammetry of $[(BPM^R)Fe(MeCN)_2]^{2+}$ in acetonitrile with 50 equiv. NH₃ using 0.05 M ammonium triflate as supporting electrolyte with BDD WE. Peak locations (indicated by vertical bars in plot) were picked using the peak picking function in EC-Lab.
D6. Catalytic controlled potential coulometry experiments

The data in the main text related to catalytic activity versus E° was obtained coulometrically. While such data is typically obtained from an assessment of catalytic current via voltammetric methods, the low faradaic efficiencies observed with some of the catalysts under our catalytic conditions make such analyses unadvisable. Lowered faradaic efficiencies are observed with more electron-donating R-substituents that also result in lower overall activity. This is partially due to the catalytic conditions required for uniform comparison of the systems, i.e., low overall ammonia concentration to prevent ligand demetallation for CF₃-substituted catalysts. Since the catalytic rate is dependent on ammonia concentration, using only 50 equiv. NH₃ results in lower rates and facilitates unproductive, reversible redox processes with early intermediates, for example Fe^{II}–NH₃ \rightleftharpoons Fe^{III}–NH₂, thus lowering the faradaic efficiency for N₂.

Coulometric analysis of turnover frequency can accurately reflect the intrinsic turnover frequency, provided that catalyst decomposition is negligible or equivalent between systems. Employing a high catalyst concentration (0.4 mM) as compared to the optimal concentration (0.05 mM) and a relatively short reaction time (8 h vs 48 h) both serve to mitigate effects related to catalyst loss.

Procedures for controlled potential coulometry (CPC)

Preparation of the BDD electrode: A 10 cm² boron-doped diamond (BDD) plate electrode (IKA) was physically attached to standard electrical wire, and the connection was covered

with Teflon tape. The surface area of the electrode submerged in solution was approximately 4 cm^2 .

BDD has a surface that exists in various states of reduction (H-terminated) and oxidation (O-terminated).¹⁵ In order to remove attached nitrogen and iron generated during controlled potential coulometry experiments and to ensure a reliable electrode surface prior to CPC measurements, the BDD plate electrode was oxidatively treated prior to use. First, the electrode was soaked in dilute nitric acid (~2 M) for 5 minutes. Then, a potential of 3.0 V vs Ag/AgCl was applied to the BDD electrode in a 0.5 M H₂SO₄ solution for 10 minutes. The electrode was then thoroughly rinsed with water prior to use.

Preparation of the platinum counter electrode: In order to ensure a highly active Pt surface for HER prior to CPC experiments, the Pt mesh counter electrode was soaked in concentrated hydrochloric acid for at least 5 minutes prior to usage.

Preparation of the custom Ag/AgOTf reference electrode: To ensure reliable potential measurements, the custom Ag/AgOTf reference electrode was prepared prior to each CPC experiment then a CV of ferrocene was measured. In a glass tube fitted with a Vycor porous glass frit attached by Teflon heatshrink tubing, an acetonitrile solution containing 5 mM AgOTf and 0.1 M TBAPF₆ or 0.05 M NH₄OTf was added. A silver wire was placed inside and the electrode was sealed.

Preparation of ammonia solutions: Saturated 2 M solutions¹⁶ in acetonitrile were prepared by bubbling anhydrous ammonia through acetonitrile in a Schlenk tube under an

argon/ammonia atmosphere. More dilute stock solutions were immediately prepared from this saturated solution.

CPC: Inside an argon glovebox, a gas-tight electrochemical cell equipped with a 24/40 cap containing three tungsten rods for electrical contacts and a 14/20 joint carefully sealed with a Suba-Seal septum was prepared. A BDD plate electrode ($A = 4 \text{ cm}^2$), high surface area platinum mesh electrode, and custom Ag/AgOTf reference electrode were connected to the 24/40 cap. All chemical reagents were then rapidly added to the cell to prevent evaporation of ammonia, then the cell was sealed with the 24/40 cap. Prior to each CPC experiment, a ZIR and CV were taken. No IR compensation was applied for CPC measurements. The CPC experiment was conducted for 8–48 h, then the cell was removed from the glovebox for analysis by gas chromatography. For headspace analysis, 100 µL of the headspace was injected into a GC-TCD for quantification using a lockable Hamilton syringe with a 26S gauge needle.

Table D1. Results of catalytic CPC experiments performed at 0.85 V vs Fc/Fc⁺ for 8 h for examining the E_2 LFER with 0.4 mM [Fe] and 20 mM NH₃ (50 equivalents) in 10 mL acetonitrile total. Low ammonia concentrations were used to prevent demetallation.

Entry	Fe Source	Eq. N ₂	Charge (C)	FE N ₂ (%)	FE H ₂ (%)
1	[(TPA ^{NMe2})Fe(MeCN) ₂]OTf ₂	0.73	3.9	44	16
2	[(TPA ^{NMe2})Fe(MeCN) ₂]OTf ₂	0.51	8.2	14	14
3	[(TPA ^{OMe})Fe(MeCN) ₂]OTf ₂	1.5	24.8	14	16
4	[(TPA ^{OMe})Fe(MeCN) ₂]OTf ₂	1.4	25	13	18
5	[(TPA ^H)Fe(MeCN) ₂]OTf ₂	3.3	18.3	42	44
6	[(TPA ^H)Fe(MeCN) ₂]OTf ₂	2.9	15.6	44	42
7	[(TPA ^{CF3})Fe(MeCN) ₂]OTf ₂	4.0	14.4	65	56
8	[(TPA ^{CF3})Fe(MeCN) ₂]OTf ₂	3.7	19.7	43	51
9	[(BPM ^{NMe2})Fe(MeCN) ₂]OTf ₂	6.0	22.2	63	55
10	[(BPM ^{NMe2})Fe(MeCN) ₂]OTf ₂	6.3	24.1	61	55
11	[(BPM ^{OMe})Fe(MeCN) ₂]OTf ₂	12.8	34.4	87	75
12	[(BPM ^{OMe})Fe(MeCN) ₂]OTf ₂	12.7	39.1	76	68
13	[(BPM ^H)Fe(MeCN) ₂]OTf ₂	14.4	41.0	82	73
14	[(BPM ^H)Fe(MeCN) ₂]OTf ₂	12.6	39.2	75	67
15	[(BPM ^{CF3})Fe(MeCN) ₂]OTf ₂	7.4	18.5	94	82
16	[(BPM ^{CF3})Fe(MeCN) ₂]OTf ₂	9.4	24.3	90	84

Table D2. Results of catalytic CPC experiments performed at 0.85 V vs Fc/Fc⁺ for 48 h with 0.05 mM [Fe] and 100 mM NH₃ (2000 equivalents) in 10 mL acetonitrile total. Reload experiments are listed as x.1 and x.2 for the first and second experiment, respectively. The reload was stopped after 24 h when current ceased. LOQ indicates that too little gas was produced to exceed the limit of quantification.

Entry	Fe Source	Eq. N ₂	Charge (C)	FE N ₂ (%)	FE H ₂ (%)
1	[(BPM ^{OMe})Fe(MeCN) ₂]OTf ₂	383	108.9	102	83
1.1	[(BPM ^{OMe})Fe(MeCN) ₂]OTf ₂	52	17.8	85	75
2	[(BPM ^{OMe})Fe(MeCN) ₂]OTf ₂	381	113.9	98	86
3	[(BPM ^{OMe})Fe(MeCN) ₂]OTf ₂	rinse test	113.7	rinse test	rinse test
3 – rinse test	[(BPM ^{OMe})Fe(MeCN) ₂]OTf ₂	LOQ	1.5	LOQ	LOQ

Rinse test procedure: After performing 48 h of CPC (entry 3), the electrochemical cell was purged and opened inside of the glovebox to prevent exposure of the working electrode to ambient conditions. The working electrode was thoroughly rinsed with acetonitrile. Then, fresh acetonitrile containing 0.05 M NH₄OTf and 100 mM NH₃ (2000 equivalents) was added to the electrochemical cell, and the cell was resubjected to a potential of 0.85 V vs Fc/Fc^+ for 48 h (entry 3 – rinse test).



Figure D7. Cyclic voltammogram of 0.05 mM $[(BPM^{OMe})Fe(N')_2]^{2+}$ in acetonitrile containing 0.05 M ammonium triflate electrolyte and 2000 equiv. NH₃ (0.1 M). A BDD plate working electrode, platinum mesh counter electrode, and Ag/AgOTf reference electrode were used.

D8. NMR spectra of iron complexes used for Evans method



Figure D8. ¹H NMR spectrum of [(TPA^{NMe2})Fe(MeCN)₂]OTf₂ in CD₃CN at 25 °C.



6.3 6.2 6.1 6.0 5.9 5.8 5.7 5.6 5.5 5.4 5.3 5.2 5.1 5.0 4.9 4.8 4.7 4.6 4.5 4.4 4.3 4.2 4.1 4.0 3.9 3.8 3.7 3.8 3.5 3.4 3.3 3.2 ft (ppm)

Figure D9. Trimethoxybenzene signals used for Evan's method for $[(TPA^{NMe2})Fe(MeCN)_2]OTf_2$ in CD₃CN at 25 °C. [Fe] = 0.011 M, $\Delta f = 145$ Hz, f = 400.15 MHz, $\mu_B = 4.4$.



Figure D10. ¹H NMR spectrum of $[(TPA^{NMe2})Fe(L)_2]OTf_2$ (L = MeCN, NH₃), formed by mixing $[(TPA^{NMe2})Fe(MeCN)_2]OTf_2$ with 75 equivalents of NH₃ in CD₃CN at 25 °C.



Figure D11. Trimethoxybenzene signals used for Evan's method for $[(TPA^{NMe2})Fe(L)_2]OTf_2$ (L = MeCN, NH₃), formed by mixing $[(TPA^{NMe2})Fe(MeCN)_2]OTf_2$ with 75 equivalents of NH₃ in CD₃CN at 25 °C. [Fe] = 0.011 M, $\Delta f = 182$ Hz, f = 400.15 MHz, $\mu_B = 4.9$.



Figure D12. ¹H NMR spectrum of [(TPA^{OMe})Fe(MeCN)₂]OTf₂ in CD₃CN at 25 °C.



6.3 6.2 6.1 6.0 5.9 5.8 5.7 5.6 5.5 5.4 5.3 5.2 5.1 5.0 4.9 4.8 4.7 4.6 4.5 4.4 4.3 4.2 4.1 4.0 3.9 3.8 3.7 3.6 3. f1 (ppm)

Figure D13. Trimethoxybenzene signals used for Evan's method for $[(TPA^{OMe})Fe(MeCN)_2]OTf_2$ in CD₃CN at 25 °C. [Fe] = 0.011 M, $\Delta f = 62.9$ Hz, f = 400.15 MHz, $\mu_B = 2.9$.



Figure D14. ¹H NMR spectrum of $[(TPA^{OMe})Fe(L)_2]OTf_2$ (L = MeCN, NH₃), formed by mixing $[(TPA^{OMe})Fe(MeCN)_2]OTf_2$ with 75 equivalents of NH₃ in CD₃CN at 25 °C.



Figure D15. Trimethoxybenzene signals used for Evan's method for $[(TPA^{OMe})Fe(L)_2]OTf_2$ (L = MeCN, NH₃), formed by mixing $[(TPA^{OMe})Fe(MeCN)_2]OTf_2$ with 75 equivalents of NH₃ in CD₃CN at 25 °C. [Fe] = 0.011 M, $\Delta f = 212$ Hz, f = 400.15 MHz, $\mu_B = 5.4$.



Figure D16. ¹H NMR spectrum of [(TPA)Fe(MeCN)₂]OTf₂ in CD₃CN at 25 °C.



65 64 63 62 61 60 59 58 57 56 55 54 53 52 51 50 49 48 47 46 45 44 43 42 41 40 39 38 37 36 35 f1(ppm)

Figure D17. Trimethoxybenzene signals used for Evan's method for $[(TPA)Fe(MeCN)_2]OTf_2$ in CD₃CN at 25 °C. [Fe] = 0.011 M, Δf = 3.46 Hz, f = 400.15 MHz, $\mu_B = 0.68$.



Figure D18. ¹H NMR spectrum of $[(TPA)Fe(L)_2]OTf_2$ (L = MeCN, NH₃), formed by mixing $[(TPA)Fe(MeCN)_2]OTf_2$ with 75 equivalents of NH₃ in CD₃CN at 25 °C.



Figure D19. Trimethoxybenzene signals used for Evan's method for $[(TPA)Fe(L)_2]OTf_2$ (L = MeCN, NH₃), formed by mixing $[(TPA)Fe(MeCN)_2]OTf_2$ with 75 equivalents of NH₃ in CD₃CN at 25 °C. [Fe] = 0.011 M, $\Delta f = 200$ Hz, f = 400.15 MHz, $\mu_B = 5.2$.



Figure D20. ¹H NMR spectrum of [(TPA^{CF3})Fe(MeCN)₂]OTf₂ in CD₃CN at 25 °C.



6.3 6.2 6.1 6.0 5.9 5.8 5.7 5.6 5.5 5.4 5.3 5.2 5.1 5.0 4.9 4.8 4.7 4.6 4.5 4.4 4.3 4.2 4.1 4.0 3.9 3.8 3.7 3.6 3. ff (ppm)

Figure D21. Trimethoxybenzene signals used for Evan's method for $[(TPA^{CF3})Fe(MeCN)_2]OTf_2$ in CD₃CN at 25 °C. [Fe] = 0.011 M, $\Delta f = 4.74$ Hz, f = 400.15 MHz, $\mu_B = 0.78$.

Note: At the total NH₃/[Fe] concentrations required for NMR experiments, the TPA^{CF3} ligand dissociates, thus NMR data for $[(TPA^{CF3})Fe(L)_2]OTf_2$ is unavailable.



Figure D22. ¹H NMR spectrum of [(BPM^{NMe2})Fe(MeCN)₂]OTf₂ in CD₃CN at 25 °C.



6.4 6.3 6.2 6.1 6.0 5.9 5.8 5.7 5.6 5.5 5.4 5.3 5.2 5.1 5.0 4.9 4.8 4.7 4.6 4.5 4.4 4.3 4.2 4.1 4.0 3.9 3.8 3.7 3.6 3.5 3.4 f1 (com)

Figure D23. Trimethoxybenzene signals used for Evan's method for $[(BPM^{NMe2})Fe(MeCN)_2]OTf_2$ in CD₃CN at 25 °C. [Fe] = 0.011 M, $\Delta f = 1.4$ Hz, f = 400.15 MHz, $\mu_B = 0.43$.



Figure D24. ¹H NMR spectrum of $[(BPM^{NMe2})Fe(L)_2]OTf_2$ (L = MeCN, NH₃), formed by mixing $[(BPM^{NMe2})Fe(MeCN)_2]OTf_2$ with 75 equivalents of NH₃ in CD₃CN at 25 °C.



Figure D25. Trimethoxybenzene signals used for Evan's method for [(BPM^{NMe2})Fe(L)₂]OTf₂ (L = MeCN, NH₃), formed by mixing [(BPM^{NMe2})Fe(MeCN)₂]OTf₂ with 75 equivalents of NH₃ in CD₃CN at 25 °C. [Fe] = 0.011 M, $\Delta f = 7.0$ Hz, f = 400.15 MHz, $\mu_B = 0.96$.



Figure D26. ¹H NMR spectrum of [(BPM^{OMe})Fe(MeCN)₂]OTf₂ in CD₃CN at 25 °C.



3.5 6.4 6.3 6.2 6.1 6.0 5.9 5.8 5.7 5.6 5.5 5.4 5.3 5.2 5.1 5.0 4.9 4.8 4.7 4.6 4.5 4.4 4.3 4.2 4.1 4.0 3.9 3.8 3.7 3.6 3.5 3.4 3.3 fl(nom)

Figure D27. Trimethoxybenzene signals used for Evan's method for $[(BPM^{OMe})Fe(MeCN)_2]OTf_2$ in CD₃CN at 25 °C. [Fe] = 0.011 M, $\Delta f = 1.8$ Hz, f = 400.15 MHz, $\mu_B = 0.48$.



Figure D28. ¹H NMR spectrum of $[(BPM^{OMe})Fe(L)_2]OTf_2$ (L = MeCN, NH₃), formed by mixing $[(BPM^{OMe})Fe(MeCN)_2]OTf_2$ with 75 equivalents of NH₃ in CD₃CN at 25 °C.



Figure D29. Trimethoxybenzene signals used for Evan's method for $[(BPM^{OMe})Fe(L)_2]OTf_2$ (L = MeCN, NH₃), formed by mixing $[(BPM^{OMe})Fe(MeCN)_2]OTf_2$ with 75 equivalents of NH₃ in CD₃CN at 25 °C. [Fe] = 0.011 M, $\Delta f = 88.9$ Hz, f = 400.15 MHz, $\mu_B = 3.4$.



Figure D30. ¹H NMR spectrum of [(BPM^{CF3})Fe(MeCN)₂]OTf₂ in CD₃CN at 25 °C.



6.7 6.6 6.5 6.4 6.3 6.2 6.1 6.0 5.9 5.8 5.7 5.6 5.5 5.4 5.3 5.2 5.1 5.0 4.9 4.8 4.7 4.6 4.5 4.4 4.3 4.2 4.1 4.0 3.9 3.8 3.7 3.6 3.5 f1 (pm)

Figure D31. Trimethoxybenzene signals used for Evan's method for $[(BPM^{CF3})Fe(MeCN)_2]OTf_2$ in CD₃CN at 25 °C. [Fe] = 0.011 M, $\Delta f = 1.5$ Hz, f = 400.15 MHz, $\mu_B = 0.45$.



Figure D32. ¹H NMR spectrum of $[(BPM^{CF3})Fe(L)_2]OTf_2$ (L = MeCN, NH₃), formed by mixing $[(BPM^{CF3})Fe(MeCN)_2]OTf_2$ with 5 equivalents of NH₃ in CD₃CN at 25 °C. Demetallation occurs with 75 equivalents.



Figure D33. Trimethoxybenzene signals used for Evan's method for $[(BPM^{CF3})Fe(L)_2]OTf_2$ (L = MeCN, NH₃), formed by mixing $[(BPM^{CF3})Fe(MeCN)_2]OTf_2$ with 5 equivalents of NH₃ in CD₃CN at 25 °C. Demetallation occurs with 75 equivalents. [Fe] = 0.011 M, $\Delta f = 1.2$ Hz, f = 400.15 MHz, $\mu_B = 0.40$.

D9. UV-vis titration data for stability against demetallation



Figure D34. UV-vis spectra of acetonitrile solution containing 0.12 mM $[(TPA^{NMe2})Fe(MeCN)_2]OTf_2$ and varying equivalents NH₃ in a 1 cm cuvette.



Figure D35. Selected UV-vis spectra used for determining the onset of demetallation for acetonitrile solution containing 0.12 mM $[(TPA^{NMe2})Fe(MeCN)_2]OTf_2$ and varying equivalents NH₃ in a 1 cm cuvette.



Figure D36. UV-vis spectra of acetonitrile solution containing 0.12 mM $[(TPA^{OMe})Fe(MeCN)_2]OTf_2$ and varying equivalents NH₃ in a 1 cm cuvette.



Figure D37. Selected UV-vis spectra used for determining the onset of demetallation for acetonitrile solution containing $0.12 \text{ mM} [(TPA^{OMe})Fe(MeCN)_2]OTf_2$ and varying equivalents NH₃ in a 1 cm cuvette.



Figure D38. UV-vis spectra of acetonitrile solution containing 0.12 mM [(TPA^{CF3})Fe(MeCN)₂]OTf₂ and varying equivalents NH₃ in a 1 cm cuvette.



Figure D39. Selected UV-vis spectra used for determining the onset of demetallation for acetonitrile solution containing $0.12 \text{ mM} [(TPA^{CF3})Fe(MeCN)_2]OTf_2$ and varying equivalents NH₃ in a 1 cm cuvette.



Figure D40. UV-vis spectra of acetonitrile solution containing 0.12 mM $[(BPM^{NMe2})Fe(MeCN)_2]OTf_2$ and varying equivalents NH₃ in a 1 cm cuvette.



Figure D41. Selected UV-vis spectra used for determining the onset of demetallation for acetonitrile solution containing 0.12 mM $[(BPM^{NMe2})Fe(MeCN)_2]OTf_2$ and varying equivalents NH₃ in a 1 cm cuvette.



Figure D42. UV-vis spectra of acetonitrile solution containing 0.12 mM [(BPM^{OMe})Fe(MeCN)₂]OTf₂ and varying equivalents NH₃ in a 1 cm cuvette.



Figure D43. Selected UV-vis spectra used for determining the onset of demetallation for acetonitrile solution containing 0.12 mM $[(BPM^{OMe})Fe(MeCN)_2]OTf_2$ and varying equivalents NH₃ in a 1 cm cuvette.



Figure D44. UV-vis spectra of acetonitrile solution containing 0.12 mM [(BPM^{CF3})Fe(MeCN)₂]OTf₂ and varying equivalents NH₃ in a 1 cm cuvette.



Figure D45. Selected UV-vis spectra used for determining the onset of demetallation for acetonitrile solution containing 0.12 mM $[(BPM^{CF3})Fe(MeCN)_2]OTf_2$ and varying equivalents NH₃ in a 1 cm cuvette.



Figure D47. ESI mass spectrum of [(TPA^{NMe2})Fe(MeCN)₂]OTf₂ in acetonitrile.



Figure D48. ESI mass spectrum of [(TPA^{OMe})Fe(MeCN)₂]OTf₂ in acetonitrile.



Figure D49. ESI mass spectrum of [(TPA^{CF3})Fe(MeCN)₂]OTf₂ in acetonitrile.



Figure D50. ESI mass spectrum of [(BPM^{CF3})Fe(MeCN)₂]OTf₂ in acetonitrile.



Figure D51. ESI mass spectrum of [(BPM^{OMe})Fe(MeCN)₂]OTf₂ in acetonitrile.



Figure D52. ESI mass spectrum of [(BPM^{NMe2})Fe(MeCN)₂]OTf₂ in acetonitrile.

D11. NMR spectra

TPA^{CF3} (tris(4-(trifluoromethyl)-2-picolyl)amine)

¹H NMR:



2,2'-(ethene-1,1-diyl)bis(4-(dimethylamino)pyridine)

¹H NMR:



2,2'-(ethane-1,1-diyl)bis(4-(dimethylamino)pyridine)

¹H NMR:



BPM^{NMe2} (6-(1,1-bis(4-(dimethylamino)pyridin-2-yl)ethyl)-2,2'-bipyridine)

¹H NMR:



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)





2.5

2.0 1.5

1.0 0.5 0.0

40 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 -20 -30 -4 f1 (ppm)


210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)



^{210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10} f1 (ppm)

2,2',2'',2'''- 2,2',2'''-(butane-1,1,3,3-tetrayl)tetrakis(4-methoxypyridine) ¹H NMR:



¹³C{¹H} NMR:



BPM^{OMe} (6-(1,1-bis(4-methoxypyridin-2-yl)ethyl)-2,2'-bipyridine)



2,2'-(ethane-1,1-diyl)bis(4-(trifluoromethyl)pyridine)







BPM^{CF3} (6-(1,1-bis(4-(trifluoromethyl)pyridin-2-yl)ethyl)-2,2'-bipyridine)







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