

**CROSTALK BETWEEN SOLUBLE FACTORS AND  
CELL-CELL INTERACTIONS: IMPLICATIONS FOR  
CELL CYCLE CONTROL AND TUMOR DEVELOPMENT**

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## ABSTRACT

Crosstalk between Soluble Factors and Cell-Cell Interactions:  
Implications for Cell Cycle Control and Tumor Development

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Precise and dynamic control of cell behaviors, including proliferation, adhesion, and migration, is required for proper tissue organization and homeostasis. A key element to understanding how cellular functions are controlled lies in uncovering the topology of the molecular signaling networks that couple environmental signals to cellular responses. In this study, we have parsed the signaling networks involved in cell cycle regulation and tumor development and uncovered novel mechanisms of crosstalk between soluble factors and cell-cell interactions.

Our findings demonstrate that extracellular cues, including the epidermal growth factor (EGF), stimulate proliferative signaling through  $\beta$ -catenin, an intracellular protein that participates in both cell adhesion and transcription of cell cycle genes. In fact, EGF-mediated  $\beta$ -catenin transcriptional activity is an essential signal for proliferation of normal epithelial cells. Additionally, in a cancer cell system, we discover that EGF cooperates with Wnt 3a, a classical agonist of  $\beta$ -catenin transcriptional activity, to induce greater signaling than either ligand alone. Notably, EGF and Wnt 3a activate

transcription using different sub-cellular pools of  $\beta$ -catenin. Because hyperactive  $\beta$ -catenin signaling drives proliferation in cancer, this suggests that attenuation of  $\beta$ -catenin signaling may require different therapeutic strategies for EGF- and Wnt-driven tumors.

Since  $\beta$ -catenin signaling can be antagonized by sequestration with the cell-cell contact protein E-cadherin at the plasma membrane, proliferative signals mediated by  $\beta$ -catenin may regulate growth suppression at high cell density, a property of normal cells that is often lost during tumorigenesis. Indeed, in non-tumorigenic epithelial cells, we demonstrate that E-cadherin is upregulated in contexts where  $\beta$ -catenin signaling and DNA synthesis are suppressed. Additionally, exogenous E-cadherin suppresses proliferation with a strict requirement for  $\beta$ -catenin binding. Future studies to test the hypothesis that E-cadherin regulates the growth of normal cells will benefit from a quantitative assay developed to measure E-cadherin: $\beta$ -catenin complexes. Such quantitative measurements are likely to be important because contact-mediated growth suppression by E-cadherin is coupled with a density-dependent, ligand-depletion mechanism that concomitantly regulates proliferation.

Finally, we demonstrate that EGF and other soluble factors synergistically control cell-cell interactions governing organization of normal epithelial cells into multicellular structures. Notably, this behavior resembles the program initiated during metastatic cancer, thus illustrating the flexibility of the epithelial phenotype even in non-cancerous cells. Together, these studies illustrate how the topology of molecular signaling networks can couple environmental cues including soluble extracellular factors and cell-cell interactions to regulate fundamental cellular functions.

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## ABBREVIATIONS

<b>Ab</b>	Antibody
<b>APC</b>	<i>Adenomatous polyposis coli</i> gene product
<b>BrdU</b>	Bromodeoxyuridine
<b>cAMP</b>	Cyclic adenosine monophosphate
<b>ChT</b>	Cholera toxin
<b>CREB</b>	cAMP-responsive element-binding protein
<b>DAPI</b>	4',6-diamidino-2'-phenylindole-dihydrochloride
<b>E-</b>	Epithelial
<b>EGF</b>	Epidermal growth factor
<b>EGFR</b>	EGF receptor
<b>EGTA</b>	Ethylene glycol tetraacetic acid
<b>ELISA</b>	Enzyme-linked immunosorbent assay
<b>EMT</b>	Epithelial-mesenchymal transition
<b>ERK</b>	Extracellular signal-regulated kinase
<b>Frz</b>	Frizzled
<b>GM</b>	Growth medium
<b>GSK3<math>\beta</math></b>	Glycogen synthase kinase 3 $\beta$
<b>HEPES</b>	4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid
<b>HGF</b>	Hepatocyte growth factor
<b>IgG</b>	Immunoglobulin G
<b>IP</b>	Immunoprecipitation
<b>Lef</b>	Lymphoid enhancer factor
<b>LRP</b>	Low-density lipoprotein receptor
<b>MOI</b>	Multiplicity of infection
<b>PAGE</b>	Poly-acrylamide gel electrophoresis
<b>pAkt</b>	Phospho-serine 473 Akt
<b>PBS</b>	Phosphate-buffered saline
<b>ppERK</b>	Dually-phosphorylated (Thr202/Tyr204) ERK
<b>PI3K</b>	Phosphatidylinositol 3-kinase

<b>PCR</b>	Polymerase chain reaction
<b>PKA</b>	cAMP-dependent protein kinase
<b>PKC</b>	Protein kinase C
<b>pRb</b>	Retinoblastoma protein
<b>RTK</b>	Receptor tyrosine kinase
<b>RT-PCR</b>	Reverse transcription PCR
<b>shRNA</b>	Short hairpin RNA
<b>siRNA</b>	Short interfering RNA
<b>SDS</b>	Sodium dodecyl sulfate
<b>S.E.</b>	Standard error
<b>TBS</b>	Tris-buffered saline
<b>TBST</b>	TBS plus 0.5% (v/v) Tween-20
<b>Tcf</b>	T-cell factor
<b>TGF<math>\beta</math></b>	Transforming growth factor $\beta$
<b>VE</b>	Vascular endothelial
<b>VEGF</b>	VE growth factor
<b>VEGFR</b>	VEGF receptor
<b>v/v</b>	Volume / volume