

Chapter 3

The bHLH Transcription Factor Olig2 Promotes Oligodendrocyte Differentiation
in Collaboration with Nkx2.2

The bHLH Transcription Factor *Olig2* Promotes Oligodendrocyte Differentiation in Collaboration with *Nkx2.2*

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Summary

Olig2, a basic helix-loop-helix (bHLH) transcription factor, is expressed in a restricted domain of the spinal cord ventricular zone that sequentially generates motoneurons and oligodendrocytes. Just prior to oligodendrocyte precursor formation, the domains of *Olig2* and *Nkx2.2* expression switch from being mutually exclusive to overlapping, and *Neurogenins1* and *2* are extinguished within this region. Coexpression of *Olig2* with *Nkx2.2* in the spinal cord promotes ectopic and precocious oligodendrocyte differentiation. Both proteins function as transcriptional repressors in this assay. This effect is blocked by forced expression of *Neurogenin1*. By contrast, misexpression of *Olig2* alone derepresses *Neurogenins* and promotes motoneuron differentiation. *Olig2* therefore functions sequentially in motoneuron and oligodendrocyte fate specification. This dual action is enabled by spatio-temporal changes in the expression domains of other transcription factors with which *Olig2* functionally interacts.

Introduction

A fundamental problem in neural development is to understand how multipotential progenitor cells generate the various classes of neurons and glia of the central nervous system (CNS). The production of these different cell types is highly organized in space and in time. Accordingly, two general strategies have evolved to orchestrate the generation of such cellular diversity. In one strategy, different classes of cells are generated from different regions of the ventricular zone at the same time, such as occurs in the ventral spinal cord (Tanabe and Jessell, 1996; Jessell, 2000). In the second strategy, a given region of the ventricular zone sequentially produces different cell types according to a particular schedule, such as occurs in the retina and cortex (Cepko, 1999; Desai and McConnell, 2000). These two strategies are not mutually exclusive. The molecular mechanisms underlying the spatial strategy have been shown to involve patterning processes that convert initially graded morphogen signals into discrete domains of transcription factor gene expression within the neuroepithelium, from which distinct neuronal subtypes arise (Tanabe and Jessell, 1996; Jessell, 2000). Much

less is known, however, about the corresponding mechanisms that underlie temporal switches in the generation of particular cell types from a common progenitor domain.

One system in which such a temporal switch occurs is the sequential production of neurons and oligodendrocytes from the ventral spinal cord. Oligodendrocytes, the myelinating glial cell type of the central nervous system, emerge from a discrete domain in the ventral spinal cord, beginning on or about E12.5–E13.5 in the mouse (Miller, 1996). This domain can be visualized by staining for PDGF receptor- α (PDGFR α), a specific marker of oligodendrocyte precursors (Pringle and Richardson, 1993; Hall et al., 1996). At earlier stages of development, this region of the ventricular zone generates one or more neuronal subtypes, including motoneurons and/or interneurons (Jessell, 2000). The molecular mechanisms that control the switch from the production of these neuronal subtypes to oligodendrocytes are not understood.

Studies of the spatial relationship of oligodendroglialogenesis to neurogenesis have been aided by the discovery that the ventricular zone of the ventral spinal cord is subdivided into five distinct progenitor domains, which generate different neuronal subtypes (Briscoe et al., 2000). These progenitor domains are defined by the boundaries of expression of various homeodomain transcription factors, which constitute a combinatorial code for the specification of neuronal progenitor identity (Briscoe et al., 2000). Motoneurons, for example, are generated from the pMN domain. This domain is bounded ventrally by the p3 domain, which is defined by expression of *Nkx2.2*, and dorsally by the p2 domain, whose ventral boundary is set by *Ir3*. It has been proposed that oligodendrocytes later emerge from the pMN domain (Richardson et al., 1997; Sun et al., 1998), but recent evidence has been interpreted to suggest that some or all oligodendroglial precursors emerge from the underlying *Nkx2.2*⁺ p3 domain (Soula et al., 2001).

Previously, we and others described two related basic helix-loop-helix (bHLH) transcription factors, called *Olig1* and *Olig2*, which delimit the oligodendrocyte precursor domain several days before such precursors can be detected by expression of PDGFR α (Lu et al., 2000; Zhou et al., 2000). Expression of the *Olig* genes persists in migratory oligodendrocyte precursors and in mature oligodendrocytes. Misexpression of *Olig2* in chick mesoderm causes ectopic expression of *Sox10*, a marker that is coexpressed with *Olig* genes in oligodendrocyte precursors (Zhou et al., 2000); however, it does not induce later oligodendrocyte markers. In addition, expression of *Olig1* in cultured CNS neural precursors increases the number of cells expressing NG2, another oligodendrocyte precursor marker (Lu et al., 2000). These data suggested that *Olig* genes might play an early role in oligodendrocyte precursor specification. Direct evidence of such a role for *Olig* genes in the spinal cord has, however, been lacking.

If the domain of *Olig* gene expression indeed determines the site of origin of oligodendrocytes, then the

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timing of *Olig* gene expression within the spinal cord might determine the time at which such precursors are specified. Paradoxically, however, *Olig2* is also expressed in the pMN domain at earlier stages, when motoneurons are being generated (Takebayashi et al., 2000). Moreover, in the accompanying manuscript, Novitsch et al. (2001 [this issue of *Neuron*]) demonstrate that misexpression of *Olig2* promotes motoneuron fate specification. These observations suggest either that *Olig* genes function in motoneuron and not oligodendrocyte fate determination, or alternatively that they play a role in both processes. To address this question, we have isolated a chicken homolog of *Olig2*, and examined its expression and function in relation to that of other transcription factors involved in neuronal differentiation in the ventral spinal cord (Briscoe et al., 2000).

At E3.0, the time that motoneurons are being generated, the ventral boundary of the *Olig2* expression domain is adjacent to, and nonoverlapping with, that of the underlying *Nkx2.2* domain. However, just before oligodendrocyte precursors start to be produced (E6–E7), these two boundaries become overlapping, and numerous *Olig2*⁺, *Nkx2.2*⁺ cells are seen migrating away from the ventricular zone in a pattern characteristic of oligodendrocyte precursors. Combined misexpression of both *Olig2* and *Nkx2.2*, but not of either gene alone, causes ectopic and precocious oligodendrocyte differentiation in the spinal cord. The normal generation of oligodendrocytes is also preceded by an extinction of the proneural bHLH factors *Ngn1* and *Ngn2* from the *Olig2*⁺ domain. Forced expression of *Ngn1* blocks both ectopic oligodendrocyte differentiation induced by coexpression of *Olig2*+*Nkx2.2* as well as normal oligodendrocyte differentiation. Taken together with the results of Novitsch et al. (2001), these data suggest that *Olig2* plays sequential roles in both motoneuron and oligodendrocyte fate specification. This dual action is enabled by spatio-temporal changes in the expression domains of other transcription factors expressed in the ventral spinal cord, with which *Olig2* functionally interacts.

Results

Identification of a Chick *Olig2* Gene

Previously, two murine *Olig* genes, *Olig1* and *Olig2*, were isolated and characterized (Lu et al., 2000; Zhou et al., 2000). Because the generation of neuronal diversity in the ventral spinal cord has been characterized most extensively in the chick, however (Jessell, 2000), we first sought to isolate chicken orthologs of the *Olig* genes in order to study their expression and function in that system. In situ hybridization to chick embryonic neural tissues, using a mouse *Olig2* cRNA probe, revealed the existence of one or more crosshybridizing species expressed at high levels in E3 and E4 spinal cord (data not shown). Accordingly, we constructed and screened a cDNA library made from dissociated E3 and E3.5 chick neural tubes (see Experimental Procedures). Several full-length chick *Olig2* clones were identified, as well as orthologs of *Olig3* (Takebayashi et al., 2000), but no *Olig1* orthologs were found. The chick and mouse *Olig2* genes have identical bHLH domains and share an overall 75% amino acid sequence identity (Supplemental Figure S1). However, several strings of alanine or glycine re-

peats present in murine *Olig2* are absent in the predicted chick *Olig2* protein. Nevertheless, the chick and mouse genes behaved indistinguishably in our functional assays (see below).

Expression of *Olig2* in Relation to Other Oligodendrocyte Markers in Embryonic Chick Spinal Cord

We compared the expression profile of chick *Olig2* with that of several other genes that were previously identified as markers of either oligodendrocyte precursors, such as *Nkx2.2* (Xu et al., 2000), *PDGFR α* (Hall et al., 1996), or *Sox10* (Zhou et al., 2000), or of differentiated, mature oligodendrocytes, such as myelin basic protein (MBP) or proteolipid protein (PLP/DM20) (Pfeiffer et al., 1993). This analysis was carried out at axial levels just posterior to the forelimb. At E6, the time at which the first oligodendrocyte precursors begin to arise in the spinal cord (Ono et al., 1995), both *Olig2* and *Nkx2.2* are strongly expressed in bilateral ventral foci in the ventricular zone, while the other markers are not yet detectable (Figures 1A–1E, arrows). Beginning on E7, individual *Olig2*- or *Nkx2.2*-positive cells can be seen migrating away from this zone into the gray matter, both laterally and dorsally (Figures 1F and 1G, arrowheads). By E8, many *Olig2*⁺ and *Nkx2.2*⁺ cells have already moved into the white matter (Figures 1K and 1L, arrowheads). Throughout the stages examined, from E6 to E9.5, the ventral foci of *Olig2* and *Nkx2.2* expression in the ventricular zone were maintained (Figures 1F, 1G, 1K, 1L, 1P, and 1Q; arrows).

In contrast to *Olig2* and *Nkx2.2*, the first *PDGFR α* -positive cells do not appear in the spinal cord at this axial level until E6–E6.5 (Figures 1C and 1H, arrow). Over the ensuing 2 days, these cells subsequently disperse throughout the spinal cord so that by E9.5, there is a relatively even distribution of *PDGFR α* ⁺ cells (Figure 1R). *Sox10*, which is strongly expressed in murine oligodendrocyte precursors (Zhou et al., 2000), is expressed at high levels only in mature oligodendrocytes in chick (Figure 1S, arrowheads). However, weak expression of *Sox10* in the oligodendrocyte precursor domain of the ventricular zone can be detected starting around E7 (Figures 1I and 1N, arrows). Mature oligodendrocyte markers, such as MBP and PLP/DM20, are not detectable until E9.5 (Figure 1T, arrowheads and data not shown). Our observations of *Nkx2.2*, *PDGFR α* , and MBP mRNA expression in the chick spinal cord are consistent with those described in previous reports (Xu et al., 2000). The spatio-temporal pattern of *Olig2* expression is consistent with that reported by Novitsch et al. (2001). However, the precise timing of the onset of migration of *Olig2*⁺ cells is shifted later in development in our study, due to the more caudal levels we have examined and reflecting the strong caudal-to-rostral gradient of oligodendroglialogenesis (Perez Villegas et al., 1999).

A Developmental Switch from Mutually Exclusive to Coincident Expression of *Olig2* and *Nkx2.2* Correlates with the Emergence of Oligodendrocyte Precursors

Since *Olig2* and *Nkx2.2* are both expressed in restricted ventricular zone foci, as well as in presumptive migrating

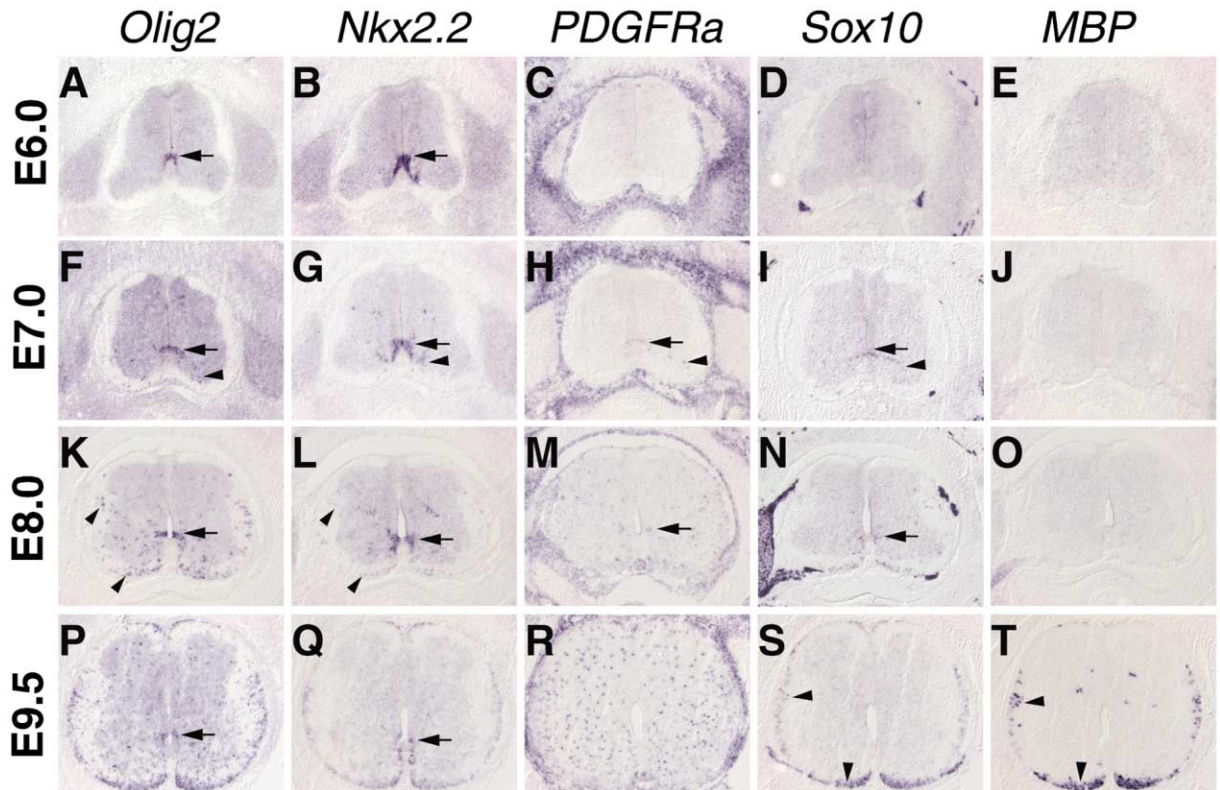


Figure 1. Expression of Olig2 in Relation to that of Oligodendrocyte Markers in Chick Spinal Cord

Nonisotopic *in situ* hybridizations were performed with the indicated probes at the indicated developmental ages. (A)–(E), (F)–(J), etc. are adjacent serial sections. Arrows indicate the oligodendrocyte precursor domain. Note that migrating oligodendrocyte precursors detected by Olig2, Nkx2.2, or PDGFR α are detected by E7.0 (F, G, and H, arrowheads), while mature oligodendrocyte markers such as MBP and PLP/DM20 are not detected until E9.5 in the white matter (T, arrowheads, and data not shown). Sox10 is first expressed at low levels at E7.0 in the ventricular zone (I and N, arrows) and in some migrating cells (I, arrowhead), but is not expressed at high levels in white matter until E9.5 (S, arrowheads).

oligodendrocyte precursor cells (Figure 1 and Lu et al., 2000; Xu et al., 2000; Zhou et al., 2000; Soula et al., 2001), we sought to compare their expression more directly. To this end, we raised a mouse antibody to chick Olig2 and performed double labeling with a rabbit anti-Nkx2.2 antibody (see Experimental Procedures). From E3 to E4, during the period of neurogenesis, the Olig2 and Nkx2.2 expression domains are adjacent and nonoverlapping (Figures 2A and 2B, open and filled arrowheads). As detailed in the accompanying manuscript (Novitsch et al., 2001), the domain of Olig2 expression precisely corresponds to the pMN domain, which overlies the Nkx2.2⁺ p3 domain. In some cases, a low level of Nkx2.2 expression can be observed to extend dorsally into the pMN domain (B. Novitsch, personal communication). Beginning on E5, however, a few cells in the ventricular zone at the pMN/p3 interface are observed to coexpress the two transcription factors at high levels (Figure 2C, arrow). The extent of this coexpression remains largely unchanged at E6 (Figure 2D, arrow), but subsequently expands so that by E7, the majority of Olig2⁺ cells coexpress Nkx2.2 at levels comparable to that in p3 (Figure 2E, arrow; yellow cells). (There are, however, one or two layers of Olig2⁺ cells at the dorsal-most aspect of pMN, which do not express detectable levels of Nkx2.2 at all stages examined.)

In addition to this ventricular coexpression, streams of Olig2⁺, Nkx2.2⁺ cells appear to delaminate and migrate away from the ventricular foci where Olig2 and Nkx2.2 overlap (Figure 2E, white arrowheads), as well as perhaps from the Nkx2.2⁺ domain. Nkx2.2⁺, Olig2⁻ cells are also observed to migrate from the Nkx2.2-expressing zone (Figure 2E, open arrowheads), beginning on around E5 (Figure 2C, open arrowheads) and continuing until the latest stage examined (E12; data not shown). By E8.0, the number of migrating Olig2⁺Nkx2.2⁺ and Nkx2.2⁺Olig2⁻ cells has increased dramatically and many of the cells have moved into the ventral white matter (Figure 2F, lower arrowheads). Strikingly, at these stages, no migrating Olig2⁺Nkx2.2⁻ cells were observed in either the gray or white matter.

In mouse, Olig2⁺ cells migrating from the ventricular zone coexpress oligodendrocyte precursor markers such as PDGFR α and Sox10 (Lu et al., 2000; Zhou et al., 2000). The migrating Olig2⁺Nkx2.2⁺ and Nkx2.2⁺Olig2⁻ cells in chick spinal cord have a similar distribution, and are therefore likely to be oligodendrocyte precursors. Consistent with this, some of the migrating Olig2⁺ cells are actively dividing (data not shown), and the only proliferating cells in the spinal cord at these late stages are glial precursors (Altman and Bayer, 1984). To confirm this identification, E8.5 chick spinal cords were dissoci-

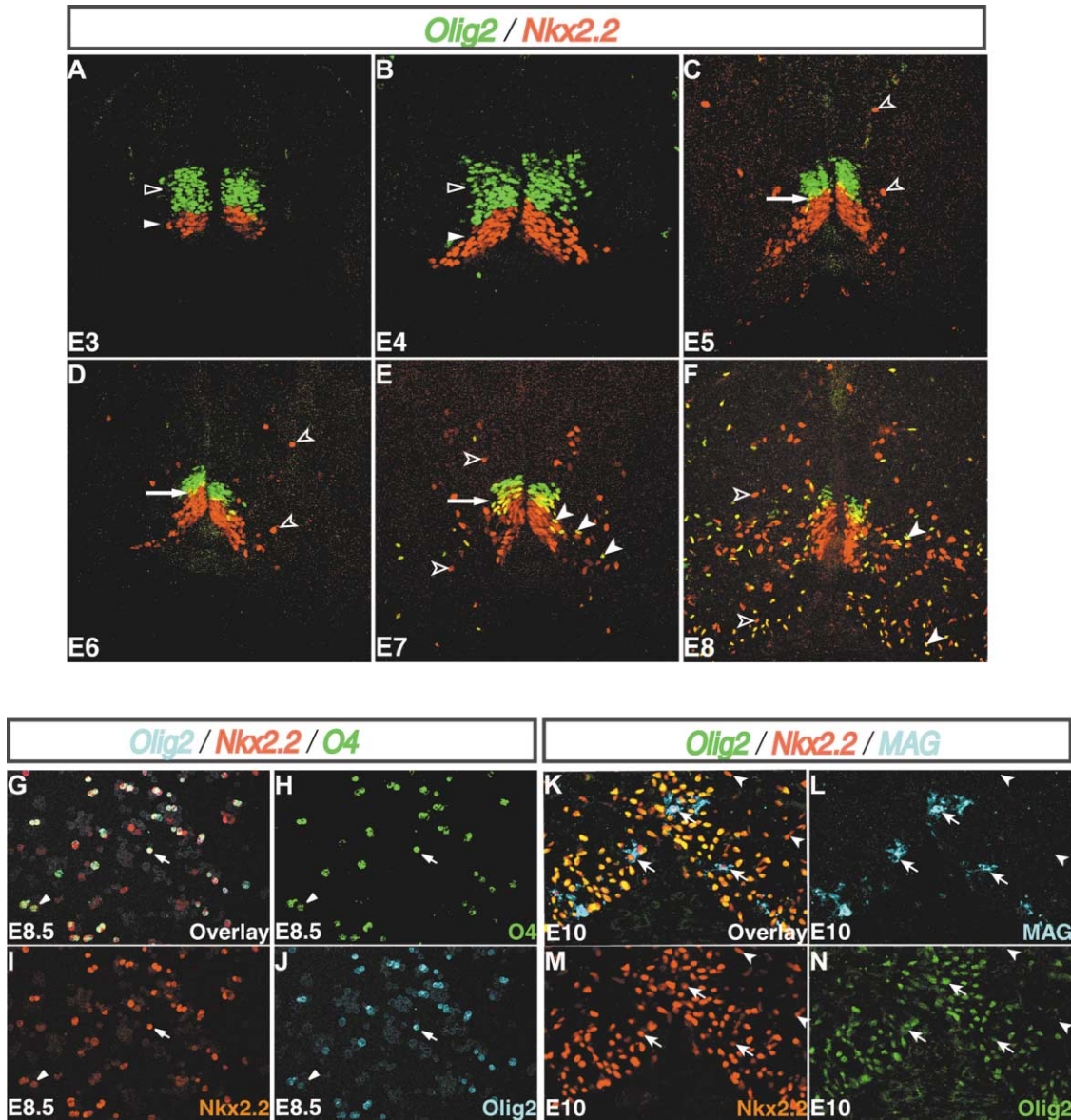


Figure 2. Coexpression of Olig2 and Nkx2.2 in Oligodendrocyte Precursors

(A–F) Double label immunofluorescence confocal microscopy was performed with rabbit anti-Nkx2.2 (red) and mouse anti-Olig2 (green) antibodies at the indicated stages. Double-positive cells (yellow) are first detected at the pMN/p3 boundary at E5.0 (C, arrow). By E7.0, double-positive cells appear to be emigrating in streams from the ventricular zone into the gray matter (E, filled arrowheads), where they later disperse and localize to the white matter (F, lower filled arrowheads). Nkx2.2 single-positive cells begin emigrating at E5.0 (C–F, open arrowheads), but Olig2 single-positive cells are never observed to migrate. (G–J) Olig2⁺, Nkx2.2⁺ cells are oligodendrocyte precursors. E8.5 spinal cord was dissociated and triple labeled with antibodies to O4, Olig2, and Nkx2.2. The four panels are from the same field. Most or all (98 ± 1.4%) Olig2⁺, Nkx2.2⁺ cells coexpress O4 (arrow). A subset of Nkx2.2⁺ cells are weak or negative for Olig2, and some of these (20.5 ± 9.3%) coexpress O4 (arrowhead). (K–N) Expression of a mature oligodendrocyte marker by Olig2⁺, Nkx2.2⁺ cells in white matter. Triple label immunofluorescence on sections through the ventral region of E10 spinal cord was performed using antibodies to Olig2, Nkx2.2 and myelin-associated glycoprotein (MAG). The four panels are from the same field. Note that all MAG⁺ cells are Olig2⁺, Nkx2.2⁺ (arrows), while Nkx2.2⁺, Olig2⁻ cells are MAG⁻ (arrowhead). At later stages, virtually all Nkx2.2⁺ cells are Olig2⁺ (not shown).

ated and triple-labeled with antibodies to Olig2, Nkx2.2, and the oligodendrocyte precursor cell surface marker O4 (Gard and Pfeiffer, 1990). The results indicated that most (98 ± 1.4%) Olig2⁺, Nkx2.2⁺ cells were also O4⁺ (Figures 2G–2J, arrow). In addition, some (20.5 ± 9.3%) Nkx2.2⁺ cells that were weak or negative for Olig2 expressed O4 (Figures 2G–2J, arrowhead). Furthermore, at later stages (E10), at least some Olig2⁺Nkx2.2⁺ cells in

the white matter coexpress the mature oligodendrocyte marker myelin-associated glycoprotein (MAG) (Philippe et al., 1986) (Figures 2K–2M, arrows). Taken together, these data suggest that from E7.0 onward, both Olig2⁺Nkx2.2⁺ and Nkx2.2⁺Olig2⁻ cells in the ventral spinal cord are likely to be oligodendrocyte precursors. Our results are consistent with recent studies showing that Nkx2.2 is expressed by oligodendrocyte precursors (Xu et al.,

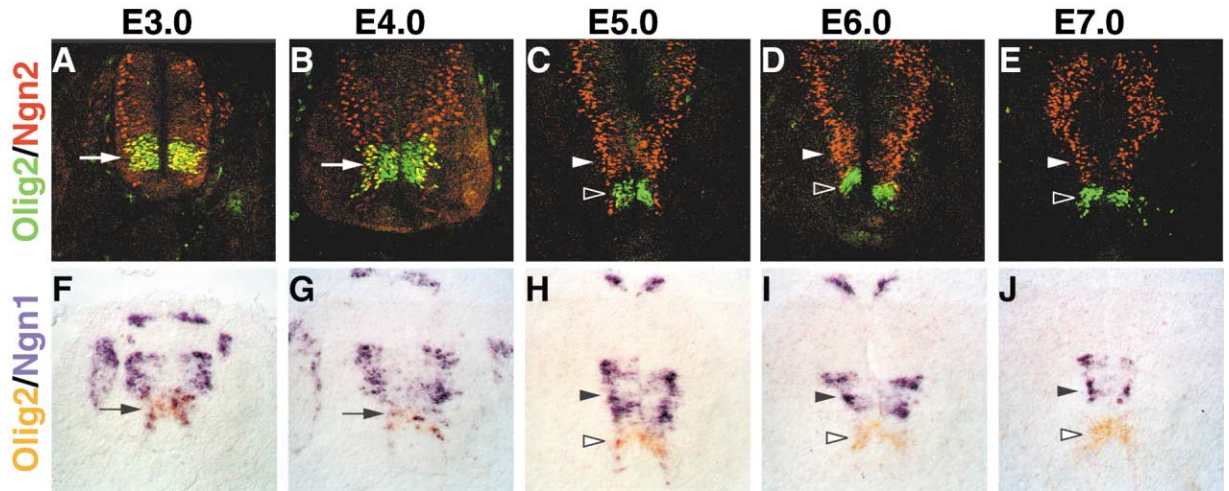


Figure 3. Extinction of Ngns from the Olig2-Expressing Domain Precedes Oligodendrocyte Precursor Formation

(A–E) Double label immunofluorescence microscopy with rabbit anti-Ngn2 (red) and mouse anti-Olig2 (green) antibodies. Arrows in (A) and (B) indicate double-positive (yellow) cells. Note that by E5.0 (C), most Ngn2 expression has already been extinguished from the Olig2⁺ domain (open arrowhead). (F–J) Double label in situ hybridization with Olig2 and Ngn1 cRNA probes. The domains become well separated by E6.0 (I). Note the gap between the dorsal boundary of Olig2 (J, white arrowhead) and the ventral boundary of Ngn1 (J, black arrowhead) at E7.0.

2000; Soula et al., 2001), but indicate that these precursors are heterogeneous with respect to Olig2 coexpression.

Emigration of Olig2⁺ Oligodendrocyte Precursors from the Ventricular Zone Is Preceded by Extinction of Neurogenin Expression

The observation that Olig2 is expressed in migrating and dividing presumptive oligodendrocyte precursors seems paradoxical, in light of the demonstration that this bHLH factor functions to promote motoneuron differentiation and cell cycle arrest at earlier stages of spinal cord development (Novitch et al., 2001). The induction of motoneuron differentiation by Olig2 in part involves the derepression of *Ngn1* and *Ngn2*, vertebrate proneural genes that activate a program of generic neuronal differentiation and cell cycle withdrawal in uncommitted precursor cells (Ma et al., 1996, 1998; Fode et al., 1998; Farah et al., 2000). We were therefore interested to know whether *Ngn1/2* expression is maintained at later stages when oligodendrocytes are being generated, or rather is extinguished at that time.

Double labeling with antibodies to Ngn2 and Olig2 revealed a strong colocalization of these bHLH proteins in the pMN domain at E3.0–E4.0 (Figures 3A and 3B, arrows; see also Novitch et al., 2001). A similar overlap with Ngn1 in this region was detectable by in situ hybridization (Figures 3F and 3G, arrows), although single cell resolution was more difficult to achieve with the nonfluorescent probes. Between E4.5 and E5.0, however, there was a striking downregulation of Ngn2 within the Olig2⁺ region of the ventricular zone, although a few Ngn2⁺ cells persisted laterally to the pMN domain (Figures 3C and 3D, open arrowheads). By E7.0, the stage at which the first Olig2⁺ oligodendrocyte precursors emigrate from the ventricular zone, Ngn2 expression was no longer detectable either within, or lateral to, the Olig2⁺ domain of the ventricular zone (Figure 3E, open arrowhead). However, it was still expressed immediately dor-

sal to this region (Figure 3E, closed arrowhead). A clear separation between the domains of *Ngn1* and *Olig2* mRNA expression was visible by this stage as well (Figure 3J, arrowheads). Thus, the emigration of Olig2⁺ oligodendrocyte precursors from the ventricular zone is preceded by the extinction of *Ngn1* and *Ngn2* expression within the *Olig2*-expressing domain, as well as by the acquisition of overlap with the Nkx2.2⁺ domain (Figure 2).

Comisexpression of Olig2 and Nkx2.2 Cell-Autonomously Induces Ectopic and Precocious Oligodendrocyte Differentiation

The coincidence of Olig2 and Nkx2.2 expression within the ventricular zone, together with the observation that migrating Olig2⁺Nkx2.2⁺ oligodendrocyte precursors are derived from this domain of overlap, suggested that the combined activity of both transcription factors might be important for the specification and/or differentiation of oligodendroglia. We therefore tested whether comisexpression of these two genes could promote ectopic oligodendrocyte generation. The spinal cord of E2 chick embryos was electroporated in ovo with replication-competent RCASBP(B) retroviral vectors (Morgan and Fekete, 1996) harboring the chick *Olig2* or *Nkx2.2* genes, either singly or in combination. The embryos were harvested 4 days later (E6), at which time endogenous oligodendrocyte precursors are just beginning to appear in the spinal cord, but when overt oligodendrocyte differentiation has not yet occurred (Ono et al., 1995) (Figures 1A–1E).

Coelectroporation of *Olig2* plus *Nkx2.2* yielded ectopic and precocious oligodendrocyte differentiation on the electroporated side of the spinal cord, as assessed by the robust induction of Sox10, PDGFR α , MBP, and PLP/DM20 in embryos harvested at E6 (Figures 4E, 4I, and 4M, arrows; and data not shown). Many of these ectopic oligodendrocytes appeared to be migrating away from the ventricular zone toward the white matter.

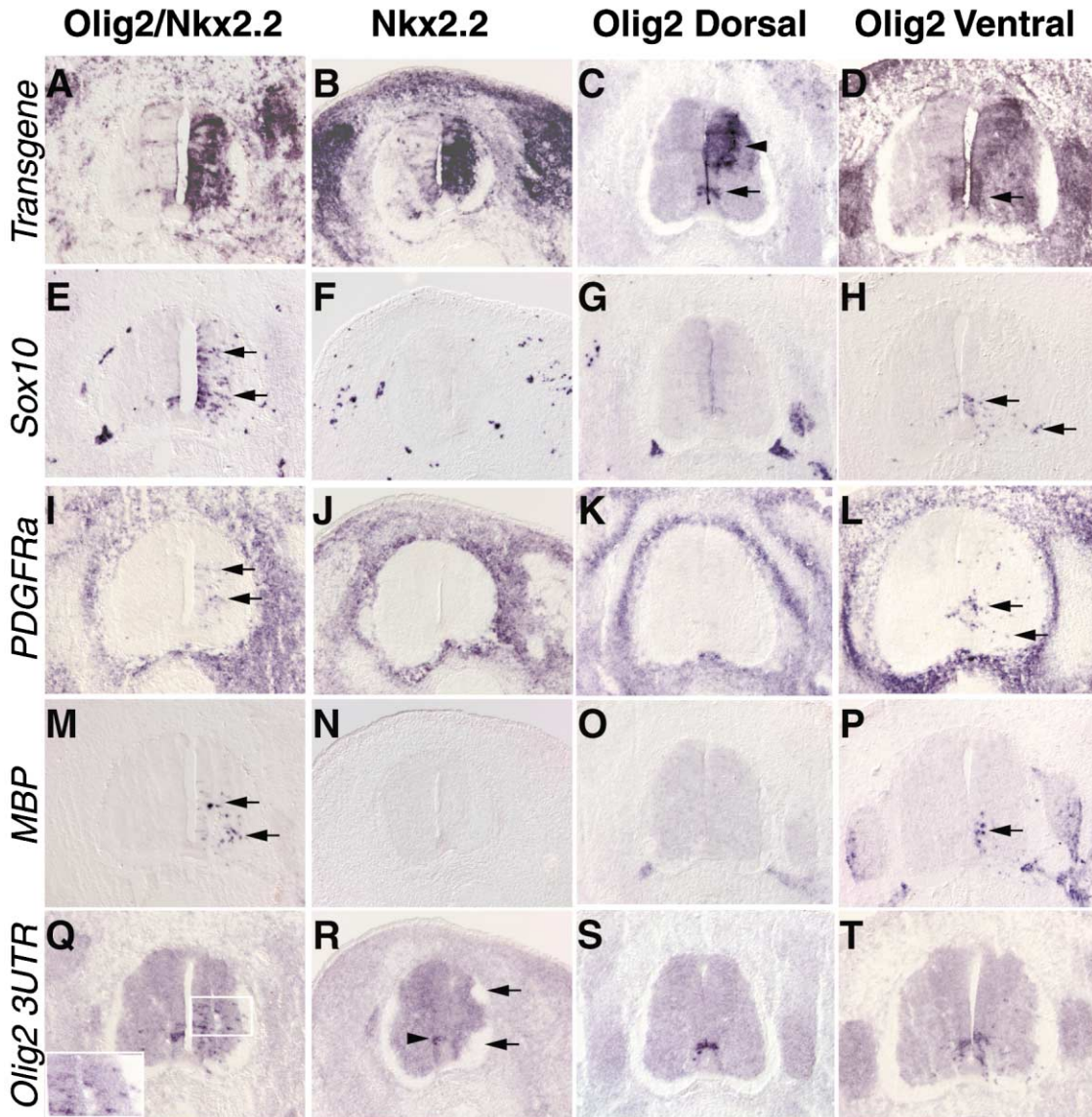


Figure 4. Coelectroporation of Olig2+Nkx2.2 Promotes Ectopic and Precocious Oligodendrocyte Differentiation

E2 embryos were electroporated with the RCASBP(B) plasmids indicated above the columns, harvested 4 days later (E6) and hybridized in situ with the probes indicated to the left of the rows. Note the strong induction of Sox10, PDGFR α , and MBP in embryos coelectroporated with Olig2+Nkx2.2 (E, I, and M, arrows). At this stage (E6), none of these oligodendrocyte markers is normally expressed in the oligodendrocyte precursor region (Figures 2C–2E), thus their induction by Olig2+Nkx2.2 is precocious as well as spatially ectopic. Neither Olig2 alone nor Nkx2.2 alone induces such oligo markers in the dorsal region (F, G, J, K, N, and O). However, electroporation of Olig2 in the ventral spinal cord overlapping the endogenous Nkx2.2⁺ domain results in premature differentiation of oligodendrocytes in this region (H, L, and P, arrows). Nkx2.2 alone, in contrast, potentially represses endogenous Olig2 expression (R, arrows). However, this repression is overcome by inclusion of exogenous Olig2 (Q, boxed region and inset).

In contrast, neither Olig2 nor Nkx2.2 alone caused such ectopic oligodendrocyte differentiation in the dorsal spinal cord (Figures 4B, 4C, 4F, 4G, 4J, 4K, 4N, and 4O). However, in cases where the Olig2 transgene was electroporated in the ventral region of the spinal cord (Figure 4D, arrow), oligodendrocytes were observed to differentiate precociously where the transgene overlapped the domain of endogenous Nkx2.2 expression (Figures 4H, 4L, and 4P, arrows). (The expression of endogenous Nkx2.2 is not affected by ectopic expression of Olig2 in the p3 domain [Novitsch et al., 2001].) Thus, Olig2 can

collaborate with either exogenous or endogenous Nkx2.2 to promote ectopic and/or precocious oligodendrocyte differentiation.

We were not able to determine whether, conversely, exogenous Nkx2.2 can collaborate with endogenous as well as exogenous Olig2 because electroporation of Nkx2.2 in the pMN domain potentially represses endogenous Olig2 expression (Figure 4R, arrows; see also Novitsch et al., 2001). This repression in turn raised the question of how Nkx2.2 and Olig2 can normally be coexpressed in oligodendrocyte precursors (Figure 2). Inter-

estingly, such repression of endogenous *Olig2* was not observed when *Olig2*+*Nkx2.2* were coelectroporated; to the contrary, the combination of these two transcription factors activated ectopic expression of endogenous *Olig2* (Figure 4Q, boxed area and inset). This ectopic expression of the bHLH factor was not activated by forced expression of *Olig2* alone (Figure 4S). Taken together, these data suggest that *Olig2*+*Nkx2.2* in combination activate a program of oligodendrocyte differentiation that is not triggered by either gene alone, and that this interaction converts *Nkx2.2* from a repressor to a coinducer of *Olig2* expression.

In the accompanying paper, Novitch et al. demonstrate that misexpression of *Olig2* alone promotes motoneuron differentiation. This finding raised the possibility that the cells ectopically expressing oligodendrocyte markers in response to combined misexpression of *Olig2*+*Nkx2.2* might coexpress some motoneuron markers, and therefore be phenotypically hybrid cells rather than true oligodendrocytes. To test this, we performed double labeling with motoneuron and oligodendrocyte markers in embryos electroporated with *Olig2*, in either the absence or presence of *Nkx2.2*. Consistent with the results of Novitch et al. (2001), a large number of ectopic motoneurons were generated following electroporation of *Olig2* alone (Figures 5A and 5D, arrowheads), while *Sox10* was not induced. In contrast, both *Sox10*⁺ cells and *MNR2*⁺ or *LIM3*⁺ motoneurons (albeit in smaller numbers) were observed when *Olig2* and *Nkx2.2* were coelectroporated (Figures 5B and 5E). There was, however, little or no overlap between *Sox10*⁺ and *MNR2*⁺ cells, or between *Sox10*⁺ and *Lim3*⁺ cells (Figures 5C and 5F, arrows). Double labeling with the pan-neuronal marker NeuN produced similar results (data not shown). These NeuN⁺ cells likely include some v3 interneurons as well as motoneurons, as coelectroporation of *Olig2*+*Nkx2.2* yields some cells ectopically expressing *Sim1* (not shown), a v3 interneuron marker whose expression is promoted by ectopic *Nkx2.2* expression (Briscoe et al., 1999). Thus, the cells ectopically expressing oligodendrocyte markers in embryos coelectroporated with *Olig2*+*Nkx2.2* are not phenotypic hybrids coexpressing neuronal markers, but rather are glial cells.

If *Olig2*+*Nkx2.2* together promote oligodendrocyte differentiation, why are there any ectopic motoneurons or v3 interneurons in embryos coelectroporated with both transcription factors? In addition to cells ectopically coexpressing *Olig2* and *Nkx2.2*, there are also many cells expressing *Olig2* or *Nkx2.2* alone in such embryos (Figure 5J, right, green and red arrowheads). This may reflect the fact that some cells incorporate only one of the two plasmids during the initial electroporation. In addition, the RCASBP(B) vectors used to express the transgenes encode replication-competent retroviruses (Morgan and Fekete, 1996). Therefore, following electroporation and expression, infectious virus particles will form and neural tube cells will become secondarily infected within 2–3 days (Morgan and Fekete, 1996). Because the *Nkx2.2*- and *Olig2*-expressing viruses are both of the (B) type, they will not coinfect the same cells (Morgan and Fekete, 1996). Thus, cells expressing either *Nkx2.2* or *Olig2*, but not both transgenes, will progressively accumulate at later stages. The existence of such single-positive secondarily infected

cells could, therefore, also explain the intermingling of some ectopic motoneurons and v3 interneurons with the ectopic oligodendrocytes, in *Olig2*+*Nkx2.2*-electroporated embryos.

The induction of ectopic motor and interneurons in embryos coelectroporated with *Olig2*+*Nkx2.2* raised the possibility that induction of ectopic oligodendrocyte differentiation might not reflect a cell-autonomous collaboration between these two transcription factors, but rather a non-cell-autonomous interaction between the ectopic neurons also generated. To address this question, we deliberately coexpressed *Olig2* and *Nkx2.2* in nonoverlapping cell populations. To do this, we packaged infectious B-type retroviral particles expressing either *Olig2* or *Nkx2.2*, and coinjected E2 chick embryos with a 1:1 mixture of these particles, then harvested and analyzed them 4 days later (E6). Double labeling with antibodies to *Nkx2.2* and *Olig2* confirmed that there were many ectopic *Nkx2.2*⁺ or *Olig2*⁺ cells in such coinjected embryos (Figure 5G, red and green arrowheads) compared to controls (e.g., see Figure 5J, left side), but as expected there were very few double-positive cells (Figure 5G, yellow arrowhead). In such coinjected embryos, no ectopic oligodendrocytes were produced (Figures 5H and 5I), whereas in coelectroporated embryos, many such ectopic cells were seen at this time (Figures 5K and 5L, arrows). As expected, in retrovirally infected embryos where the *Olig2* transgene overlapped the endogenous *Nkx2.2*⁺ domain (Figure 5G, yellow arrow), premature oligodendrocyte differentiation was observed (Figures 5H and 5I, arrows). These data suggest that *Olig2* and *Nkx2.2* co-operate in a cell-autonomous manner to promote ectopic oligodendrocyte differentiation. We cannot exclude, however, that feedback signals from motoneurons (and/or v3 interneurons) may contribute to this process, even if they are not sufficient to promote it.

Neurogenins Interfere with the Promotion of Oligodendrocyte Differentiation by *Olig2*+*Nkx2.2*

Recently, *Ngn1* has been shown to negatively regulate glial (astrocyte) differentiation, in addition to promoting neuronal differentiation (Sun et al., 2001). The downregulation of *Ngns1/2* in the *Olig2*⁺ domain prior to the emergence of oligodendrocyte precursors (Figure 3) therefore raised the possibility that these proneural genes might similarly inhibit oligodendrocyte differentiation, and that their downregulation might be required for the specification of these glial cells. Paradoxically, however, misexpression of *Olig2* alone promotes ectopic induction of *Ngns*, as part of its activity to induce motoneuron differentiation (Novitch et al., 2001). As *Nkx2.2* potently inhibits motoneuron differentiation (Ericson et al., 1997), we reasoned that one essential function of *Nkx2.2* in promoting ectopic oligodendrocyte differentiation might be to prevent the induction of *Ngns* by *Olig2*, thereby suppressing motoneuron differentiation. Consistent with this hypothesis, ectopic expression of *Nkx2.2* repressed endogenous *Ngn1* and *2* expression (Figures 6A and 6C, arrow and data not shown). It also prevented the ectopic induction of these proneural genes by *Olig2* (Figures 6B and 6D, arrow and data not shown).

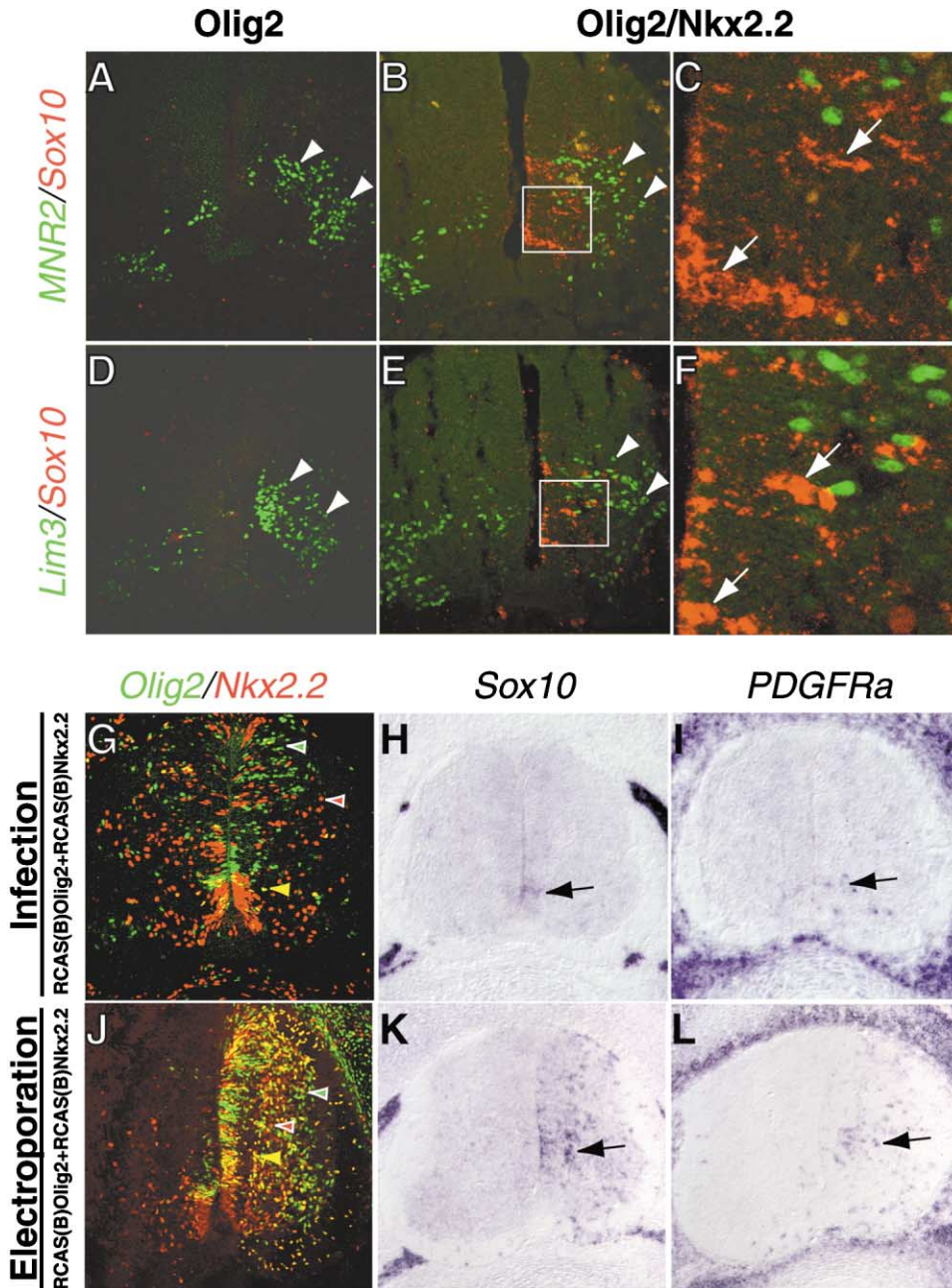


Figure 5. Collaboration between Olig2 and Nkx2.2 Cell-Autonomously Promotes Oligodendrocyte Differentiation at the Expense of Motoneuron Induction

(A–F) Cells expressing the oligo marker Sox10 (C and F, arrows) induced by Olig2+Nkx2.2 do not coexpress the motoneuron markers MNR2 (A and B, green, arrowheads) or Lim3 (D and E, green, arrowheads). The induction of ectopic motoneurons likely reflects cells singly expressing exogenous Olig2 but not Nkx2.2 (Novitsch et al., 2001; see panel [J], green arrowhead). (G–L) Promotion of ectopic oligodendrocytes by Olig2 plus Nkx2.2 is cell autonomous. (G–I) Embryos infected with a 1:1 mixture of RCAS(B)-Olig2 and RCAS(B)-Nkx2.2 retroviruses, which do not coinfect the same cells (G, red and green arrowheads), do not produce ectopic oligodendrocyte differentiation (H and I). However, precocious expression of oligo markers is observed where exogenous Olig2 overlaps the endogenous Nkx2.2⁺ domain (G, yellow arrowhead and H and I, arrows). (J–L) Coelectroporation of RCAS(B)-Olig2 and RCAS(B)-Nkx2.2 plasmids results in coexpression of the two transgenes in many cells (J, yellow arrowhead) and induction of ectopic oligodendrocytes (K and L, arrows).

Given that ectopic coexpression of Olig2+Nkx2.2 indeed represses endogenous *Ngns*, we next asked whether such repression is essential for the induction of ectopic oligodendrocytes. To address this question, we triple-electroporated E2 embryos with *Ngn1*, *Olig2*,

and *Nkx2.2* constructs. In the majority (7/10) of such embryos, the induction of ectopic oligodendrocytes by Olig2+Nkx2.2 was potently suppressed (compare Figures 6F and 6H with Figures 6E and 6G, arrow). To determine whether the repression of *Ngns* was also a

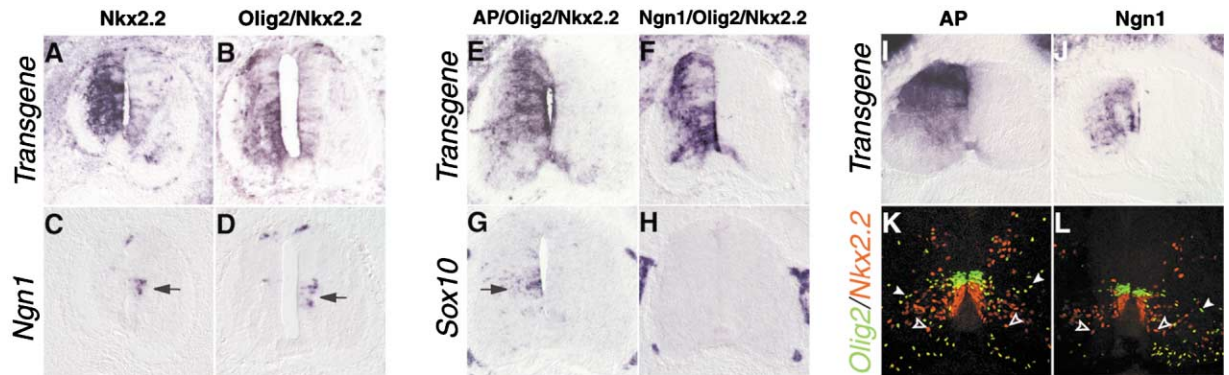


Figure 6. Repression of *Ngn*s Is Required for Both Ectopic and Normal Oligodendrocyte Differentiation

(A–D) *Nkx2.2* potentially represses expression of *Ngn1* (C) and *Ngn2* (not shown), and prevents the induction of ectopic *Ngn1* expression by *Olig2* (D and Novitsch et al., 2001). (E–H) Inclusion of *Ngn1* blocks ectopic oligodendrocyte differentiation induced by misexpression of *Olig2*+*Nkx2.2*, as assayed by induction of ectopic *Sox10* expression (G, arrow and H) and other oligo markers (not shown). AP, control alkaline phosphatase-expressing plasmid. (I–L) *Ngn1* alone blocks emigration of *Olig2*⁺, *Nkx2.2*⁺ oligodendrocyte precursors from their normal domain. Closed arrowheads in (K) and (L) indicate *Olig2*⁺, *Nkx2.2*⁺ cells, open arrowheads indicate *Nkx2.2*⁺, *Olig2*⁻ cells. Note the absence of double-positive cells on the electroporated (left) side of *Ngn1*-expressing embryos (L). (A)–(J) are in situ hybridizations, (K) and (L) double label immunofluorescence.

prerequisite for the specification of oligodendrocyte precursors in their normal location, we electroporated E2 embryos with the *Ngn1*-expressing vector alone (Figure 6J). In such embryos, most migrating oligodendrocyte precursor cells, identified by coexpression of endogenous *Olig2* and *Nkx2.2* (Figure 6K, solid arrowheads), were missing on the electroporated side (Figure 6L, left side), as were *Olig2*⁺*Nkx2.2*⁺ cells that had settled in the white matter. No such reduction in migrating *Olig2*⁺, *Nkx2.2*⁺ cells was observed in embryos electroporated with a control alkaline phosphatase (AP)-encoding construct (Figures 6I and 6K, filled arrowheads). Taken together, these data indicate that downregulation of *Ngn*s is an essential prerequisite for both normal oligodendrocyte differentiation, as well as for ectopic oligodendrocyte differentiation elicited by coelectroporation of *Olig2*+*Nkx2.2*.

Misexpression of *Olig2* in the Absence of *Ngn*s and *Nkx2.2* Promotes Development of Oligodendrocyte Precursors

The foregoing data implied that one essential function of *Nkx2.2* in collaborating with *Olig2* is to repress endogenous *Ngn*s, which otherwise interfere with oligodendrocyte precursor specification. However, during normal development, the clearance of *Ngn*s from the *Olig2*⁺ domain of the ventricular zone precedes the overlap with *Nkx2.2* in this region (compare Figures 2 and 3). Thus, *Nkx2.2* is unlikely to be responsible for repressing *Ngn*s within the *Olig2*⁺ domain during normal oligodendrocyte development. This observation raised the question of whether the sole function of *Nkx2.2* in collaborating with ectopically expressed *Olig2* is to repress the *Ngn*s and thereby prevent motoneuron differentiation. If this were the case, then *Olig2* should be sufficient to promote ectopic oligodendrocyte differentiation in the absence of *Nkx2.2*, if the expression or function of *Ngn*s could be prevented by other means.

To repress *Ngn* expression and function, we misexpressed components of the Notch signaling pathway

(Artavanis-Tsakonas et al., 1999). Notch signaling is known to be necessary and sufficient to repress expression of *Ngn*s in a number of vertebrate systems (Ma et al., 1996, 1998; de la Pompa et al., 1997). Repression of such proneural genes by Notch involves proteolytic cleavage and nuclear translocation of its intracellular domain (ICD), which then interacts with Suppressor of Hairless (Su(H); RBPJ_c) to promote expression of transcriptional effectors such as *Hes* genes (reviewed in Artavanis-Tsakonas et al., 1999; Mumm and Kopan, 2000). In several systems, Notch^{CD} has been shown to be a constitutively active form of Notch (Struhl et al., 1993; Chitnis et al., 1995). We therefore asked whether repression of *Ngn*s by expression of Notch^{CD} would be sufficient to allow *Olig2* to promote ectopic oligodendrocytes in the absence of exogenous *Nkx2.2*. To ensure that levels of endogenous *Su(H)* were not limiting in this experiment, we coelectroporated a construct expressing that gene as well (Wettstein et al., 1997).

Expression of Notch^{CD}+*Su(H)* indeed repressed *Ngn1* on the electroporated side of the neural tube, in either the absence (Figure 7F, arrow) or presence (Figure 7D, arrow) of coelectroporated *Olig2*. Similar results were obtained for *Ngn2* (data not shown). Such repression was by itself insufficient to promote ectopic oligodendrocyte differentiation (Figures 7I, 7L, and 7O; the small number of bilaterally migrating oligodendrocyte precursors in [L] reflects the fact that these particular embryos were analyzed at E6.5 rather than E6.0). In contrast, when embryos were triple electroporated with Notch^{CD}, *Su(H)* and *Olig2*, ectopic expression of the oligodendrocyte precursor markers *Sox10* and *PDGFR α* was clearly detected in the dorsal neural tube (Figures 7G and 7J, black arrowheads). Positive cells were observed in both the ventricular zone and in the gray matter, suggesting that migration of such ectopic precursors had occurred. However, expression of mature oligodendrocyte markers, such as *MBP*, was not detected (Figure 7M).

These data suggested that *Olig2* is able, in the absence of *Nkx2.2*, to promote ectopic oligodendrocyte

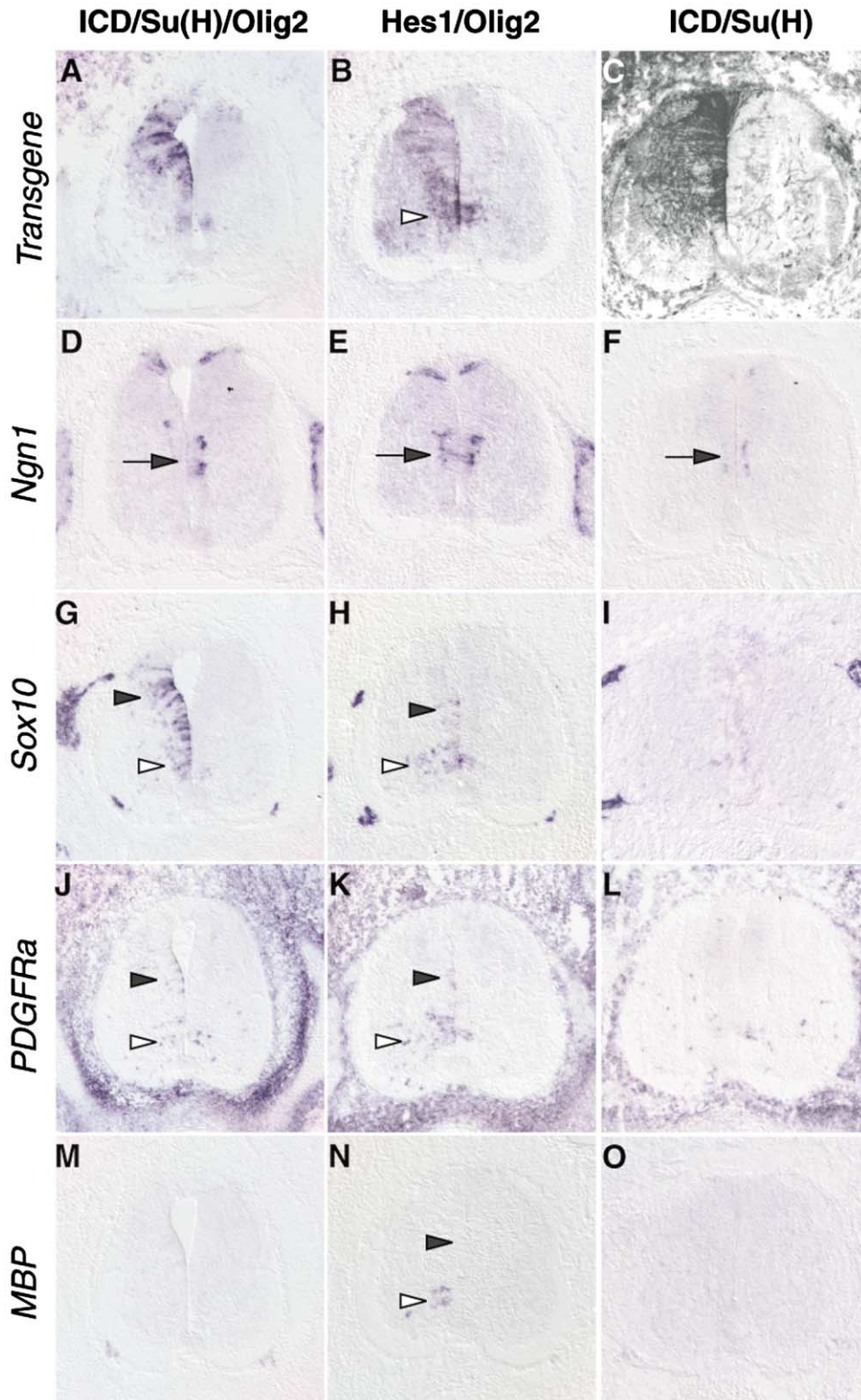


Figure 7. *Olig2* Is Sufficient to Induce Ectopic Oligodendrocyte Precursor Markers, but Not MBP⁺ Oligodendrocytes, in the Absence of *Ngn*s and *Nkx2.2*

Embryos were electroporated with the mixture of plasmids indicated above the columns, and hybridized with the probes indicated next to the rows. The combination of Notch^{CD} (“ICD”) and Su(H) represses expression of *Ngn1* (F, arrow) and *Ngn2* (not shown), but does not induce ectopic oligo precursor markers (I and L) or MBP (O). In the presence of *Olig2*, induction of *Ngn1* (D) and *Ngn2* (not shown) is still blocked, and Sox10 (G, closed arrowheads) and PDGFR α (J, closed arrowheads) are ectopically induced. However, no MBP is detected in the dorsal

precursor specification, but not differentiation, if endogenous *Ngns* are repressed. The lack of ectopic MBP⁺ cells under these circumstances might, therefore, reflect an additional requirement for Nkx2.2 in oligodendrocyte differentiation, independent of its function to repress *Ngns*. Consistent with this, Olig2 alone promoted precocious differentiation of MBP⁺ cells when expressed in the endogenous Nkx2.2⁺ p3 domain, where *Ngns* are normally not expressed (Figure 3 and Figure 4P). Alternatively, the lack of ectopic mature oligodendrocyte markers in embryos coelectroporated with Olig2 and Notch^{ICD}+Su(H) might instead reflect an effect of constitutive Notch activation to inhibit terminal differentiation of oligodendrocyte precursors, as described in other studies (Wang et al., 1998).

To try to distinguish these alternatives, we sought a less potent way of mimicking constitutive Notch activation. To this end, we employed *Hes1* (Akazawa et al., 1992), a downstream transcriptional effector of Notch that mediates some, but not all, of its biological actions (Ishibashi et al., 1995; Kageyama and Nakanishi, 1997). Consistent with this assumption, electroporation of *Hes1* in either the presence or absence of exogenous *Olig2* produced a detectable, but less pronounced, repression of *Ngn1* (Figure 7E, arrow and data not shown) than was observed using Notch^{ICD}+Su(H) (cf. Figure 7D, arrow). Despite this relatively modest effect on *Ngn* expression, coelectroporation of *Olig2*+*Hes1* also promoted ectopic expression of Sox10 and PDGFR α (Figures 7H and 7K, black arrowheads). Electroporation of *Hes1* alone did not have this effect (not shown). Moreover, in those embryos where *Hes1*+*Olig2* were coelectroporated in the Nkx2.2⁺ ventral region of the neural tube (Figure 7B, open arrowhead), not only migrating oligodendrocyte precursors (Figures 7H and 7K, open arrowheads), but also MBP⁺ oligodendrocytes (Figure 7N, open arrowhead) were observed precociously. In contrast, ectopic MBP⁺ cells were not detected when exogenous *Olig2* and *Hes1* were coexpressed in more dorsal regions where endogenous *Nkx2.2* is not expressed (Figure 7N, black arrowhead). These data suggest that exogenous *Hes1* does not inhibit precocious expression of mature oligodendrocyte markers promoted by exogenous *Olig2* plus endogenous Nkx2.2. Taken together, these results suggest that repression of *Ngn* expression by activated Notch (or its downstream effectors) allows specification of ectopic oligodendrocyte precursors by *Olig2*. Progression of such precursors to MBP⁺ oligodendrocytes may require additional functions provided by Nkx2.2, as well as abrogation of Notch signaling (Wang et al., 1998).

Olig2 and Nkx2.2 Both Function as Transcriptional Repressors in Promoting Oligodendrocyte Differentiation

Basic helix-loop-helix proteins of the “B” class, such as MyoD, typically function as transcriptional activators

(Weintraub et al., 1991). Surprisingly, fusion of *Olig2* to the yeast Gal4 DNA binding domain indicated that it acts instead as a repressor (Novitsch et al., 2001). Consistent with this observation, a fusion of the *Olig2* bHLH domain with the transcriptional repressor domain of *Drosophila* engrailed (*Olig2*-EnR) mimics the activity of intact *Olig2* to promote motoneuron differentiation (Novitsch et al., 2001). Conversely, an activator form of *Olig2*, *Olig2*-VP16, antagonized motoneuron differentiation (Novitsch et al., 2001). These results led us to test whether *Olig2*-EnR or *Olig2*-VP16 would phenocopy the activity of intact *Olig2* to promote ectopic oligodendrocyte differentiation in combination with Nkx2.2. We found that the *Olig2*bHLH-EnR fusion phenocopied the intact protein (Figures 8A, 8D, and 8G), while the *Olig2*-VP16 fusion did not (data not shown). Such an activity was not observed by coelectroporating either a construct containing just the *Olig2* bHLH domain, or a *Phox2a*-EnR fusion, together with Nkx2.2 (data not shown). Thus the effect of *Olig2*-EnR requires both the EnR domain, and correct DNA binding specificity conferred by the *Olig2* bHLH domain. In support of the latter conclusion, a full-length *Olig2* construct lacking the basic region was inactive in this assay, confirming that the activity of this protein requires DNA binding.

We also examined whether the *Olig2*bHLH-EnR fusion could function in the absence of Nkx2.2 to specify oligodendrocyte precursors, using Notch^{ICD}+Su(H) to repress endogenous *Ngns*. Triple electroporation of these constructs indeed promoted formation of ectopic Sox10⁺ oligodendrocyte precursors (Figure 8F, arrow). These results suggest that *Olig2* also acts as a repressor in its Nkx2.2-independent oligodendrocyte precursor-specification function. Importantly, in such triply electroporated embryos, precocious MBP⁺ oligodendrocytes were still detected in the ventral region where endogenous Nkx2.2 is expressed (Figure 8I, open arrowhead), but not in more dorsal regions (Figure 8I, arrow). These data provide further evidence that the inability of *Olig2* alone to promote expression of mature oligodendrocyte markers reflects a requirement for Nkx2.2 in this later differentiation step, and not simply an inhibition of such differentiation by constitutive Notch activation. Nevertheless, we cannot exclude that exogenous *Olig2* promoted MBP expression in the Nkx2.2⁺ domain in only a subset of cells which did not coexpress Notch^{ICD} and/or Su(H).

Previous studies have shown that Nkx2.2 functions as a repressor at earlier stages of neural tube development when it promotes v3 interneuron specification (Muhr et al., 2001). We were therefore curious to know whether Nkx2.2 similarly functions as a repressor in the ectopic oligodendrocyte differentiation assay. When coelectroporated with either the *Olig2*bHLH-EnR fusion (Figures 8B, 8E, and 8H) or intact *Olig2* (not shown), an Nkx2.2 homeodomain-EnR fusion (Nkx2.2EnR; Muhr et al., 2001) also promoted ectopic oligodendrocyte differenti-

domain (M). Similar results were obtained using *Hes1* instead of ICD+Su(H) (E, H, K, and N), although the extent of *Ngn* repression was not as pronounced (E, arrow). Note that in the region where exogenous *Hes1*+*Olig2* overlap the ventral domain where endogenous Nkx2.2 is expressed (B, open arrowhead; cf. Figure 1B), not only oligo precursor markers (H and K, open arrowheads), but also MBP (N, open arrowhead) are expressed. This suggests that *Hes1* does not block induction of MBP by the combination of exogenous *Olig2* plus endogenous Nkx2.2.

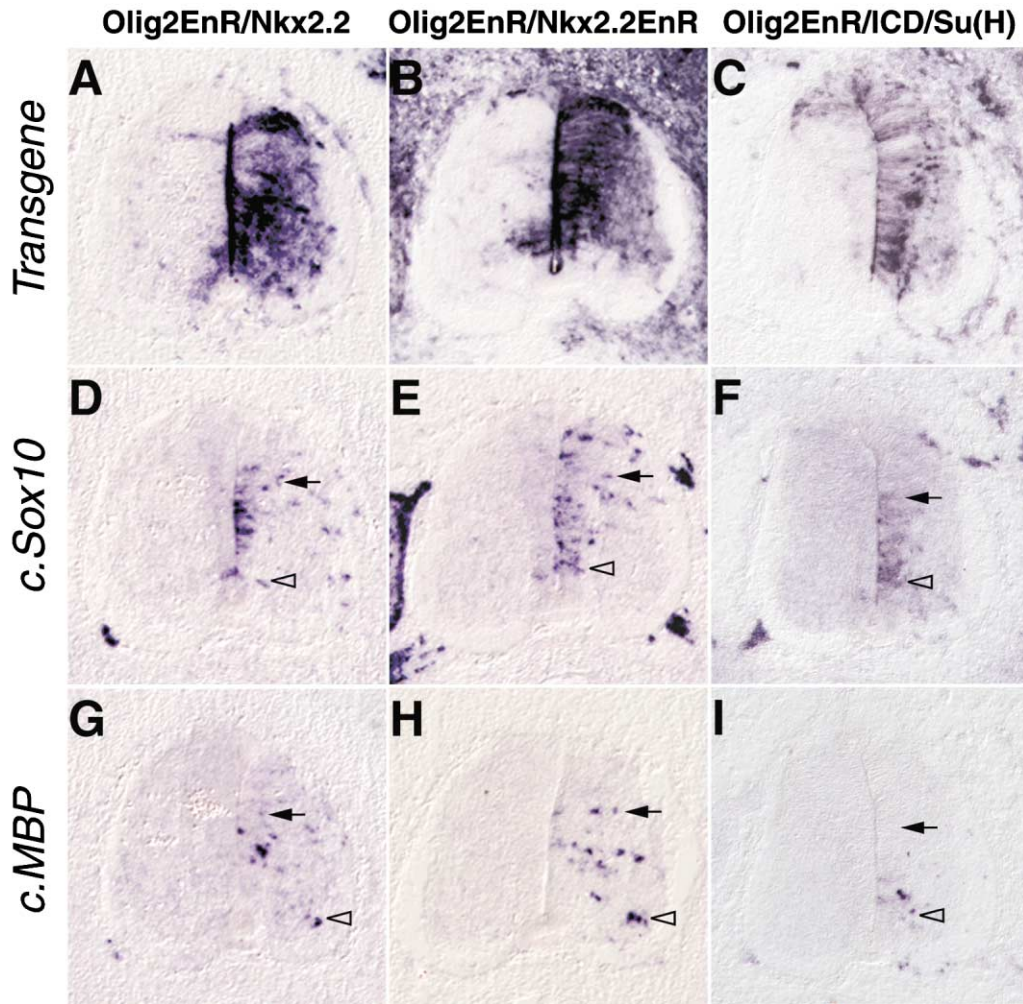


Figure 8. Olig2 and Nkx2.2 Act as Repressors in Promoting Ectopic Oligodendrocyte Differentiation

Coelectroporation of an Olig2bHLH-EnR fusion with either intact Nkx2.2 (A, D, and G) or an Nkx2.2 homeodomain-EnR fusion (Nkx2.2HD-EnR) (B, E, and H) results in ectopic expression of both Sox10 (D and E) and MBP (G and H). Similar results were obtained when EnR fusions of both proteins were coelectroporated (not shown). (C, F, and I) Triple electroporation of Olig2bHLH-EnR+ICD+Su(H) induces ectopic and precocious expression of Sox10 dorsally (F, arrow), but dorsal expression of MBP is not detected (I, arrow). However, precocious MBP expression is detected on the electroporated side in the ventral region where the transgenes overlap the domain of endogenous Nkx2.2 expression. This result again suggests that Nkx2.2 is required for progression to mature oligodendrocytes.

ation. In contrast, the corresponding activator form, Nkx2.2-VP16, did not promote ectopic oligodendrocytes when coelectroporated with full-length Olig2 (data not shown). Thus, EnR fusion proteins phenocopied the activities of both intact Olig2 and Nkx2.2 in this assay. This suggests that both Olig2 and Nkx2.2 function as repressors, rather than as activators, in promoting ectopic oligodendrocyte differentiation.

Olig2-VP16 Inhibits Oligodendrocyte Formation

The finding that Olig2bHLH-EnR phenocopies the activity of intact Olig2 suggested in turn that the corresponding activator form, Olig2bHLH-VP16, might conversely function as a dominant inhibitor of endogenous Olig2 function. Consistent with this notion, the Olig2bHLH-VP16 fusion suppresses motoneuron differentiation (Novitsch et al., 2001). Accordingly, we tested whether this Olig2bHLH-VP16 fusion would also inhibit oligodendrocyte precursor formation. Electroporation of this con-

struct caused an approximately 40% reduction in the average number of Olig2⁺, Nkx2.2⁺ oligodendrocyte precursors in gray and white matter, in embryos harvested at E8 (Figures 9A, 9B, arrowheads, and 9D). There was also a reduction in the number of PDGFR α ⁺ cells on the electroporated side (Figure 9C, arrowheads). By contrast, the number of Nkx2.2⁺, Olig2⁻ progenitors was unaffected (Figure 9D). (Since such single-positive cells constitute only ~25% of the oligodendrocyte precursor population at this stage, their persistence does not mask the reduction of PDGFR α ⁺ cells.)

In addition, among cells in the gray or white matter expressing the Olig2-VP16 transgene, only 8 \pm 2% coexpressed endogenous Olig2. In contrast, cells coexpressing endogenous Olig2 and the Olig2-VP16 transgene could be easily observed in the ventricular zone (Figure 9A, yellow arrow). These data suggest that the reduction in Olig2⁺ cells in the gray and white matter reflects an inhibition of oligodendrocyte precursor generation or

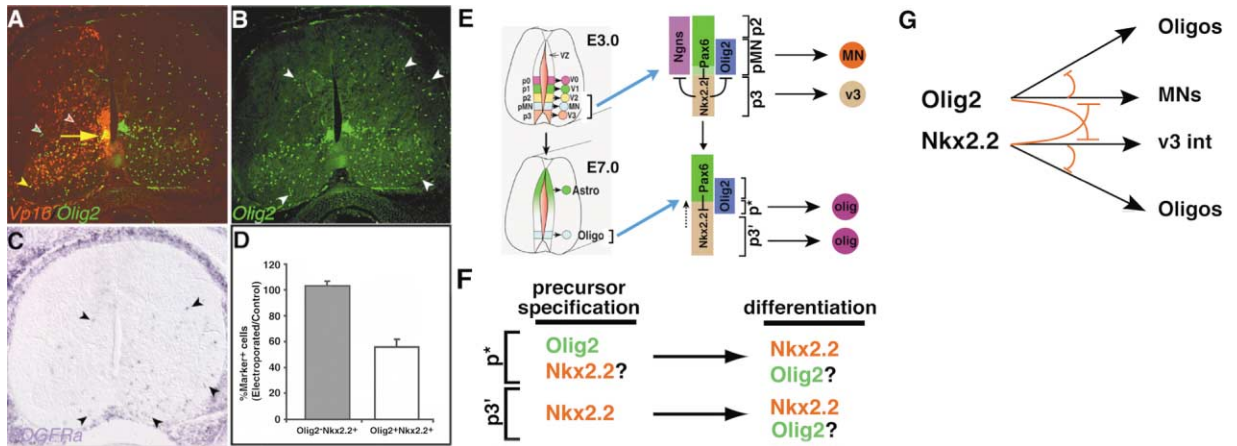


Figure 9. Olig2-VP16 Inhibits Oligodendrocyte Precursor Formation

(A)–(C) show sections from an embryo electroporated with an Olig2-VP16 fusion at E2 and harvested at E8. Migrating oligo precursors are marked by expression of Olig2 (B, arrowheads). Olig2-VP16 causes a reduction in the number of migrating Olig2⁺ cells (A, green arrowheads and B), as well as of PDGFR α ⁺ cells (C) on the electroporated (left) side relative to the contralateral control (right) side. Quantification (D) shows an approximately 40% reduction in the number of Olig2⁺, Nkx2.2⁺ oligodendrocyte precursors (white bar) on the electroporated versus the contralateral control side. In contrast, the number of Olig2⁻, Nkx2.2⁺ cells (shaded bar) is unchanged. Numbers represent the average of six sections from a single experiment; the experiment was performed twice with similar results. (E) Summary of spatio-temporal changes in progenitor domains and their relationship to oligodendrocyte production. Between E3.0 and E7.0, the ventral-most expression domain of Pax6 (light green) disappears, and Nkx2.2 expands dorsally into this region to overlap Olig2. In this way, a new progenitor domain (p*) becomes intercalated dorsally to the erstwhile p3 domain (p3'). (F) Olig2 and Nkx2.2 may act simultaneously and/or sequentially in precursor specification and differentiation. The ventral-most oligodendrocyte precursors that emerge from p3' initially express Nkx2.2 but not Olig2. However, these cells appear to subsequently coexpress Olig2 in white matter. (G) Cooperation of Olig2 and Nkx2.2 in p*-derived oligodendrocyte precursors may involve mutual antagonism of the neuron-promoting activities of each factor. Activation of motoneuron (MN) and v3 interneuron differentiation by Olig2 and Nkx2.2, respectively, is assumed to be mutually exclusive with oligodendrocyte differentiation. Note that this model applies only to p*-derived oligodendrocyte precursors which coexpress Nkx2.2 and Olig2.

survival, and not simply negative regulation of endogenous *Olig2* expression by Olig2-VP16. The inhibition of oligodendrocyte formation obtained with Olig2-VP16 was not observed using just the bHLH domain of Olig2, with the Olig2-EnR fusion, or with a Phox2a-VP16 fusion (not shown). Thus the inhibition reflects transcriptional activation functions conferred by the VP16 domain, and not simply the deletion of the Olig2 domain that it replaces.

That oligo precursor formation was not completely inhibited by the Olig2bHLH-VP16 fusion likely reflects the fact that some ventricular zone cells escaped the initial electroporation. Consistent with this interpretation, most (~66%) of the Olig2⁺ precursors that did migrate on the electroporated side did not coexpress the Olig2-VP16 transgene (Figure 9A, green arrowheads), although a few did (yellow arrowhead). These latter cells may represent precursors that expressed the Olig2-VP16 fusion only after they had committed to the oligodendrocyte fate, perhaps because they acquired the transgene by secondary infection rather than during the initial electroporation. The increasing proportion of such secondarily infected cells, and our inability to distinguish them from the cells that initially incorporated the transgene by electroporation, precluded an analysis at later stages (E9.5 and older) when mature oligodendrocyte markers such as MBP are first expressed (Figure 1T). Thus these experiments do not address whether Olig2-VP16 inhibits oligodendrocyte maturation. However, they do indicate that this activator form of Olig2 inhibits earlier stages of oligodendrocyte precursor formation or survival.

Discussion

Progenitor cells in the ventricular zone of the ventral spinal cord undergo a developmental transition from generating neurons to producing oligodendrocytes (Sun et al., 1998; Richardson et al., 2000). The evidence presented in this and in the companion paper (Novitsch et al., 2001) together indicate that the same bHLH transcription factor, Olig2, plays a determinative and sequential role in both motoneuron and oligodendrocyte fate specification. This dual function is controlled by spatio-temporal changes in the domains of expression of several other transcription factors in relation to the domain of *Olig2* expression. These changes include both the extinction of *Ngn1* and *2* expression within the Olig2⁺ domain and the acquisition of a partial overlap with the underlying Nkx2.2⁺ domain. That such changes in expression are functionally important is suggested by the fact that Olig2 can collaborate with Nkx2.2 to promote ectopic oligodendrocyte differentiation, and that this effect is blocked by constitutive expression of Ngn1. In this way, the landscape of gene expression in the spinal cord ventricular zone changes with time, to enable sequential utilization of common transcription factors in the production of different cell types.

Multiple Populations of Oligodendrocyte Precursors

Our double labeling experiments with antibodies to Olig2 and Nkx2.2 suggest the existence of at least two different oligodendrocyte precursor populations, with distinct spatial origins in the chick ventricular zone. One population coexpresses Olig2 and Nkx2.2, and emigrates from

the zone of overlap between these two factors, while the other population expresses *Nkx2.2* but not *Olig2*, and emigrates more ventrally (Figures 9E and 9F). These data are surprising since there was no reason previously to think that spinal cord oligodendrocyte precursors were anything other than a homogenous population.

The observation of oligodendrocyte precursor heterogeneity has several important implications. First, it raises the question of whether it serves to generate different subpopulations of oligodendrocytes with distinct functional properties. Second, it suggests that some oligodendrocyte precursors (i.e., those that are *Nkx2.2*⁺ and *Olig2*⁻) can be specified in the absence of *Olig2*. How this occurs is not clear; however, we note that *Olig3*, which is closely related to *Olig2*, is coexpressed with *Nkx2.2* in the p3 domain (data not shown). Third, it may help to resolve disagreements about whether oligodendrocyte precursors arise from the erstwhile motoneuron progenitor domain (pMN) (Richardson et al., 1997; Sun et al., 1998), or rather the underlying p3 domain (Soula et al., 2001); our discovery raises the possibility that they may arise from both locations (Figure 9E).

Temporal Roles of *Olig2* and *Nkx2.2* in Oligodendrocyte Fate Determination

Our results demonstrate that exogenous *Olig2* and *Nkx2.2* can collaborate to promote ectopic oligodendrocyte development. This functional interaction is well correlated with the fact that coexpression of the endogenous proteins in oligodendrocyte precursors coincides with their emergence from the ventricular zone. Nevertheless, it is not completely clear whether this collaboration reflects a simultaneous, or rather sequential, function for the two transcription factors in oligodendrocyte development (Figure 9F). When misexpressed without *Nkx2.2* in the context of constitutive Notch signaling, *Olig2* is able to ectopically induce early oligodendrocyte precursor markers such as *Sox10* and *PDGFR α* , but not later differentiation markers such as *MBP*. Expression of *MBP* is only detected when *Olig2* is coexpressed with *Nkx2.2* (either exogenous or endogenous). Consistent with these data, misexpression of *Olig2* in chick limb bud (which does not express *Nkx2.2*) induced ectopic expression of *Sox10*, but not of later markers such as *MBP* (Zhou et al., 2000). In addition, expression of *MBP* is lost in *Nkx2.2*^{-/-} mice, while the number of oligodendrocyte precursors does not appear to be diminished (M. Qiu, personal communication). These observations suggest that *Olig2* can on its own induce oligodendrocyte precursor formation under some circumstances, but that *Nkx2.2* is required to permit such precursors to progress to later stages of differentiation. Nevertheless, the observation that some oligodendrocyte precursors are initially *Nkx2.2*⁺ and *Olig2*⁻ raises the possibility that the homeodomain factor may also function in precursor specification in some circumstances (Figure 9F).

Is *Olig2*, conversely, important only for oligodendrocyte precursor specification, or also for later stages of differentiation? *Olig2* expression persists in mature oligodendrocytes (Lu et al., 2000 and this study). In addition, most or all *Nkx2.2*⁺, *Olig2*⁻ oligodendrocyte precursors appear to subsequently express *Olig2* after they settle in the white matter (Supplemental Figure S2G).

Furthermore, at intermediate stages of development, *Nkx2.2*⁺ cells in the white matter that are *Olig2*⁻ do not yet express *MAG*, a marker of mature oligodendrocytes (Figures 2K–2N, arrowheads), while all neighboring cells that express *MAG* are positive for both *Nkx2.2* and *Olig2* (Figure 2K–2N, arrows). This circumstantial evidence is consistent with the idea that *Olig2* may function in oligodendrocyte differentiation as well as in precursor specification. Such a late function may have been obscured in our experiments by the use of constitutive Notch pathway components to examine *Olig2* activities in the absence of *Nkx2.2* (Wang et al., 1998).

One caveat to our conclusion that *Olig2* and *Nkx2.2* collaborate in oligodendrocyte fate determination is that in rats and mice, *Olig2*⁺ oligodendrocyte precursors do not detectably coexpress *Nkx2.2* when they first migrate, although some coexpression within the ventricular zone can be detected (Supplemental Figures S2B–S2D). Nevertheless, many *Olig2*⁺ cells do eventually coexpress *Nkx2.2* after they have settled in the white matter (Supplemental Figure S2F). Thus, the most general implication of our results is that *Olig2* and *Nkx2.2* collaborate in oligodendrocyte fate determination, but that there may be species and/or lineage differences in the precise time at which these two factors are brought into play.

In addition to the collaboration between *Olig2* and *Nkx2.2* and the extinction of *Ngn* expression, there may be other time-dependent contextual changes required for oligodendrocyte generation. This is suggested by the observation that the induction of ectopic oligodendrocyte markers by comisexpression of *Olig2*+*Nkx2.2* is detectable at E6, but not at E4. In contrast, ectopic motoneuron differentiation promoted by *Olig2* alone is already detectable at E4 (Novitsch et al., 2001). Apparently, the dorsal spinal cord is not competent to express oligodendrocyte markers in response to coexpression of *Olig2*+*Nkx2.2* until after E4. This may reflect the fact that neurogenesis continues in the dorsal spinal cord after it has ceased in the motoneuron domain (E5.5–E6), and precludes oligodendrocyte formation. Consistent with this notion, quadruple electroporation of *Olig2*+*Nkx2.2* together with *Notch*^{CD}+*Su(H)* (to inhibit neurogenesis) yielded some ectopic *Sox10*⁺ cells as early as E4.5 (data not shown).

Olig2 and *Nkx2.2* Act as Repressors in Oligodendrocyte Fate Determination

Olig2-EnR, but not *Olig2*-VP16, can substitute for wild-type *Olig2* in collaborating with *Nkx2.2* to promote oligodendrocyte differentiation. These data suggest that *Olig2* acts as a repressor in oligodendrocyte, as well as in motoneuron (Novitsch et al., 2001), fate determination. What genes does *Olig2* repress to promote oligodendrocyte development? As this function involves collaboration with *Nkx2.2*, and as *Nkx2.2* on its own normally promotes v3 interneuron specification, one possibility is that *Olig2* represses this interneuron-promoting function of *Nkx2.2*. Consistent with this idea, induction of ectopic *Sim1*⁺ v3 interneurons by *Nkx2.2* is inhibited by coelectroporation of *Olig2* (data not shown). If the v3 interneuron-promoting function of *Nkx2.2* were incompatible with a concurrent function in gliogenesis, then

antagonism of this neurogenic function by Olig2 might allow Nkx2.2 to promote oligodendrocyte generation (Figure 9G).

Nkx2.2 also appears to function as a repressor in oligodendrocyte differentiation, as it does in v3 interneuron generation (Muhr et al., 2001). One function of this repressor activity may be to antagonize the promotion of motoneuron differentiation by Olig2. Consistent with this, Nkx2.2 strongly inhibits motoneuron differentiation when ectopically expressed in the Olig2⁺ pMN domain (Briscoe et al., 1999, 2000). Thus, one way of viewing the collaboration between Olig2 and Nkx2.2 in oligodendrocyte differentiation is that each functions to antagonize the other's ability to promote differentiation of a particular neuronal subtype. In this way, oligodendrocytes would emerge when the convergent activity of Olig2 and Nkx2.2 prevents differentiation of either motoneurons or v3 interneurons (Figure 9G). This view does not preclude the possibility that Nkx2.2 plays a dual role as a transcriptional activator (Watada et al., 2000) and repressor in oligodendrocyte differentiation.

An Activator Form of Olig2 Inhibits Oligodendrocyte Development

Our gain-of-function experiments demonstrate that Olig2 is sufficient to promote ectopic oligodendrocytes when misexpressed together with Nkx2.2, but is this gene required during normal oligodendrocyte development? Since a repressor form of Olig2, Olig2-EnR, phenocopied the function of intact Olig2 to promote ectopic oligodendrocytes in combination with Nkx2.2, we reasoned that the corresponding activator form, Olig2-VP16, might conversely inhibit endogenous oligodendrocyte formation. Consistent with this expectation, the number of oligodendrocyte precursors was reduced by the Olig2-VP16 fusion. The extent of the reduction (~40%) was similar to the extent of reduction of Isl1/2⁺ motoneurons by Olig2-VP16 (~50%; Novitch et al., 2001).

The inhibition of motoneuron differentiation by Olig2-VP16 raises the possibility that its activity to inhibit oligodendrocyte precursor formation is a secondary, non-cell-autonomous consequence of this earlier effect. Several lines of evidence argue against this possibility. First, the majority (~66%) of oligo precursors that do migrate in Olig2-VP16-electroporated embryos do not coexpress the transgene. Second, only 8 ± 2% of Olig2-VP16⁺ cells in gray and white matter were oligodendrocyte precursors. Third, oligodendrocytes are generated normally in the absence of motoneurons, in spinal cord explants from *Isl1*^{-/-} embryos (Sun et al., 1998) (in which motoneuron differentiation and survival is prevented; Pfaff et al., 1996). These data suggest that Olig2-VP16 inhibits oligodendrocyte formation cell autonomously. On the assumption (Muhr et al., 2001) that Olig2-VP16 behaves as a dominant-negative (antimorphic) form of Olig2, this inhibition would suggest that endogenous Olig2 (and/or other Olig proteins) is necessary for oligodendrocyte development in vivo. However, we cannot formally exclude that Olig2-VP16 behaves instead as a neomorph that prevents oligodendrocyte formation indirectly, via induction of unknown inhibitory factors. Nevertheless, preliminary analysis of *Olig* knockout mice

supports the idea that *Olig* genes are indeed required for oligodendrocyte formation in vivo (Q.Z. and D.J.A., unpublished observations).

Temporal Control of Neural Fate Specification Involves Alterations in Progenitor Domains

At earlier stages of spinal cord development (E3.0–E4.0), motoneurons arise from the pMN domain. This domain is congruent with the zone of *Olig2* expression (Novitch et al., 2001). It is bounded ventrally by the domain of expression of Nkx2.2, which potently represses *Olig2*, and dorsally by *lrx3* (Briscoe et al., 2000). Our results indicate that the pMN domain no longer exists at later stages when oligodendrocytes are being generated. Rather, the ventral part of the Olig2⁺ domain now overlaps the dorsal part of the Nkx2.2⁺ domain. In this way, a new progenitor domain (Figure 9E, p*) becomes established above p3.

This overlap appears to reflect a dorsal-ward expansion of Nkx2.2 into the Olig2⁺ domain, rather than vice-versa. At earlier stages, the Olig2⁺ pMN domain expresses a lower level of Pax6 expression than does the overlying p2 domain (Ericson et al., 1997; Figure 9E, upper, and Supplemental Figures S3A–S3C). However, at later stages (E6–E8), this zone of low Pax6 expression progressively contracts dorsally, and the Nkx2.2 boundary rises until it abuts the more dorsal domain of intermediate-level Pax6 expression (Figure 9E, lower and Supplemental Figures S3D–S3O). Since Pax6 represses Nkx2.2 expression (Briscoe et al., 2000; Muhr et al., 2001), the extinction of Pax6 expression within the ventral part of the Olig2⁺ domain would permit Nkx2.2 to expand dorsally into this region. Such a change might occur via a temporal increase in *Sonic Hedgehog* (*Shh*) expression, which positively regulates Nkx2.2 (Briscoe et al., 1999) and negatively regulates Pax6 (Ericson et al., 1997). In this respect, it is interesting that *Shh* has recently been shown to be important at later stages for oligodendrocyte differentiation (Soula et al., 2001), in addition to its earlier role in neural tube ventralization and motoneuron induction (Ericson et al., 1996).

The observation of overlap between the Olig2⁺ and Nkx2.2⁺ domains is surprising, given that Nkx2.2 is a potent repressor of Olig2 at earlier developmental stages (Novitch et al., 2001). Evidently, this repressor activity must be abrogated, at least in the domain where Nkx2.2 and Olig2 are coexpressed. How might this occur? Our data indicate that in the presence of constitutive Olig2, Nkx2.2 is converted from an inhibitor to a coinducer of endogenous *Olig2* expression. Since the overlap between Nkx2.2 and Olig2 occurs by dorsal expansion of *Nkx2.2* expression into the Olig2⁺ domain, the pre-existing Olig2 protein could conceivably protect *Olig2* from repression by Nkx2.2. Alternatively, other unknown factors might serve to protect *Olig2* from such repression until the positive-feedback loop with Nkx2.2 could be established. Similarly, Olig2 must also lose its erstwhile ability to derepress *Ngn2* (Novitch et al., 2001) in order to promote oligodendrocyte generation. The mechanisms that control these temporal changes in regulatory circuitry will provide interesting topics for further study.

Our results, taken together with those of Novitch et

al. (2001), indicate that the switch from motoneuron to oligodendrocyte generation involves changes in the crossregulatory interactions that position some of the boundaries of transcription factor gene expression in the ventricular zone of the spinal cord (Briscoe et al., 2000). These changes alter the landscape of the progenitor domains established at earlier stages, and permit the same transcription factor to function sequentially in neuronal and glial fate determination. Such dynamic changes in spatial patterning may represent a general mechanism for sequentially generating different neural cell types from a common progenitor domain.

Experimental Procedures

Chick Spinal Cord Library Construction and Isolation of Chick *Olig2* cDNA Clones

Trunk neural tubes of E3 and E3.5 chick embryos were dissociated using a modified enzymatic treatment method based on the protocol of Stemple et al. (Stemple and Anderson, 1992). Total RNA was extracted with TriZol reagent (Gibco), and cDNA synthesis performed with the Superscript Choice System (Gibco) according to the manufacturer's instructions, using an Oligd(T) primer. After ligation into the ZipLox vector (Gibco), the library was packaged with Gigapack (Stratagene), and amplified. 1.5×10^6 clones of the library were screened with the bHLH region of the mouse *Olig2* gene and 3 full-length cDNA clones corresponding to the chick *Olig2* gene were identified. Sequence alignment was performed with the CLUSTALW program of MacVector.

In Situ Hybridization

Nonradioactive in situ hybridization on frozen sections of chick embryos was performed as previously described (Zhou et al., 2000). The following chick gene probes were used: *Olig2*, *Olig3*, *Nkx2.2* (a gift of Dr. Mengsheng Qiu), *Sox10*, *PDGFR α* (a gift of Dr. Mark Mecola), *MBP*, *PLP/DMD20*, *Neurogenin1*, *Neurogenin2*, *Olig2* 3' UTR, and *Olig3* 3' UTR.

Double label in situ hybridization was performed as previously described (White and Anderson, 1999). The two different probes were labeled with either digoxigenin or fluorescein. The first probe was detected with alkaline phosphatase (AP)-conjugated anti-digoxigenin antibody, developed with Nitro blue tetrazolium (NBT) and 5-bromo-4-chloro-3-indolyl phosphate (BCIP), which yields a purple precipitate. After the NBT/BCIP reaction, the AP-conjugated anti-digoxigenin antibody was inactivated at 85°C, the slides were subsequently incubated with AP-conjugated anti-fluorescein antibody, and detected with 2-[4-iodophenyl]-3-[4-nitrophenyl]-5-phenyl tetrazolium chloride (INT) and BCIP, which yields a reddish brown precipitate. For fluorescent in situ hybridization, the basic protocol is essentially the same except Fast Red was used as the color substrate.

Immunohistochemistry

The peptide APLPAHPGHPASHPAHHPILPPA near the C terminus of the chick *Olig2* protein was synthesized and coupled to ovalbumin by ImmunoVison Technologies, Inc. Antibody production was carried out by the Caltech monoclonal facility.

Chick and rat embryos were fixed by immersion in 4% paraformaldehyde from 1 hr to overnight at 4°C depending on the age. The following primary antibodies were used: c.*Olig2* (1:100, mouse polyclonal), m.*Olig2* (1:2000, Rabbit Polyclonal, kind gift of Dr. Takebayashi Hirohide), *Nkx2.2* (1:10, hybridoma, Developmental Studies Hybridoma Bank (DSHB)), c.*Neurogenin2* (1:1000, Rabbit polyclonal, Dr. Sharon Perez), *Lim3* (1:100, hybridoma, DSHB), and *MNR2* (1:100, hybridoma, DSHB). Secondary antibodies used were: goat-anti-rabbit-IgG-Alexa 568 and Goat-anti-mouse-IgG-Alexa 488 (Molecular Probes).

Electroporation and Infection of Chick Embryos

All expression constructs were cloned into either the RCASBP(B) or RCASBP(A) (Morgan and Fekete, 1996). The constructs *Olig2*bHLH-EnR and *Olig2*bHLH-VP16 are the kind gift of Dr. Ben Novitch. The

control virus used in this study is RCASBP(B) encoding alkaline phosphatase. Viral production was performed essentially as described in Perez et al. (1999). Electroporation was carried out with a BTX Electro Squar Porator T820 at HH 13–15 (somites 15–19) (Christ and Ordahl, 1995).

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References

- Akazawa, C., Sasai, Y., Nakanishi, S., and Kageyama, R. (1992). Molecular characterization of a rat negative regulator with a basic helix-loop-helix structure predominantly expressed in the developing nervous system. *J. Biol. Chem.* **267**, 21879–21885.
- Altman, J., and Bayer, S.A. (1984). The development of the rat spinal cord. *Adv. Anat. Embryol. Cell Biol.* **85**, 1–166.
- Artavanis-Tsakonas, S., Rand, M.D., and Lake, R.J. (1999). Notch signaling: cell fate control and signal integration in development. *Science* **284**, 770–776.
- Briscoe, J., Sussel, L., Serup, P., Hartigan-O'Connor, D., Jessell, T.M., Rubenstein, J., and Ericson, J. (1999). Homeobox gene *Nkx2.2* and specification of neuronal identity by graded Sonic hedgehog signalling. *Nature* **398**, 622–627.
- Briscoe, J., Pierani, A., Jessell, T.M., and Ericson, J. (2000). A homeo-domain protein code specifies progenitor cell identity and neuronal fate in the ventral neural tube. *Cell* **101**, 435–445.
- Cepko, C.L. (1999). The roles of intrinsic and extrinsic cues and bHLH genes in the determination of retinal cell fates. *Curr. Opin. Neurobiol.* **9**, 37–46.
- Chitnis, A., Henrique, D., Lewis, J., Ish-Horowicz, D., and Kintner, C. (1995). Primary neurogenesis in *Xenopus* embryos regulated by a homologue of the *Drosophila* neurogenic gene *Delta*. *Nature* **375**, 761–766.
- Christ, B., and Ordahl, C.P. (1995). Early stages of chick somite development. *Anat. Embryol.* **191**, 381–396.
- de la Pompa, J.L., Wakeham, A., Correia, K.M., Samper, E., Brown, S., Aguilera, R.J., Nakano, T., Honjo, T., Mak, T.W., Rossant, J., and Conlon, R.A. (1997). Conservation of the Notch signalling pathway in mammalian neurogenesis. *Development* **124**, 1139–1148.
- Desai, A.R., and McConnell, S.K. (2000). Progressive restriction in fate potential by neural progenitors during cerebral cortical development. *Development* **127**, 2863–2872.
- Ericson, J., Morton, S., Kawakami, A., Roelink, H., and Jessell, T.M. (1996). Two critical periods of sonic hedgehog signaling required for the specification of motor-neuron identity. *Cell* **87**, 661–673.
- Ericson, J., Rashbass, P., Schedl, A., Brenner-Morton, S., Kawakami, A., Van Heynigen, V., Jessell, T.M., and Briscoe, J. (1997). *Pax6* controls progenitor cell identity and neuronal fate in response to graded *Shh* signaling. *Cell* **90**, 169–180.
- Farah, M.H., Olson, J.M., Sucic, H.B., Hume, R.I., Tapscott, S.J., and Turner, D.L. (2000). Generation of neurons by transient expression of neural bHLH proteins in mammalian cells. *Development* **127**, 693–702.
- Fode, C., Gradwohl, G., Morin, X., Dierich, A., LeMour, M., Goridis, C., and Guillemot, F. (1998). The bHLH protein NEUROGENIN 2 is a determination factor for epibranchial placode-derived sensory neurons. *Neuron* **20**, 483–494.
- Gard, A.L., and Pfeiffer, S.E. (1990). Two proliferative stages of the

- oligodendrocyte lineage (A2B5+O4⁻ and O4+GalC⁻) under different mitogenic control. *Neuron* 5, 615–625.
- Hall, A., Giese, N.A., and Richardson, W.D. (1996). Spinal cord oligodendrocytes develop from ventrally derived progenitor cells that express PDGF alpha-receptors. *Development* 122, 4085–4094.
- Ishibashi, M., Ang, S.-L., Shiota, K., Nakanishi, S., Kageyama, R., and Guillemot, F. (1995). Targeted disruption of mammalian *hairy* and *Enhancer of split* homolog-1 (*HES-1*) leads to up-regulation of neural helix-loop-helix factors, premature neurogenesis, and severe neural tube defects. *Genes Dev.* 9, 3136–3148.
- Jessell, T.M. (2000). Neuronal specification in the spinal cord: inductive signals and transcriptional codes. *Nat. Rev. Genet.* 1, 20–29.
- Kageyama, R., and Nakanishi, S. (1997). Helix-loop-helix factors in growth and differentiation of the vertebrate nervous system. *Curr. Opin. Genet. Dev.* 7, 659–665.
- Lu, Q.R., Yuk, D., Alberta, J.A., Zhu, Z., Pawlitzky, I., Chan, J., McMahon, A.P., Stiles, C.D., and Rowitch, D.H. (2000). Sonic hedgehog-regulated oligodendrocyte lineage genes encoding bHLH proteins in the mammalian central nervous system. *Neuron* 25, 317–329.
- Ma, Q., Kintner, C., and Anderson, D.J. (1996). Identification of *neurogenin*, a vertebrate neuronal determination gene. *Cell* 87, 43–52.
- Ma, Q., Chen, Z.F., Barrantes, I.B., de la Pompa, J.L., and Anderson, D.J. (1998). *Neurogenin 1* is essential for the determination of neuronal precursors for proximal cranial sensory ganglia. *Neuron* 20, 469–482.
- Miller, R. (1996). Oligodendrocyte origins. *Trends Neurosci.* 19, 92–96.
- Morgan, B.A., and Fekete, D.M. (1996). Manipulating gene expression with replication-competent retroviruses. In *Methods in Avian Embryology*, M.E. Bronner-Fraser, ed. (San Diego: Academic Press), pp. 185–218.
- Muhr, J., Anderson, E., Persson, M., Jessell, T., and Ericson, J. (2001). Groucho-mediated transcriptional repression establishes progenitor cell pattern and neuronal fate in the ventral neural tube. *Cell* 104, 861–873.
- Mumm, J.S., and Kopan, R. (2000). Notch signaling: from the outside in. *Dev. Biol.* 228, 151–165.
- Novitsch, B.G., Chen, A.I., and Jessell, T.M. (2001). Coordinate regulation of motor neuron subtype identity and pan-neuronal properties by the bHLH repressor Olig2. *Neuron* 31, this issue, 773–789.
- Ono, K., Bansal, R., Payne, J., Rutishauser, U., and Miller, R.H. (1995). Early development and dispersal of oligodendrocyte precursors in the embryonic chick spinal cord. *Development* 121, 1743–1754.
- Perez, S.E., Rebelo, S., and Anderson, D.J. (1999). Early specification of sensory neuron fate revealed by expression and function of neurogenins in the chick embryo. *Development* 126, 1715–1728.
- Perez Villegas, E.M., Olivier, C., Spassky, N., Poncet, C., Cochard, P., Zalc, B., Thomas, J.-L., and Martinez, S. (1999). Early specification of oligodendrocytes in the chick embryonic brain. *Dev. Biol.* 216, 98–113.
- Pfaff, S.L., Mendelsohn, M., Stewart, C.L., Edlund, T., and Jessell, T.M. (1996). Requirement for LIM homeobox gene *Is1* in motor-neuron generation reveals a motor neuron-dependent step in interneuron differentiation. *Cell* 84, 309–320.
- Pfeiffer, S.E., Warrington, A.E., and R., B. (1993). The oligodendrocyte and its many cellular processes. *Trends Cell Bio.* 3, 191–197.
- Philippe, E., Omlin, F.X., and Droz, B. (1986). Myelin-associated glycoprotein immunoreactive material: an early neuronal marker of dorsal root ganglion cells during chick development. *Brain Res.* 392, 275–277.
- Pringle, N.P., and Richardson, W.D. (1993). A singularity of PDGF alpha-receptor expression in the dorsoventral axis of the neural tube may define the origin of the oligodendrocyte lineage. *Development* 117, 525–533.
- Richardson, W.D., Pringle, N.P., Yu, W.-P., and Hall, A.C. (1997). Origins of spinal cord oligodendrocytes: possible developmental and evolutionary relationships with motor neurons. *Dev. Neurosci.* 19, 58–68.
- Richardson, W.D., Smith, H.K., Sun, T., Pringle, N.P., Hall, A., and Woodruff, R. (2000). Oligodendrocyte lineage and the motor neuron connection. *Glia* 29, 136–142.
- Soula, C., Danesin, C., Kan, P., Grob, M., Poncet, C., and Cochard, P. (2001). Distinct sites of origin of oligodendrocytes and somatic motoneurons in the chick spinal cord: oligodendrocytes arise from Nkx2.2-expressing progenitors by a Shh-dependent mechanism. *Development* 128, 1369–1379.
- Stemple, D.L., and Anderson, D.J. (1992). Isolation of a stem cell for neurons and glia from the mammalian neural crest. *Cell* 71, 973–985.
- Struhl, G., Fitzgerald, K., and Greenwald, I. (1993). Intrinsic activity of the Lin-12 and Notch intracellular domains in vivo. *Cell* 74, 331–345.
- Sun, T., Pringle, N., Hardy, A.P., Richardson, W.D., and Smith, H.K. (1998). Pax6 influences the time and site of origin of glial precursors in the ventral neural tube. *Mol. Cell. Neurosci.* 12, 228–239.
- Sun, Y., Nadal-Vicens, M., Misono, S., Lin, M.Z., Zubiaga, A., Hua, X., Fan, G., and Greenberg, M.E. (2001). Neurogenin promotes neurogenesis and inhibits glial differentiation by independent mechanisms. *Cell* 104, 365–376.
- Takebayashi, H., Yoshida, S., Sugimori, M., Kosako, H., Kominami, R., Nakafuku, M., and Nabeshima, Y. (2000). Dynamic expression of basic helix-loop-helix Olig family members: implication of Olig2 in neuron and oligodendrocyte differentiation and identification of a new member, Olig3. *Mech. Dev.* 99, 143–148.
- Tanabe, Y., and Jessell, T.M. (1996). Diversity and pattern in the developing spinal cord. *Science* 274, 1115–1123.
- Wang, S., Schrulla, A.D., diSibio, G., Bush, G., Nofziger, D., Hicks, C., Weinmaster, G., and Barres, B.A. (1998). Notch receptor activation inhibits oligodendrocyte differentiation. *Neuron* 21, 63–75.
- Watada, H., Mirmira, R.G., Kalamaras, J., and German, M.S. (2000). Intramolecular control of transcriptional activity by the NK2-specific domain in NK-2 homeodomain proteins. *Proc. Natl. Acad. Sci. USA* 97, 9443–9448.
- Weintraub, H., Dwarki, V.J., Verma, I., Davis, R., Hollenberg, S., Snider, L., Lassar, A., and Tapscott, S.J. (1991). Muscle-specific transcriptional activation by MyoD. *Genes Dev.* 5, 1377–1386.
- Wettstein, D.A., Turner, D.L., and Kintner, C. (1997). The *Xenopus* homolog of *Drosophila Suppressors of Hairless* mediates NOTC signaling during primary neurogenesis. *Development* 124, 693–702.
- White, P.M., and Anderson, D.J. (1999). In vivo transplantation of mammalian neural crest cells into chick hosts reveals a new autonomic sublineage restriction. *Development* 126, 4351–4363.
- Xu, X., Cai, J., Fu, H., Wu, R., Qi, Y., Modderman, G., Liu, R., and Qiu, M. (2000). Selective expression of Nkx-2.2 transcription factor in chicken oligodendrocyte progenitors and implications for the embryonic origin of oligodendrocytes. *Mol. Cell. Neurosci.* 16, 740–753.
- Zhou, Q., Wang, S., and Anderson, D.J. (2000). Identification of a novel family of oligodendrocyte lineage-specific basic helix-loop-helix transcription factors. *Neuron* 25, 331–343.

Accession Numbers

The chick Olig2 sequence reported in this paper has been deposited in GenBank with accession number AF411041.

Note Added in Proof

While our paper was in press, a report appeared showing a requirement for Nkx2.2 in oligodendrocyte differentiation: Qi, Y., Cai, J., Wu, Y., Wu, R., Lee, J., Fu, H., Rao, M., Sussel, L., Rubenstein, J., and Qiu, M. (2001). Control of oligodendrocyte differentiation by the Nkx2.2 homeodomain transcription factor. *Development* 128, 2723–2733.