# THE TOTAL SYNTHESIS OF DRAGMACIDINS D AND F

Thesis by

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To my family

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#### ABSTRACT

The dragmacidins are an emerging class of bis(indole) natural products isolated from deep-water marine organisms. Although there has been a substantial effort to prepare the simple piperazine dragmacidins, little synthetic work has been done in the area of the pyrazinone-containing family members, dragmacidins D, E, and F. These compounds are particularly interesting due to their complex structures and broad range of biological activity.

A highly convergent strategy to access dragmacidin D has been developed. In this approach, sequential halogen-selective Suzuki couplings were used to assemble the carbon scaffold of the natural product. After executing a highly optimized sequence of final events, the first completed total synthesis of dragmacidin D was achieved.

An enantiodivergent strategy for the total chemical synthesis of both (+)- and (–)dragmacidin F from a single enantiomer of quinic acid has been developed and successfully implemented. Although unique, the synthetic routes to these antipodes share a number of key features, including novel reductive isomerization reactions, Pd(II)-mediated oxidative carbocyclization reactions, halogen-selective Suzuki couplings, and high-yielding latestage Neber rearrangements.

The formal total syntheses of dragmacidin B, *trans*-dragmacidin C, and dihydrohamacanthin A are described. In addition, preliminary studies involving a novel approach for the preparation of dragmacidin E are reported.

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## LIST OF ABBREVIATIONS

$[\alpha]_{\rm D}$	specific rotation at wavelength of sodium D line
Ac	acetyl, acetate
app.	apparent
aq.	aqueous
atm	atmosphere
Bn	benzyl
Boc	<i>tert</i> -butyloxycarbonyl
br	broad
Bu	butyl
<i>n</i> -Bu	butyl
<i>t</i> -Bu	<i>tert</i> -Butyl
С	concentration for specific rotation measurements
°C	degrees Celsius
calc'd	calculated
CCDC	Cambridge Crystallographic Data Centre
CDI	1,1'-carbonyldiimidazole
CI	chemical ionization
Су	cyclohexyl
d	doublet
dba	dibenzylideneacetone
dppb	1,4-bis(diphenylphosphino)butane
dec	decomposition
DMAP	4-dimethylaminopyridine
DMF	N,N-dimethylformamide
DMSO	dimethyl sulfoxide
EC <sub>50</sub>	50% effective concentration
ee	enantiomeric excess
eNOS	endothelial nitric oxide synthase
equiv	equivalent
ESI	electrospray ionization
Et	ethyl
FAB	fast atom bombardment

g	gram(s)
gCOSY	gradient-selected Correlation Spectroscopy
h	hour(s)
HIV	human immunodeficiency virus
HRMS	high resolution mass spectroscopy
HPLC	high performance liquid chromatography
HSV	herpes simplex virus
hv	light
Hz	hertz
iNOS	inducible nitric oxide synthase
IR	infrared (spectroscopy)
J	coupling constant
λ	wavelength
L	liter
m	multiplet or milli
т	meta
m/z	mass to charge ratio
μ	micro
Me	methyl
MHz	megahertz
min	minute(s)
mol	mole(s)
mp	melting point
Ms	methanesulfonyl (mesyl)
MS	molecular sieves
nbd	norbornadiene
NBS	N-bromosuccinimide
NMO	N-methylmorpholine N-oxide
NMR	nuclear magnetic resonance
NOE	Nuclear Overhauser Effect
NOESY	Nuclear Overhauser Enhancement Spectroscopy
NOS	nitric oxide synthase
nNOS	neural nitric oxide synthase
[0]	oxidation
р	para

PDC	pyridinium dichromate
Ph	phenyl
pН	hydrogen ion concentration in aqueous solution
PhH	benzene
ppm	parts per million
PP	protein phosphatase
Pr	propyl
<i>i</i> -Pr	isopropyl
pyr	pyridine
q	quartet
rt	room temperature
$\mathbf{R}_{f}$	retention factor
S	singlet or strong
SEM	(trimethylsilyl)ethoxymethyl
t	triplet
TBAF	tetrabutylammonium fluoride
TBS	tert-butyldimethylsilyl
Tf	trifluoromethanesulfonyl (trifyl)
TFA	trifluoroacetic acid
THF	tetrahydrofuran
TIPS	triisopropylsilyl
TLC	thin layer chromatography
TMS	trimethylsilyl
Tr	triphenylmethyl (trityl)
Ts	<i>p</i> -toluenesulfonyl (tosyl)
UV	ultraviolet
W	weak

#### **CHAPTER ONE**

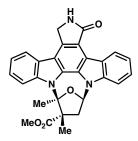
## The Dragmacidins: A Family of Biologically Active Marine Alkaloids

#### **1.1 Introduction**

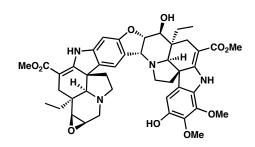
#### 1.1.1 Bis(indole) Alkaloids

Over the past several decades, the search for natural products in marine and terrestrial environments has led to the discovery of a number of biologically active bis(indole) alkaloids.<sup>1</sup> These compounds, as well as their unnatural analogs, have shown promise as leads for the development of novel therapeutics. Several representative bis(indole) compounds are shown below in Figure 1.1.1.<sup>2</sup>

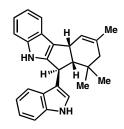
Figure 1.1.1



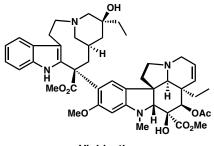
Staurosporine protein kinase C inhibition



Conophylline pancreatic β-cell regeneration



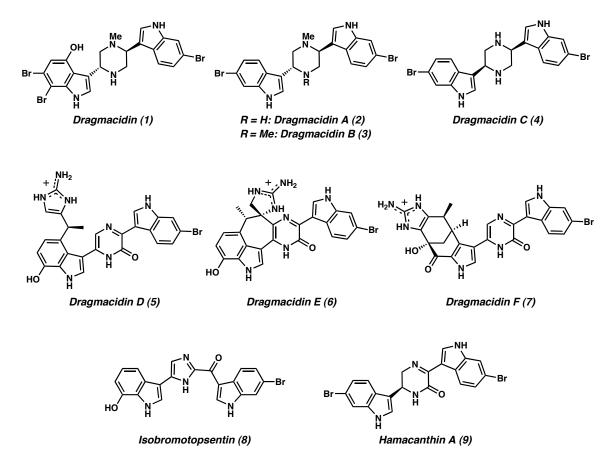




Vinblastine anti-neoplastic

Although many bis(indole) alkaloids have been found in nature, relatively few have been discovered in marine environments.<sup>3</sup> Of those, the dragmacidins have received considerable attention from the scientific community over the past decade due to their broad range of biological activity and complex structures (**1**-**7**, Figure 1.1.2).<sup>4,5</sup> Several closely related bis(indole) natural products have been discovered in similar environments, such as the topsentins (e.g., **8**)<sup>4b,6</sup> and the hamacanthins (e.g., **9**).<sup>7</sup>





#### 1.1.2 The Dragmacidins

The dragmacidins are an emerging class of novel bis(indole) natural products isolated from the deep-water marine sponges *Dragmacidon*, *Halicortex*, *Spongosorites*, and *Hexadella*, and the tunicate *Didemnum candidum*. The four dragmacidins initially identified (**1-4**) contain a piperazine linker and display modest antifungal, antiviral, and cytotoxic activities.<sup>4a-c</sup> However, our interest in these natural products was piqued by the structurally complex pyrazinone-containing family members, dragmacidins D (**5**), E (**6**), and F (**7**).<sup>4d-g</sup> Although the relative stereochemistry of **1-7** was known, at the onset of our investigations, the absolute stereochemistry of **1-7** had not been established.

#### 1.2 Biological Activity of Pyrazinone-Containing Dragmacidins

The following section describes the wide range of biological activity associated with the pyrazinone-containing dragmacidins, D (5), E (6), and F (7). Preliminary studies suggest that these compounds are interesting from a biological standpoint and are therefore attractive targets for total synthesis. Synthetic routes to the pyrazinone dragmacidins could facilitate the production of sufficient quantities of material needed for advanced biological studies.

#### **1.2.1** Inhibitors of Protein Phosphatases

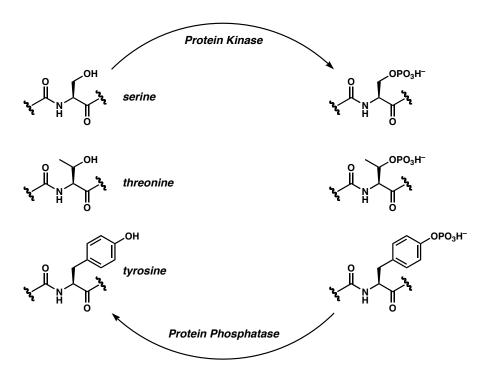
#### 1.2.1.1 Activity of Dragmacidins

In 1998, Capon et al. reported that dragmacidins D (5) and E (6) are potent inhibitors of serine-threonine protein phosphatases (PP).<sup>4e</sup> In addition, preliminary testing showed that dragmacidin D (5) selectively inhibited PP1 over the PP2A isozyme.

#### **1.2.1.2** About Protein Phosphatases

The reversible phosphorylation of proteins containing serine, threonine, and tyrosine residues is widely recognized as a mechanism by which many cellular events are regulated (Figure 1.2.1).<sup>8</sup> While phosphorylation is catalyzed by protein kinases, dephosphorylation is carried out by protein phosphatases. To date, many phosphatase enzymes have been discovered; however, discerning which phosphatase is responsible for controlling particular cellular pathways has remained an elusive goal. In particular, distinguishing the action of the PP1 and PP2A isozymes has been extremely difficult. Ultimately, the discovery of small molecules that display selective PP inhibition could help elucidate the mechanism of many physiological processes including cell division, gene expression, neurotransmission, and muscle contraction.<sup>8c</sup>





#### 1.2.2 Inhibitors of Neural Nitric Oxide Synthase

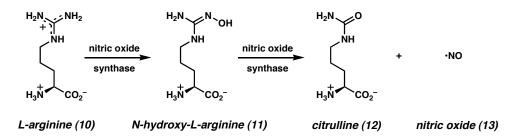
#### 1.2.2.1 Activity of Dragmacidins

Dragmacidin D (**5**) has also been shown to selectively inhibit neural nitric oxide synthase (nNOS) in the presence of inducible nitric oxide synthase (iNOS).<sup>9</sup> The ability to efficiently prepare dragmacidin D, and related derivatives thereof, could be extremely valuable for the discovery of novel drugs that target neurodegenerative disorders.

#### **1.2.2.2** About Nitric Oxide Synthase

The production of nitric oxide (NO) in the human body is known to be associated with the regulation of a number of physiological properties.<sup>10</sup> NO (**13**) arises from the decomposition of L-arginine (**10**) by an enzyme known as nitric oxide synthase (NOS)  $(10 \rightarrow 11 \rightarrow 12 + 13)$ , Scheme 1.2.1). This enzyme occurs in three main isoforms: a) inducible NOS (iNOS), which generates NO during the immune response where NO acts as a cytotoxic molecule, b) endothelial NOS (eNOS), which produces NO for vasodilatation, and c) neuronal NOS (nNOS), which provides NO involved in neuronal physiology. Although NO provides many beneficial functions, the overproduction of NO in the brain has been linked to a number of neurodegenerative disorders. Thus, the ability to selectively inhibit nNOS may be useful for the treatment of related illnesses, including Alzheimer's, Parkinson's, and Huntington's diseases.<sup>11</sup>

Scheme 1.2.1



#### **1.2.2.3** Aminoimidazoles as Inhibitors

Compounds bearing aminoimidazole functionality are an attractive class of NOS inhibitors since, when protonated, they resemble the guandinium system present in arginine (10).<sup>12</sup> Therefore, the aminoimidazole moiety of dragmacidin D (5) could potentially be responsible for its reported NOS activity through competitive inhibition. It is possible that the other aminoimidazole-containing dragmacidins (6 and 7) could display similar activity, although studies in this area have not appeared in the literature.

#### 1.2.3 Miscellaneous Biological Activity

#### 1.2.3.1 Cytotoxicity

Many bis(indole) compounds discovered in nature have shown promise as leads in the search for new anti-cancer medicines. Although its mechanism of action is not known, dragmacidin D (**5**) shows cytotoxicity against several human lung tumor cell lines.<sup>4d</sup> Dragmacidins E (**6**) and F (**7**), on the other hand, have not yet been evaluated for anti-neoplastic activity.

#### 1.2.3.2 Antiviral and Anti-Inflammatory Properties

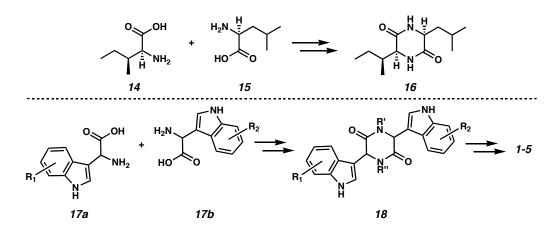
Dragmacidin F (7) is reported to exhibit in vitro antiviral activity against herpes simplex virus (HSV-I;  $EC_{50} = 95.8$  mM) and human immunodeficiency virus (HIV-I;  $EC_{50} = 0.91$  mM).<sup>4f</sup> In addition, dragmacidins D (5) and F (7) display anti-inflammatory activity in resiniferatoxin-induced inflammation of the mouse ear.<sup>13,4g</sup>

## **1.3 Biosynthesis of Dragmacidins**

## 1.3.1 Biosynthesis of Piperazine Dragmacidins and Dragmacidin D

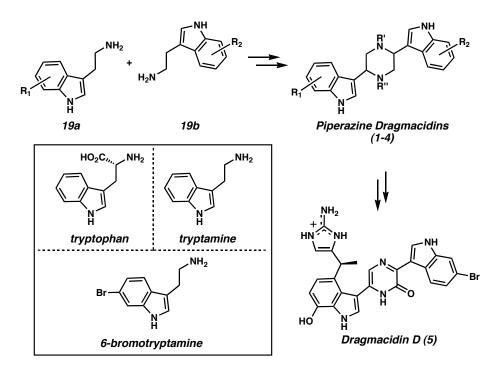
The biosynthesis of the dragmacidins has not been studied in detail.<sup>14</sup> However, in the 1960s, MacDonald and co-workers examined the origin of simple diketopiperazine natural products.<sup>15</sup> It was found that disubstituted piperazine derivatives could form via the condensation of two amino acids, L-isoleucine and L-leucine ( $14 + 15 \rightarrow 16$ , Scheme 1.3.1). Based on this work, one could propose that the dragmacidins arise by a related pathway ( $17a + 17b \rightarrow 18 \rightarrow 1$ -5). However, the necessary indole-containing amino acids (17a and 17b) for this biosynthesis are not known to be naturally occurring.

Scheme 1.3.1



Tryptophan and tryptamine (Scheme 1.3.2), on the other hand, are both commonly found in nature. In fact, 6-bromotryptamine was found in the same marine sponge from which dragmacidin C was isolated.<sup>4c</sup> It seems plausible that the dragmacidins could be biosynthetically derived from building blocks of this type (i.e., **19a** and **19b**). Various oxidations could take place before or after the dimerization event occurs,<sup>16</sup> eventually leading to formation of the piperazine dragmacidins (**1**-**4**). These molecules, or a related derivative, could perhaps be biosynthetically transformed into dragmacidin D (**5**).

Scheme 1.3.2

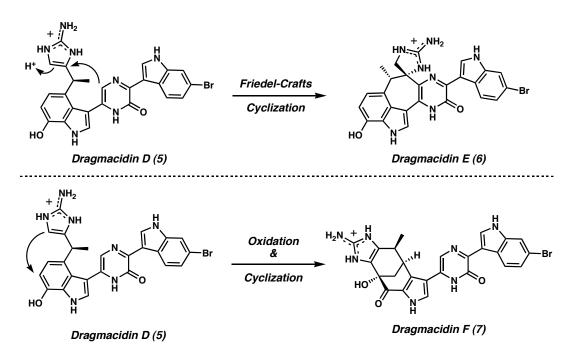


#### 1.3.2 Biosynthesis of Dragmacidins E and F

Dragmacidins D, E, and F are likely biosynthetically related.<sup>4f</sup> Of the possible biosynthetic scenarios, most probable is that dragmacidins E (6) and F (7) are derived by

cyclization of either dragmacidin D (5) or a closely related congener (Scheme 1.3.3). For example, dragmacidins D and E are isomers that differ by a single C–C bond. In nature, it is likely that a Friedel-Crafts cyclization between the pyrazinone and aminoimidazole groups of dragmacidin D occurs in order to construct the seven-membered ring of dragmacidin E (i.e.,  $5 \rightarrow 6$ ). Dragmacidins D (5) and F (7) also differ in connectivity by one C–C bond; however, in this case, there is also a difference in oxidation state between the two natural products. Thus, oxidative dearomatization with concomitant cyclization could facilitate the formation of the unique polycyclic framework present in dragmacidin F (i.e.,  $5 \rightarrow 7$ ). Related oxidation pathways for tryptophan derivatives have been observed in nature.<sup>17</sup>





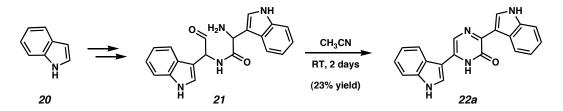
### **1.4** Synthetic Studies Relating to the Pyrazinone-Containing Dragmacidins

At the onset of our studies, there was a single report related to the synthesis of the pyrazinone-containing dragmacidins (**5**-**7**) by Jiang and Gu.<sup>5g</sup> Although the authors claimed to have prepared the bis(indole)pyrazinone scaffold of **5** and **6**, this work was clearly erroneous but was never retracted.<sup>18</sup>

### 1.4.1 Jiang's Approach to the Pyrazinone Core

In 2000, shortly after we began work in the area of the dragmacidin natural products, Jiang and co-workers reported a successful synthetic route to the bis(indole)pyrazinone core of dragmacidins D and E (Scheme 1.4.1).<sup>5h</sup> Their strategy involved the elaboration of indole (**20**) to bis(indole)amide **21** via a series of functional group manipulations. Then, in the final step, intramolecular condensation of amide **21** produced pyrazinone **22a** in 23% yield. Although Jiang's route produced the desired bis(indole) core (**22a**), it is lengthy and hampered by low yields.



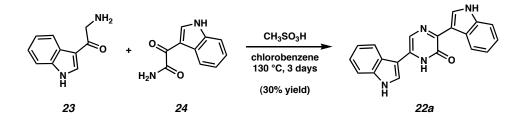


## 1.4.2 Horne's Approach to the Pyrazinone Core

In 2002, during the course of our own investigations, a convergent strategy for constructing pyrazinone **22a** was reported by Horne (Scheme 1.4.2).<sup>5j</sup> Upon exposure to

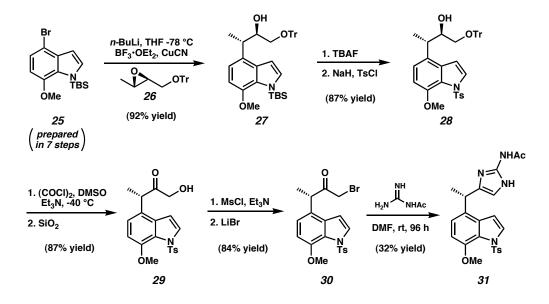
methanesulfonic acid at 130 °C, aminoketone **23** underwent a cyclocondensation reaction with ketoamide **24** to afford the desired product **22a** in 30% yield. Further work in this area using substituted indoles has yet to be reported.

Scheme 1.4.2



### 1.4.3 Jiang's Approach to the Aminoimidazole Segment of Dragmacidin D

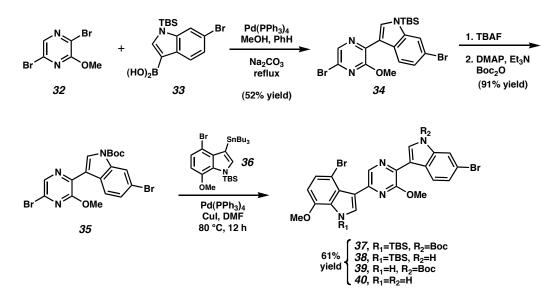
Concurrent with our own work, in 2002 Jiang described a synthesis of the aminoimidazole segment of dragmacidin D.<sup>5i</sup> The 4,7-disubstituted indole (**25**) was prepared in 7 steps from commercially available compounds via a Leimgruber-Batcho indole synthesis (Scheme 1.4.3).<sup>19,20</sup> Subsequent metallation and quenching with epoxide **26** afforded alcohol **27** in good yield. After manipulations of the indole nitrogen protecting group (**27**  $\rightarrow$  **28**), the 2° alcohol was oxidized, and the trityl group was removed to produce hydroxyketone **29**. Elaboration to bromide **30**, followed by exposure to acetylguanidine for 4 days, installed the desired aminoimidazole segment (**31**) in 32% yield. At the time of Jiang's publication, our group had already independently prepared **31** and determined that it was not a productive route to dragmacidin D (see Chapter 2, reference 33).



## 1.4.4 Jiang's Second Generation Approach to the Pyrazinone Core

Following our publication describing the first total synthesis of dragmacidin D, Jiang reported a similar approach to construct the core of the natural product in the form of a bis(indole)pyrazine (Scheme 1.4.4).<sup>5k</sup> First, dibromopyrazine **32** was cross-coupled with boronic acid **33** to afford indolopyrazine **34**. After switching protecting groups on the indole nitrogen ( $34 \rightarrow 35$ ), indolopyrazine **35** was coupled with stannane **36** to produce a mixture of pyrazine products (**37-40**) in 61% combined yield. An account describing the elaboration of **37-40** to the natural product (**5**) has yet to appear in the literature.

# Scheme 1.4.4



## **1.5** Conclusion

The dragmacidin alkaloids are a unique class of molecules that are interesting from both a biological and structural standpoint. Although there has been synthetic work aimed at the piperazine dragmacidins (1-4), the pyrazinone-containing dragmacidins, D (5), E (6), and F (7), have received little attention from the synthetic community. Ultimately, synthetic routes to these natural products could be extremely valuable in the search for new medicines.

#### **1.6 Notes and References**

- (a) Blunt, J. W.; Copp, B. R.; Munro, M. H. G.; Northcote, P. T.; Prinsep, M. R. Nat. Prod. Rep. 2004, 21, 1-49. (b) Aygün, A.; Pindur, U. Curr. Med. Chem.
   2003, 10, 1113-1127. (c) Faulkner, D. J. Nat. Prod. Rep. 2002, 19, 1-48. (d) Pindur, U.; Lemster, T. Curr. Med. Chem. 2001, 8, 1681-1698.
- (2) (a) For staurosporine, see: Omura, S.; Iwai, Y.; Hirano, A.; Nakagawa, A.; Awaya, J.; Tsuchiya, H.; Takahashi, Y.; Masuma, R. J. Antibiot. 1977, 30, 275-289. (b) For conophylline, see: Kam, T.-S.; Pang, H.-S.; Lim, T.-M. Org. Biomol. Chem. 2003, 1, 1292-1297. (c) For yuehchukene, see: Kong, Y. C.; Cheng, K. F.; Cambie, R. C.; Waterman, P. G. J. Chem. Soc., Chem. Commun. 1985, 47-48. (d) For vinblastine, see: Noble, R. L.; Beer, C. T.; Cutts, J. H. Ann. N.Y. Acad. Sci. 1958, 76, 882-894.
- (3) (a) Yang, C.-G.; Huang, H.; Jiang, B. Curr. Org. Chem. 2004, 8, 1691-1720. (b) Jin, Z. Nat. Prod. Rep. 2003, 20, 584-605. (c) Hibino, S.; Choshi, T. Nat. Prod. Rep. 2002, 19, 148-180. (d) Sasaki, S.; Ehara, T.; Sakata, I.; Fujino, Y.; Harada, N.; Kimura, J.; Nakamura, H.; Maeda, M. Bioorg. Med. Chem. Lett. 2001, 11, 583-585.
- (4) For the isolation of the piperazine-containing dragmacidins, see: (a) Kohmoto, S.;
  Kashman, Y.; McConnell, O. J.; Rinehart, K. L., Jr.; Wright, A.; Koehn, F. J. Org. *Chem.* 1988, 53, 3116-3118. (b) Morris, S. A.; Andersen, R. J. Tetrahedron 1990,

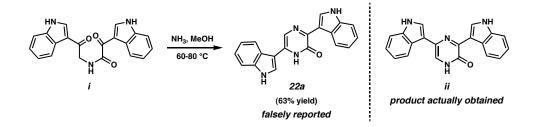
46, 715-720. (c) Fahy, E.; Potts, B. C. M.; Faulkner, D. J.; Smith, K. J. Nat. Prod.
1991, 54, 564-569. For the isolation of the pyrazinone-containing dragmacidins, see: (d) Wright, A. E.; Pomponi, S. A.; Cross, S. S.; McCarthy, P. J. Org. Chem.
1992, 57, 4772-4775. (e) Capon, R. J.; Rooney, F.; Murray, L. M.; Collins, E.; Sim, A. T. R.; Rostas, J. A. P.; Butler, M. S.; Carroll, A. R. J. Nat. Prod. 1998, 61, 660-662. (f) Cutignano, A.; Bifulco, G.; Bruno, I.; Casapullo, A.; Gomez-Paloma, L.; Riccio, R. Tetrahedron 2000, 56, 3743-3748. (g) Wright, A. E.; Pomponi, S. A.; Jacobs, R. S. PCT Int. Appl. WO 9942092, August 26, 1999.

(5) For synthetic work aimed toward the piperazine-containing dragmacidins, see: (a) Jiang, B.; Smallheer, J. M.; Amaral-Ly, C.; Wuonola, M. A. J. Org. Chem. 1994, 59, 6823-6827. (b) Whitlock, C. R.; Cava, M. P. Tetrahedron Lett. 1994, 35, 371-374. (c) Kawasaki, T.; Enoki, H.; Matsumura, K.; Ohyama, M.; Inagawa, M.; Sakamoto, M. Org. Lett. 2000, 2, 3027-3029. (d) Miyake, F. Y.; Yakushijin, K.; Horne, D. A. Org. Lett. 2000, 2, 3185-3187. (e) Yang, C.-G.; Wang, J.; Tang, X.-X.; Jiang, B. Tetrahedron: Asymmetry 2002, 13, 383-394. (f) Kawasaki, T.; Ohno, K.; Enoki, H.; Umemoto, Y.; Sakamoto, M. Tetrahedron Lett. 2002, 43, 4245-4248. For studies targeting dragmacidins D, E, or F, see: (g) Jiang, B.; Gu, X.-H. Bioorg. Med. Chem. 2000, 8, 363-371. (h) Jiang, B.; Gu, X.-H. Heterocycles 2000, 53, 1559-1568. (i) Yang, C.-G.; Wang, J.; Jiang, B. Tetrahedron Lett. 2002, 4, 941-943. (k) Yang, C.-G.; Liu, G.; Jiang, B. J. Org. Chem. 2002, 67, 9392-9396.

- (6) For the isolation of topsentin natural products, see: (a) Bartik, K.; Braekman, J.-C.; Daloze, D.; Stoller, C.; Huysecom, J.; Vendevyver, G.; Ottinger, R. *Can. J. Chem.* **1987**, *65*, 2118-2121. (b) Tsujii, S.; Rinehart, K. L.; Gunasekera, S. P.; Kashman, Y.; Cross, S. S.; Lui, M. S.; Pomponi, S. A.; Diaz, M. C. *J. Org. Chem.* **1988**, *53*, 5446-5453. (c) Murray, L. M.; Lim, T. K.; Hooper, J. N. A.; Capon, R. J. *Aust. J. Chem.* **1995**, *48*, 2053-2058. (d) Shin, J.; Seo, Y.; Cho, K. W.; Rho, J.-R.; Sim, C. J. *Nat. Prod.* **2000**, *62*, 647-649.
- (7) For the isolation of hamacanthins and dihydrohamacanthins, see: (a) Gunasekera,
  S. P.; McCarthy, P. J.; Kelly-Borges, M. J. Nat. Prod. 1994, 57, 1437-1441. (b)
  Casapullo, A.; Bifulco, G.; Bruno, I.; Riccio, R. J. Nat. Prod. 2000, 63, 447-451.
- (8) For reviews regarding protein phosphatases, see: (a) McCluskey, A.; Sim. A. T. R.; Sakoff, J. A. J. Med. Chem. 2002, 45, 1151-1175. (b) Burke, T. R.; Zhang, Z.-Y. Biopolymers 1998, 47, 225-241. (c) Sheppeck, J. E.; Gauss, C.-M.; Chamberlin, A. R. Bioorg. Med. Chem. 1997, 5, 1739-1750.
- (9) Longley, R. E.; Isbrucker, R. A.; Wright, A. E. U.S. Patent 6,087,363, July 11, 2000.
- (10) (a) Kerwin, J. F., Jr.; Lancaster, J. R., Jr.; Feldman, P. L. J. Med. Chem. 1995, 38, 4343-4362. (b) Schmidt, H. H. W.; Walter, U. Cell 1994, 78, 919-925. (c) Mocada, S.; Palmer, R. M. J.; Higgs, E. A. Pharmacol. Rev. 1991, 43, 109-142.

- (11) (a) Marletta, M. A. J. Med. Chem. 1994, 37, 1899-1907. (b) Molina, J. A.; Jimenez-Jimenez, F. J.; Orti-Pareja, M.; Navarro, J. A. Drugs Aging 1998, 12, 251-259. (c) Thorns, V.; Hansen, L. M. Exp. Neurol. 1998, 150, 14-20.
- (12) (a) Ulhaq, S.; Naylor, M. A.; Chinje, E. C.; Threadgill, M. D.; Stratford, I. J. Anti-Cancer Drug Des. 1997, 12, 61-65. (b) Aoki, S.; Ye, Y.; Higuchi, K.; Takashima, A.; Tanaka, Y.; Kitagawa, I.; Kobayashi, M. Chem. Pharm. Bull. 2001, 49, 1372-1374.
- (13) Jacobs, R. S.; Pomponi, S.; Gunasekera, S.; Wright, A. PCT Int. Appl. WO 9818466, May 7, 1998.
- (14) For a review regarding the biosynthesis of indole alkaloids derived from plants, see: Kutney, J. P. *Nat. Prod. Rep.* **1990**, *7*, 85-103.
- (15) (a) MacDonald, J. C. J. Biol. Chem. 1961, 236, 512-514. (b) Leete, E.; Bjorklund, J. A.; Reineccius, G. A.; Chen, T.-B. Spec. Publ.–R. Soc. Chem. 1992, 95, 75-95.
- (16) For a review regarding the biosynthesis of halogenated marine natural products, see: Butler, A.; Carter-Franklin, J. N. Nat. Prod. Rep. 2004, 21, 180-188.
- (17) McIntire, W. S.; Wemmer, D. E.; Chistoserdov, A.; Lidstrom, M. E. Science 1991, 252, 817-824.

(18) Jiang reported the synthesis of bis(indole)pyrazinone 22a by the amination/ cyclization of i. It is likely that *m*-substituted product ii actually formed during this reaction. See reference 5h.



- (19) For the Leimgruber-Batcho indole synthesis, see: Batcho, A. D.; Leimgruber, W. Org. Synth. 1985, 63, 214-225, and references therein.
- (20) We also explored a Leimgruber-Batcho route to 4,7-disubstituted indoles. See Chapter 2, section 2.3.3 and references 22 and 28.

### **CHAPTER TWO**

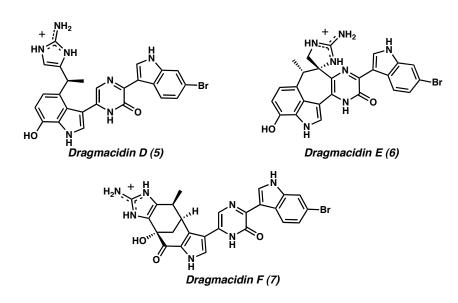
# The Total Synthesis of Dragmacidin $D^{\dagger}$

# 2.1 Background

# 2.1.1 Introduction

In 2000, the pyrazinone-containing dragmacidins, namely, dragmacidins D, E, and F, were selected as formidable synthetic targets for our laboratory (Figure 2.1.1).<sup>1</sup> Initially, we chose to pursue the total synthesis of dragmacidin D (**5**),<sup>1a,b</sup> predominantly because it was believed to be the biosynthetic precursor to dragmacidins E (**6**) and F (**7**).<sup>1c</sup> In addition, **5** appeared to be the simplest of the pyrazinone-containing family members. Thus, we hoped to develop a strategy for the preparation of dragmacidin D (**5**) that would be amenable to the synthesis of the other complex dragmacidin natural products.<sup>2</sup>





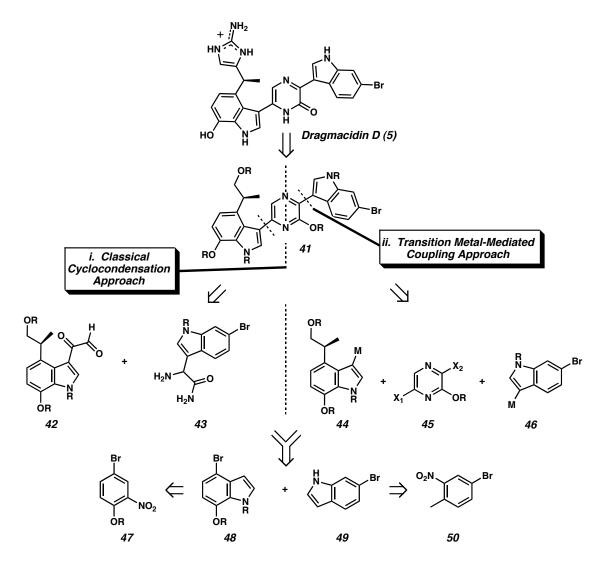
<sup>&</sup>lt;sup>†</sup> This work was performed in collaboration with Dr. Richmond Sarpong, a postdoctoral scholar in the Stoltz group.

When considering the structure of dragmacidin D (**5**), several synthetic challenges become apparent. Dragmacidin D possesses a total of seven nitrogen atoms, three of which are incorporated in the aminoimidazole moiety, while two are within the pyrazinone core. The compound contains an unusual bis(indole) architecture featuring a 3,4,7-trisubsituted indole and a 3,6-disubstituted indole. Both of these indole substitution patterns are known to be synthetically challenging targets.<sup>3</sup> It was predicted that dragmacidin D, as well as many of its synthetic precursors, would be highly polar, extremely reactive, and perhaps difficult to handle in a laboratory setting.

### 2.1.2 Retrosynthetic Analysis of Dragmacidin D

Two retrosynthetic strategies for the synthesis of dragmacidin D are presented in Scheme 2.1.1. As a critical maneuver, we chose to introduce the aminoimidazole moiety at a late stage in the synthesis in order to facilitate the handling of key precursors. Thus, disconnection of the aminoimidazole in the natural product (**5**) provided ether **41**. We then targeted **41** through two complementary routes: *i*) a classical cyclocondensation approach<sup>4</sup> and *ii*) a more modern transition metal-mediated cross-coupling approach.<sup>5</sup> In approach *i*, the pyrazinone system would be constructed through the linkage of two functionalized indole units (**42** + **43**), while in route *ii*, the dragmacidin core was envisioned to arise by a stepwise three-component coupling sequence (**44** + **45** + **46**). Both routes relied on the same indole building blocks (**48** and **49**), which were readily available from simple aromatic starting materials **47** and **50**, respectively.

Scheme 2.1.1

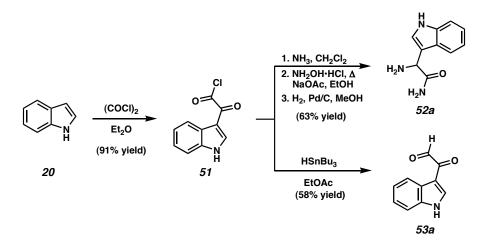


## 2.2 The Cyclocondensation Approach to Access the Bis(indole) Framework

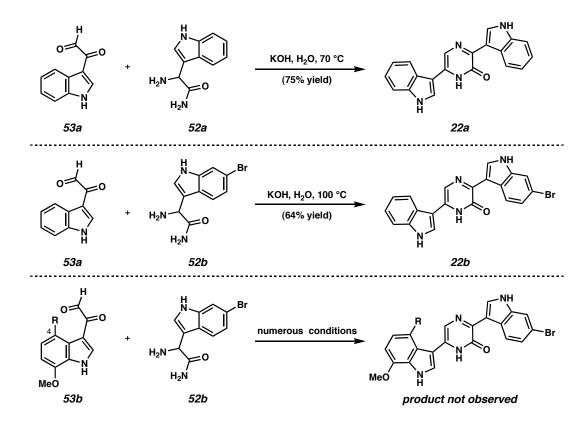
Our initial efforts toward the total synthesis of dragmacidin D (5) focused on the cyclocondensation approach (*i*). A model system for the preparation of the pyrazinone core (i.e., **22a**) was explored. Treatment of indole (**20**) with oxalyl chloride produced **51** in high yield (Scheme 2.2.1).<sup>6</sup> This compound was then employed as a common intermediate for the synthesis of the unsubstituted coupling fragments **52a** and **53a**. The synthesis of aminoamide **52a** proceeded via elaboration of **51** by a sequence involving: a)

amidation using ammonia, b) oxime formation, and c) reduction using hydrogen, catalyzed by palladium on carbon. Ketoaldehyde **53a** was prepared directly by reduction of **51** with tributyltin hydride.<sup>7</sup>

Scheme 2.2.1



With the key fragments in hand, we investigated the viability of the cyclocondensation reaction (Scheme 2.2.2). Upon exposure to heated aqueous potassium hydroxide, compounds **52a** and **53a** underwent smooth conversion to the desired pyrazinone **22a** in good yield, as the only observed product of the reaction. Under similar conditions, bromide **52b**<sup>8</sup> also participated in the pyrazinone-forming reaction  $(53a + 52b \rightarrow 22b)$ . However, under our optimized conditions, as well as a variety of others (Bronsted acids and bases, Lewis acids), we were unable to effect cyclocondensative coupling with any C(4)-substituted ketoaldehyde derivative (i.e., **53b**).<sup>9</sup>



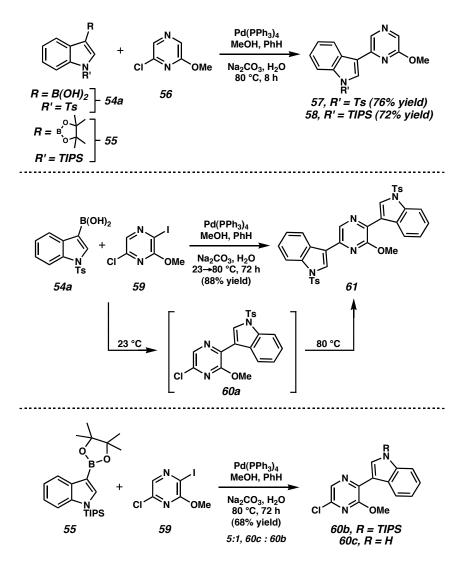
#### 2.3 The Metal-Mediated Strategy to Construct the Bis(indole) Framework

We turned our attention to the alternative retrosynthetic strategy, the transition metal-catalyzed cross-coupling route *ii*. The ability to carry the 6-bromoindole moiety directly through the reaction sequence would be critical for the success of our plan. The synthesis clearly became an issue of strategy involving not only the exact order of the coupling reactions, but also the specific identity of each substrate involved. The appropriate selection of halides, metals, and protecting groups would be crucial. We thus turned our attention toward experiments that would delineate suitable conditions for coupling.

### **2.3.1** The Development of Suitable Conditions for Selective Cross-Couplings

We initially surveyed a variety of coupling reactions involving model indoles and various halogenated pyrazine derivatives in order to assess the relative reactivity of such systems as well as the suitability of the protective groups on the indole nitrogen. It was quickly established that halogenated pyrazines are highly reactive toward palladiummediated couplings to metalated indoles. Furthermore, the oxidative addition of palladium(0) to pyrazinyl halides is more facile than to simple aromatic halides.<sup>10</sup> For example, reaction of borylated indoles 54a and 55 with readily available chloropyrazine 56<sup>11</sup> proceeded smoothly at 80 °C under standard Suzuki conditions to afford coupled products 57 and 58 (Scheme 2.3.1). Under identical conditions, simple aryl chlorides do not readily participate in such couplings.<sup>12</sup> Additionally, treatment of chloroiodopyrazine 59 with 2 equiv of indole 54a at 23 °C produced indolylpyrazine 60a exclusively, while raising the temperature to 80 °C resulted in the formation of the bis(indole)pyrazine 61. A more surprising development was observed upon treatment of pyrazine 59 with an excess of silvlated boronic ester 55 (2.3 equiv) at 80 °C. Under these conditions, exclusively monocoupled product was obtained as a mixture of silvlated and desilvlated compounds (60b and 60c). This difference in reactivity points to a remote electronic effect of the indole protecting group on the activation of the intermediate chloroindolylpyrazine (60) toward coupling.

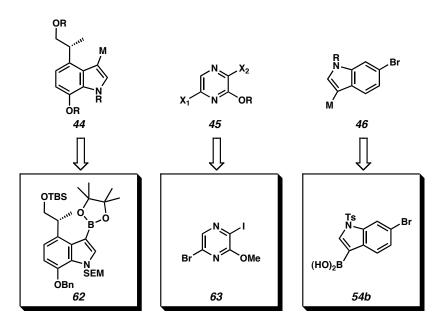
Scheme 2.3.1



Having conducted these simple experiments, we began to identify appropriate substitution patterns for building blocks 44, 45, and 46 (Figure 2.3.1). It was immediately clear that the use of Suzuki couplings would be favorable due to our success in the model systems. Thus, boron substituents were employed as the metal species for both indole substrates (44 and 46). In addition, the protecting groups for indoles 44 and 46 were chosen in a manner that optimized orthogonality with respect to deprotection, which would facilitate control during late-stage manipulations. In particular, the *N*-

protecting groups were very carefully selected. The SEM group<sup>13</sup> of **62** was considered ideal due to its marked stability and electron-donating nature, while the Ts group of **54b** was preferred mainly because of its success in the model system studies. Perhaps the most important decision was the selection of halogens  $X_1$  and  $X_2$  of pyrazine **45**. Although chloroiodopyrazine **59** was utilized in the model systems described above, it was believed that replacing the chloride with a bromide would allow for better position selectivity.<sup>14</sup> Thus, bromoiodopyrazine **63** was selected as the key synthetic fragment. We then proceeded to develop rapid syntheses of the three essential pieces (**62**, **63**, and **54b**).



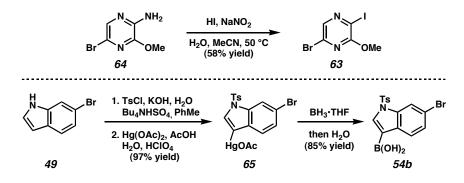


## 2.3.2 Synthesis of Pyrazine and Bromoindole Fragments

The key differentially halogenated pyrazine fragment **63** was readily prepared via iodide displacement of the in situ prepared diazonium salt of aminopyrazine **64**<sup>15</sup>

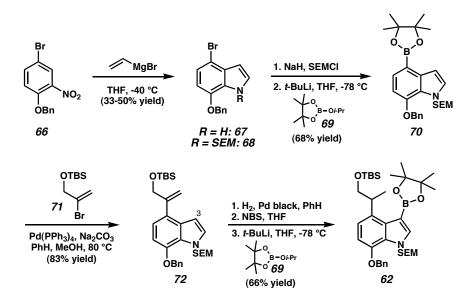
(Scheme 2.3.2).<sup>16</sup> Bromoindole boronic acid derivative **54b** was synthesized from parent indole **49**<sup>17</sup> by protection of the indole nitrogen,<sup>18</sup> treatment with mercuric acetate, and reaction of the resulting organomercurial (**65**) with borane•THF complex followed by hydrolytic work-up (82% yield, 3 steps).<sup>19</sup>





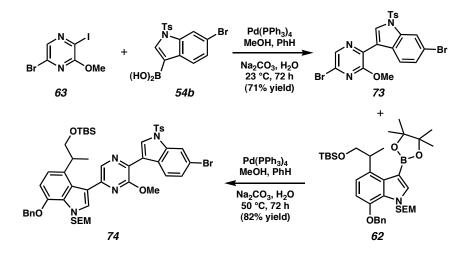
## 2.3.3 Synthesis of the 3,4,7-Trisubstituted Indole Fragment

The 3,4,7-trisubstituted indole fragment (62) was synthesized by the Bartoli indolization reaction (Scheme 2.3.3).<sup>20</sup> Treatment of nitroaromatic  $66^{21}$  with vinyl Grignard produced the highly functionalized indole 67 directly. Although the yield of this reaction was variable and modest, we were able to prepare 67 on multigram scale.<sup>22</sup> Following protection of the indole nitrogen by a 2-(trimethylsilyl)ethoxymethyl (SEM) group (67  $\rightarrow$  68),<sup>13</sup> halogen-metal exchange and trapping with dioxaborolane reagent 69 produced metalloindole 70.<sup>23</sup> Suzuki coupling of 70 with the known vinyl bromide 71,<sup>24</sup> smoothly provided olefin 72.<sup>25</sup> Final conversion of 72 to the coupling fragment 62 was accomplished using a sequence involving selective hydrogenation of the terminal olefin,<sup>26</sup> bromination at the C(3) position,<sup>27</sup> and halogen-metal exchange/trapping with the dioxaborolane reagent (69).<sup>28</sup>



### 2.3.4 Construction of the Fully Substituted Bis(indole)pyrazinone Core

With the appropriate fragments in hand (**62**, **63**, and **54b**), the critical threecomponent coupling reaction sequence was explored. Suzuki coupling of dihalopyrazine **63** and indole **54b** proceeded selectively to afford the coupled indolylpyrazine **73** (Scheme 2.3.4). In the second Suzuki coupling of dibromide **73** with boronic ester **62**, we were delighted to find that the desired bis(indole)pyrazine **74** formed in good yield and with complete selectivity for coupling of the pyrazinyl bromide in the presence of the indolyl bromide. Precise temperature control was critical for the success of both coupling reactions (23 °C and 50 °C, respectively). Importantly, the selectivity of the second Suzuki reaction depended not only on temperature, but also on the exact identity of each coupling substrate. In fact, varying protective groups on the indole nitrogen in **54a** had a dramatic effect on halide reactivity, as competitive coupling of the indolyl bromide



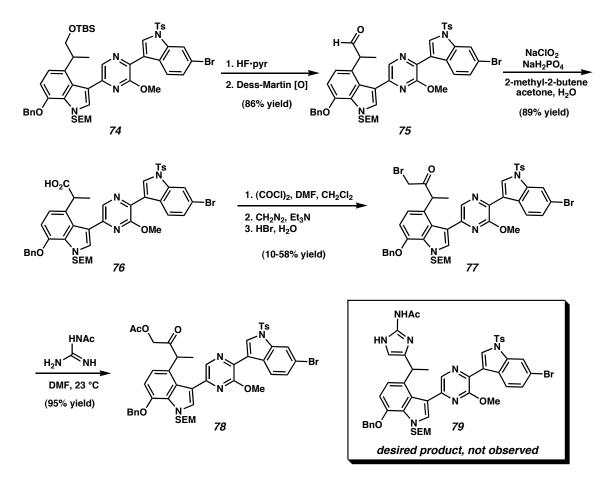
## 2.4 End-Game Studies

# 2.4.1 End-Game Strategy 1

Having established a viable route to the core structure of dragmacidin D (**5**), we focused our efforts on completing the natural product synthesis. Selective cleavage of the silyl ether in **74**,<sup>29</sup> followed by Dess-Martin oxidation<sup>30</sup> furnished aldehyde **75**, which was further oxidized to its carboxylic acid derivative **76** (Scheme 2.4.1). Conversion of **76** to bromoketone **77** was accomplished by an Arndt-Eistert-type homologation, followed by treatment with aqueous HBr. The extreme sensitivity of the intermediate acid chloride was particularly troublesome and required that the diazomethane used in the reaction sequence be dried thoroughly over both potassium hydroxide and sodium metal immediately before use.<sup>31</sup> In addition, chemical yields for this homologation varied to a large extent and caused substantial material throughput problems. Nonetheless, with bromide **77** in hand, we explored installation of the aminoimidazole functionality. Reaction of bromide **79** based on model studies.<sup>32,33</sup> However, the only product observed in

the reaction was acetoxyketone **78**, in quantitative yield. Although a variety of guanidine sources, solvents, and temperatures were explored to promote the synthesis of aminoimidazole **79**, all of our efforts resulted in the formation of the same undesired product (**78**).

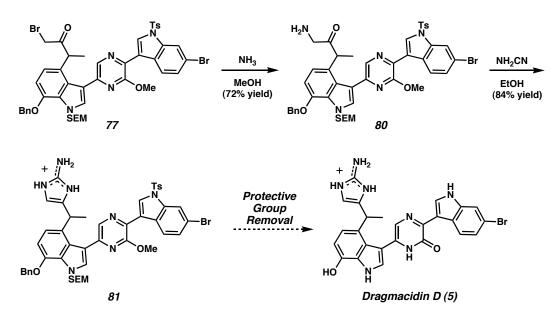
Scheme 2.4.1



# 2.4.2 End-Game Strategy 2

Through our studies it was observed that nucleophilic displacement of the alkyl bromide in **77** was actually a facile process. Thus, we considered an alternative endgame strategy that would exploit this reactivity (Scheme 2.4.2). Treatment of **77** with ammonia afforded aminoketone **80** which, in turn, underwent facile condensation with cyanamide to produce the desired aminoimidazole product (**81**).<sup>34</sup> At this point, all that remained in order to complete the total synthesis of dragmacidin D (**5**) was the removal of the four protective groups from **81**. Despite several months of experimentation, our efforts to complete the total synthesis of (**5**) were accompanied by decomposition of the aminoimidazole moiety, which was exceptionally unstable to the basic conditions needed to remove the protective groups that we had strategically chosen (vide supra).

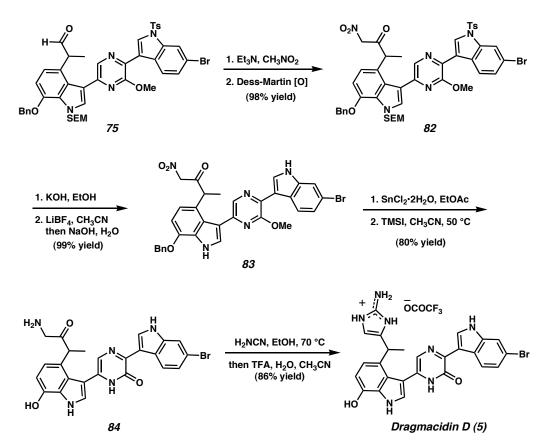


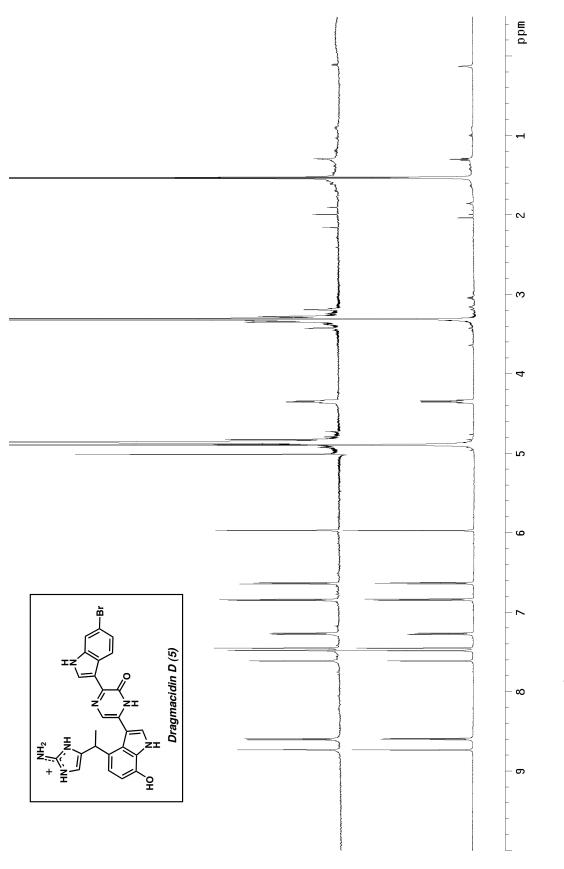


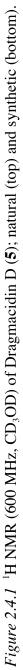
# 2.4.3 End-Game Strategy 3: The Total Synthesis of Dragmacidin D

The possibility of installing the aminoimidazole as the last step of the total synthesis, after the full deprotection of a late-stage intermediate, was explored next. In addition, we sought an alternative one-carbon homologation reaction in place of the unreliable Arndt-Eistert sequence. After extensive experimentation, we found that nitromethane addition<sup>35</sup> to aldehyde **75** and subsequent oxidation produced **82** in high yield (Scheme 2.4.3).<sup>36</sup> Deoxygenated ethanolic potassium hydroxide facilitated removal of the *N*-tosyl group,<sup>37</sup> while lithium tetrafluoroborate followed by aqueous sodium hydroxide effected complete hydrolysis of the SEM group (**82**  $\rightarrow$  **83**). Reduction of nitroketone **83** with stannous chloride,<sup>38</sup> then cleavage of the benzyl and methyl ethers with iodotrimethylsilane, revealed fully deprotected aminoketone **84**.<sup>39</sup> Final installation of the aminoimidazolium unit occurred by treatment of **84** with cyanamide followed by trifluoroacetic acid workup to produce the natural product (**5**) in 86% yield. Synthetic dragmacidin D (**5**) was spectroscopically identical to samples obtained from natural sources (Figure 2.4.1).

Scheme 2.4.3





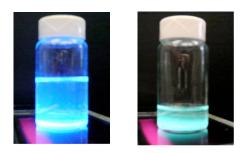


### 2.4.4 Subtleties of Late-Stage Manipulations

The exact order of final synthetic events presented herein was essential for the completion of dragmacidin D. In particular, intermediates **82** through **84** were highly labile when treated under a variety of other conditions. For example, attempts to reduce nitroketone **82** or to remove the SEM group prior to detosylation resulted in substantial nonspecific decomposition. Likewise, efforts to deprotect **83** prior to reduction of the nitro group led to decomposition of the nitroketone moiety. Finally, reversing the order of the final two steps (i.e., aminoimidazole formation followed by treatment with TMSI) afforded only a low yield of dragmacidin D (ca. 5%).

Also noteworthy is the brilliant fluorescent nature of most of the bis(indole) pyrazine/pyrazinone intermediates. By shining longwave UV light from a benchtop UV lamp ( $\lambda = 365$  nm), we were able to monitor and isolate extremely small amounts of compounds in large quantitites of solvent during necessary reversed-phase chromatography (i.e., ca. 1 mg/30-50 mL solvent). Two typical examples that demonstrate the fluorescent behavior of these compounds are shown below (Figure 2.4.2).<sup>40</sup>

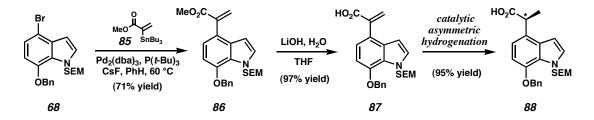
*Figure* 2.4.2



## 2.5 An Asymmetric Route to Dragmacidin D

With the racemic synthesis of dragmacidin D (5) completed, our attention turned to the development of an asymmetric route to the natural product. It was envisioned that the stereocenter present in 5 could arise from an asymmetric hydrogenation<sup>41</sup> of 87, a compound prepared readily from 68 via Fu-modified Stille coupling<sup>42</sup> with 85<sup>43</sup> followed by saponification (Scheme 2.5.1). Dr. Richmond Sarpong investigated the ruthenium-catalyzed asymmetric reduction of 87 by varying several reaction parameters including solvent, pressure, temperature, and additive effects (Table 2.5.1). Ultimately, hydrogenation of 87 in the presence of chiral ruthenium complex 89 under optimized conditions (78 atm of hydrogen, at -10 °C in MeOH) resulted in the formation of enantioenriched carboxylic acid 88 in 90% ee.

Scheme 2.5.1

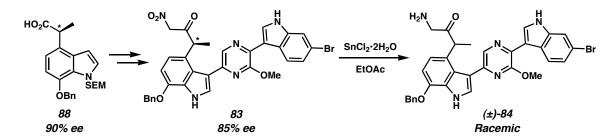


# Table 2.5.1

catalyst	solvent	temp	additive	$H_2$ pressure	time	conversion	ee
	MeOH	25 °C	-	30 atm	14 h	100%	83%
	MeOH/ CH <sub>2</sub> Cl <sub>2</sub> (1:1)	25 °C	-	30 atm	14 h	100%	59%
PPh <sub>2</sub>	CH <sub>2</sub> Cl <sub>2</sub>	25 °C	-	30 atm	14 h	40%	0%
	MeOH	25 °C	Et <sub>3</sub> N (1 equiv)	30 atm	14 h	100%	80%
PPh <sub>2</sub>	MeOH	5 °C	Et <sub>3</sub> N (1 equiv)	30 atm	72 h	100%	86%
	MeOH	5 °C	-	30 atm	72 h	100%	87%
89	MeOH	-10°C	-	78 atm	72 h	100%	90%

Carboxylic acid **88** could be elaborated to nitroketone **83** without substantial loss in enantiomeric excess (Scheme 2.5.2). However, upon Sn(II)-promoted reduction to aminoketone **84**, racemization occurred. Alternative conditions to access enantiopure **84** were explored but were also unsuccessful at promoting reduction without epimerization of the benzylic  $\alpha$ -keto stereocenter. The enantiopurity of natural dragmacidin D (**5**) is also somewhat uncertain. During the first isolation of **5**, no optical rotation could be detected.<sup>1a</sup> Subsequently, a very small rotation value was reported ( $[\alpha]_D + 12^\circ$  (*c* 0.95, EtOH)).<sup>1b</sup> Nonetheless, an asymmetric total synthesis of dragmacidin D remains an elusive goal.





## 2.6 Conclusion

In summary, we have completed the first total synthesis of the important bis(indole) alkaloid dragmacidin D (5). The concise route that we have developed (longest linear sequence of 17 steps from 66) relies on a series of halogen-selective Suzuki couplings and a meticulous late-stage sequence to complete the natural product synthesis.

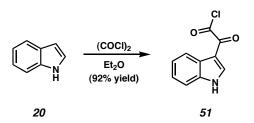
### 2.7 Experimental Section

### 2.7.1 Materials and Methods

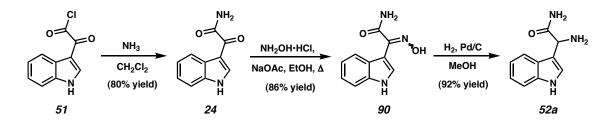
Unless stated otherwise, reactions were performed in flame-dried glassware under a nitrogen or argon atmosphere using dry, deoxygenated solvents. All other commercially obtained reagents were used as received. Solvents were dried by passage through an activated alumina column under argon. Reaction temperatures were controlled by an IKAmag temperature modulator. Thin-layer chromatography (TLC) was performed using E. Merck silica gel 60 F254 precoated plates (0.25 mm) and visualized using a combination of UV, anisaldehyde, ceric ammonium molybdate, and potassium permanganate staining. ICN Silica gel (particle size 0.032-0.063 mm) was used for flash chromatography. Disposable Sep-Pak C<sub>18</sub> Vac Cartridges were purchased from Waters and used for all reversed-phase filtrations. HPLC analysis was performed on a Beckman Gold system using a Rainin C<sub>18</sub>, Microsorb MV, 5µm, 300 x 4.6 mm reversed-phased column in 0.1% (w/v) TFA with acetonitrile/H<sub>2</sub>O as eluent and a flow rate of 1.0 ml/min, gradient elution of 1.25% acetonitrile/min. Preparatory reversedphase HPLC was performed on a Beckman HPLC with a Waters DeltaPak 25 x 100 mm, 100 mm C<sub>18</sub> column equipped with a guard, 0.1% (w/v) TFA with acetonitrile/H<sub>2</sub>O as eluent, and gradient elution of 0.50% acetonitrile/min. For all reversed-phase purifications,  $H_2O$  (18M $\Omega$ ) was obtained from a Millipore MiliQ water purification system and TFA from Halocarbon, Inc. <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on either a Varian Mercury 300 (at 300 MHz and 75 MHz, respectively), Varian Mercury 500 (at 500 MHz and 125 MHz, respectively), or on a Varian Mercury 600 (600 MHz for proton only) spectrometer and are reported relative to Me<sub>4</sub>Si ( $\delta$  0.0). Data for <sup>1</sup>H NMR spectra

are reported as follows: chemical shift ( $\delta$  ppm), multiplicity, coupling constant (Hz), and integration. Data for <sup>13</sup>C NMR spectra are reported in terms of chemical shift. IR spectra were recorded on a Perkin Elmer Paragon 1000 spectrometer and are reported in frequency of absorption (cm<sup>-1</sup>). UV spectra were measured on a Hewlett-Packard Model 8452A diode array spectrophotometer. High resolution mass spectra were obtained from the UC Irvine Mass Spectral Facility. Chiral HPLC was performed on a Chiralcel AD column (4.6 mm x 25 cm) obtained from Daicel Chemical Industries, Ltd.

#### **2.7.2 Preparative Procedures**



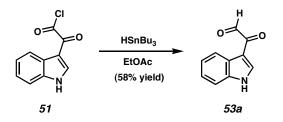
**Glyoxal chloride 51.** To a solution of indole **20** (20.0 g, 171 mmol) in anhydrous  $Et_2O$  (340 mL) at 0 °C, was added oxalyl chloride (17.3 mL, 198 mmol) dropwise over 30 min. The reaction mixture was stirred at 0 °C for 3 h, then allowed to warm to 23 °C over 1 h. The resulting yellow crystals were collected by filtration, washed with anhydrous  $Et_2O$  (100 mL), and dried under vacuum to yield **51** (32.52 g, 92% yield), which was used without further purification.



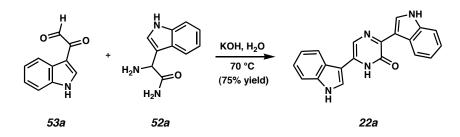
Aminoamide 52a. Gaseous ammonia was bubbled through a suspension of 51 (12.4 g, 59.7 mmol) in  $CH_2Cl_2$  (300 mL) for 10 min. After stirring for 30 min, the solvent was removed under reduced pressure. Addition of  $H_2O$  (600 mL) was followed by extraction of the resulting heterogeneous mixture with EtOAc (2 x 600 mL). The combined organic layers were washed with brine (300 mL), dried over magnesium sulfate, and evaporated under reduced pressure to afford the crude amide 24 (9.0 g, 80% yield), which was used without further purification.

To a suspension of **24** (500 mg, 5.32 mmol) in CH<sub>3</sub>OH (7.8 mL) was added hydroxylamine hydrochloride (2.0 g, 39.9 mmol) in H<sub>2</sub>O (3.8 mL) and sodium acetate (1.64 g, 39.9 mmol) in H<sub>2</sub>O (3.8 mL). The resulting heterogeneous mixture was heated under reflux for 10 h and allowed to cool to 23 °C. The solvent was removed under reduced pressure, and the remaining crude residue was extracted with EtOAc (3 x 20 mL). The combined organic layers were washed with brine (20 mL) and dried over sodium sulfate. After removal of solvent under reduced pressure, the crude residue was purified by flash chromatography (3:1 CH<sub>2</sub>Cl<sub>2</sub>/hexanes eluent) to afford oxime **90** (454 mg, 86% yield), which was used without further purification.

To a solution of **90** (4.07g, 20 mmol) in CH<sub>3</sub>OH in a stainless steel bomb was added 10% palladium on charcoal (450 mg). The bomb was then purged with hydrogen and pressurized to 450 psi. After stirring for 14 h at 23 °C, the palladium on carbon was removed via filtration, and the solvent was removed under reduced pressure. Passage through a plug of celite (CH<sub>3</sub>OH eluent) afforded the desired aminoamide **52a** (3.5 g, 92% yield) as a yellow oil:  $R_f$  0.10 (5:1 EtOAc:CH<sub>3</sub>OH); <sup>1</sup>H NMR (300 MHz, DMSO- $d_6$ )  $\delta$  10.93 (s, 1H), 7.71 (d, J = 7.7 Hz, 1H), 7.43 (s, 1H), 7.33 (d, J = 7.7 Hz, 1H), 7.23 (s, 1H), 7.07-6.93 (comp. m, 3H), 4.56 (s, 1H), 2.38 (br s, 2H); <sup>13</sup>C NMR (75 MHz, DMSO- $d_6$ )  $\delta$  175.9, 136.2, 125.7, 122.8, 120.9, 119.5, 118.2, 116.2, 111.3, 52.5; IR (film) 3176, 1660, 1592 cm<sup>-1</sup>; HRMS-NH<sub>3</sub>CI (*m*/*z*): [M + H]<sup>+</sup> calc'd for C<sub>10</sub>H<sub>12</sub>N<sub>3</sub>O, 190.0980; found, 190.0978.

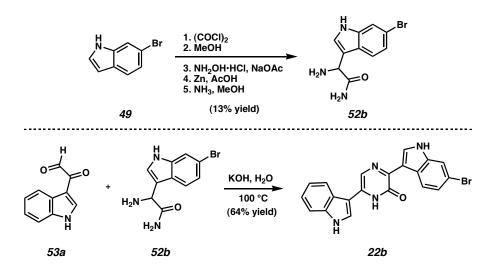


**Ketoaldehyde 53a.** To a suspension of **51** (22.0 g, 106 mmol) in EtOAc (106 mL) at 0 °C, was added a solution of tributyltin hydride (28.5 mL, 106 mmol) in EtOAc (158 mL). The reaction mixture was stirred at 0 °C for 30 min, warmed to 23 °C, then stirred for an additional 15 h. Hexanes (150 mL) was added, and the resulting yellow powder was collected by filtration. Washing with copious amounts of hexanes (1 L) and drying under vacuum, gave ketoaldehyde **53a** (10.6 g, 58% yield):  $R_f$  0.76 (13:7 CHCl<sub>3</sub>:CH<sub>3</sub>OH); <sup>1</sup>H NMR (300 MHz, acetone- $d_6$ )  $\delta$  9.54 (s, 1H), 8.65 (s, 1H), 8.36-8.33 (comp. m, 1H), 7.61-7.58 (comp. m, 1H), 7.33-7.29 (comp. m, 2H); <sup>13</sup>C NMR (75 MHz, acetone- $d_6$ )  $\delta$  192.9, 183.2, 138.1, 125.3, 124.6, 124.1, 123.5, 123.1, 113.6, 113.3; IR (film) 3117, 1628, 1580, 1518, 1234 cm<sup>-1</sup>; HRMS-NH<sub>3</sub>CI (*m/z*): [M + H]<sup>+</sup> calc'd for C<sub>10</sub>H<sub>8</sub>NO<sub>2</sub>, 174.0555; found, 174.0555.



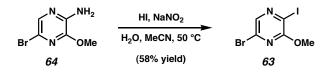
**Pyrazinone 22a.** To  $H_2O$  (17 mL) at 70 °C was added ketoaldehyde **53a** (300 mg, 1.73 mmol) and aminoamide **52a** (321 mg, 1.73 mmol), followed by powdered potassium hydroxide (487 mg, 8.67 mmol). After stirring at 70 °C for 5 h, the reaction mixture was allowed to cool to 23 °C, poured into saturated aq. ammonium chloride (75

mL), and extracted with EtOAc (4 x 75 mL). The combined organic layers were dried briefly over sodium sulfate and concentrated under reduced pressure to afford the desired pyrazinone **22a** (423 mg, 75% yield) as an orange/red solid:  $R_f 0.57$  (5:1 CH<sub>2</sub>Cl<sub>2</sub>: CH<sub>3</sub>OH); <sup>1</sup>H NMR (300 MHz, DMSO- $d_6$ )  $\delta$  12.23 (s, 1H), 11.75 (s, 1H), 11.52 (s, 1H), 8.75 (s, 1H), 8.69 (d, J = 7.3 Hz, 1H), 8.11 (d, J = 2.6 Hz, 1H,), 8.40-7.82 (comp. m, 2H), 7.51-7.45 (comp. m, 2H), 7.25-7.12 (comp. m, 4H); <sup>13</sup>C NMR (125 MHz, DMSO- $d_6$ )  $\delta$  155.5, 147.2, 136.8, 136.2, 130.1, 126.3, 125.9, 124.1, 122.7, 122.2, 122.0, 120.6, 120.1, 119.7, 112.2, 111.9, 111.6, 106.8; IR (film) 3307, 1633, 1602, 1421 cm<sup>-1</sup>; HRMS-ESI (m/z): [M + Na]<sup>+</sup> calc'd for C<sub>20</sub>H<sub>15</sub>N<sub>4</sub>ONa, 349.1065; found, 349.1070.

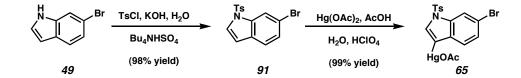


**Bromopyrazinone 22b.** To a solution of bromoindole **49**<sup>17</sup> (3.4 g, 17.3 mmol) in anhydrous  $Et_2O$  (35 mL) at 0 °C was added oxalyl chloride (1.76 mL, 20.1 mmol) dropwise over 30 min. The reaction mixture was stirred at 0 °C for 3 h, then allowed to warm to 23 °C over 1 h. The resulting yellow crystals were collected by filtration, washed with anhydrous  $Et_2O$  (15 mL), and dried under vacuum. To a suspension of these crystals at 0 °C in CH<sub>2</sub>Cl<sub>2</sub> (70 mL) was added MeOH (2.2 mL, 70 mmol). The reaction

mixture was warmed to 23 °C, stirred for 12 h, and then filtered to collect the product. To this crude material in MeOH (87 mL) was added NaOAc (8.09g, 98.4 mmol) in H<sub>2</sub>O (19 mL), followed by NH<sub>2</sub>OH•HCl (9.84 g, 98.4 mmol) in H<sub>2</sub>O (19 mL). The mixture was heated to 80 °C for 24 h then cooled to 23 °C. After removal of solvent under reduced pressure, the aqueous residue was extracted with EtOAc (3 x 50 mL). The combined organic layers were washed with brine (30 mL) and evaporated in vacuo. The resulting material was purified by flash chromatography (2:1 EtOAc:hexanes eluent) to afford an off-white solid. To this compound in MeOH (84 mL) was added 1 N HCl (84 mL) followed by zinc dust (2.75 g, 42.1 mmol). The reaction mixture was stirred for 10 min and filtered (MeOH eluent). The filtrate was evaporated under reduced pressure to afford an off-white solid that was triturated with CH<sub>2</sub>Cl<sub>2</sub>, diluted with 1 N NaOH (25 mL), and extracted with CH<sub>2</sub>Cl<sub>2</sub> (4 x 50 mL). The combined organic layers were dried over MgSO<sub>4</sub> and evaporated. The residue was dissolved in a saturated solution of NH<sub>3</sub> in MeOH (50 mL) at 23 °C and stirred for 72 h. After removal of solvents under vacuum and trituration with Et<sub>2</sub>O, bromoamide **52b** (600 mg, 13% yield, 5 steps) was obtained as an off-white solid. This product was used immediately in the subsequent reaction. To H<sub>2</sub>O (1.5 mL) at 100 °C was added ketoaldehyde 53a (26.2 mg, 0.152 mmol) and bromoamide **52b** (40 mg, 0.152 mmol), followed by powdered potassium hydroxide (43 mg, 0.76 mmol). After stirring at 100 °C for 5 h, the reaction mixture was allowed to cool to 23 °C, poured into saturated aq. ammonium chloride (10 mL), and extracted with EtOAc (4 x 10 mL). The combined organic layers were dried briefly over sodium sulfate and concentrated under reduced pressure to afford known pyrazinone **22b**<sup>44</sup> (39 mg, 64% yield) as an orange/red solid:  $R_f 0.52$  (7:1 CH<sub>2</sub>Cl<sub>2</sub>: CH<sub>3</sub>OH).



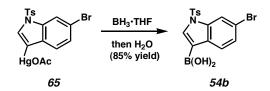
**Iodopyrazine 63.** To a thick-walled flask charged with **64**<sup>15</sup> (100 mg, 0.49 mmol) was added acetonitrile (1.0 mL), H<sub>2</sub>O (1.5 mL), and 48% aq. HI (1.3 mL). The resulting solution was cooled in an ice bath, and a solution of sodium nitrite (600 mg, 8.7 mmol) in H<sub>2</sub>O (1.0 mL) was added in a dropwise fashion. The reaction mixture was sealed, allowed to warm to 23 °C, then stirred at 50 °C for 30 h. After cooling, the solution was poured into 20% aq. NaOH and extracted with Et<sub>2</sub>O (3 x 20 mL). The combined organic layers were washed with saturated aq. sodium metabisulfite (20 mL) and brine (20 mL), dried over anhydrous sodium sulfate, then evaporated under reduced pressure. The crude product was then dissolved in a CH<sub>2</sub>Cl<sub>2</sub>/hexanes mixture (1:1) and filtered over silica gel (1:1 CH<sub>2</sub>Cl<sub>2</sub>:hexanes eluent) to provide iodopyrazine **63** (90 mg, 58% yield) as a white powder:  $R_f$  0.52 (1:1 CH<sub>2</sub>Cl<sub>2</sub>:hexanes); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.07 (s, 1H), 4.05 (s, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  158.8, 139.2, 136.1, 104.4, 56.2; IR (KBr) 1357, 1150 cm<sup>-1</sup>; HRMS-NH<sub>4</sub>CI (*m*/*z*): [M]<sup>+</sup> calc'd for C<sub>4</sub>H<sub>2</sub>RIN<sub>2</sub>O, 313.8552; found, 313.8553.



**Organomercurial 65.** To a solution of **49**<sup>17</sup> (4.35 g, 22.2 mmol) in toluene (22 mL) was added tetrabutylammonium hydrogensulfate (520 mg, 1.54 mmol), KOH (50% aq. solution, 28 mL), and a solution of *p*-toluenesulfonyl chloride (5.08 g, 26.6 mmol) in toluene (44 mL). After stirring for 4 h, H<sub>2</sub>O (40 mL) was added and the layers separated. The organic layer was washed with H<sub>2</sub>O (2 x 20 mL) and brine (20 mL), dried over

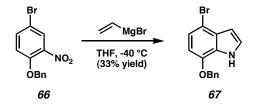
magnesium sulfate, and concentrated under reduced pressure to afford 6-bromo-*N*-tosylindole **91** (7.6 g, 98% yield) as an off-white powder:  $R_f 0.25$  (9:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.19 (s, 1H), 7.75 (d, *J* = 8.2 Hz, 2H), 7.53 (d, *J* = 3.3 Hz, 1H), 7.33 (comp. m, 2H), 7.21 (d, *J* = 7.7 Hz, 2H), 6.61 (d, *J* = 3.3 Hz, 1H), 2.32 (s, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  145.5, 135.6, 135.1, 130.2, 129.7, 126.9, 126.8, 122.6, 118.3, 116.7, 108.9, 21.7; IR (film) 1364, 1169 cm<sup>-1</sup>; HRMS-NH<sub>3</sub>CI (*m/z*): [M]<sup>+</sup> calc'd for C<sub>15</sub>H<sub>12</sub>BrNO<sub>2</sub>S, 348.9772; found, 348.9773.

To a solution of 6-bromo-*N*-tosylindole **91** (7.6 g, 21.7 mmol) in acetic acid (145 mL) was added mercuric acetate (6.92 g, 21.7 mmol). After stirring at 23 °C for 15 min, perchloric acid (5 drops) was added. The mixture was stirred for 24 h, poured into H<sub>2</sub>O (200 mL), then filtered. The resulting white solid was washed with copious amounts of H<sub>2</sub>O and dried under vacuum for 12 h to afford organomercurial derivative **65** (13.05 g, 99% yield) as an unstable white powder that was used immediately without further purification:  $R_f 0.57$  (2:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, DMSO- $d_6$ )  $\delta$  8.02 (d, J = 1.1 Hz, 1H), 7.76 (d, J = 8.4 Hz, 2H), 7.71 (d, J = 8.4 Hz, 1H), 7.52 (s, 1H), 7.42-7.39 (comp. m, 3H), 2.32 (s, 3H), 1.96 (s, 3H).

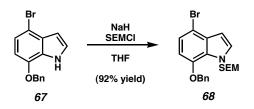


**Boronic acid 54b.** To a solution of **65** (3.91 g, 6.4 mmol) in THF (128 mL) at 23 °C was added borane (1 M in THF, 32 mL, 32 mmol). The resulting solution was stirred for 1 h, then carefully quenched by the dropwise addition of  $H_2O$  (38 mL). After filtration, the organic solvent was evaporated under reduced pressure, and the residue was

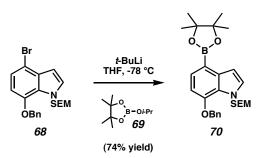
extracted with EtOAc (2 x 60 mL). The combined organic layers were washed with brine (30 mL) and concentrated under reduced pressure. Trituration of the crude product with hexanes (4x) afforded boronic acid **54b** (2.15 g, 85% yield) as an unstable off-white solid that was used immediately without further purification.



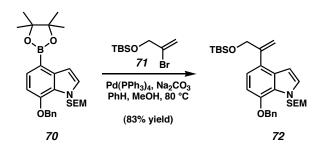
**Indole 67.** To a solution of **66**<sup>21</sup> (14.0 g, 45.5 mmol) in THF (455 mL) at -40 °C was added vinylmagnesium bromide (1.0 M in THF, 159 mL, 159 mmol) in a dropwise fashion. The reaction mixture was stirred at -40 °C for 4 h and then poured into a saturated aq. ammonium chloride (350 mL). The reaction mixture was extracted with Et<sub>2</sub>O (2 x 200 mL), and the combined organic layers were washed with brine (200 mL), dried over magnesium sulfate, and evaporated under reduced pressure. CH<sub>2</sub>Cl<sub>2</sub> (50 mL) was added, and the resulting suspension was filtered over a pad of silica gel topped with celite. Removal of solvent under reduced pressure afforded the crude product as a red oil, which was further purified by flash chromatography (8:1 hexanes:Et<sub>2</sub>O eluent) to yield 7-benzyloxy-4-bromoindole 67 (4.54 g, 33% yield) as a yellow oil:  $R_f 0.30$  (4:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 8.46 (br s, 1H), 7.50-7.40 (comp. m, 5H), 7.20 (d, J = 8.2 Hz, 1H), 7.16 (app.t, J = 2.7 Hz, 1H), 6.61-6.58 (comp. m, 2H), 5.17 (s, 2H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 145.0, 136.8, 129.7, 128.8, 128.5, 128.1, 126.9, 124.4, 122.6, 106.1, 104.4, 103.4, 70.6; IR (film) 3426, 1228 cm<sup>-1</sup>; HRMS-NH<sub>3</sub>CI (m/z): [M]<sup>+</sup> calc'd for C<sub>15</sub>H<sub>12</sub>BrNO, 301.0101; found, 301.0102.



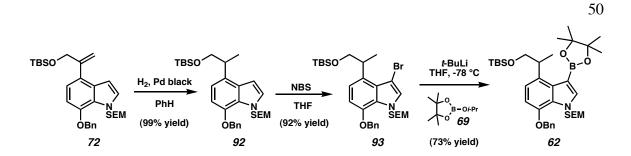
Indole 68. To a stirred suspension of NaH (60% dispersion in mineral oil, 154 mg, 4.0 mmol) in THF (15 mL) at 0 °C was added a solution of 67 (930 mg, 3.08 mmol) in THF (15 ml). The reaction mixture was allowed to warm to 23 °C and stirred for 30 min. The solution was cooled to 0 °C, SEMCl (600 µL, 3.4 mmol) was added, and the mixture was stirred at 23 °C for 6 h. The reaction mixture was poured into saturated aq. ammonium chloride (20 mL) and extracted with Et<sub>2</sub>O (2 x 30 mL). The combined organic layers were washed with brine (20 mL), dried over magnesium sulfate, and The crude residue was purified by flash evaporated under reduced pressure. chromatography (14:1 hexanes: EtOAc eluent) to afford 68 (1.22 g, 92% yield) as a yellow oil:  $R_f 0.51$  (4:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.53-7.50 (comp. m, 2H), 7.46-7.37 (comp. m, 3H), 7.21 (d, J = 3.3 Hz, 1H), 7.18 (d, J = 8.2 Hz, 1H), 6.63 (d, J = 8.2 Hz, 1H), 6.59 (d, J = 3.3 Hz, 1H), 5.73 (s, 2H), 5.20 (s, 2H), 3.45 (t, J = 8.2 Hz)Hz, 2H), 0.84 (t, J = 8.2 Hz, 2H), -0.06 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  146.3, 136.8, 131.6, 129.8, 128.8, 128.3, 127.8, 126.2, 123.0, 106.4, 105.7, 103.6, 77.7, 70.8, 65.5, 17.9, -1.2; IR (film) 1244, 1054 cm<sup>-1</sup>; HRMS-NH<sub>3</sub>CI (m/z): [M]<sup>+</sup> calc'd for C<sub>21</sub>H<sub>26</sub>BrNO<sub>2</sub>Si, 431.0916; found, 431.0919.



Boronic ester 70. To a solution of 68 (3.81 g, 8.8 mmol) in THF (147 mL) at -78 °C was added t-BuLi (1.7 M in pentane, 11.4 mL, 19.4 mmol). Following addition, the reaction mixture was stirred for 15 min at -78 °C, then borolane 69 (3.6 mL, 17.6 mmol) was added. The mixture was stirred at -78 °C for 1.5 h, allowed to warm to 23 °C, then quenched with saturated aq. ammonium chloride (75 mL). The layers were separated, and the aqueous portion was extracted with Et<sub>2</sub>O (3 x 75 mL). The combined organic layers were washed with brine (100 mL), briefly dried over magnesium sulfate, and evaporated under reduced pressure. The crude residue was purified by flash chromatography (14:1 hexanes: EtOAc eluent) to give boronic ester 70 (3.11 g, 74%) yield) as a yellow oil:  $R_f 0.32$  (9:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.67 (d, J = 7.7 Hz, 1H), 7.59-7.56 (comp. m, 2H), 7.48-7.39 (comp. m, 3H), 7.25 (d, J = 3.3 Hz, 1H), 7.12 (d, J = 3.3 Hz, 1H), 6.83 (d, J = 7.7 Hz, 1H), 5.81 (s, 2H), 5.28 (s, 2H), 3.49 (t, J = 8.2 Hz, 2H), 1.44 (s, 12H), 0.87 (t, J = 8.2 Hz, 2H), -0.02 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 149.3, 137.0, 136.5, 129.9, 129.7, 128.7, 128.1, 127.8, 125.2, 105.1, 104.2, 83.2, 77.5, 70.3, 65.2, 25.1, 17.9, -1.3; IR (film) 1371, 1330, 1251 cm<sup>-1</sup>; HRMS-NH<sub>3</sub>CI (m/z): [M]<sup>+</sup> calc'd for C<sub>27</sub>H<sub>38</sub>BNO<sub>4</sub>Si, 479.2668; found, 479.2673.



Olefin 72. A solution containing boronic ester 70 (3.17 g, 6.62 mmol) and bromide 71 (3.32 g, 13.2 mmol) in benzene (130 mL), CH<sub>3</sub>OH (30 mL), and aq. sodium carbonate (2 M, 11 mL) was deoxygenated by bubbling a stream of argon through the reaction mixture for 5 min. Tetrakis(triphenylphosphine)palladium(0) (1.15 g, 0.99 mmol) was then added and the flask was equipped with a reflux condenser. The mixture was heated to 80 °C for 2 h and allowed to cool to 23 °C. To the reaction vessel was added sodium sulfate (10 g), which was allowed to stand for 30 min. After filtration over a pad of silica gel (CH<sub>2</sub>Cl<sub>2</sub> eluent) and concentrating to dryness under reduced pressure, the resulting residue was purified by flash chromatography (1:1 CH<sub>2</sub>Cl<sub>2</sub>:hexanes eluent) to provide olefin **72** (2.87 g, 83% yield) as a yellow oil:  $R_f 0.53$  (9:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 7.55-7.53 (comp. m, 2H), 7.46-7.37 (comp. m, 3H), 7.18 (d, J = 3.3 Hz, 1H), 6.94 (d, J = 7.7 Hz, 1H), 6.74 (d, J = 8.2 Hz, 1H), 6.67 (d, J = 3.3 Hz, 1H), 5.78 (s, 2H), 5.62 (m, 1H), 5.40 (m, 1H), 5.24 (s, 2H), 4.55 (s, 2H), 3.48 (t, J = 8.2 Hz, 2H), 0.99 (s, 9H), 0.85 (t, J = 8.2 Hz, 2H), 0.15 (s, 6H), -0.06 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 146.9, 146.3, 137.2, 130.0, 129.3, 128.8, 128.2, 127.8, 126.3, 126.0, 118.9, 111.9, 104.2, 103.0, 77.6, 70.6, 65.6, 65.4, 26.2, 18.7, 18.0, -1.2, -5.1; IR (film) 1250, 1088 cm<sup>-1</sup>; HRMS-ESI (m/z):  $[M + H]^+$  calc'd for C<sub>30</sub>H<sub>46</sub>NO<sub>3</sub>Si<sub>2</sub>, 524.3016; found, 524.3019.

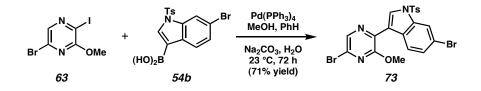


**Boronic ester 62.** To a solution of olefin **72** (545 mg, 1.04 mmol) in benzene (12 mL), saturated with hydrogen, was added palladium black (35 mg, 0.33 mmol). The reaction vessel was purged with hydrogen and kept under a hydrogen atmosphere (1 atm) with vigorous stirring for 1 h. Palladium black was removed via filtration through a pad of silica gel (benzene eluent) to afford the reduced silyl ether **92** (542 mg, 99% yield) as a yellow oil:  $R_f$  0.53 (9:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.53-7.50 (comp. m, 2H), 7.43-7.34 (comp. m, 3H), 7.16 (d, *J* = 3.3 Hz, 1H), 6.86 (d, *J* = 7.7 Hz, 1H), 6.71 (d, *J* = 7.7 Hz, 1H), 6.60 (d, *J* = 3.3 Hz, 1H), 5.75 (s, 2H), 5.20 (s, 2H), 3.87 (dd, *J* = 9.9, 4.9 Hz, 1H), 3.62 (app.t, *J* = 9.3 Hz, 1H), 3.45 (t, *J* = 8.2 Hz, 2H), 3.30 (m, 1H), 1.40 (d, *J* = 6.6 Hz, 3H), 0.89 (s, 9H), 0.82 (t, *J* = 8.2 Hz, 2H), 0.00 (s, 6H), -0.09 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  145.3, 137.3, 130.7, 129.5, 128.8, 128.7, 128.1, 127.8, 125.7, 117.3, 104.4, 101.7, 77.6, 70.6, 69.0, 65.4, 39.1, 26.3, 18.7, 18.0, 17.5, -1.1, -4.99, -5.04; IR (film) 1249, 1076 cm<sup>-1</sup>; HRMS-ESI (*m*/*z*): [M + Na]<sup>+</sup> calc'd for C<sub>30</sub>H<sub>47</sub>NO<sub>3</sub>Si<sub>2</sub>Na, 548.2992; found, 548.2997.

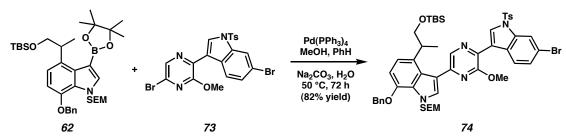
To a solution of the crude silyl ether **92** (270 mg, 0.51 mmol) in THF (5 mL) was added *N*-bromosuccinimide (92.2 mg, 0.51 mmol). After stirring for 5 min, the reaction mixture was poured into a saturated aq. solution of sodium metabisulfite (3 ml), extracted with Et<sub>2</sub>O (3 x 2 mL), and dried by passage through a plug of silica gel (Et<sub>2</sub>O eluent). After concentrating to dryness under reduced pressure, the crude residue was purified by flash chromatography (1:1 CH<sub>2</sub>Cl<sub>2</sub>:hexanes eluent) to furnish the 3-bromoindole derivative **93** (285 mg, 92% yield) as a yellow oil:  $R_f 0.47$  (1:1 CH<sub>2</sub>Cl<sub>2</sub>:hexanes); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.57-7.54 (comp. m, 2H), 7.49-7.40 (comp. m, 3H), 7.24 (s, 1H), 7.02 (d, J = 8.2 Hz, 1H), 6.80 (d, J = 7.7 Hz, 1H), 5.77 (d, J = 9.9 Hz, 1H), 5.73 (d, J = 10.4 Hz, 1H), 5.23 (s, 2H), 4.36 (m, 1H), 4.02 (dd, J = 9.6, 4.7 Hz, 1H), 3.65 (dd, J = 9.3, 8.3 Hz, 1H), 3.50 (t, J = 8.0 Hz, 2H), 1.46 (d, J = 7.1 Hz, 3H), 0.96 (s, 9H), 0.89 (t, J = 8.0 Hz, 2H), 0.08 (s, 3H), 0.06 (s, 3H), 0.00 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  145.0, 136.9, 130.1, 129.6, 128.7, 128.2, 127.8, 126.1, 125.9, 118.7, 105.2, 90.1, 77.7, 70.7, 69.3, 65.6, 34.7, 26.2, 18.6, 18.0, -1.1, -5.0, -5.1; IR (film) 1250 cm<sup>-1</sup>; HRMS-ESI (m/z): [M + Na]<sup>+</sup> calc'd for C<sub>30</sub>H<sub>40</sub>BrNO<sub>3</sub>Si<sub>2</sub>Na, 626.2097; found, 626.2079.

To a solution of the 3-bromoindole derivative **93** (2.5 g, 4.1 mmol) in THF (69 mL) at -78 °C was added *t*-BuLi (1.7 M in pentane, 5.4 mL, 9.1 mmol). The reaction mixture was stirred for 15 min at -78 °C and borolane **69** (1.69 mL, 8.3 mmol) was added. The mixture was stirred at -78 °C for 1 h, quenched with saturated aq. ammonium chloride (20 mL), and allowed to warm to 23 °C. The layers were separated, and the aqueous layer was extracted with Et<sub>2</sub>O (3 x 50 mL). The combined organic layers were washed with brine (75 mL), briefly dried over magnesium sulfate, and concentrated in vacuo. The crude residue was purified by flash chromatography (14:1 hexanes:EtOAc eluent) to afford boronic ester **62** (1.96 g, 73% yield) as an unstable colorless oil that was used immediately in the coupling reaction that follows:  $R_f$  0.38 (9:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz,  $C_6D_6$ )  $\delta$  7.79 (s, 1H), 7.35 (d, J = 7.7 Hz, 1H), 7.20-7.05 (comp. m, 5H), 6.65 (d, J = 8.0 Hz, 1H), 5.50 (d, J = 10.6 Hz, 1H), 5.46 (d, J = 10.3 Hz, 1H), 4.86 (s, 2H), 4.70 (m, 1H), 4.09 (dd, J = 9.5, 4.8 Hz, 1H), 3.91 (dd, J = 9.5, 7.3 Hz, 1H), 3.28

(t, *J* = 7.7 Hz, 2H), 1.62 (d, *J* = 6.6 Hz, 3H), 1.21 (s, 6H), 1.19 (s, 6H), 0.98 (s, 9H), 0.64 (t, *J* = 7.7 Hz, 2H), 0.00 (s, 3H), -0.01 (s, 3H), -0.19 (s, 9H).

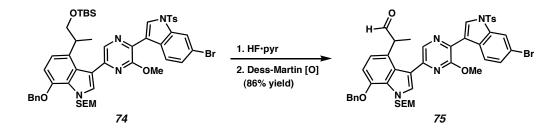


Indolylpyrazine 73. A solution containing iodopyrazine 63 (133 mg, 0.42 mmol) and boronic acid 54b (200 mg, 0.51 mmol) in benzene (10 mL), CH<sub>3</sub>OH (2 mL), and aq. sodium carbonate (2 M, 0.70 mL) was deoxygenated by bubbling a stream of argon through the reaction mixture for 5 min. Tetrakis(triphenylphosphine)palladium(0) (73 mg, 0.06 mmol) was then added, the flask was evacuated, and purged with N<sub>2</sub>. The reaction mixture was stirred at 23 °C for 72 h and guenched by addition of sodium sulfate (500 mg). Filtration over a pad of silica gel (CH<sub>2</sub>Cl<sub>2</sub> eluent) and concentration to dryness under reduced pressure, followed by trituration of the remaining residue with  $Et_{2}O(3x)$ further purification by flash chromatography (CH<sub>2</sub>Cl<sub>2</sub> eluent) afforded and indolylpyrazine **73** (161 mg, 71% yield) as an off-white powder:  $R_f 0.13$  (3:1 hexanes:  $CH_2Cl_2$ ); <sup>1</sup>H NMR (300 MHz,  $CDCl_3$ )  $\delta$  8.52 (d, J = 8.2 Hz, 1H), 8.45 (s, 1H), 8.33 (s, 1H), 8.18 (d, J = 2.2 Hz, 1H), 7.81 (d, J = 8.8 Hz, 2H), 7.43 (dd, J = 8.5, 2.2 Hz, 1H), 7.27 (d, J = 8.8 Hz, 2H), 4.19 (s, 3H), 2.36 (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  156.5, 145.9, 137.6, 137.0, 135.8, 135.1, 132.7, 130.4, 129.3, 128.2, 127.6, 127.2, 125.2, 119.3, 116.5, 116.1, 55.2, 21.8; IR (film) 1374, 1165 cm<sup>-1</sup>; HRMS-ESI (*m/z*): [M + H]<sup>+</sup> calc'd for C<sub>20</sub>H<sub>16</sub>Br<sub>2</sub>N<sub>3</sub>O<sub>3</sub>S, 535.9279; found, 535.9272.



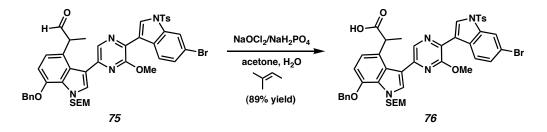
**Bis(indole)** 74. In a Schlenk flask, a solution containing dibromide 73 (82 mg, 0.15 mmol) and boronic ester 62 (129 mg, 0.20 mmol) in benzene (4 mL),  $CH_3OH$  (0.80 mL), and aq. sodium carbonate (2 M, 0.25 mL) was deoxygenated by bubbling a stream reaction 5 of argon through the mixture for min. Tetrakis(triphenylphosphine)palladium(0) (27 mg, 0.02 mmol) was then added, and the flask was evacuated, purged with N<sub>2</sub>, and sealed. The reaction mixture was heated to 50 °C for 72 h, cooled to 23 °C, then quenched by addition of sodium sulfate (300 mg). Following filtration through a pad of silica gel (CH<sub>2</sub>Cl<sub>2</sub> eluent) and evaporation to dryness in vacuo, the remaining residue was purified by flash chromatography (2:1 CH<sub>2</sub>Cl<sub>2</sub>:hexanes eluent) to give a crude product, which was further purified by flash chromatography (7:1 hexanes: EtOAc eluent) to afford bis(indole) 74 (122 mg, 82%) yield) as a yellow oil:  $R_f 0.2$  (9:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.71 (d, J = 8.4 Hz, 1H), 8.52 (s, 1H), 8.50 (s, 1H), 8.27 (d, J = 1.8 Hz, 1H), 7.86 (d, J = 8.4 Hz, 2H), 7.56-7.38 (comp. m, 7H), 7.28 (d, J = 8.4 Hz, 2H), 7.03 (d, J = 8.1 Hz, 1H), 6.85 (d, J = 8.4 Hz, 1H), 5.84 (s, 2H), 5.25 (s, 2H), 4.25 (s, 3H), 4.07 (m, 1H), 3.62 (dd, J = 9.2, 4.4 Hz, 1H), 3.55 (t, J = 8.1, 2H), 3.35 (app.t, J = 9.2 Hz, 1H), 2.37 (s, 3H), 1.33 (d, J =7.0 Hz, 3H), 0.89 (t, J = 8.4 Hz, 2H), 0.69 (s, 9H), -0.04 (s, 9H), -0.16 (s, 3H), -0.28 (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 156.1, 145.7, 145.6, 145.4, 137.2, 135.8, 135.6, 135.5, 135.1, 131.3, 130.9, 130.3, 128.8, 128.7, 128.3, 127.8, 127.7, 127.3, 127.1, 127.0,

125.5, 119.0, 118.8, 117.3, 116.4, 115.6, 105.5, 78.0, 70.8, 69.2, 65.8, 54.2, 36.8, 25.9, 21.8, 18.2, 18.0, 17.7, -1.2, -5.5, -5.6; IR (film) 1374, 1178, 1087 cm<sup>-1</sup>; HRMS-ESI (m/z): [M + H]<sup>+</sup> calc'd for C<sub>50</sub>H<sub>62</sub>BrN<sub>4</sub>O<sub>6</sub>SSi<sub>2</sub>, 981.3112; found, 981.3097.



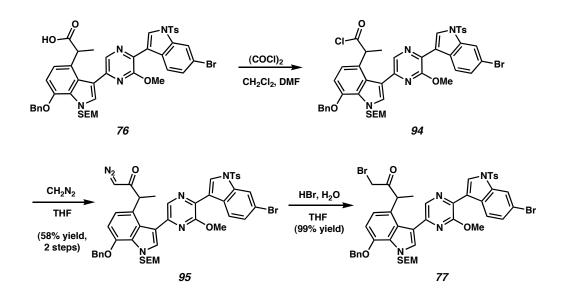
Aldehyde 75. To a Falcon tube containing a THF (5 mL) solution of bis(indole) 74 (70 mg, 0.07 mmol) at 0 °C was added HF•pyridine (800  $\mu$ L) in a dropwise fashion. The reaction mixture was stirred at 0 °C for 1.5 h until the reaction was judged complete by TLC. After dilution of the mixture with Et<sub>2</sub>O (10 mL), saturated aq. sodium bicarbonate (10 mL) was added in a dropwise manner at 0 °C. The layers were separated, and the organic portion was further washed with saturated aq. sodium bicarbonate (3 x 10 mL), dried over magnesium sulfate, and concentrated under reduced pressure.

The crude residue prepared above was dissolved in anhydrous  $CH_2Cl_2$  (5 mL), and Dess-Martin periodinane (91 mg, 0.214 mmol) was introduced. The reaction mixture was stirred at 23 °C for 20 min, poured into a saturated aq. solution of sodium bicarbonate/sodium thiosulfate (1:1, 5 mL), and extracted with  $CH_2Cl_2$  (3 x 10 mL). The organic layers were washed with brine (5 mL), dried over magnesium sulfate, and evaporated under reduced pressure to provide the crude product, which was purified by flash chromatography (2:1 hexanes:EtOAc) to furnish aldehyde **75** (53 mg, 86% yield) as a yellow oil:  $R_f$  0.67 (2:1 hexanes:EtOAc); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.73 (s, 1H), 8.69 (d, J = 8.3 Hz, 1H), 8.52 (s, 1H), 8.46 (s, 1H), 8.24 (d, J = 1.5 Hz, 1H), 7.86 (d, J = 8.3 Hz, 2H), 7.56-7.39 (comp. m, 7H), 7.30 (d, J = 7.8 Hz, 2H), 6.89 (d, J = 8.3 Hz, 1H), 6.83 (d, J = 8.3 Hz, 1H), 5.85 (s, 2H), 5.27 (s, 2H), 4.76 (q, J = 6.8 Hz, 1H), 4.18 (s, 3H), 3.55 (t, J = 8.0 Hz, 2H), 2.39 (s, 3H), 1.37 (d, J = 6.8 Hz, 3H), 0.89 (t, J = 8.0 Hz, 2H), -0.05 (s, 9H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) & 201.9, 156.1, 146.4, 145.7, 144.8, 136.8, 136.2, 135.8, 135.6, 135.0, 131.8, 130.4, 129.1, 128.9, 128.6, 128.5, 128.0, 127.9, 127.5, 127.4, 127.2, 125.4, 124.3, 121.0, 119.1, 116.9, 116.5, 115.2, 105.9, 78.2, 70.9, 66.0, 54.3, 48.5, 21.8, 18.0, 15.0, -1.2; IR (film) 1720, 1374, 1177, 1086 cm<sup>-1</sup>; HRMS-ESI (*m*/*z*): [M + H]<sup>+</sup> calc'd for C<sub>44</sub>H<sub>46</sub>BrN<sub>4</sub>O<sub>6</sub>SSi, 865.2090; found, 865.2103.



Acid 76. A solution of aldehyde 75 (53 mg, 0.061 mmol) in acetone (12 mL) was treated with a saturated solution of NaH<sub>2</sub>PO<sub>4</sub> that had been acidified to pH 2 with 1 N HCl (1.4 mL) and cooled to 0 °C. After the addition of 2-methyl-2-butene (32.5  $\mu$ L, 0.31 mmol), a solution of NaClO<sub>2</sub> (13.9 mg, 0.123 mmol) in H<sub>2</sub>O (1.4 mL) was added dropwise over 5 min. The reaction mixture was poured into cold H<sub>2</sub>O (2 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 x 5 mL). The combined organic layers were dried over magnesium sulfate and evaporated to dryness. The crude residue was passed through a short plug of silica gel (EtOAc eluent), and the solvent was evaporated to afford acid **76** (49 mg, 89% yield): R<sub>f</sub> 0.22 (2:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.68 (d, *J* = 8.3 Hz, 1H), 8.52 (s, 1H), 8.44 (s, 1H), 8.23 (d, *J* = 1.5 Hz, 1H), 7.85 (d, *J* = 8.3 Hz, 2H), 7.53-7.37 (comp. m, 7H), 7.28 (d, *J* = 9.2 Hz, 2H), 7.08 (d, *J* = 8.3 Hz, 1H), 6.84 (d,

 $J = 8.3 \text{ Hz}, 1\text{H}, 5.84 \text{ (d, } J = 10.3 \text{ Hz}, 1\text{H}, 5.78 \text{ (d, } J = 10.3 \text{ Hz}, 1\text{H}, 5.24 \text{ (s, } 2\text{H}), 4.86 \text{ (q, } J = 6.8 \text{ Hz}, 1\text{H}), 4.20 \text{ (s, } 3\text{H}), 3.53 \text{ (t, } J = 8.3 \text{ Hz}, 2\text{H}), 2.36 \text{ (s, } 3\text{H}), 1.42 \text{ (d, } J = 6.8 \text{ Hz}, 3\text{H}), 0.86 \text{ (t, } J = 8.3 \text{ Hz}, 2\text{H}), -0.07 \text{ (s, } 9\text{H}); {}^{13}\text{C} \text{ NMR} (125 \text{ MHz}, \text{CDCl}_3) \delta 178.2, 156.2, 146.2, 145.7, 145.0, 136.9, 136.2, 135.8, 135.7, 135.1, 131.7, 130.4, 129.1, 128.9, 128.6, 128.5, 128.4, 127.9, 127.5, 127.3, 127.2, 126.3, 125.4, 119.8, 119.1, 116.9, 116.5, 114.9, 105.8, 78.1, 70.9, 66.0, 54.5, 40.6, 21.8, 18.4, 18.0, -1.2; \text{ IR (film) 2948, 1703, 1373, 1177, 1139, 1088 cm<sup>-1</sup>; HRMS-ESI ($ *m*/*z*): [M + Na]<sup>+</sup> calc'd for C<sub>44</sub>H<sub>45</sub>BrN<sub>4</sub>O<sub>7</sub>SSiNa, 881.2040; found, 881.2009.



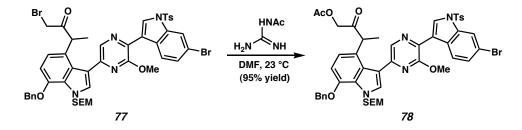
**Bromoketone 77.** To a solution of **76** (49 mg, 0.0559 mmol) in  $CH_2Cl_2$  (400 mL) at 0 °C was added oxalyl chloride (6.3 µL, 0.0727 mmol), followed by DMF (2 drops). After stirring at 0 °C for 30 min, all solvents were removed in vacuo. The crude acid chloride **94** was allowed to dry under vacuum for an additional 1 h and was used in the next step without further purification.

Note: In the next step, diazomethane was dried by storing the ethereal diazomethane solution over potassium hydroxide pellets. Immediately before use, the diazomethane was further dried over sodium metal for approximately 15 minutes.

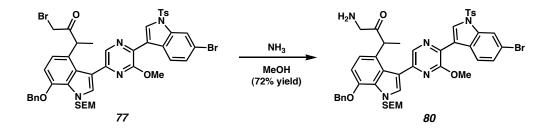
To crude **94** in THF (400  $\mu$ L) at 0 °C was added an ethereal solution of thoroughly dried diazomethane (1.5 mL) via a flamed glass pipette. The reaction mixture was allowed to warm to 23 °C, poured into saturated aq. sodium bicarbonate (1 mL), and extracted with Et<sub>2</sub>O (3 x 3 mL). The combined organic layers were dried over magnesium sulfate and evaporated under reduced pressure. The crude residue was purified by flash chromatography (4:1 hexanes:EtOAc eluent) to give diazoketone **95** (29.5 mg, 58% yield) as a yellow oil.

To **95** (19 mg, 0.021 mmol) in THF (2 mL) at 0 °C, 48% aq. HBr (3 drops) was added slowly down the walls of the flask. After stirring for 5 min, the reaction mixture was poured into a saturated aq. solution of sodium bicarbonate and extracted with Et<sub>2</sub>O (3 x 4 mL). The combined organic layers were dried over magnesium sulfate and evaporated under reduced pressure to afford bromoketone **77** (20 mg, 99% yield) as a yellow oil:  $R_f$  0.68 (2:1 hexanes:EtOAc); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.69 (d, *J* = 8.6 Hz, 1H), 8.53 (s, 1H), 8.47 (s, 1H), 8.24 (d, *J* = 1.5 Hz, 1H), 7.87 (d, *J* = 8.2 Hz, 2H), 7.55-7.38 (comp. m, 7H), 7.31 (d, *J* = 8.2 Hz, d), 6.85 (d, 2H), 5.83 (s, 2H), 5.25 (s, 2H), 5.01 (q, *J* = 6.7 Hz, 1H), 4.21 (s, 3H), 3.76 (d, *J* = 12.8 Hz, 1H), 3.67 (d, *J* = 13.1 Hz, 1H), 3.55 (t, *J* = 8.2 Hz, 2H), 2.39 (s, 3H), 1.48 (d, *J* = 6.7 Hz, 3H), 0.88 (t, *J* = 8.1 Hz, 2H), -0.052 (s, 9H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  156.3, 146.5, 145.8, 144.8, 136.7, 136.5, 135.8, 135.7, 135.1, 140.0, 130.4, 129.2, 128.9, 128.6, 128.5, 128.0, 127.7, 127.5, 125.5)

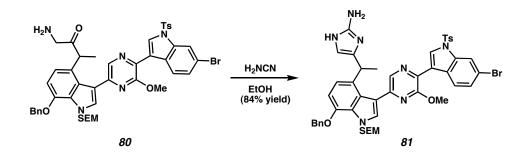
127.3, 127.2, 125.5, 125.4, 120.7, 119.1, 116.8, 116.5, 114.9, 106.0, 78.2, 71.0, 66.1, 54.3, 46.0, 34.2, 21.8, 18.0, 17.8, -1.2; IR (film) 2949, 1724, 1374, 1246, 1178, 1141, 1088 cm<sup>-1</sup>; HRMS-ESI (m/z): [M + H]<sup>+</sup> calc'd for C<sub>45</sub>H<sub>47</sub>Br<sub>2</sub>N<sub>4</sub>O<sub>6</sub>SSi, 957.1353; found, 957.1376.



Acetoxyketone 78. To a solution of 77 (10 mg, 0.0104 mmol) in DMF (350  $\mu$ L) was added acetylguanidine (32 mg, 0.316 mmol). After stirring at 23 °C for 48 h, H<sub>2</sub>O (1 mL) and EtOAc (1 mL) were added. The layers were separated, and the organic layer was washed with  $H_2O$  (3 x 500 µL) and brine (500 µL), then dried by passage through a plug of silica gel, and evaporated to afford acetoxyketone 78 (9.5 mg, 95% yield):  $R_f 0.53$ (2:1 hexanes:EtOAc); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.69 (d, J = 8.5 Hz, 1H), 8.54 (d, J = 0.9 Hz, 1H), 8.46 (d, J = 1.2 Hz, 1H), 8.24 (s, 1H), 7.87 (d, J = 7.6 Hz, 2H), 7.55-7.39 (comp. m, 7H), 7.31 (d, J = 8.2 Hz, 2H), 6.86 (s, 2H), 5.83 (s, 2H), 5.25 (s, 2H), 4.83 (q, J = 6.7 Hz, 1H), 4.47 (s, 2H), 4.21 (s, 3H), 3.54 (t, J = 8.2 Hz, 2H), 2.39 (s, 3H), 2.01 (s, 3H), 1.42 (d, J = 7.0 Hz, 3H), 0.88 (t, J = 8.2 Hz, 2H), -0.05 (s, 9H); <sup>13</sup>C NMR (125) MHz, CDCl<sub>3</sub>) δ 204.5, 170.3, 156.3, 146.4, 145.8, 144.9, 136.8, 136.5, 135.9, 135.6, 135.1, 131.9, 130.4, 129.2, 128.9, 128.6, 128.5, 128.0, 127.6, 127.5, 127.3, 127.2, 125.7, 125.4, 120.6, 119.1, 116.9, 116.5, 114.9, 106.0, 78.2, 71.0, 67.2, 66.1, 54.3, 45.6, 21.9, 20.6, 18.1, 17.3, -1.2; IR (film) 2949, 1750, 1728, 1373, 1244, 1178, 1141, 1088 cm<sup>-1</sup>; HRMS-ESI (m/z):  $[M + H]^+$  calc'd for  $C_{47}H_{50}BrN_4O_8SSi$ , 937.2302; found, 937.2290.



Aminoketone 80. Bromoketone 77 (6 mg, 0.0062 mmol) was dissolved in a saturated solution of ammonia in CH<sub>3</sub>OH (1 mL). After stirring for 6 h at 23 °C, the reaction mixture was filtered through a plug of silica gel (CH<sub>3</sub>OH eluent), and the solvent was evaporated. The crude residue was then purified by preparative thin layer chromatography (7:1 CH<sub>2</sub>Cl<sub>2</sub>:CH<sub>3</sub>OH) to afford aminoketone 80 (4 mg, 72% yield):  $R_f$  0.67 (7:1 CH<sub>2</sub>Cl<sub>2</sub>:CH<sub>3</sub>OH); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.19 (d, 8.8, 1H), 8.53 (s, 1H), 8.46 (s, 1H), 8.23 (s, 1H), 7.85 (d, J = 8.4 Hz, 2H), 7.54-7.37 (comp. m, 7H), 7.29 (d, J = 8.4 Hz, 2H), 6.84-6.83 (comp. m, 2H), 5.82 (s, 2H), 5.24 (s, 2H), 4.71 (q, J = 6.6 Hz, 1H), 4.19 (s, 3H), 3.54 (t, J = 8.1 Hz, 2H), 3.31 (d, J = 19.3 Hz, 1H), 3.07 (d, J = 19.1 Hz, 1H), 2.38 (s, 3H), 1.44 (d, J = 7.0 Hz, 3H), 0.87 (t, J = 8.1 Hz, 2H), -0.06 (s, 9H).



**Aminoimidazole 81.** To a solution of aminoketone **80** (7 mg, 0.0078 mmol) in ethanol (700  $\mu$ L) was added cyanamide (15 mg, 0.36 mmol). The reaction vessel was sealed and heated to 70 °C for 10 h. After cooling to 23 °C, the reaction mixture was

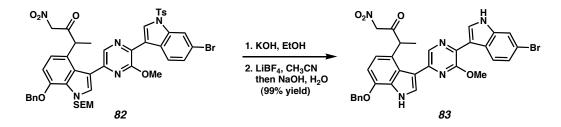
purified by reversed-phase filtration through a Sep-Pak column: first 10% acetonitrile, then 100% acetonitrile to collect the product. After removal of solvent under reduced pressure, **81** (6 mg, 84% yield) was isolated as an orange/red oil:  $R_f$  0.27 (7:1 CH<sub>2</sub>Cl<sub>2</sub>:CH<sub>3</sub>OH); <sup>1</sup>H NMR (500 MHz, CD<sub>3</sub>OD)  $\delta$  8.68 (d, *J* = 8.8 Hz, 1H), 8.52 (s, 1H), 8.38 (s, 1H), 8.16 (d, *J* = 1.5 Hz, 1H), 7.87 (d, *J* = 8.1 Hz, 2H), 7.60-7.33 (comp. m, 9H), 6.95 (d, *J* = 8.4 Hz, 1H), 6.90 (d, *J* = 8.4 Hz, 1H), 6.10 (s, 1H), 5.85 (s, 2H), 5.28 (s, 2H), 5.09 (q, *J* = 6.8 Hz, 1H), 4.19 (s, 3H), 3.57 (t, *J* = 7.9 Hz, 2H), 2.37 (s, 3H), 1.41 (d, *J* = 7.0 Hz, 3H), 0.82 (t, *J* = 7.9 Hz, 2H), -0.10 (s, 9H); HRMS-ESI (*m/z*): [M + H]<sup>+</sup> calc'd for C<sub>46</sub>H<sub>49</sub>BrN<sub>7</sub>O<sub>5</sub>SSi, 918.2468; found, 918.2467.



Ketone 82. To aldehyde 75 (20 mg, 0.023 mmol) in nitromethane (1 mL) was added triethylamine (75  $\mu$ L, 0.54 mmol). The reaction mixture was stirred at 23 °C for 15 h. The excess nitromethane was removed by evaporation under reduced pressure to afford the crude nitroaldol product, which was used without further purification.

The crude residue was dissolved in anhydrous  $CH_2Cl_2$  (1.5 mL) and treated with Dess-Martin periodinane (15% solution in  $CH_2Cl_2$ , 200 µL, 0.099 mmol). The reaction mixture was stirred at 23 °C for 5 min and quenched by addition of saturated aq. sodium bicarbonate/sodium thiosulfate (1:1, 2 mL). The layers were separated, and the aqueous layer was extracted with EtOAc (8 x 1 mL). The combined organic layers were washed with brine (2 mL), dried by passage through a plug of silica gel (EtOAc eluent), and

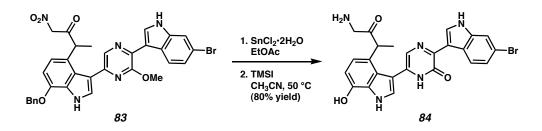
evaporated under reduced pressure to afford ketone **82** (21 mg, 98% yield) as a yellow oil:  $R_f 0.20$  (3:1 hexanes:EtOAc); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.62 (d, J = 8.3 Hz, 1H), 8.44 (s, 1H), 8.39 (s, 1H), 8.18 (s, 1H), 7.80 (d, J = 8.3 Hz, 2H), 7.49-7.33 (comp. m, 7H), 7.24 (d, J = 8.3 Hz, 2H), 6.83 (d, J = 8.3 Hz, 1H), 6.81 (d, J = 7.8 Hz, 1H), 5.77 (s, 2H), 5.21 (s, 2H), 5.03 (d, J = 14.6 Hz, 1H), 4.98 (d, J = 14.6 Hz, 1H), 4.90 (q, J = 6.8Hz, 1H), 4.14 (s, 3H), 3.49 (t, J = 7.8 Hz, 2H), 2.32 (s, 3H), 1.44 (d, J = 6.8 Hz, 3H), 0.82 (t, J = 8.0 Hz, 2H), -0.12 (s, 9H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  197.0, 156.3, 146.9, 145.8, 144.3, 136.9, 136.5, 135.8, 135.6, 135.0, 132.3, 130.4, 129.3, 129.0, 128.6, 128.5, 128.0, 127.6, 127.5, 127.4, 127.2, 125.4, 123.7, 121.0, 119.2, 116.7, 116.5, 114.6, 106.2, 82.2, 78.3, 71.0, 66.2, 54.4, 47.6, 21.8, 18.0, 16.9, -1.2; IR (film) 1732, 1559, 1376, 1178, 1080 cm<sup>-1</sup>. HRMS-FAB (m/z): [M + H]<sup>+</sup> calc'd for C<sub>45</sub>H<sub>47</sub>BrN<sub>5</sub>O<sub>8</sub>SiSBr, 926.2077; found, 926.2080.



**Nitroketone 83.** To a suspension of ketone **82** (30 mg, 0.032 mmol) in EtOH (2 mL, deoxygenated by sparging with argon for 2 min) was added powdered KOH (100 mg, 1.8 mmol). The reaction vessel was equipped with a reflux condenser and heated to 40 °C for 2 h. After cooling to 23 °C, the reaction mixture was quenched with saturated aq. ammonium chloride (2 mL) and extracted with EtOAc (8 x 1 mL). The combined organic layers were washed with brine (2 mL), dried by passage through a plug of silica

gel (EtOAc eluent), and evaporated under reduced pressure to afford the crude detosylated ketone, which was used without further purification.

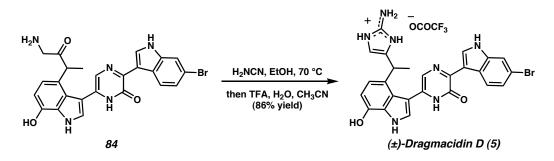
To the crude detosylated ketone prepared above in acetonitrile (3 mL, deoxygenated by sparging with argon for 2 min) and water (30  $\mu$ L) was added lithium tetrafluoroborate (120 mg, 0.13 mmol). The reaction vessel was equipped with a reflux condenser and heated to 70 °C for 1.5 h. After cooling to 40 °C, sodium hydroxide (20% aq., 2 mL) was added. The resulting mixture was stirred for 10 min, allowed to cool to 23 °C, quenched with saturated aq. ammonium chloride (2 mL), and extracted with EtOAc (8 x 1 mL). The combined organic layers were washed with brine (2 mL), dried by passage through a plug of silica gel (EtOAc eluent), and evaporated under reduced pressure to afford nitroketone 83 (20.5 mg, 99% yield) as a yellow oil:  $R_f$  0.59 (1:1) hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, acetone- $d_6$ )  $\delta$  11.14 (br s, 1H), 10.86 (br s, 1H), 8.82 (d, J = 8.8 Hz, 1H), 8.54 (s, 1H), 8.43 (m, 1H), 7.75-7.72 (comp. m, 2H), 7.62-7.59 (comp. m, 2H), 7.47-7.30 (comp. m, 4H), 6.95 (d, J = 8.1 Hz, 1H), 6.85 (d, J = 8.1 Hz, 1H), 5.41-5.22 (comp. m, 5H), 4.19 (s, 3H), 1.47 (d, J = 7.0 Hz, 3H); <sup>13</sup>C NMR (125) MHz, acetone- $d_6$ )  $\delta$  198.7, 156.2, 146.3, 143.6, 139.4, 138.6, 138.3, 136.1, 130.7, 129.4, 129.4, 128.9, 128.8, 128.3, 126.9, 126.8, 125.8, 124.9, 124.2, 120.5, 116.3, 116.0, 115.3, 112.5, 105.1, 83.4, 70.9, 54.1, 47.7, 17.3; IR (film) 3410 (br), 1728, 1697, 1557, 1450 cm<sup>-1</sup>; HRMS-ESI (m/z): [M + H]<sup>+</sup> calc'd for C<sub>32</sub>H<sub>27</sub>BrN<sub>5</sub>O<sub>5</sub>, 640.1196; found, 640.1180.



Aminoketone 84. To a solution of deprotected ketone 83 (5.5 mg, 0.0086 mmol) in EtOAc (600  $\mu$ L, deoxygenated by bubbling with argon for 1 min), was added SnCl<sub>2</sub>•2 H<sub>2</sub>O (30 mg, 0.13 mmol). The reaction vessel was equipped with a reflux condenser and heated at 80 °C for 3 h. After cooling to 23 °C, the solvent was removed under reduced pressure to leave an orange residue, which was purified by reversed-phase filtration through a Sep-Pak column: first 10% acetonitrile containing 0.1% (*w*/*v*) TFA to remove salts, then 90% acetonitrile containing 0.1% (*w*/*v*) TFA to collect the crude product. After removal of solvent in vacuo, the compound was filtered through silica gel (5:1 CH<sub>2</sub>Cl<sub>2</sub>/CH<sub>3</sub>OH eluent) to provide the reduced compound, which was used without further purification.

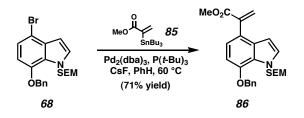
To the crude aminoketone in acetonitrile (700 µL) at 0 °C, in a Schlenk tube, was added iodotrimethylsilane (150 µL, 1.05 mmol). The reaction mixture was heated at 50 °C for 2 h, cooled to 0 °C, then quenched with saturated aq. sodium metabisulfite. The reaction mixture was purified by reversed-phase filtration through a Sep-Pak column: first 10% acetonitrile containing 0.1% (*w*/*v*) TFA to remove salts, then 90% acetonitrile containing 0.1% (*w*/*v*) TFA to collect the crude product. After removal of solvent under reduced pressure, the compound was further purified by reversed-phase HPLC. Concentration under reduced pressure provided the fully deprotected aminoketone **84** (3.5 mg, 80% yield) as an orange/red oil: <sup>1</sup>H NMR (500 MHz, CD<sub>3</sub>OD)  $\delta$  8.75 (s, 1H), 8.60 (d, *J* = 8.4 Hz, 1H), 7.67 (s, 1H), 7.62 (d, *J* = 1.7 Hz, 1H), 7.54 (s, 1H), 7.27 (dd, *J* 

= 8.5, 1.8 Hz, 1H), 6.69 (d, J = 8.1 Hz, 1H), 6.63 (d, J = 7.7 Hz, 1H), 4.15 (q, J = 6.9 Hz, 1H), 3.64 (d, J = 1.7 Hz, 2H), 1.44 (d, J = 6.8 Hz, 3H); <sup>13</sup>C NMR (125 MHz, CD<sub>3</sub>OD)  $\delta$  204.4, 157.4, 151.0, 145.3, 139.1, 132.6, 131.7, 129.0, 128.3, 127.7, 126.7, 126.1, 125.6, 124.9, 122.6, 121.0, 117.1, 115.5, 113.7, 108.5, 107.9, 46.8, 17.2; IR (film) 3140 (br), 1671, 1200, 1140 cm<sup>-1</sup>; HRMS-ESI (*m*/*z*) calc'd for C<sub>24</sub>H<sub>21</sub>BrN<sub>5</sub>O<sub>3</sub>, 506.0828; found, 506.0827.



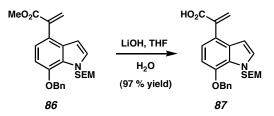
**Dragmacidin D (5).** To a solution of aminoketone **84** (2 mg, 0.0039 mmol) in ethanol (700  $\mu$ L, deoxygenated by bubbling with argon for 5 min) was added cyanamide (15 mg, 0.36 mmol). The reaction vessel was purged with argon, sealed, and heated to 70 °C for 3 h. After cooling to 23 °C, the reaction mixture was purified by reversed-phase filtration through a Sep-Pak column: first 10% acetonitrile containing 0.1% (*w*/*v*) TFA to remove salts, then 60% acetonitrile containing 0.1% (*w*/*v*) TFA to collect the product. After removal of solvent under reduced pressure, dragmacidin D (**5**, 1.8 mg, 86% yield) was isolated as an orange/red oil: <sup>1</sup>H NMR (600 MHz, CD<sub>3</sub>OD)  $\delta$  8.74 (s, 1H), 8.6 (d, *J* = 8.7 Hz, 1H), 7.62 (d, *J* = 1.8 Hz, 1H), 7.49 (s, 1H), 7.46 (s, 1H), 7.27 (dd, *J* = 8.2, 1.8 Hz, 1H), 6.84 (d, *J* = 8.2 Hz, 1H), 6.64 (d, *J* = 7.8 Hz, 1H), 5.98 (s, 1H), 4.35 (q, *J* = 6.9 Hz, 1H), 1.52 (d, *J* = 7.3 Hz, 3H); <sup>13</sup>C NMR (1645 MHz, CD<sub>3</sub>OD)  $\delta$  157.1, 150.3, 148.7, 144.8, 139.1, 134.2, 132.4, 132.2, 128.7, 127.9, 127.3, 126.7, 126.3, 125.6, 124.8, 120.2,

117.1, 115.4, 113.7, 110.2, 108.9, 107.3, 33.2, 20.8; IR (film) 3200 (br), 1667, 1204, 1138 cm<sup>-1</sup>; UV  $\lambda_{max}$  (EtOH) 216, 274, 389 nm. After addition of 1 drop concentrated HCl to 1 mL cell:  $\lambda_{max}$  (EtOH) 219, 277, 460 nm; HRMS-ESI (*m/z*): [M + H]<sup>+</sup> calc'd for C<sub>25</sub>H<sub>21</sub>BrN<sub>7</sub>O<sub>2</sub>, 530.0940; found, 530.0943.

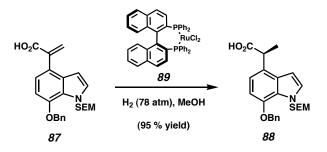


**Ester 86.** To **68** (50 mg, 0.116 mmol) in a flame-dried Schlenk flask was added tris(dibenzylideneacetone)dipalladium(0) (1.5 mg, 0.0017 mmol), stannane **85** (52 μL, 0.15 mmol), dry cesium fluoride (46 mg, 0.302 mmol), and benzene (1.3 mL), followed by tri-*t*-butylphosphine (0.78 mg, 0.0038 mmol) under an N<sub>2</sub> atmosphere in a dry box. The sealed reaction vessel was then heated at 60 °C for 14 h. The reaction mixture was cooled to 23 °C and filtered through a plug of silica gel (EtOAc eluent). After removal of solvent under reduced pressure, the crude product was purified by flash chromatography (4:1 hexanes:EtOAc) to afford **86** (42.6 mg, 71% yield) as a yellow oil: R<sub>*f*</sub> 0.64 (4:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 7.54-7.35 (comp. m, 5H), 7.16 (d, *J* = 3.3 Hz, 1H), 7.00 (d, *J* = 8.1 Hz, 1H), 6.76 (d, *J* = 8.1 Hz, 1H), 6.48 (d, *J* = 1.8 Hz, 1H), 6.41 (d, *J* = 2.9 Hz, 1H), 5.93 (d, *J* = 1.8 Hz, 1H), 5.75 (s, 2H), 5.23 (s, 2H), 3.80 (s, 3H), 3.46 (t, *J* = 8.1 Hz, 2H), 0.83 (t, *J* = 8.2 Hz, 2H), -0.08 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 168.2, 146.9, 140.3, 137.0, 130.2, 129.6, 128.8, 128.2, 127.8, 127.3, 125.6, 122.9, 121.3, 104.2, 102.4, 77.6, 70.6, 52.4, 18.0, -1.1; IR (film) 2951, 1724, 1502, 1290,

1258, 1175, 1073 cm<sup>-1</sup>; HRMS-ESI (m/z): [M + H]<sup>+</sup> calc'd for C<sub>25</sub>H<sub>32</sub>NO<sub>4</sub>Si, 460.1920; found, 460.1913.



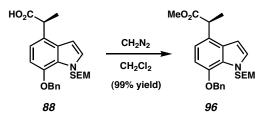
Acid 87. To a solution of 86 (970 mg, 1.88 mmol) in THF (10 mL) and  $H_2O$  (2 mL) was added lithium hydroxide monohydrate (394 mg, 9.40 mmol). The resulting solution was heated under reflux to 80 °C for 24 h, then allowed to cool to 23 °C. The reaction mixture was diluted with  $H_2O$  (5 mL) and extracted with  $Et_2O$  (2 x 10 mL). The organic layer was discarded, and the aqueous layer was acidified to pH=2 by the dropwise addition of 6 N HCl at 0 °C. After extracting the aqueous layer with  $CH_2Cl_2$  (3 x 20 mL), the combined organic layers were washed with  $H_2O$  (10 mL) and brine (10 mL), dried over magnesium sulfate, and evaporated under reduced pressure to yield 87 (919 mg, 97% yield) as a yellow oil, which was used in the subsequent step without further purification.



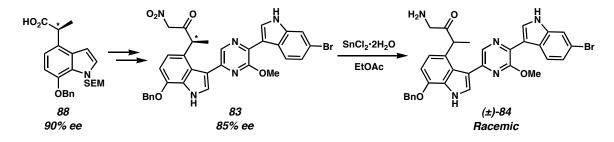
To **87** (578 mg, 1.15 mmol) in a thick-walled glass container under  $N_2$  in a dry box was added dichloro[(S)-(-)-2,2]-bis(diphenylphosphino)-1,1]-binaphthyl]ruthenium

(II) (89, 46 mg, 0.0576 mmol) and  $CH_3OH$  (11.5 mL). The reaction vessel was sealed under nitrogen, removed from the dry box, and placed in a stainless steel bomb, which was purged with argon. After removing the seal of the reaction vessel, the bomb was sealed under an argon atmosphere, cooled to -10 °C over 1 h, then purged and pressurized with  $H_2$  to 1200 psi with vigorous stirring. The  $H_2$  pressure was carefully released after 72 h of stirring, and the reaction vessel was warmed to 23 °C. After filtering the reaction mixture through a plug of celite (CH<sub>3</sub>OH eluent), the solvent was evaporated under reduced pressure to yield 88 (550 mg, 95% yield, 90% ee\*) as a yellow oil:  $R_t 0.63$  (1:1 hexanes: EtOAc); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.17 (d, J = 3.3 Hz, 1H), 6.98 (d, J = 8.1 Hz, 1H), 6.73 (d, J = 8.1 Hz, 1H), 6.61 (d, J = 3.3 Hz, 1H), 5.76 (d, J = 3.3 Hz, 1H), 5.7610.6 Hz, 1H), 5.72 (d, J = 10.3 Hz, 1H), 5.19 (s, 2H), 4.08 (q, J = 7.1 Hz, 1H), 3.45 (t, J= 8.2 Hz, 2H), 1.60 (d, J = 7.0 Hz, 3H), 0.82 (t, J = 8.2 Hz, 2H), -0.09 (s, 9H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 180.6, 146.3, 137.2, 130.3, 129.4, 128.8, 128.2, 127.8, 125.9, 124.9, 118.7, 104.6, 101.6, 77.6, 70.6, 65.5, 42.4, 18.0, 17.5, -1.2; IR (film) 2952, 1707, 1503, 1251, 1072 cm<sup>-1</sup>; HRMS-ESI (m/z): [M + Na]<sup>+</sup> calc'd for C<sub>24</sub>H<sub>31</sub>NO<sub>4</sub>SiNa, 448.190; found, 448.1918.

\*The enantiopurity of 88 was determined by derivatization to methyl ester 97 via treatment with diazomethane as described below, followed by chiral HPLC analysis:



Ester 96. To a sample of 88 (2 mg, 0.004 mmol) in  $CH_2Cl_2$  (1 mL) was added ethereal diazomethane (0.2 M, ca. 1 mL) until a bright green/yellow color persisted. The solvent was removed under reduced pressure to afford ester 96 (2 mg, 99% yield) as a yellow oil. Chiral HPLC was performed on a Chiralcel AD column using 2% *i*-PrOH in hexanes as eluent.

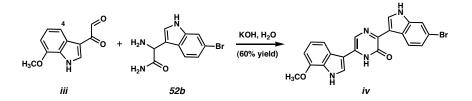


The enantiopurity of **83** was determined by chiral HPLC performed on a Chiralcel AD column using 20% EtOH in hexanes as eluent. The enantiopurity of **84** was determined by chiral HPLC performed on a Chiralcel AD column using 60% EtOH in hexanes as eluent.

## 2.8 Notes and References

- (a) Wright, A. E.; Pomponi, S. A.; Cross, S. S.; McCarthy, P. J. Org. Chem. 1992, 57, 4772-4775.
   (b) Capon, R. J.; Rooney, F.; Murray, L. M.; Collins, E.; Sim, A. T. R.; Rostas, J. A. P.; Butler, M. S.; Carroll, A. R. J. Nat. Prod. 1998, 61, 660-662.
   (c) Cutignano, A.; Bifulco, G.; Bruno, I.; Casapullo, A.; Gomez-Paloma, L.; Riccio, R. Tetrahedron 2000, 56, 3743-3748.
   (d) Wright, A. E.; Pomponi, S. A.; Jacobs, R. S. PCT Int. Appl. WO 9942092 August 26, 1999.
- (2) Portions of this work have been published, see: Garg, N. K.; Sarpong, R.; Stoltz, B.
  M. J. Am. Chem. Soc. 2002, 124, 13179-13184.
- (3) Sundberg, R. J., Ed.; *Indoles*; Academic Press: San Diego, 1996.
- (4) For a reference on biomimetic cyclocondensations to produce pyrazinones, see: Yaylayan, V. A. J. Agric. Food. Chem. 1996, 44, 2511-2516.
- (5) For comprehensive reviews on transition metal-catalyzed cross-coupling reactions, see: (a) Diederich, F.; Stang, P. J.; Eds.; *Metal-Catalyzed Cross-Coupling Reactions*; Wiley-VCH: Weinheim, 1998. (b) Geissler, H. In *Transition Metals for Organic Synthesis*; Beller, M.; Bolm, C., Eds.; Wiley-VCH: Weinheim, 1998; Chapter 2.10, p 158. (c) Tsuji, J. In *Transition Metal Reagents and Catalysts*; Wiley: Chichester, U.K., 2000; Chapter 3, p 27.

- (6) (a) Kharasch, M. S.; Kane, S. S.; Brown, H. C. J. Am. Chem. Soc. 1940, 62, 2242-2243.
  (b) Hashem, M. A.; Sultana, I.; Hai, M. A. Indian J. Chem. Sect. B 1999, 38, 789-794.
- (7) (a) Vereshchagin, A. L.; Branskii, O. V.; Semenov, A. A. Chem. Heterocycl. Compd. (Engl. Transl.) 1983, 19, 40-42. (b) Kuivila, H. G. J. Org. Chem. 1960, 25, 284-285.
- (8) Bromide 52b was prepared in a manner similar to that used for the synthesis of 52a. See Section 2.7 for details.
- (9) The steric component of C(4) substitution appears to be the determining factor as illustrated by the reaction of iii and 52b to produce pyrazinone iv in good yield.



 (10) For an excellent discussion of the cross-coupling chemistry of pyrazines, see: Li, J.
 J.; Gribble, G. W. In *Palladium in Heterocyclic Chemistry: A Guide for the Synthetic Chemist*; Pergamon: Amsterdam, 2000; pp 355-373.

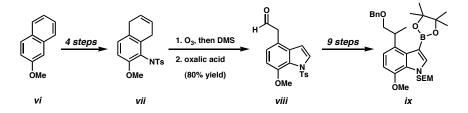
- (11) Turck, A.; Ple, N.; Dognon, D.; Harmoy, C.; Queguiner, G. J. Heterocycl. Chem. **1994**, *31*, 1449-1454.
- (12) For a discussion regarding the reactivity of aryl chlorides toward transition-metal complexes, see: Gushin, V. V.; Alper, H. *Chem. Rev.* **1994**, *94*, 1047-1062.
- (13) (a) Edwards, M. P.; Ley, S. V.; Lister, S. G.; Palmer, B. D. J. Chem. Soc., Chem. Commun. 1983, 630-633. (b) Piers, E.; Britton, R.; Andersen, R. J. J. Org. Chem. 2000, 65, 530-535.
- (14) Pyrazinyl chlorides coupled competitively with the indolylbromide functionality.
- (15) Barlin, G. B. Aust. J. Chem. 1983, 36, 983-992.
- (16) Zhu, Z.; Moore, J. S. J. Org. Chem. 2000, 65, 116-123.
- (17) Bromoindole 49 was prepared from 50 by a modified Leimgruber-Batcho synthesis, see: Schumacher, R. W.; Davidson, B. S. *Tetrahedron* 1999, 55, 935-942.
- (18) Wenkert, E.; Moeller, P. D.; Piettre, S. R.; McPhail, A. T. J. Org. Chem. 1988, 53, 3170-3178.

- (19) Zheng, Q.; Yang, Y.; Martin, A. R. Heterocycles 1994, 37, 1761-1772.
- (20) (a) Bartoli, G.; Palmieri, G.; Bosco, M.; Dalpozzo, R. *Tetrahedron Lett.* 1989, *30*, 2129-2132. (b) Bosco, M.; Dalpozzo, R.; Bartoli, G.; Palmieri, G.; Petrini, M. *J. Chem. Soc., Perkin Trans.* 2 1991, *5*, 657-663.
- (21) Benzyl ether 66 is a known compound prepared in a single-step involving benzyl protection of commercially available 4-bromo-2-nitrophenol. See: Auwers, K. *Justus Liebigs Ann. Chem.* 1907, 357, 85-94.
- (22) Derivatives of 67 were initially accessed via the Leimgruber-Batcho indole synthesis. These routes took several steps and were low-yielding. For the Leimgruber-Batcho indole synthesis, see: Batcho, A. D.; Leimgruber, W. Org. Synth. 1985, 63, 214-225. See also references therein.
- (23) Belletete, M.; Beaupre, S.; Bouchard, J.; Blondin, P.; Leclerc, M.; Durocher, G. J.
   *Phys. Chem. B* 2000, *104*, 9118-9125.
- (24) Li, K.; Du, W.; Que, N. L.; Liu, H. J. Am. Chem. Soc. 1996, 118, 8763-8764.

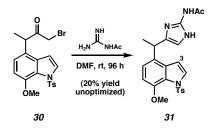
- (25) For general reviews of Suzuki couplings, see: (a) Miyaura, N.; Suzuki, A. Chem. Rev. 1995, 95, 2457-2483. (b) Suzuki, A. J. Organomet. Chem. 1999, 576, 147-168.
- (26) (a) Maki, S.; Okawa, M.; Matsui, R.; Hirano, T.; Niwa, H. Synlett 2001, 10, 1590-1592. (b) In the presence of minor impurities and/or solutions of benzene that were not pre-saturated with H<sub>2</sub>, the reaction produced a major byproduct identified as enol ether v.



- (27) (a) Ayer, W. A.; Craw, P. A.; Ma, Y. T.; Miao, S. *Tetrahedron* 1992, 48, 2919-2924. (b) Amat, M.; Hadida, S.; Sathyanarayana, S.; Bosch, J. J. Org. Chem. 1994, 59, 10-11.
- (28) A strategy involving Plieninger indolization was originally employed to prepare fragments related to 62. Unfortunately, this strategy required 15 steps to access 3,4,7-trisubstituted indoles as summarized below (vi → vii → viii → ix). For the Plieninger indole synthesis, see: (a) Plieninger, H.; Suhr, K. Chem. Ber. 1956, 89, 270-278. (b) Maehr, H.; Smallheer, J. J. Am. Chem. Soc. 1985, 107, 2943-2945.



- (29) For the cleavage of silyl ethers with HF•pyridine, see: Nicolaou, K. C.; Webber,S. E. Synthesis 1986, 6, 453-461.
- (30) Dess, D. B.; Martin, J. C. J. Org. Chem. 1983, 48, 4155-4156.
- (31) Arndt, F. Org. Synth., 1943, Coll. Vol. 2, 165.
- (32) Little, T. L.; Webber, S. E. J. Org. Chem. 1994, 59, 7299-7305.
- (33) In an early approach, we prepared aminoimidazole **31** as shown below. Attempts to selectively functionalize C(3) of **31** were unsuccessful. Thus, **31** was deemed a non-productive route and was used only for model studies. Soon afterward, a similar approach to synthesize **31** appeared in the literature. See: Yang, C.-G.; Wang, J.; Jiang, B. *Tetrahedron Lett.* **2002**, *43*, 1063-1066.



- (34) (a) Lancini, G. C.; Lazzari, E.; Sartori, G. J. Antibiot. 1968, 21, 387-392. (b)
  Howes, P. D.; Cleasby, A.; Evans, D. N.; Feilden, H.; Smith, P. W.; Sollis, S. L.;
  Taylor, N.; Wonacott, A. J. Eur. J. Med. Chem. 1999, 34, 225-234.
- (35) (a) Luzzio, F. A. *Tetrahedron* 2001, *57*, 915-945. (b) Ayerbe, M.; Arrieta, A.;
  Cossio, F. J. Org. Chem. 1998, 63, 1795-1805.
- (36) Aldehyde **75** was the last compound in the synthesis that was readily purified on preparative scale by silica gel flash chromatography. Although analytical scale purification beyond this point was performed by reversed-phase HPLC or thin layer chromatography on SiO<sub>2</sub>, all preparative scale purification was conducted by reversed-phase C<sub>18</sub> chromatography. Difficulties associated with the separation of similarly polar compounds by this method necessitated that all reactions in the sequence leading from **75**  $\rightarrow$  **5** be high yielding.
- (37) Gilbert, E. J.; Chisholm, J. D.; Van Vranken, D. L. J. Org. Chem. 1999, 64, 5670-5676.
- (38) Sakowski, J.; Bohn, M.; Sattler, I.; Dahse, H.; Schlitzer, M. J. Med. Chem. 2001, 44, 2886-2899.

- (39) Lott, R. S.; Chauhan, V. S.; Virander, S.; Stammer, C. H. J. Chem. Soc., Chem.
   *Commun.* 1979, 495-496.
- (40) The compounds depicted in Figure 2.4.2, from left to right, are aldehyde 75 and dragmacidin D (5).
- (41) For general reviews of asymmetric hydrogenation, see: (a) Takaya, H.; Ohta, T.; Noyori, R. In *Catalytic Asymmetric Synthesis*; Ojima, I., Ed.; VCH Publishers: New York, 1994; pp 1-39. (b) Noyori, R. In *Asymmetric Catalysis in Organic Synthesis*; Wiley-Interscience: New York, 1994; pp 16-94. (c) Brown, J. M. In *Comprehensive Asymmetric Catalysis*; Jacobsen, E. N.; Pfaltz, A.; Yamamoto, H., Eds.; Springer: Berlin, 1999; Vol. 1, pp 121-195.
- (42) Littke, A. F.; Schwarz, L.; Fu, G. C. J. Am. Chem. Soc. 2002, 124, 6343-6348.
- (43) Miyake, H.; Yamamura, K. Chem. Lett. 1989, 6, 981-984.
- (44) Miyake, F. Y.; Yakushijin, K.; Horne, D. A. Org. Lett. 2002, 4, 941-943.

**APPENDIX ONE** 

Synthetic Summary for Dragmacidin D (5)

## Figure A1.1 The synthesis of indolylpyrazine 73.

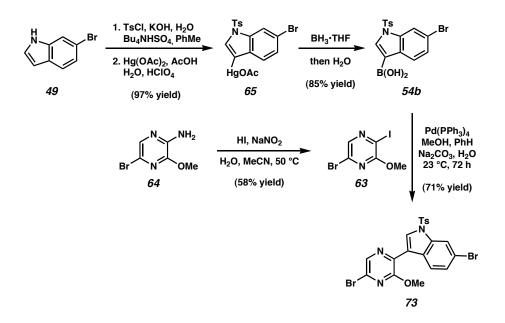
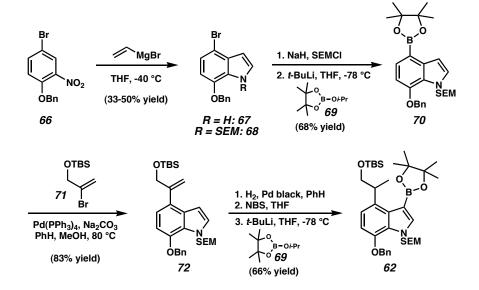
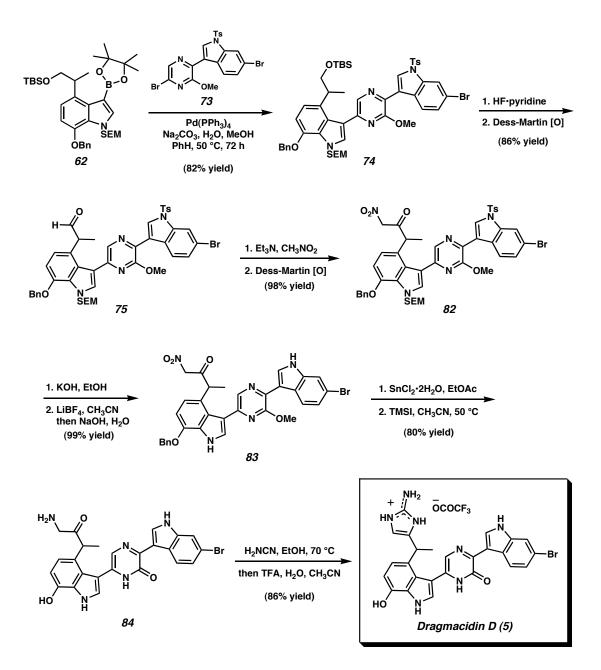


Figure A1.2 The synthesis of boronic ester 62.

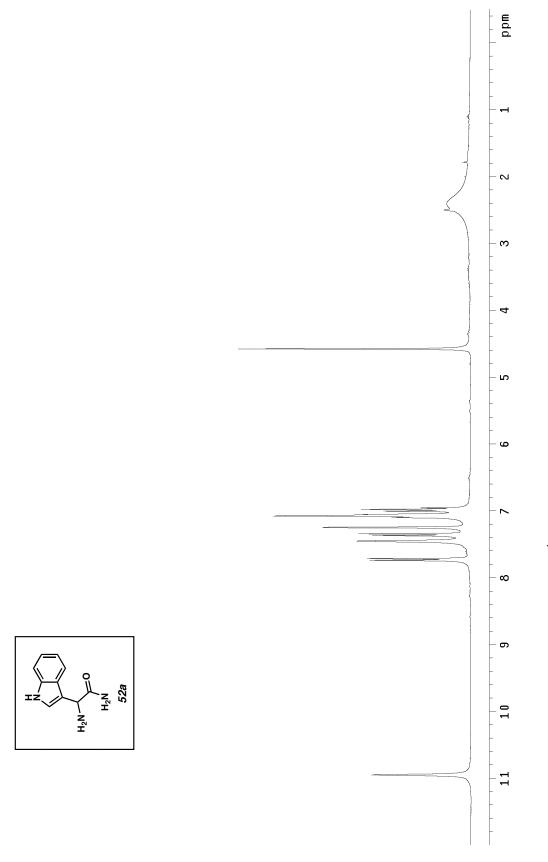




## APPENDIX TWO

Spectra Relevant to Chapter Two:

The Total Synthesis of Dragmacidin D





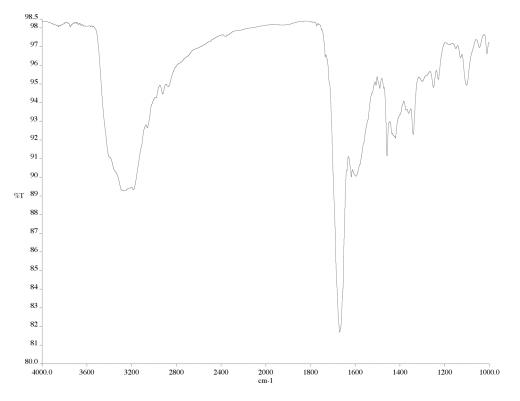
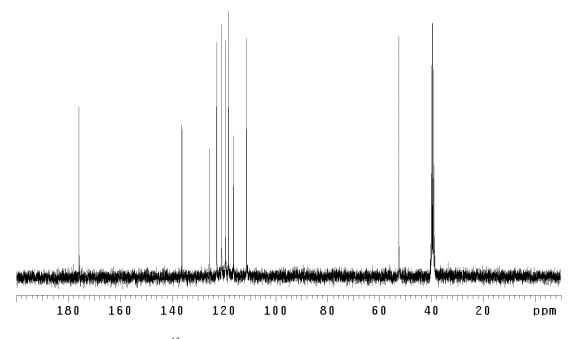
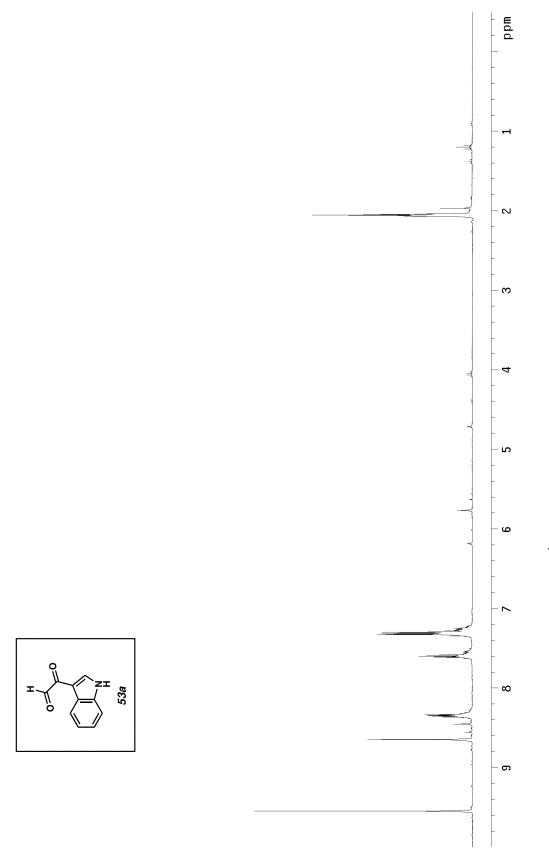


Figure A2.2 Infrared spectrum (thin film/NaCl) of compound 52a.



*Figure A2.3* <sup>13</sup>C NMR (75 MHz, DMSO- $d_6$ ) of compound **52a**.





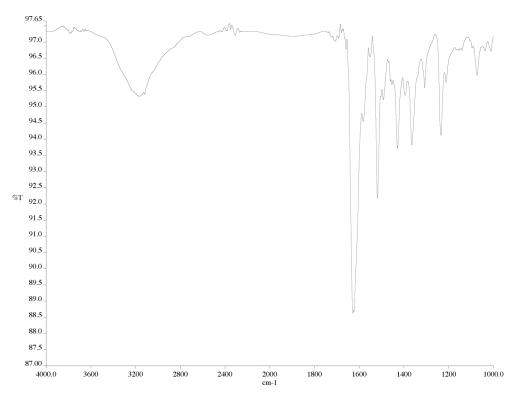
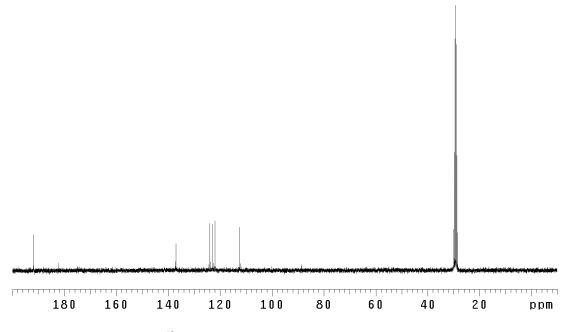
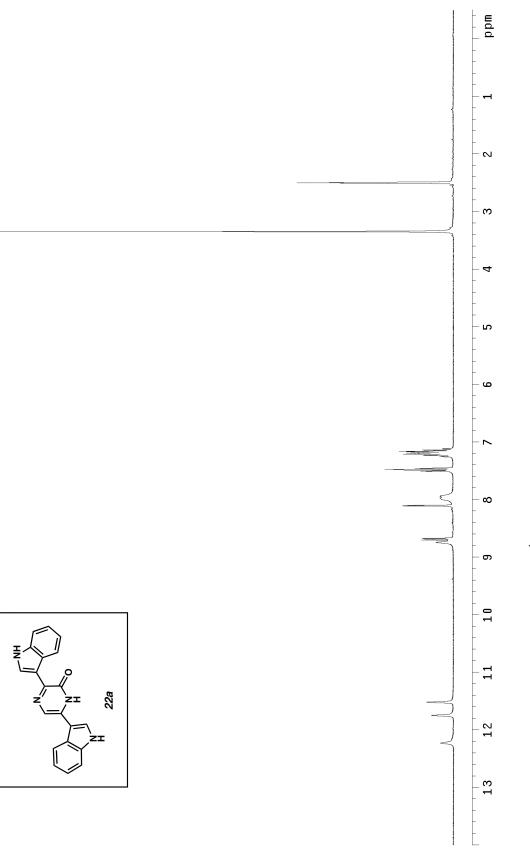


Figure A2.5 Infrared spectrum (thin film/NaCl) of compound 53a.



*Figure A2.6* <sup>13</sup>C NMR (75 MHz, acetone- $d_6$ ) of compound **53a**.





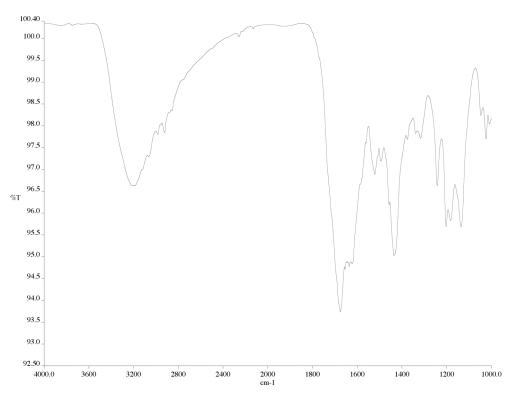
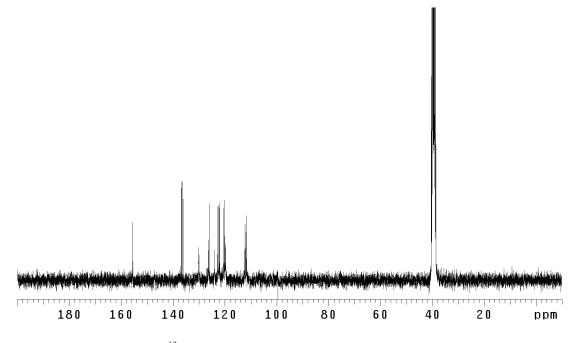
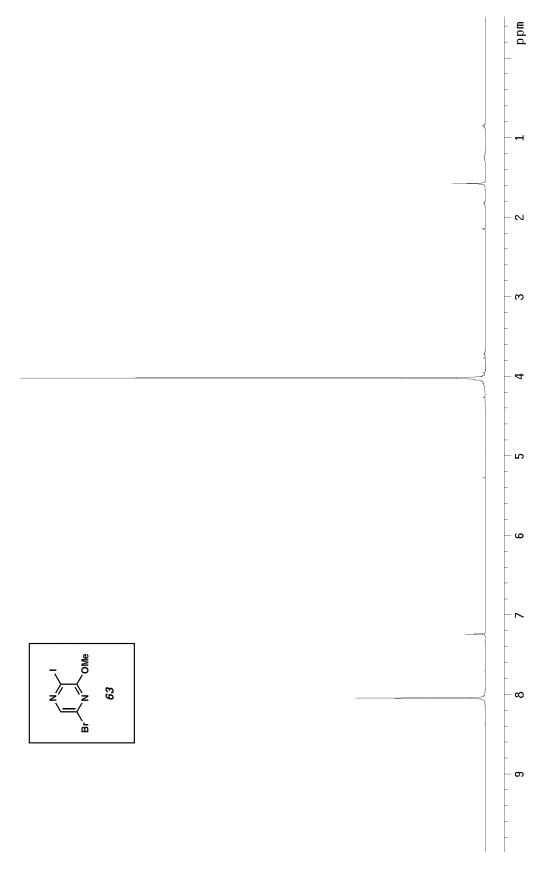


Figure A2.8 Infrared spectrum (thin film/NaCl) of compound 22a.



*Figure A2.9*  $^{13}$ C NMR (125 MHz, DMSO- $d_6$ ) of compound **22a**.





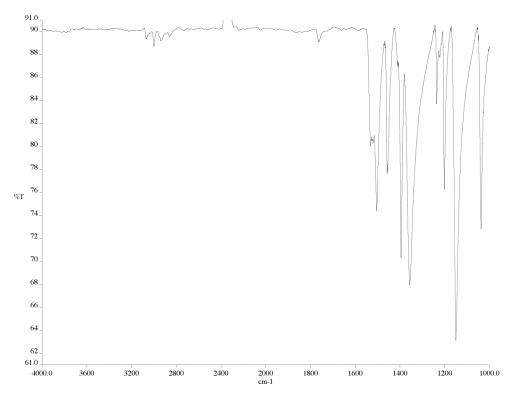
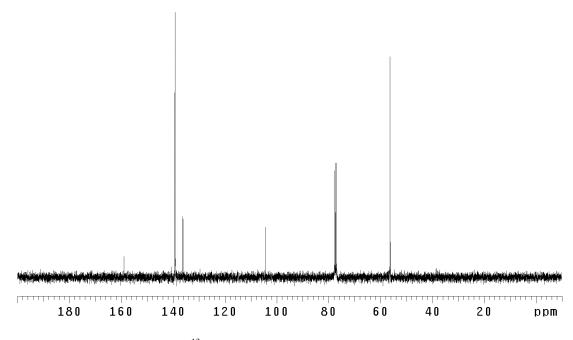
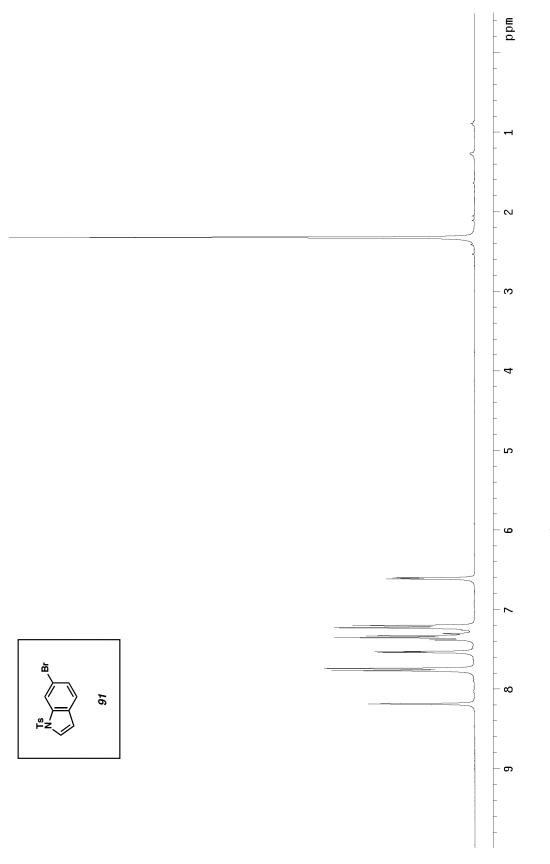


Figure A2.11 Infrared spectrum (KBr pellet) of compound 63.



*Figure A2.12* <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) of compound **63**.





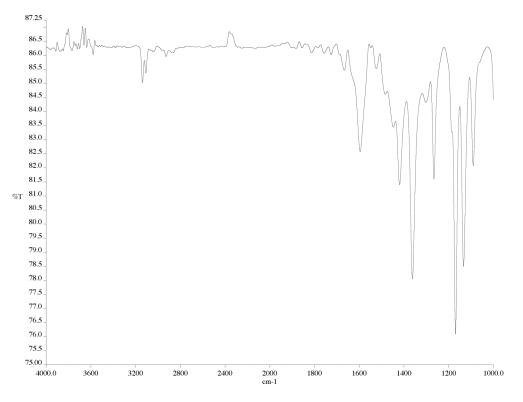
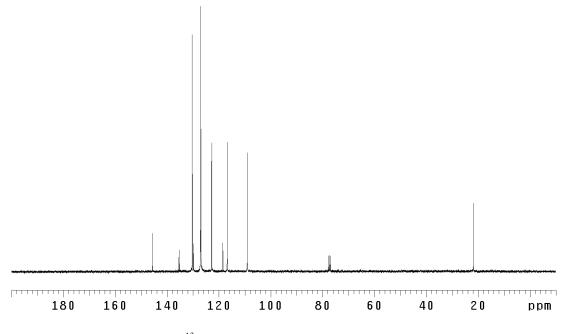
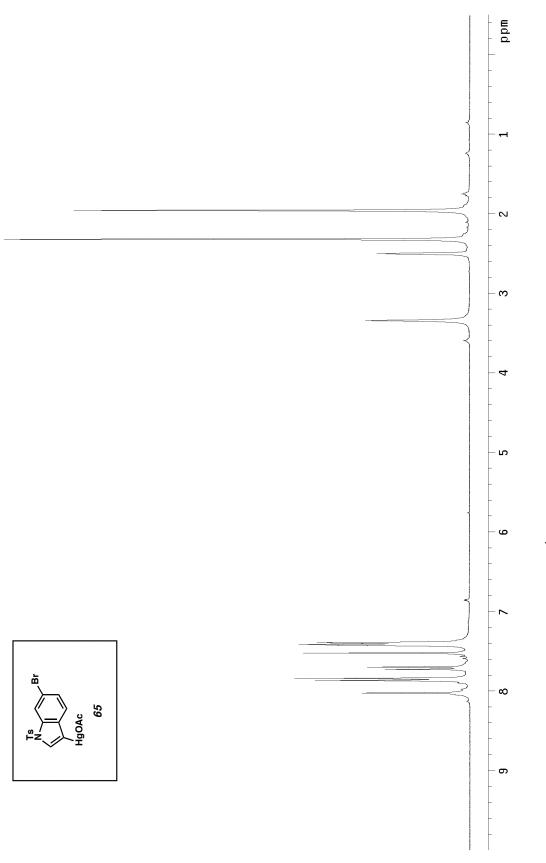


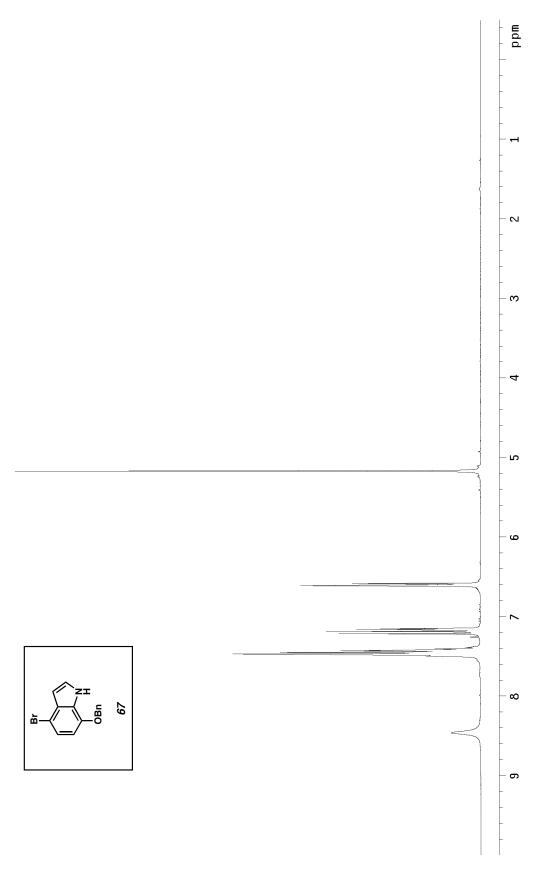
Figure A2.14 Infrared spectrum (thin film/NaCl) of compound 91.



*Figure A2.15* <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) of compound **91**.









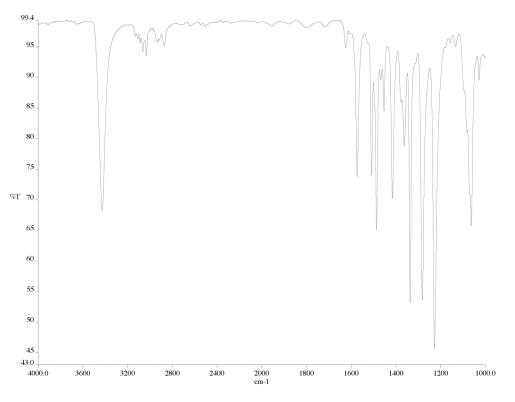
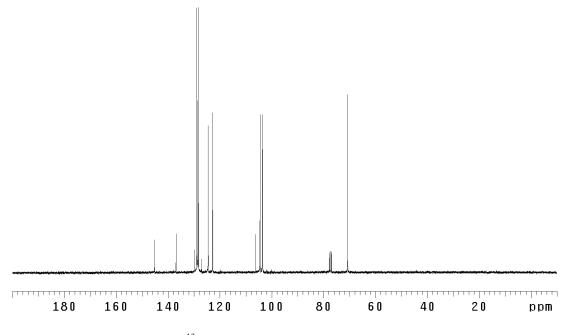
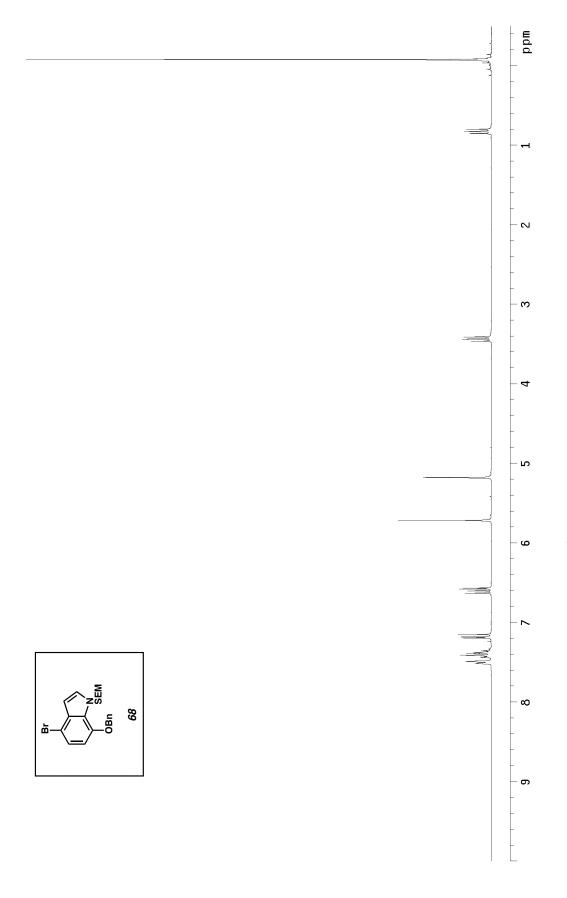


Figure A2.18 Infrared spectrum (thin film/NaCl) of compound 67.



*Figure A2.19* <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) of compound **67**.





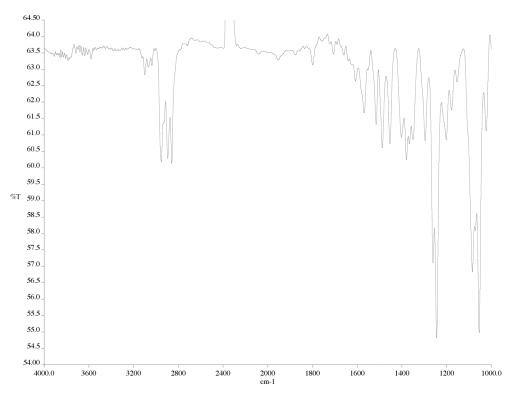
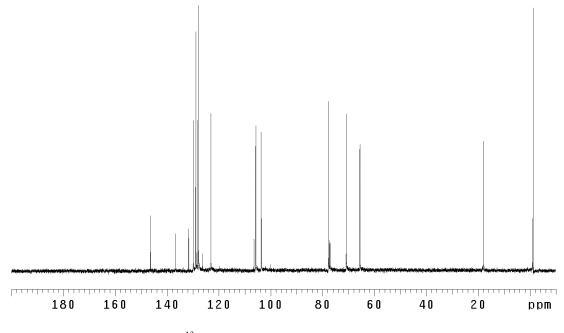
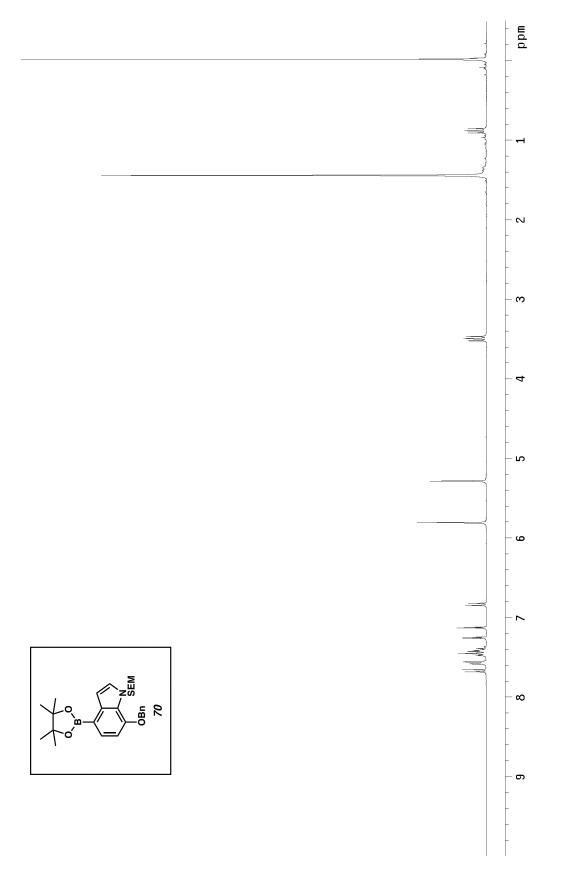


Figure A2.21 Infrared spectrum (thin film/NaCl) of compound 68.



*Figure A2.22* <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) of compound **68**.





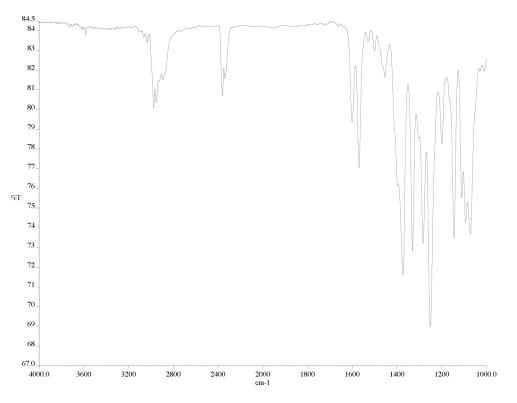
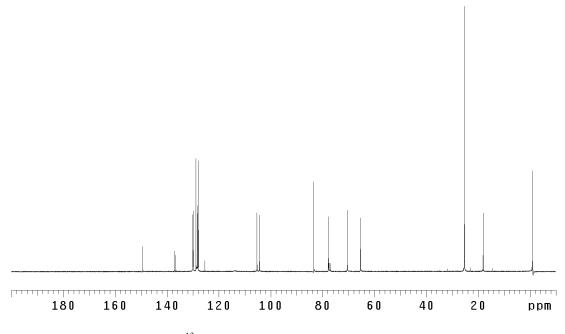
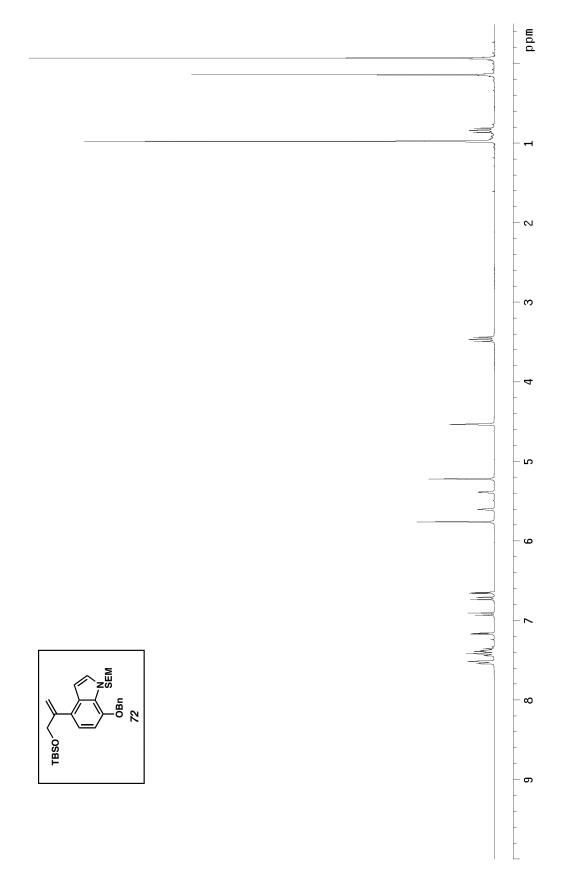


Figure A2.24 Infrared spectrum (thin film/NaCl) of compound 70.



*Figure A2.25* <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) of compound **70**.





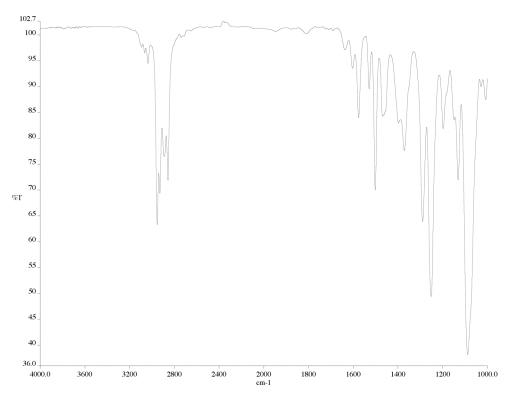
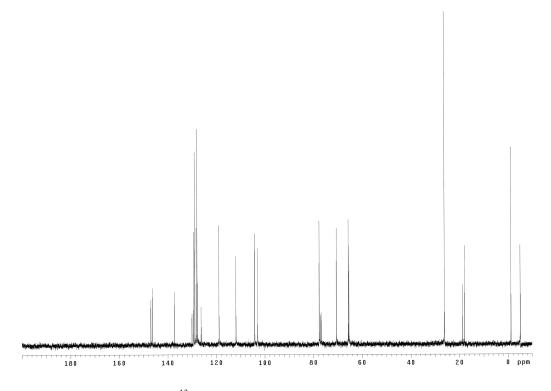
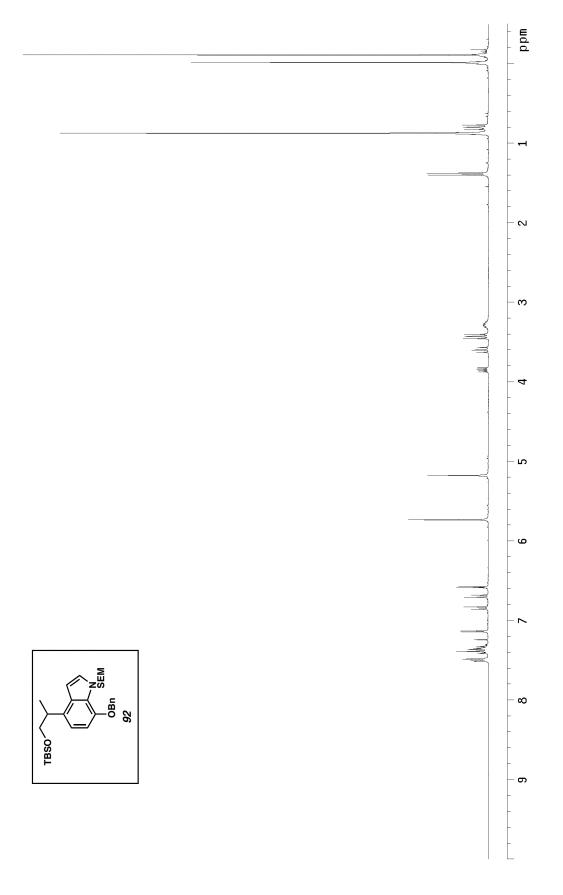


Figure A2.27 Infrared spectrum (thin film/NaCl) of compound 72.



*Figure A2.28* <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) of compound **72**.





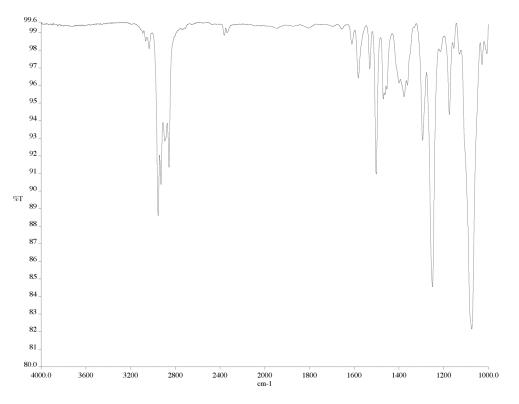
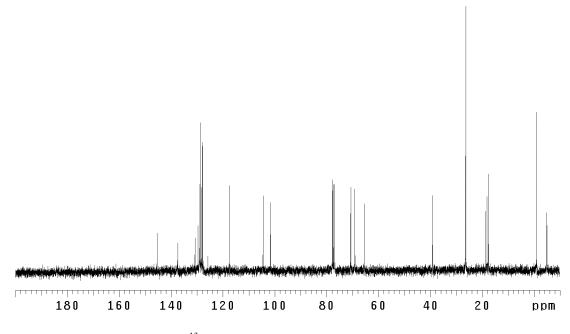
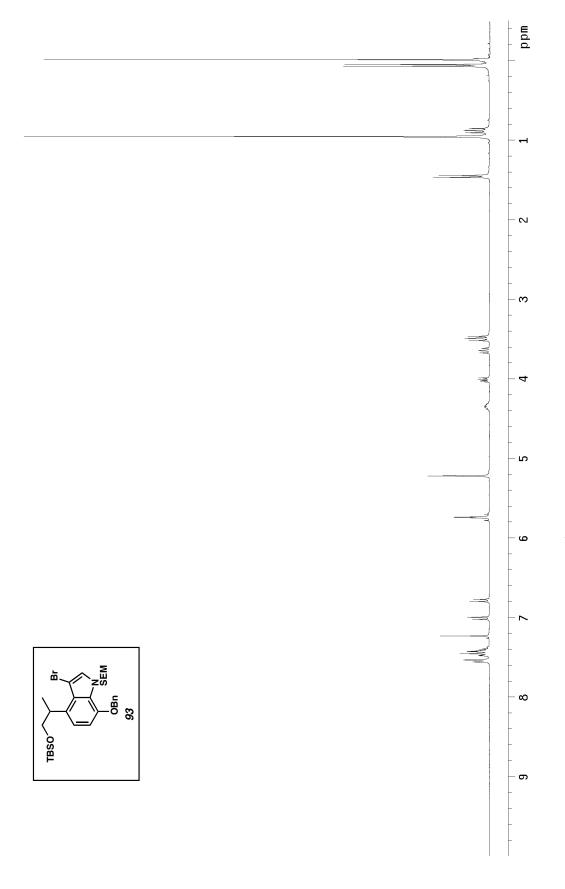


Figure A2.30 Infrared spectrum (thin film/NaCl) of compound 92.



*Figure A2.31* <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) of compound **92**.





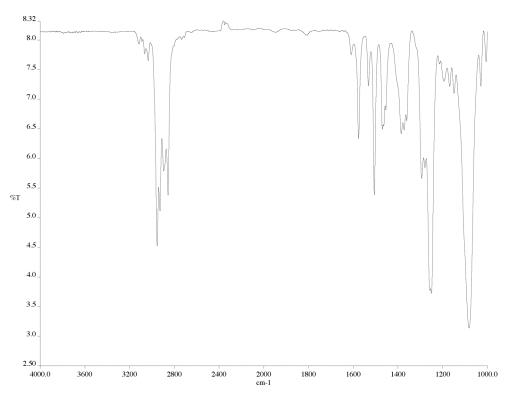
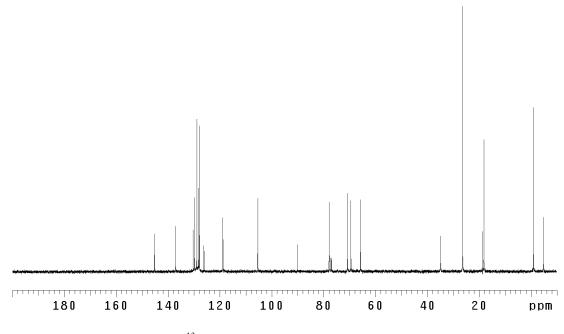
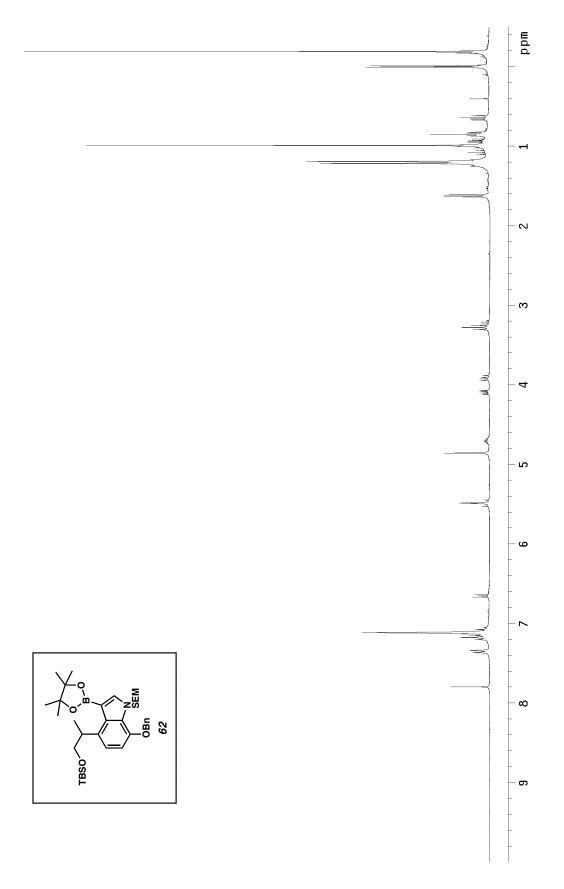


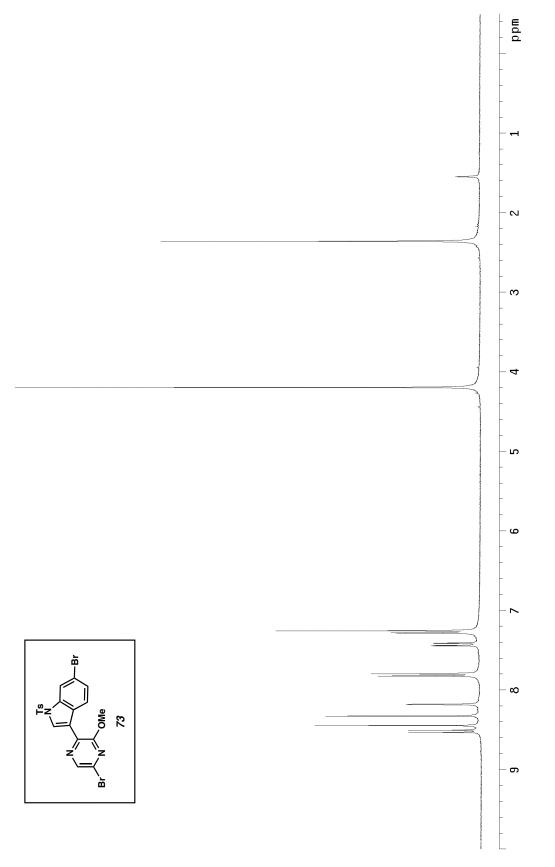
Figure A2.33 Infrared spectrum (thin film/NaCl) of compound 93.



*Figure A2.34* <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) of compound **93**.









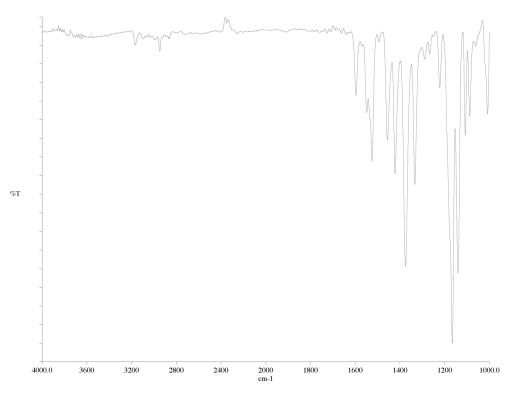
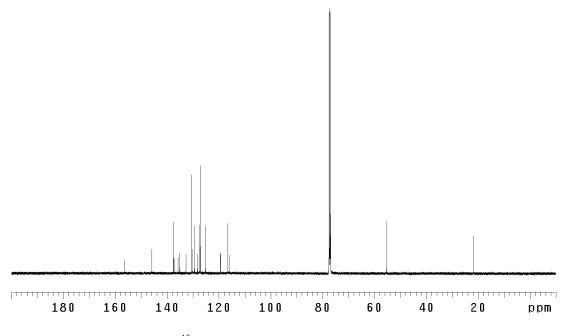
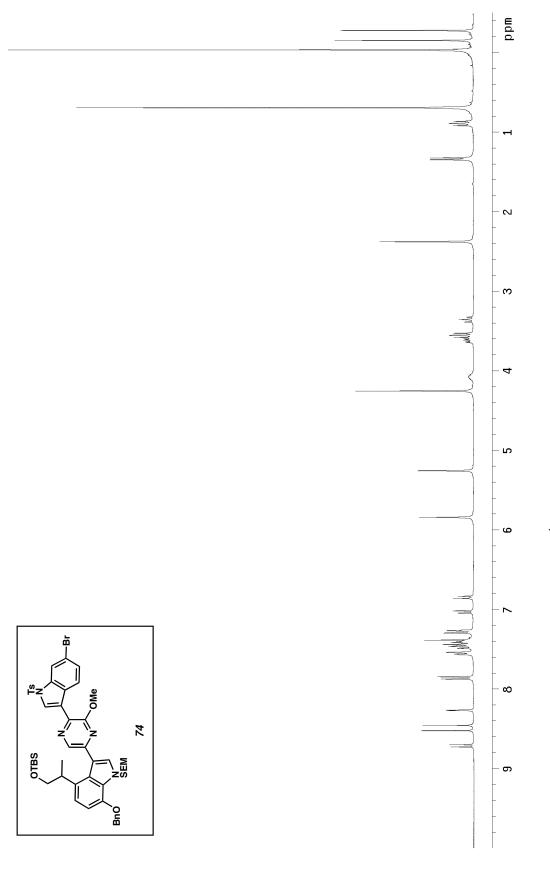


Figure A2.37 Infrared spectrum (thin film/NaCl) of compound 73.



*Figure A2.38*  $^{13}$ C NMR (125 MHz, CDCl<sub>3</sub>) of compound **73**.





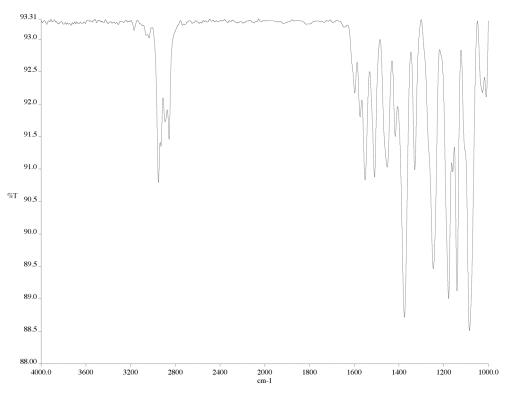
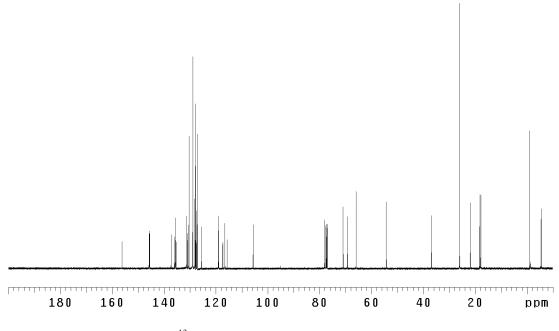
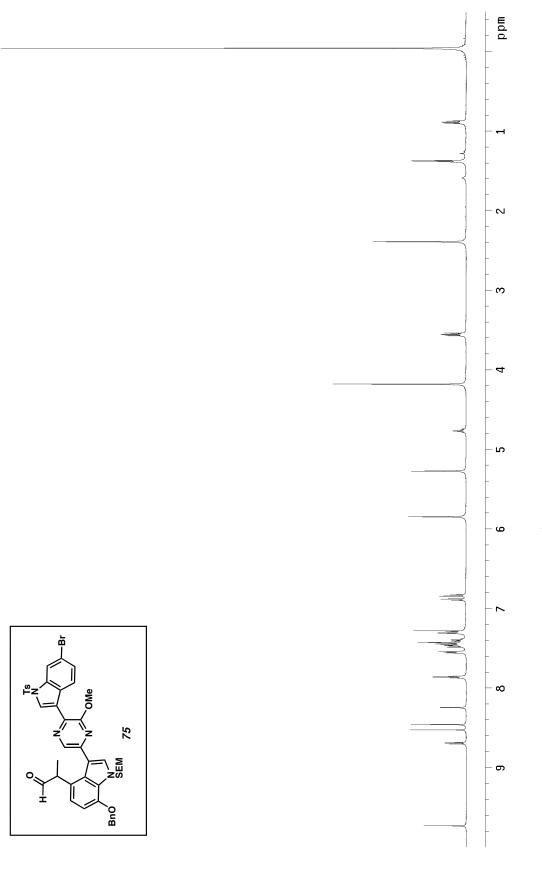


Figure A2.40 Infrared spectrum (thin film/NaCl) of compound 74.



*Figure A2.41*  $^{13}$ C NMR (125 MHz, CDCl<sub>3</sub>) of compound **74**.





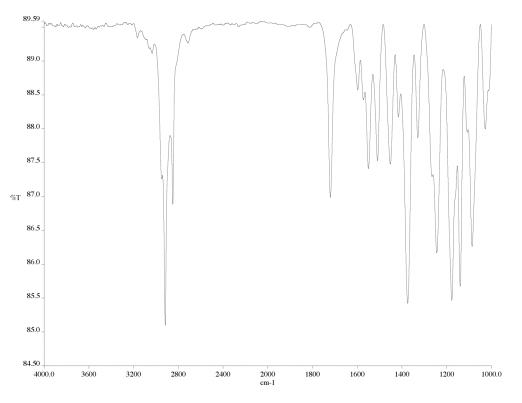
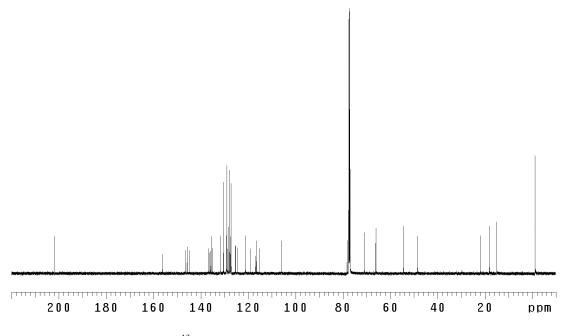
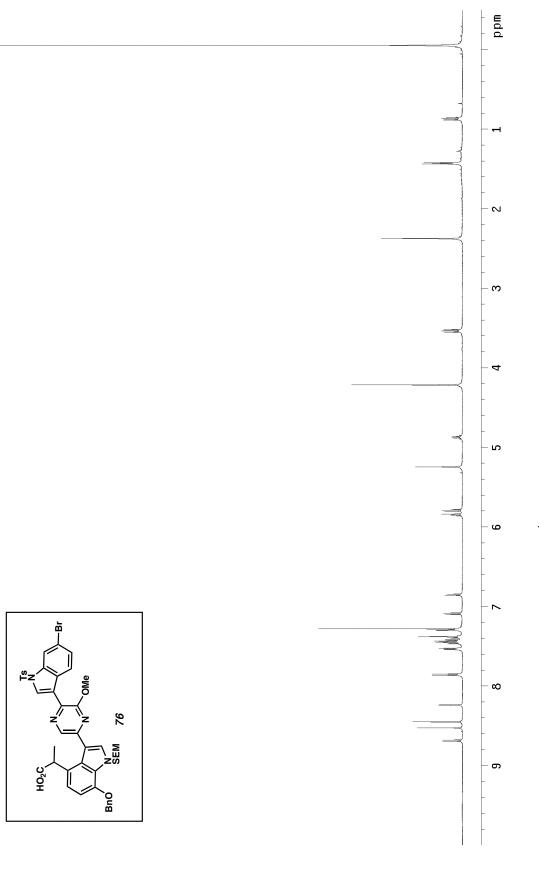


Figure A2.43 Infrared spectrum (thin film/NaCl) of compound 75.



*Figure A2.44* <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) of compound **75**.





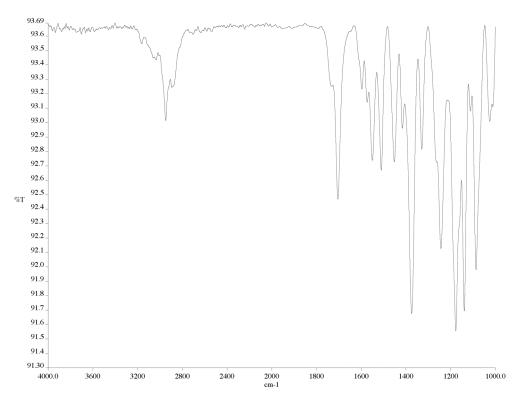
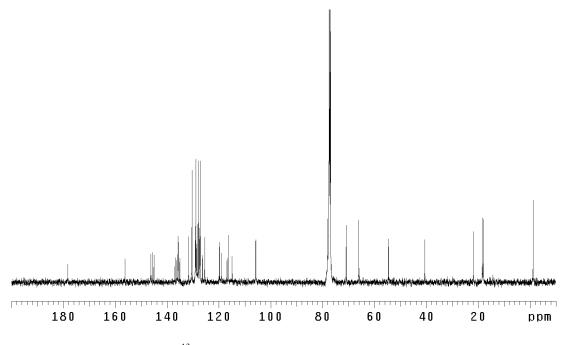
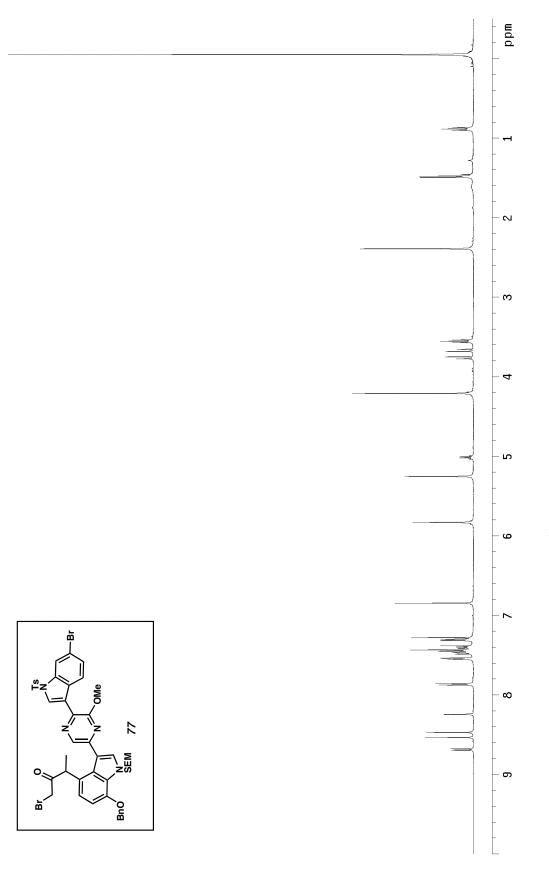


Figure A2.46 Infrared spectrum (thin film/NaCl) of compound 76.



*Figure A2.47* <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) of compound **76**.





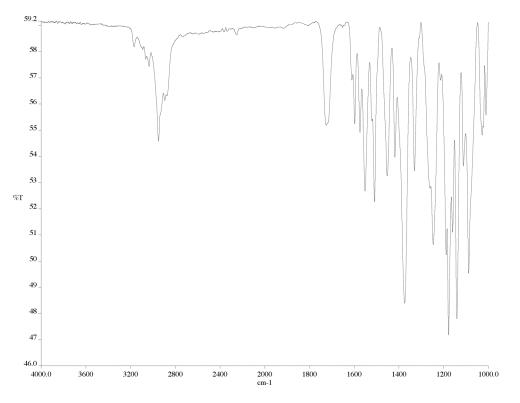
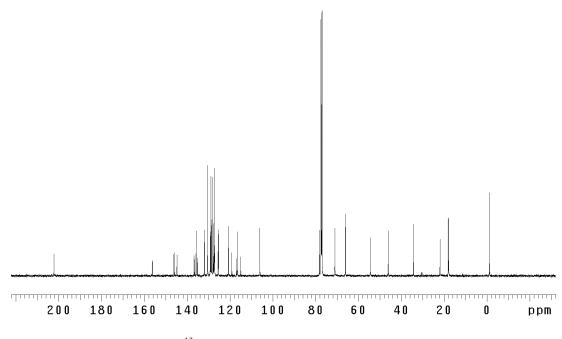
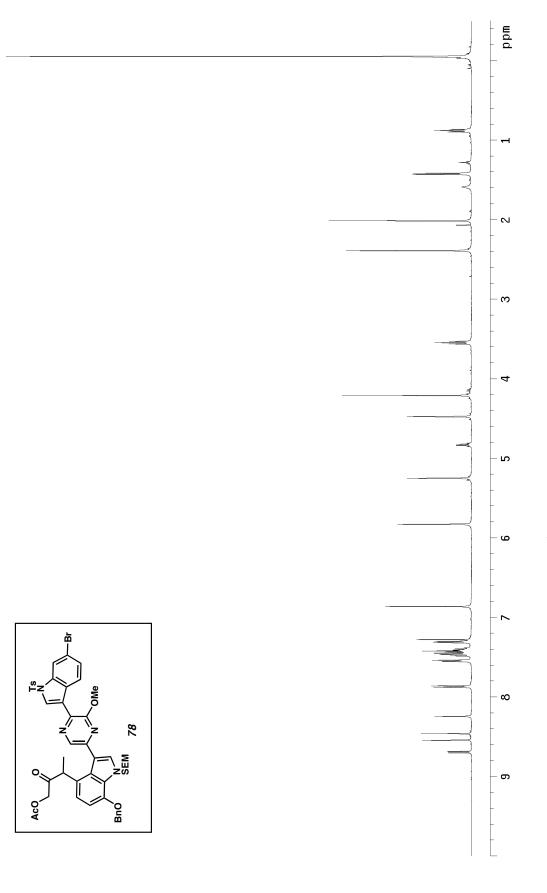


Figure A2.49 Infrared spectrum (thin film/NaCl) of compound 77.



*Figure A2.50*  $^{13}$ C NMR (125 MHz, CDCl<sub>3</sub>) of compound **77**.





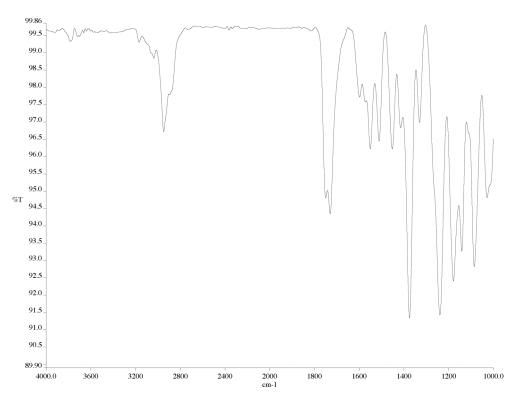
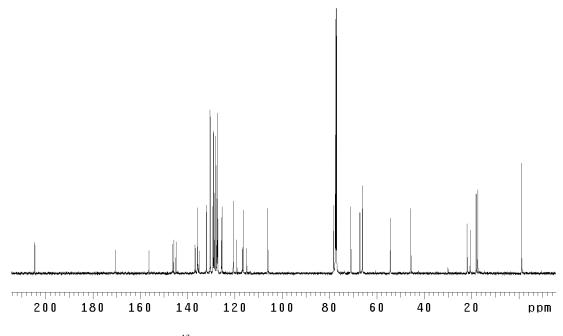
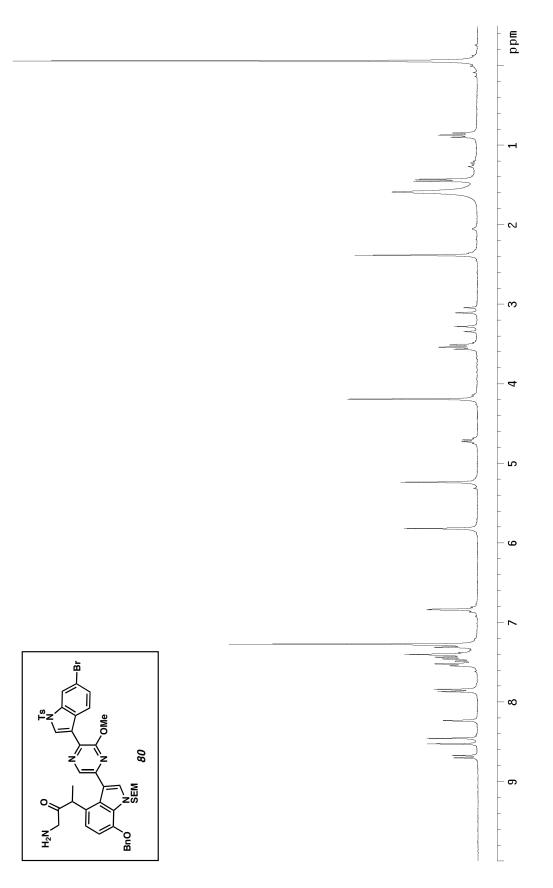


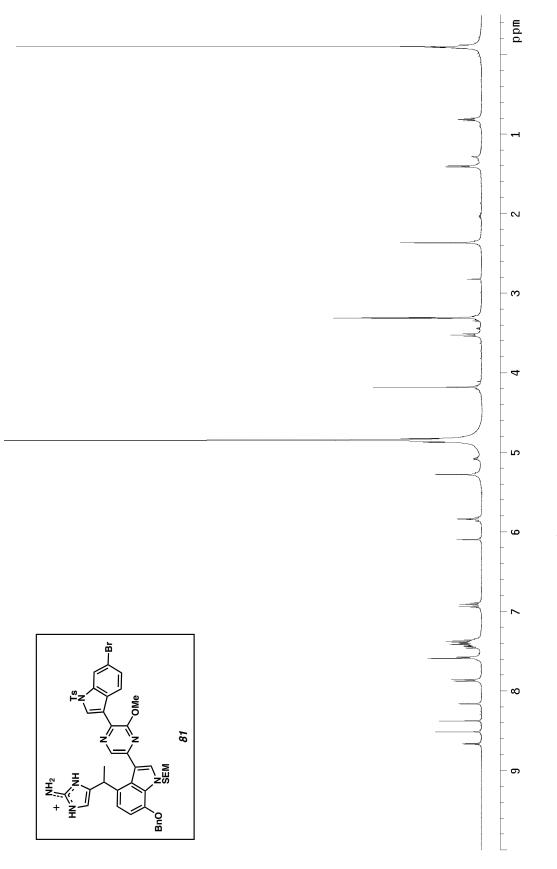
Figure A2.52 Infrared spectrum (thin film/NaCl) of compound 78.



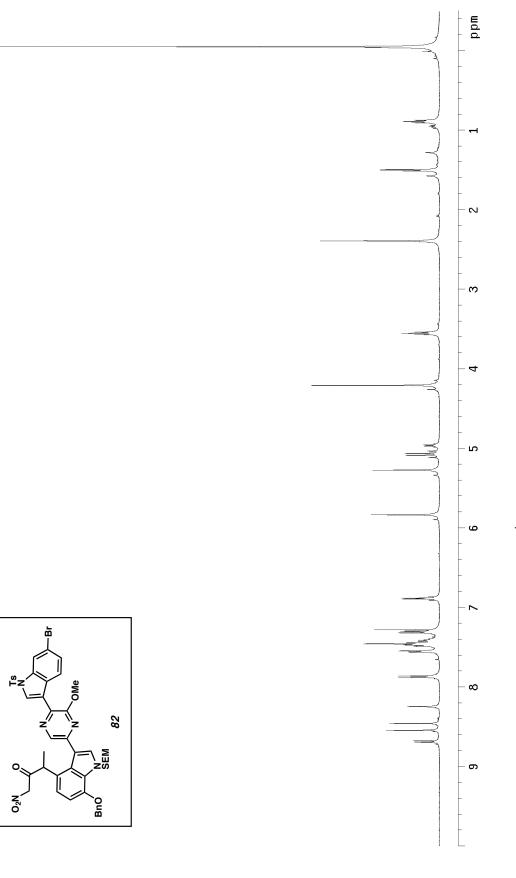
*Figure A2.53* <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) of compound **78**.













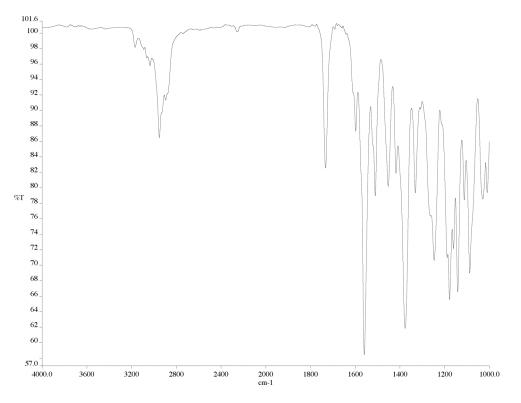
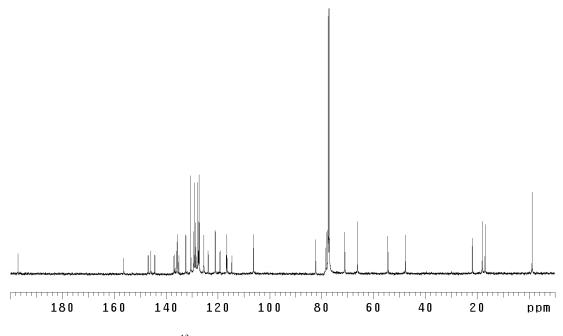
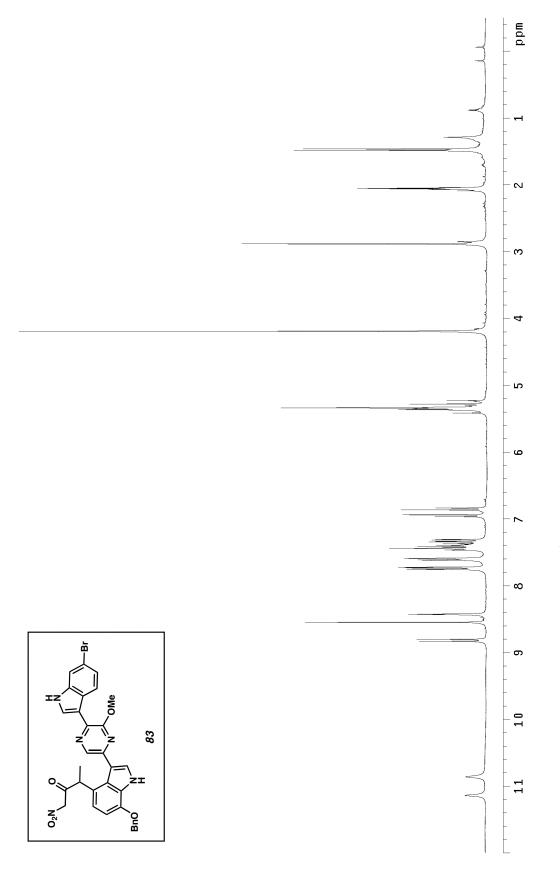


Figure A2.57 Infrared spectrum (thin film/NaCl) of compound 82.



*Figure A2.58* <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) of compound **82**.





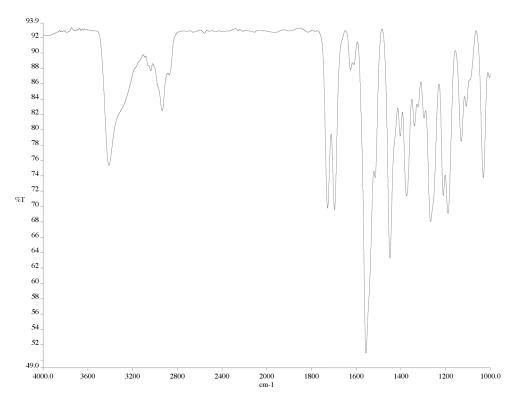
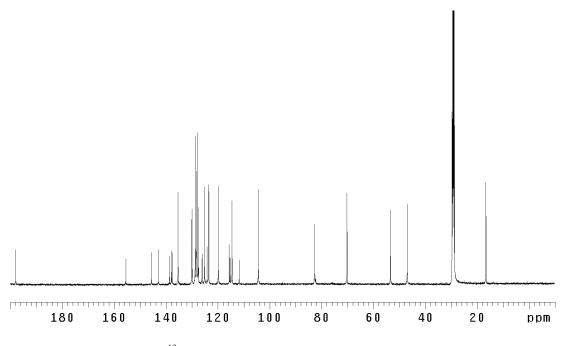
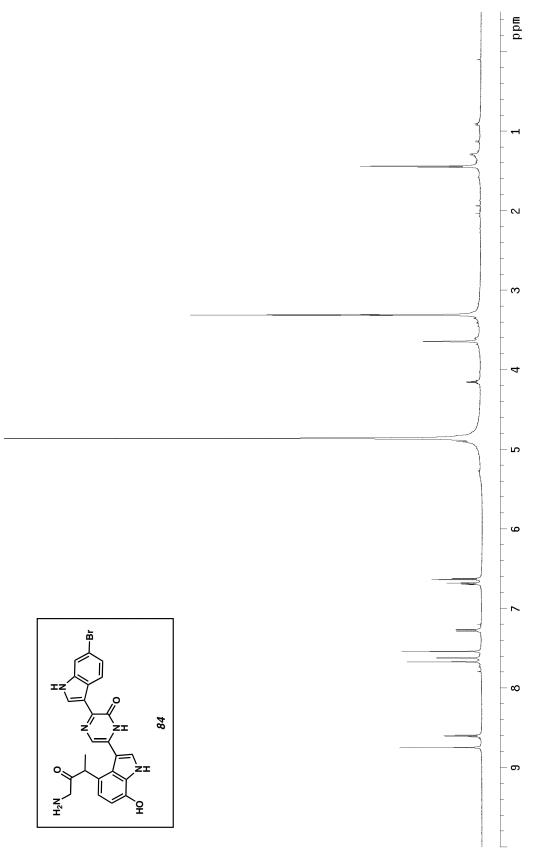


Figure A2.60 Infrared spectrum (thin film/NaCl) of compound 83.



*Figure A2.61* <sup>13</sup>C NMR (125 MHz, acetone- $d_6$ ) of compound **83**.





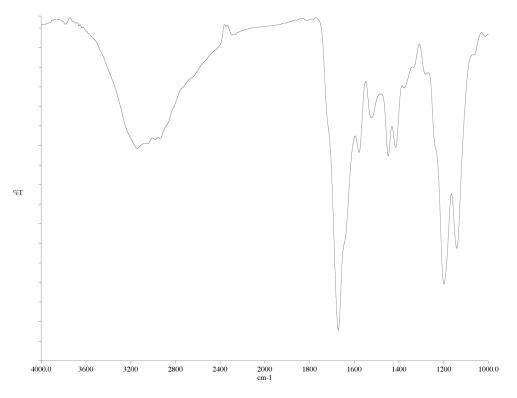
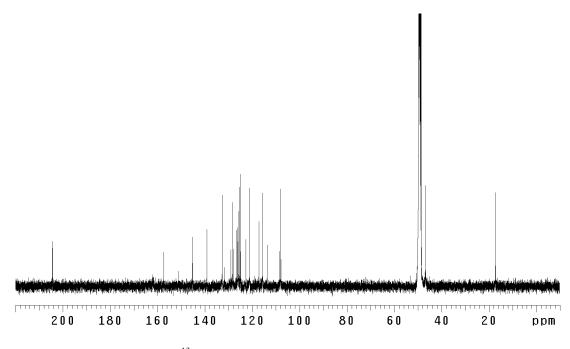
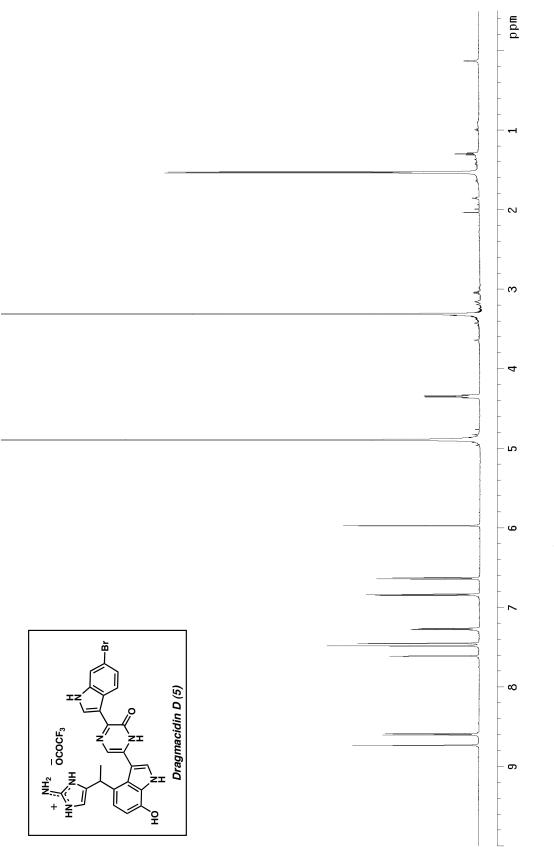


Figure A2.63 Infrared spectrum (thin film/NaCl) of compound 84.



*Figure A2.64*  $^{13}$ C NMR (125 MHz, CD<sub>3</sub>OD) of compound **84**.





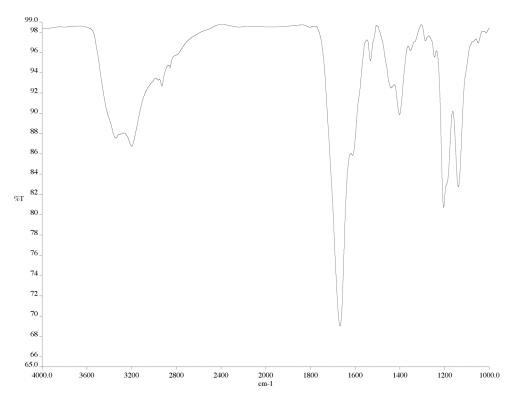
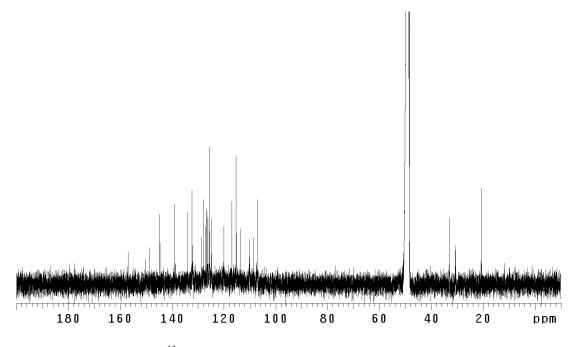
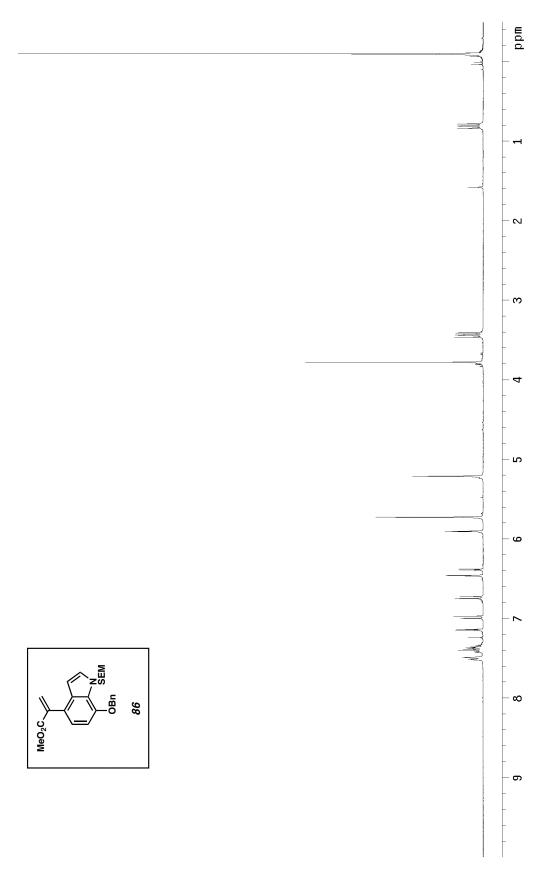


Figure A2.66 Infrared spectrum (thin film/NaCl) of dragmacidin D (5).



*Figure A2.67*  $^{13}$ C NMR (125 MHz, CD<sub>3</sub>OD) of dragmacidin D (**5**).





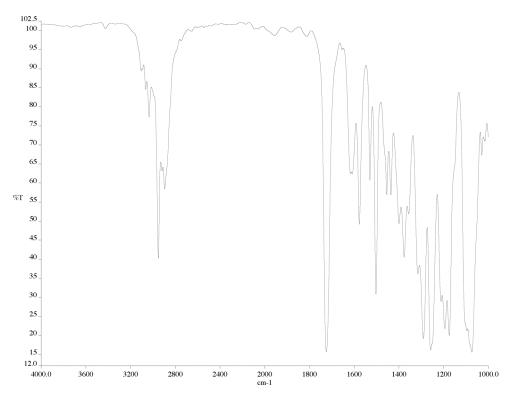
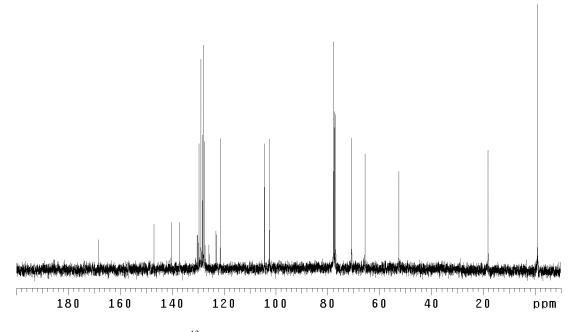
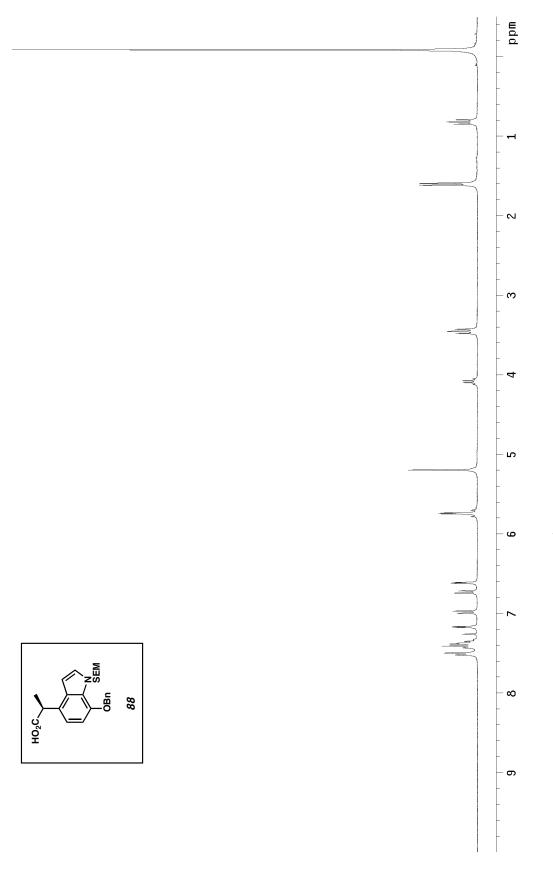


Figure A2.69 Infrared spectrum (thin film/NaCl) of compound 86.



*Figure A2.70*  $^{13}$ C NMR (75 MHz, CDCl<sub>3</sub>) of compound **86**.





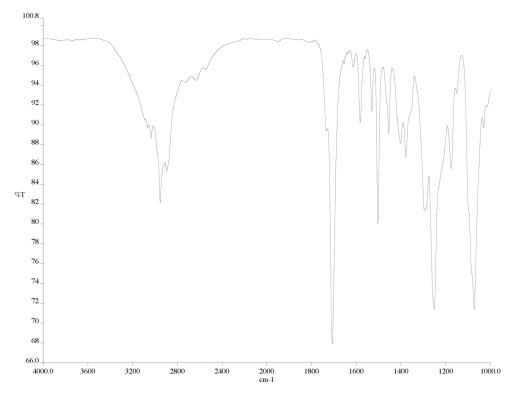
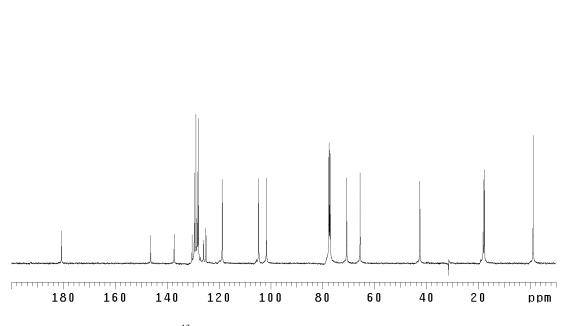


Figure A2.72 Infrared spectrum (thin film/NaCl) of compound 88.



*Figure A2.73* <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) of compound **88**.

#### **CHAPTER THREE**

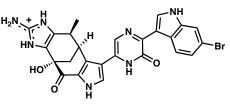
# The Total Synthesis of (+)- and (–)-Dragmacidin $\mathbf{F}^{\dagger}$

### 3.1 Background

## 3.1.1 Introduction

Having developed a strategy to construct the bis(indole)pyrazinone core of dragmacidin D (**5**, Chapter 2), we set out to extend the scope of our halogen-selective Suzuki coupling methodology to the synthesis of related natural products. We hypothesized that our approach could be amenable to the preparation of the antiviral agent dragmacidin F,<sup>1</sup> which is perhaps the most daunting target of the dragmacidin natural products (Figure 3.1.1).<sup>2</sup>

Figure 3.1.1



Dragmacidin F (7)

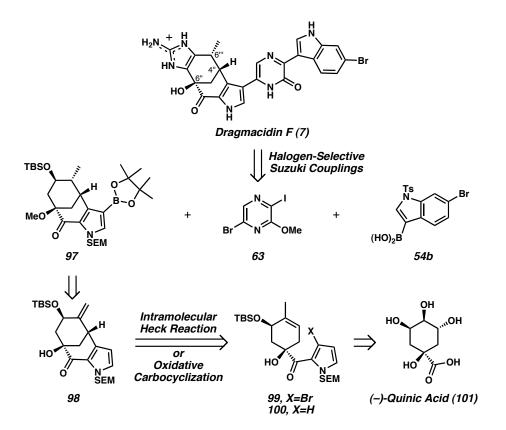
The antiviral agent dragmacidin F (7) possesses a variety of structural features that make it an attractive target for total synthesis. These synthetic challenges include the differentially substituted pyrazinone, the bridged [3.3.1] bicyclic ring system, which is

<sup>&</sup>lt;sup>†</sup> This work was performed in collaboration with Daniel D. Caspi, a graduate student in the Stoltz group.

fused to both the trisubstituted pyrrole and aminoimidazole heterocycles, and the installation and maintenance of the 6-bromoindole fragment.

#### 3.1.2 Retrosynthetic Analysis of Dragmacidin F

Our retrosynthetic analysis for dragmacidin F(7) is shown in Scheme 3.1.1. On the basis of our experience with dragmacidin D(5), we reasoned that the aminoimidazole moiety would best be incorporated at a late stage in the synthesis. The carbon skeleton of the natural product would then arise via a series of halogen-selective Suzuki crosscoupling reactions (97 + 63 + 54b). Pyrazine 63 and indolylboronic acid 54b were both readily accessible, while pyrroloboronic ester 97 perhaps could be derived from pyrrolefused bicycle 98, our key retrosynthetic intermediate. We then targeted bicycle 98 from two related directions: a Pd(0)-mediated intramolecular Heck reaction<sup>3</sup> of bromopyrrole 99 and a Pd(II)-promoted oxidative carbocyclization<sup>4</sup> involving *des*-bromopyrrole 100. The successful implementation of the latter method was particularly attractive since it is closely aligned with our interest in Pd(II)-catalyzed dehydrogenation reactions.<sup>5</sup> Both of the cyclization substrates (99 and 100) could be prepared from commercially available (-)-quinic acid (101).<sup>6</sup> At the time of this synthetic effort, the absolute stereochemistry of natural dragmacidin F (7) was not known; thus, the absolute stereochemistry of our target (7) was chosen arbitrarily.

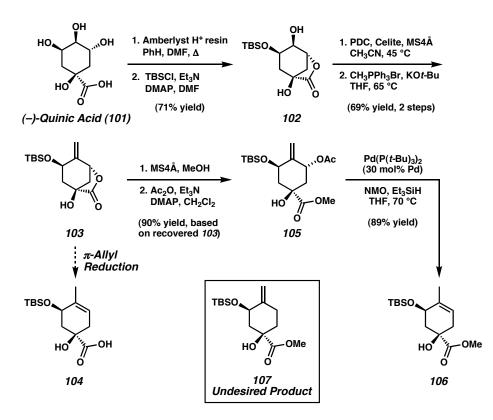


#### 3.2 The Total Synthesis of (+)-Dragmacidin F

## 3.2.1 Synthesis of Cyclization Substrates

Our synthesis of dragmacidin F (7) began with a known two-step protocol involving lactonization and silylation of (–)-quinic acid (101) to afford bicyclic lactone 102 (Scheme 3.2.2).<sup>7</sup> Subsequent oxidation and Wittig olefination of 102 produced exomethylene lactone 103 in good yield. Initially, we envisioned the direct conversion of lactone 103 to unsaturated carboxylic acid 104 by executing a homogeneous Pd(0)catalyzed  $\pi$ -allyl hydride addition reaction.<sup>8</sup> Despite considerable experimentation, however, exposure of lactone 103 to a variety of Pd and hydride sources under standard conditions<sup>8</sup> led to the formation of complex product mixtures. As a result, a more stepwise approach was tried. Methanolysis of lactone **103** followed by acetylation of the resulting 2° alcohol<sup>9</sup> gave rise to allylic acetate **105**, another potential substrate for  $\pi$ -allyl reduction chemistry. Although **105** did react under most literature protocols, undesired exocyclic olefin **107** was typically the major product observed. After substantial optimization, we were able to access **106** as the major product by employing stoichiometric Pd(P(*t*-Bu)<sub>3</sub>)<sub>2</sub><sup>10</sup> in the presence of triethylsilane as a reductant. Further refinements designed to facilitate catalysis led to a reduced Pd loading (30 mol%) when *N*-methylmorpholine-*N*-oxide (NMO) was used as an additive.<sup>11</sup> Under these conditions, cyclohexene **106** was obtained in 89% yield as a single olefin regioisomer. Unfortunately, this transformation often gave inconsistent results and was particularly sensitive to oxygen, water, and the quality of Et<sub>3</sub>SiH. These difficulties coupled with the high catalyst loading resulted in substantial material throughput problems. We therefore sought yet another method to prepare cyclohexene **106** or a closely related derivative thereof (i.e., **104**) in a more facile and preparative manner.

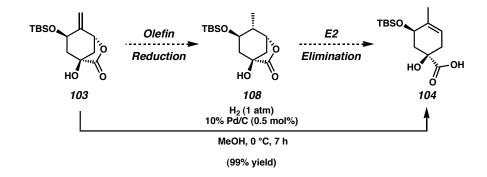
#### Scheme 3.2.2



In our revised plan, we conceived a two-step route to obtain carboxylic acid **104** via diastereoselective reduction of olefin **103** followed by base-promoted elimination of the carboxylate functionality of **108** (Scheme 3.2.3). Daniel Caspi attempted the first part of this sequence by exposing olefin **103** to standard catalytic hydrogenation conditions (Pd/C, 1 atm H<sub>2</sub>). Surprisingly, these conditions led to the production of a compound that was more polar than we expected for simple olefin hydrogenation (i.e., **108**). To our delight, the product was identified as unsaturated carboxylic acid **104**. Under our optimized reaction conditions (0.5 mol% Pd/C, 1 atm H<sub>2</sub>, MeOH, 0 °C), essentially quantitative reductive isomerization to **104** was observed. Although the mechanism of

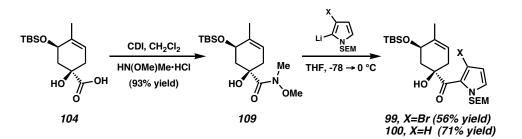
this transformation has not been studied extensively, simple control experiments suggest that stepwise reduction/elimination<sup>12</sup> or  $\pi$ -allyl reduction processes are not operative.<sup>13</sup>

Scheme 3.2.3



With facile access to cyclohexene carboxylic acid **104**, preparation of the key cyclization precursors proceeded without difficulty. Activation of acid **104** with CDI followed by the addition of HN(OMe)Me • HCl afforded Weinreb amide **109** (Scheme 3.2.4). The Weinreb amide functionality was then displaced with the appropriate lithiopyrrole<sup>14</sup> reagent to produce Heck cyclization substrate **99**<sup>15</sup> and oxidative cyclization substrate **100**.

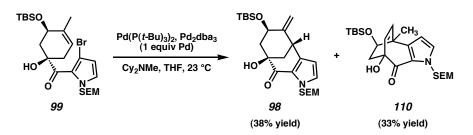
Scheme 3.2.4



## 3.2.2 Constructing the [3.3.1] Bicycle

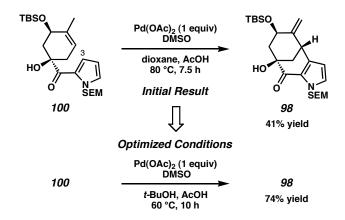
Extensive studies were carried out in order to achieve the intramolecular Heck cyclization of bromopyrrole **99**. Attempts to utilize standard procedures were unsuccessful,<sup>3</sup> likely due to the thermal instability of the bromopyrrole moiety. However, implementation of the room-temperature conditions developed by Fu<sup>16</sup> provided the desired [3.3.1] bicyclic product (**98**), albeit in low yield (Scheme 3.2.5). Unfortunately, the formation of **98** was hampered by competitive production of [3.2.2] bicycle **110**. Although efforts to optimize temperature, solvent, base, and concentration were not met with success, it was found that increased quantities of Pd improved the ratio of the desired [3.3.1] bicycle (**98**) to the undesired [3.2.2] bicycle (**110**). In addition, the ratio of **98** to **110** decreased over time,<sup>17</sup> suggesting that the active catalytic species varied during the course of the reaction or that selectivity changed as the concentration of  $R_3NH^*Br^-$  increased.





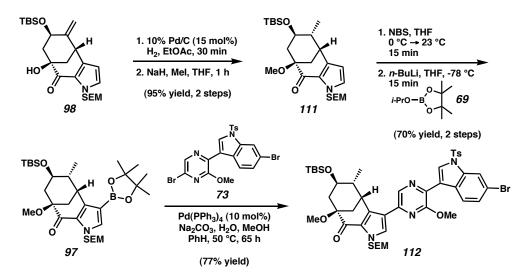
Although the Heck reaction was useful for preparing reasonable quantities of bicycle **98**, an alternative and potentially more selective route to **98** was desired. In conjunction with ongoing research in our group,<sup>5</sup> we turned to the Pd(II)-mediated C-C bond forming approach. In this scenario, C(3)-unsubstituted pyrrole **100** would undergo

intramolecular carbocyclization to afford 98 (Scheme 3.2.6). Initial experimentation revealed that pyridine and ethyl nicotinate were not effective ligands for promoting cyclization in the presence of Pd(OAc)2.5c,d However, Daniel Caspi found that by using DMSO as a ligand<sup>18</sup> the desired cyclization product could be obtained in modest yield. Subsequent optimization of solvent, temperature, and reaction time led to a set of improved conditions whereby the desired pyrrole-fused bicycle 98 was formed as a single stereo- and regioisomer in 74% yield. Interestingly, these conditions take advantage of a similar solvent mixture employed in Pd cyclization methodology from our laboratory.<sup>5c,d</sup> This transformation  $(100 \rightarrow 98)$  is particularly noteworthy since it results in functionalization of the electronically deactivated and sterically congested C(3) position of acyl pyrrole **100**.<sup>19,20</sup> Despite our best efforts, we were unable to effect catalytic turnover of Pd with a stoichiometric oxidant in this reaction, presumably due to extensive oxidative decomposition of both the starting material and the desired product.<sup>21</sup> Nonetheless, the Pd(II)-mediated strategy provided bicycle 98 in nearly twice the isolated yield as the Heck route using equivalent amounts of Pd and obviated the need for polybrominated pyrroles.<sup>15,22</sup>



## 3.2.3 Assembling the Carbon Skeleton of Dragmacidin F

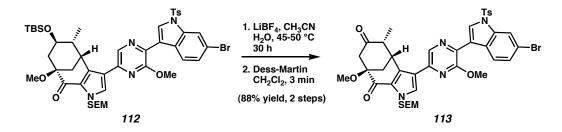
With the [3.3.1] bicyclic framework in hand (i.e., **98**), we focused our attention on constructing the full carbon skeleton of dragmacidin F (**112**, Scheme 3.2.7). The final stereocenter present in the natural product was installed via catalytic hydrogenation of olefin **108** and was followed by methylation of the 3° alcohol to produce bis(ether) **111**. The methyl protecting group was selected initially for its robustness<sup>9</sup> and would presumably allow for the exploration of late-stage chemistry in the form of a model system.<sup>23</sup> Methyl ether **111** was then elaborated via regioselective bromination of the pyrrole and metalation to boronic ester **97**. In the critical halogen-selective Suzuki fragment coupling, pyrroloboronic ester **97** was reacted with dibromide **73** (prepared from **63** + **54b**) under Pd(0) catalysis. By analogy to our dragmacidin D studies, we were pleased to find that at 50 °C, the desired C-C bond forming reaction took place to afford the fully coupled product (**112**) in 77% yield. Importantly, the indolylbromide moiety was maintained under these reaction conditions.



## 3.2.4 End-Game Studies

With the carbon framework completed, few tasks remained in order to finish the total synthesis of dragmacidin F (7), namely, removal of all protecting groups and installation of the aminoimidazole unit. Of particular note is the similarity of these synthetic challenges to those encountered in our total synthesis of dragmacidin D (5). Not surprisingly, we decided to utilize the methods that were already familiar to us in order to elaborate **112** to the desired natural product (7). To this end, we anticipated that the presence of an amino group  $\alpha$  to the ketone would allow for eventual introduction of the aminoimidazole moiety. Therefore, selective deprotection of silyl ether **112**, followed by oxidation with Dess-Martin periodinane, produced ketone **113** (Scheme 3.2.8).

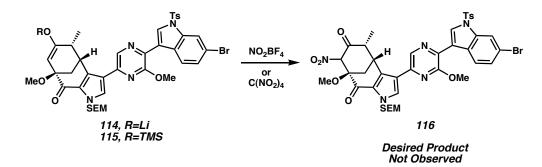
Scheme 3.2.8



# 3.2.4.1 End-Game Strategy 1

Our first effort to functionalize the ketone  $\alpha$ -position involved a nitration strategy to access a compound analogous to an intermediate employed in the dragmacidin D synthesis (Scheme 3.2.9). Both lithium enolate **114** and TMS enol ether **115** were exposed to electrophilic NO<sub>2</sub> sources.<sup>24</sup> Unfortunately, in all of these cases, formation of the desired nitroketone product (**116**) was not observed.

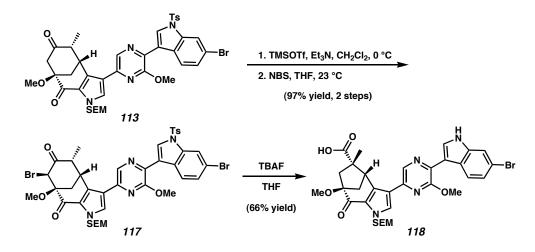
Scheme 3.2.9



#### 3.2.4.2 End-Game Strategy 2

We then turned to an alternative strategy that would involve installation of an  $\alpha$ amino substituent via nucleophilic displacement of an alkylbromide. Therefore, ketone **113** was treated with TMSOTf and then exposed to NBS to afford bromoketone **117** as a single diastereomer (Scheme 3.2.10).<sup>25</sup> Interestingly, when bromoketone **117** was treated with various nitrogenous nucleophiles, base-promoted rearrangements were observed.<sup>26</sup> In fact, reaction of bromide **117** with a basic fluoride anion source (TBAF in THF) gave [3.2.1] bicycle **118** as the major product via a Favorskii rearrangement.<sup>27</sup> The utilization of amine bases also led to the formation of related Favorskii products.

Scheme 3.2.10

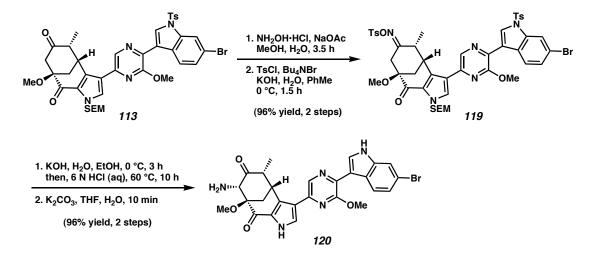


3.2.4.3 End-Game Strategy 3: The Total Synthesis of (+)-Dragmacidin F

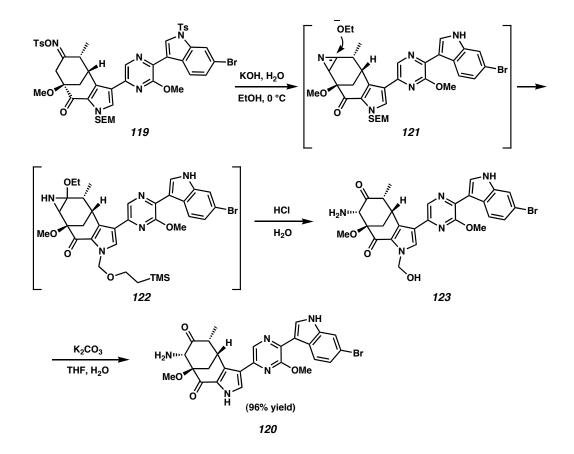
With limited options remaining, we became interested in the use of a Neber rearrangement in order to install the necessary  $\alpha$ -amino substituent.<sup>28,29</sup> In this scenario, an activated oxime derivative would undergo alkoxide-promoted rearrangement to furnish an  $\alpha$ -aminoketone. Thus, ketone **113** was converted to tosyloxime **119** via standard conditions (Scheme 3.2.11). Gratifyingly, exposure of substrate **119** to aqueous KOH in ethanol led to Neber rearrangement. After optimization, we found that simply exposing tosyloxime **119** to i) KOH, ii) HCl, and iii) K<sub>2</sub>CO<sub>3</sub> produced  $\alpha$ -aminoketone **120** as a single regio- and stereochemical isomer in excellent yield.<sup>30,31,32</sup> Furthermore,

under these reaction conditions, both the tosyl and SEM protective groups were quantitatively removed from their corresponding heterocycles. To the best of our knowledge, this is the first example of a successful Neber rearrangement in the context of natural product synthesis.<sup>33</sup>

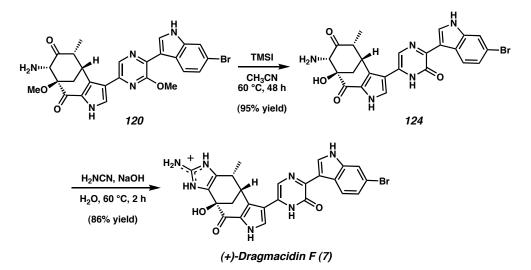
Scheme 3.2.11



А more detailed look the possible mechanism of the Neber at rearrangement/deprotection sequence is shown in Scheme 3.2.12. Exposure of tosyloxime 119 to KOH in ethanol likely leads to the formation of detosylated azirine 121, which is attacked by ethoxide to afford ethoxyaziridine 122.<sup>29a,34</sup> Following acidmediated hydrolysis, the aminoketone moiety is installed with concomitant partial cleavage of the SEM protective group  $(122 \rightarrow 123)$ .<sup>30b,35</sup> Finally, treatment of hemiaminal 123 with  $K_2CO_3$  removes the remaining portion of the SEM group, thus giving rise to the deprotected aminoketone (120).

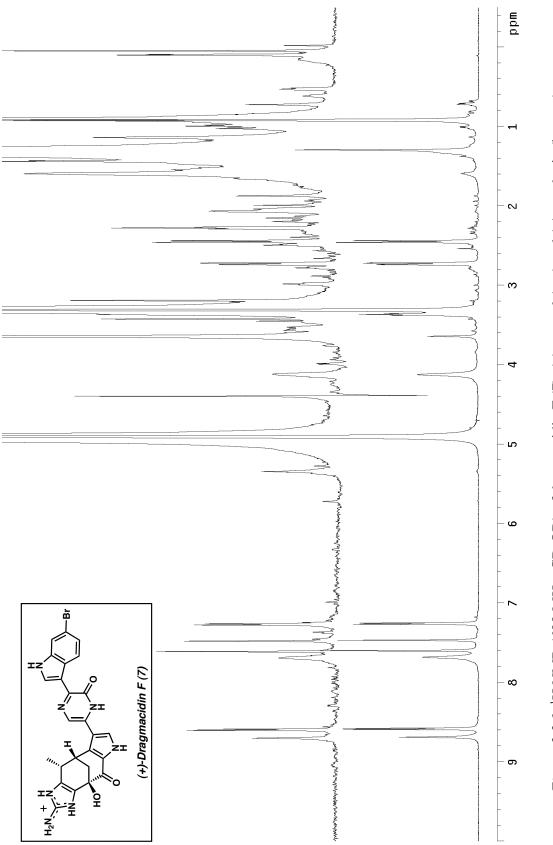


In order to unveil the masked pyrazinone functionality, Neber rearrangement product **120** was treated with TMSI at 60 °C (Scheme 3.2.13).<sup>9</sup> Fortuitously, both the pyrazinone and the 3° alcohol functionalities were revealed simultaneously (**120**  $\rightarrow$  **124**). In the final step of the synthesis, the penultimate aminoketone (**124**) was subjected to cyanamide and aqueous NaOH to produce enantiopure dragmacidin F (**7**).<sup>36</sup> Our efficient and enantiospecific route allows access to **7** in 7.8% overall yield in just 21 steps from (–)-quinic acid (**101**).

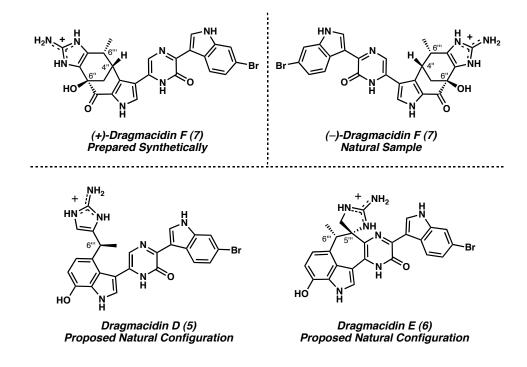


## 3.3 The Absolute Stereochemistry of the Pyrazinone-Containing Dragmacidins

Synthetic dragmacidin F (7) was spectroscopically identical (<sup>1</sup>H NMR, <sup>13</sup>C NMR, IR, UV, HPLC) to a sample obtained from natural sources (Figure 3.3.1),<sup>1b</sup> with the exception of the sign of rotation (natural:  $[\alpha]_{D}^{25}$  –159° (*c* 0.4, MeOH); synthetic:  $[\alpha]_{D}^{23}$  +146° (*c* 0.45, MeOH)). Thus, our synthesis from (–)-quinic acid (**101**) established, for the first time, the absolute configuration of natural dragmacidin F (7) to be (4"*S*, 6"*S*, 6"*S*) as shown in Figure 3.3.2.<sup>37</sup> On the basis of the hypothesis that dragmacidins D, E, and F are biosynthetically related, it is likely that the absolute stereochemical configurations of natural dragmacidins D (**5**) and E (**6**) are (6"*S*) and (5"*R*, 6"*S*), respectively. Having developed a route to the unnatural antipode of dragmacidin F ((+)-7), we set out to extend our approach to the total synthesis of (–)-7.







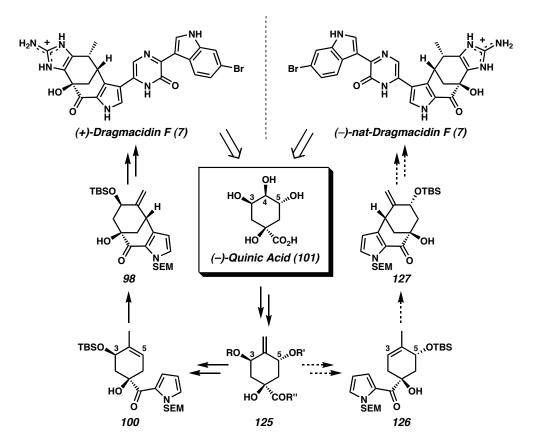
## 3.4 The Total Synthesis of (-)-Dragmacidin F

### 3.4.1 An Enantiodivergent Strategy for the Preparation of (-)-Dragmacidin F

As described above, naturally occurring and readily available (–)-quinic acid (101)<sup>6</sup> had served as the starting material for our synthetic approach to (+)-7. Unfortunately, the (+)-enantiomer of 101 is not easily accessible,<sup>38</sup> and we were confronted with the possibility that our synthesis would not be amenable to the preparation of our new target molecule, (–)-dragmacidin F ((–)-7). We reasoned, however, that it might be possible to exploit (–)-quinic acid (101) in an enantiodivergent manner that would allow access to both (+)- and (–)-7 (Scheme 3.4.1).<sup>39</sup> For such an approach to succeed, (–)-quinic acid (101) would be elaborated via selective manipulation of the C(3), C(4), and C(5) hydroxyl groups to a pseudo- $C_2$ -symmetric<sup>40</sup> derivative (125) en route to pyrrolocyclohexene 126, the diastereomer of which (i.e., 100)

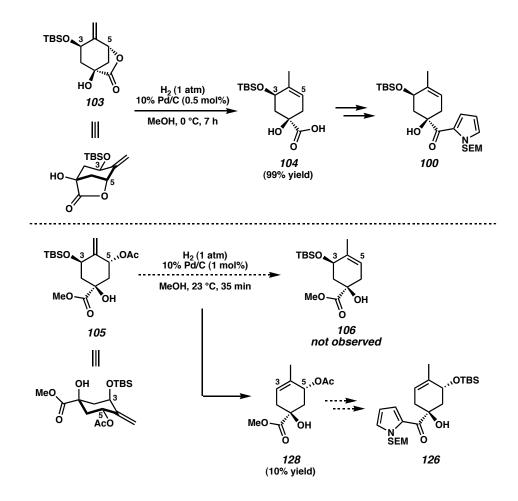
was employed in our synthesis of (+)-7. Analogous to our approach to (+)-7 (i.e., 100  $\rightarrow$  98), we anticipated that 126 could undergo oxidative carbocyclization to afford annulated pyrrole 127. Bicycle 127 would then be elaborated to (-)-dragmacidin F ((-)-7). Of the key transformations outlined in Scheme 15, we were familiar with the Pd-mediated oxidative carbocyclizations and the late-stage manipulations of related compounds; however, the successful preparation of (-)-dragmacidin F ((-)-7) would rely heavily on the identification of a suitable quinic acid derivative (125), the facile synthesis of that compound, and the rapid conversion of 125 to the requisite cyclization substrate (126).

Scheme 3.4.1



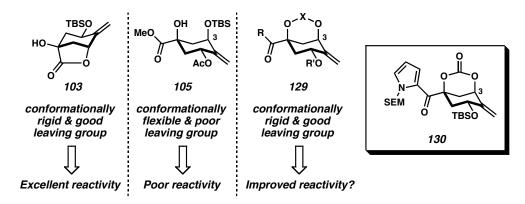
Fortunately, potential solutions to these problems had become apparent during our studies of a novel reductive isomerization reaction discovered in our synthesis of (+)dragmacidin F ((+)-7). Two critical results are shown in Scheme 3.4.2. In the first experiment, treatment of lactone 103 with Pd/C and H<sub>2</sub> in methanol at 0 °C furnished carboxylic acid 104 in essentially quantitative yield via reductive loss of the C(5)carboxylate with concomitant olefin migration (i.e., net  $S_N 2$ ' reduction). In the second experiment, a closely related derivative (105) was exposed to similar reaction conditions.<sup>41</sup> Surprisingly, the reductive isomerization reaction proceeded with loss of the C(3) silvl ether rather than the C(5) acetate, thus producing small quantities of allylic acetate 128 instead of the anticipated product (106)<sup>42</sup>. The observation that (t-Bu)Me<sub>2</sub>SiO<sup>-</sup> was preferentially ejected from compound **105** despite the clear superiority of AcO<sup>-</sup> as a leaving group led us to consider that the C(3) silvl ether moiety was positioned in an axial orientation, thereby facilitating its elimination.<sup>43</sup> This preferred conformation of 105 represents a cyclohexane ring-flip with respect to lactone 103, and thus gives rise to the reductive isomerization product (128) possessing a  $\Delta_{3,4}$  olefin. Importantly, the possibility existed that the unexpected product obtained from this reaction (i.e., 128) could be converted to cyclization substrate 126 (diastereomeric to 100).

Scheme 3.4.2

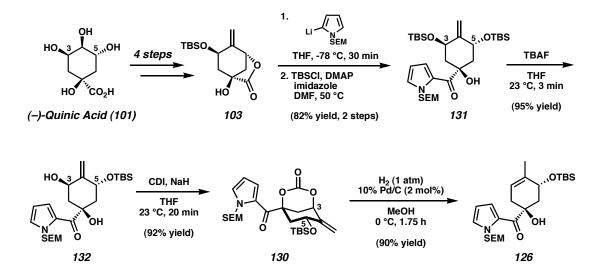


Our efforts to optimize the reductive isomerization of **105** to **128** were hampered by competitive hydrogenation of the olefin moiety of **105**, a complication not observed in the high-yielding conversion of **103** to **104**. Although both processes presumably involve the elimination of an axially disposed leaving group,<sup>43</sup> we reasoned that the successful conversion of **103** to **104** was due to the carboxylate being conformationally restricted to an axial orientation, while substrate **105** possessed a poorer leaving group  $((t-Bu)Me_2SiO^-)$  and was free to adopt alternate conformations (Figure 3.4.1).<sup>44</sup> We hypothesized that derivatives of **105** containing an axially-locked leaving group at C(3) (e.g., **129**) would be more suitable substrates for the reductive isomerization reaction. Thus, carbonate **130** was identified as the key (–)-quinic acid derived intermediate en route to the desired cyclization substrate (**126**) and became the focus of our efforts.

Figure 3.4.1

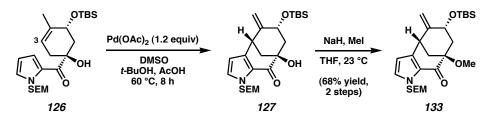


Our synthesis of carbonate **130** began with bicyclic lactone **103**, a derivative of (-)-quinic acid (**101**) that was used in our total synthesis of (+)-**7** (Scheme 3.4.3). Addition of 2-lithio-SEM-pyrrole<sup>14</sup> followed by TBS protection afforded bis(silylether) **131** in good yield. This pseudo- $C_2$ -symmetric compound then underwent rapid diastereoselective mono-desilylation upon treatment with TBAF in THF to produce the *syn* 1,3-diol **132**.<sup>45</sup> Importantly, this desymmetrization proceeded with complete selectivity and allowed us to efficiently differentiate the C(3) and C(5) positions of the cyclohexyl moiety. Diol **132** was smoothly converted to bicyclic carbonate **130** in the presence of CDI, effectively restricting the C(3) substituent to an axial disposition. Gratifyingly, exposure of carbonate **130** to our reductive isomerization conditions (2 mol% Pd/C, H<sub>2</sub>, MeOH, 0 °C) led to the selective formation of the desired cyclization substrate (**126**) in 90% yield.<sup>46</sup>



## 3.4.3 Constructing the [3.3.1] Bicycle en Route to (-)-Dragmacidin F

After assembling target substrate **126**, we turned our attention to the key Pd(II)mediated cyclization reaction (Scheme 3.4.4). Substrate **126** was treated with 1.2 equiv of Pd(OAc)<sub>2</sub> under conditions similar to those described earlier, upon which, the desired pyrrole-fused bicycle (**127**) formed as a single regio- and stereoisomer. Notably, bond formation between the pyrrole functionality and C(3) of **126** occurred even in the presence of the bulky C(5) silyl ether group positioned syn to the acyl pyrrole subunit. Following protection of the 3° alcohol, [3.3.1] bicycle **133** was obtained in 68% yield for the two-step process. Scheme 3.4.4

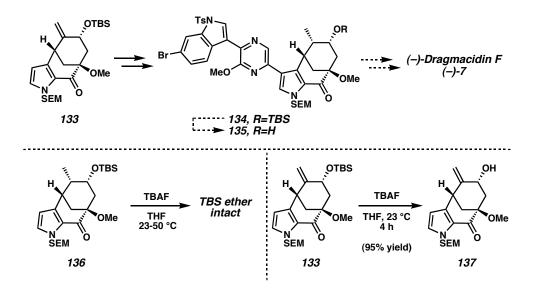


## 3.4.4 End-Game Studies

### 3.4.4.1 End-Game Strategy 1

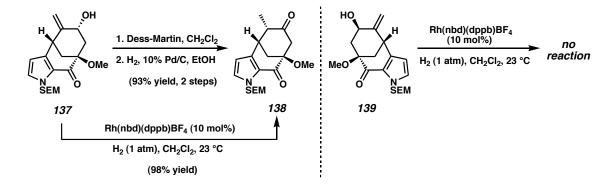
En route to (–)-dragmacidin F, cyclization product **133** was converted to pyrazine **134** (Scheme 3.4.5) by methods similar to those described above.<sup>30b</sup> Despite the similarity of **134** to its diastereomeric counterpart employed in the synthesis of (+)-7 (**112**, Scheme 3.2.8), selective desilylation of **134** to afford **135** proved to be difficult. We reasoned that the steric congestion of the axial TBS ether, positioned syn to the methyl stereocenter, was the cause of these problems. In fact, attempted TBS cleavage of parent bicycle **136** was also challenging, even at elevated temperatures.<sup>47</sup> However, in a critical reaction, the sterically less crowded TBS ether of olefinic substrate **133** underwent smooth and selective cleavage upon treatment with TBAF in THF to afford allylic alcohol **137**. With this result in hand, we conceived of a modified route that would ultimately deliver (–)-**7** in a more convergent manner.

Scheme 3.4.5

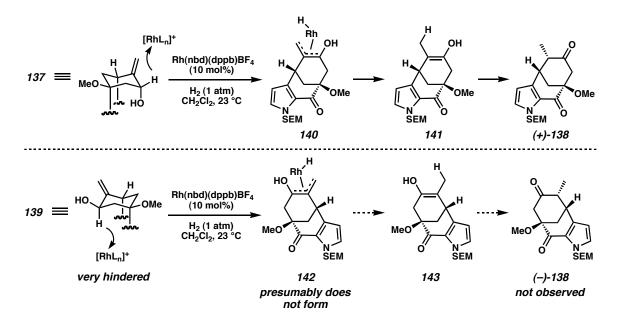


3.4.4.2 End-Game Strategy 2: Rh-Mediated Allylic Isomerization and the Total Synthesis of (–)-Dragmacidin F

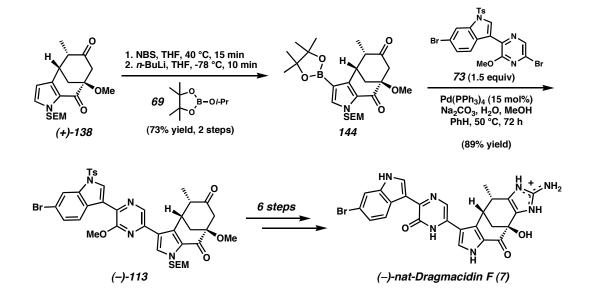
Since allylic alcohol **137** was readily accessible, we chose to employ it as an intermediate in our synthesis. Oxidation of allylic alcohol **137** followed by olefin reduction afforded ketone **138** in good overall yield (Scheme 3.4.6). However, because alcohol **137** and ketone **138** are in the same overall oxidation state, a tandem olefin isomerization/tautomerization process would be more efficient. Upon exposure of alcohol **137** to Brown's cationic rhodium catalyst Rh(nbd)(dppb)BF<sub>4</sub><sup>48</sup> and H<sub>2</sub>, ketone **138** formed directly as a single diastereomer in 98% yield. Interestingly, when diastereomer **139** (closely related to intermediates employed in the synthesis of (+)-dragmacidin F ((+)-7)) was subjected to the identical conditions, no reaction took place.

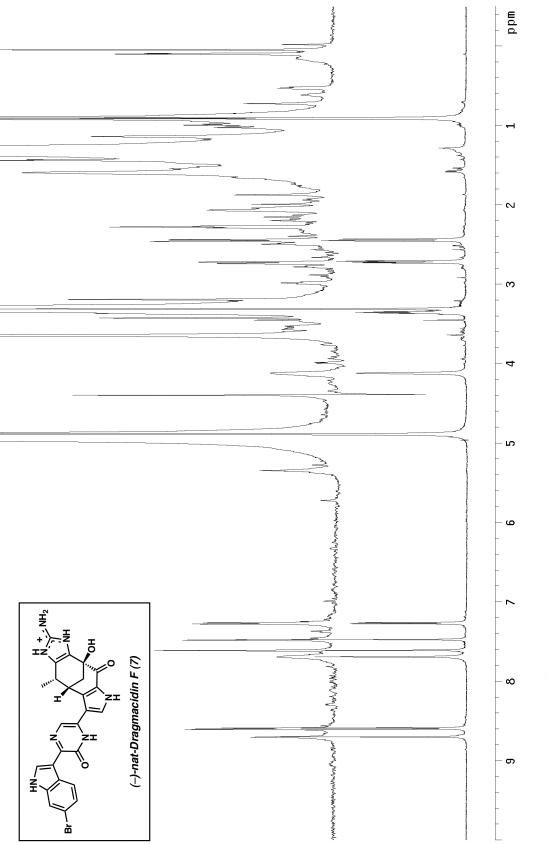


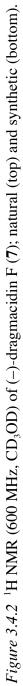
The discrepancy between the two outcomes can be rationalized after examining the mechanism of the Rh-mediated allylic isomerization reaction, which has been studied extensively.<sup>48b</sup> The process begins by interaction of the cationic rhodium complex with an allylic hydrogen atom to form a  $\pi$ -allyl rhodium complex. In the case of substrate **137**, the allylic proton necessary for isomerization is positioned on the convex face of the bicycle, pointed away from the heterocycle (Scheme 3.4.7). Thus, the  $\pi$ -allyl rhodium intermediate (**140**) can form without difficulty. **140** then undergoes reductive elimination to enol **141**, followed by tautomerization to ketone (+)-**138**. The newly formed stereocenter in **138** is presumably controlled by thermodynamics, as the methyl group rests in an equatorial position. Substrate **139**, in contrast to **137**, possesses an axially disposed allylic proton that is positioned syn to the bicycle. It is likely that this proton is severely hindered, making approach of the large cationic rhodium complex difficult. Thus, isomerization of allylic alcohol **139** to ketone (-)-**138** (via **142** and **143**) does not occur.



Elaboration of ketone (+)-138 to (-)-7 proceeded with little difficulty. Regioselective bromination and low-temperature metalation of the pyrrole in the presence of two ketones gave rise to boronic ester 144 (Scheme 3.4.8). Subsequent halogenselective cross-coupling of 144 with dibromide 73 afforded the desired Suzuki adduct (-)-113 (89% yield), the enantiomer of which had been employed in the synthesis of (+)dragmacidin F. Finally, Suzuki adduct (-)-113 was converted to (-)-dragmacidin F ((-)-7) via our previously described six-step protocol (vide supra). Synthetic and natural (-)- $7^{1b}$  were spectroscopically identical (Figure 3.4.2), including the sign of optical rotation (natural (-)-7:  $[\alpha]_{D}^{25}$  -159° (*c* 0.4, MeOH); synthetic (-)-7:  $[\alpha]_{D}^{23}$  -148° (*c* 0.2, MeOH)).<sup>30b</sup>







# 3.5 Conclusion

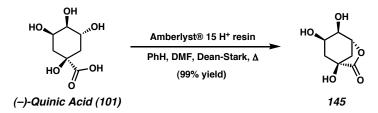
In summary, we have developed an enantiodivergent strategy to access both antipodes of dragmacidin F (7) from a single enantiomer of readily available (-)-quinic acid (101). Our highly efficient syntheses provide (+)-7 in 7.8% overall yield and (-)-7 in 9.3% overall yield beginning from 101. The routes that we have developed to (+)- and (-)-7 are concise and feature a number of key transformations, namely: a) highly efficient functionalizations of (-)-101 to differentiate C(3) and C(5), b) novel reductive isomerization reactions, sterically demanding Pd(II)-mediated oxidative c) carbocyclizations, d) halogen-selective Suzuki cross-coupling reactions, and e) highyielding late-stage Neber rearrangements. Advanced biological testing of both synthetic antipodes of dragmacidin F is currently underway.

### **3.6 Experimental Section**

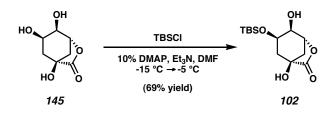
#### **3.6.1** Materials and Methods

Unless stated otherwise, reactions were conducted in flame-dried glassware under an atmosphere of nitrogen using anhydrous solvents (either freshly distilled or passed through activated alumina columns). All commercially obtained reagents were used as received. Reaction temperatures were controlled using an IKAmag temperature modulator. Thin-layer chromatography (TLC) was conducted with E. Merck silica gel 60 F254 pre-coated plates (0.25 mm) and visualized using a combination of UV, anisaldehyde, ceric ammonium molybdate, and potassium permanganate staining. ICN silica gel (particle size 0.032-0.063 mm) was used for flash column chromatography. Disposable Sep-Pak C<sub>18</sub> Vac Cartridges were purchased from Waters and used for all reversed-phase filtrations. HPLC analysis was performed on a Beckman Gold system using a Rainin C<sub>18</sub>, Microsorb MV, 5µm, 300 x 4.6 mm reversed-phased column in 0.1% (w/v) TFA with acetonitrile/H<sub>2</sub>O as eluent and a flow rate of 1.0 mL/min, gradient elution of 1.25% acetonitrile/min. Preparatory reversed-phase HPLC was performed on a Beckman HPLC with a Waters DeltaPak 25 x 100 mm, 100  $\mu m$   $C_{\rm 18}$  column equipped with a guard, 0.1% (w/v) TFA with acetonitrile/H<sub>2</sub>O as eluent, and gradient elution of 0.50% acetonitrile/min. For all reversed-phase purifications, H<sub>2</sub>O (18M $\Omega$ ) was obtained from a Millipore MiliQ water purification system and TFA from Halocarbon, Inc. <sup>1</sup>H NMR spectra were recorded on a Varian Mercury 300 (at 300 MHz), a Varian Inova 500 (at 500 MHz), or a Varian Inova 600 (at 600 MHz) and are reported relative to Me<sub>4</sub>Si (δ 0.0). Data for <sup>1</sup>H NMR spectra are reported as follows: chemical shift ( $\delta$  ppm), multiplicity, coupling constant (Hz), and integration. <sup>13</sup>C NMR spectra were recorded on

a Varian Mercury 300 (at 75 MHz), or a Varian Inova 500 (at 125 MHz) and are reported relative to Me<sub>4</sub>Si (δ 0.0). Data for <sup>13</sup>C NMR spectra are reported in terms of chemical shift. IR spectra were recorded on a Perkin Elmer Paragon 1000 spectrometer and are reported in frequency of absorption (cm<sup>-1</sup>). Optical rotations were measured with a Jasco P-1010 polarimeter. High resolution mass spectra were obtained from the California Institute of Technology Mass Spectral Facility.

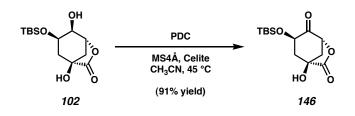


**Lactone 145.** A mixture of D-(–)-quinic acid (**101**) (50.0 g, 260.2 mmol), Amberlyst® 15 ion-exchange resin (7 g, 35 mmol), benzene (500 mL), and DMF (125 mL) was refluxed under a Dean-Stark trap for 16 h. The reaction mixture was cooled to 23 °C and filtered over a pad of Celite. The filtrate was then evaporated under reduced pressure to afford a thick oil, which was diluted with  $CH_2Cl_2$  (150 mL). Hexanes (250 mL) was added and the resulting mixture was allowed to sit at 23 °C for 2 h. The product was collected by vacuum filtration and was further dried in vacuo to afford lactone **145** (44.9 g, 99% yield) as a white powder.  $R_f$  0.40 (3:1 EtOAc:acetone); characterization data for this compound have been previously reported.<sup>7a</sup>



**TBS Lactone 102.** To a mixture of lactone **145** (90.0 g, 517 mmol), DMAP (6.31 g, 51.7 mmol), triethylamine (90 mL, 646 mmol), and DMF (345 mL) at -15 °C was added TBSC1 (84.9 g, 563 mmol) in 3 equal portions over 30 min. The temperature was maintained between -20 °C and -15 °C during the addition. The reaction mixture was

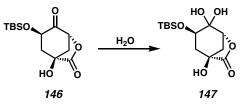
allowed to warm to -5 °C over 3 h, quenched by the addition of 5% aq. citric acid (120 mL), and then warmed to 23 °C. The solvent was removed in vacuo, and the crude product was diluted with 5% aq. citric acid (350 mL) and extracted with Et<sub>2</sub>O (1 x 500 mL, 2 x 400 mL). The combined organic layers were washed with H<sub>2</sub>O (2 x 400 mL) and brine (400 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure. The product was triturated with hexanes (750 mL) and collected by vacuum filtration. It was further dried under vacuum to afford TBS lactone **102** (102.8 g, 69% yield) as a dry white solid. R<sub>*f*</sub> 0.48 (1:1 hexanes:EtOAc); R<sub>*f*</sub> 0.28 (2:1 Et<sub>2</sub>O:hexanes); characterization data for this compound have been previously reported.<sup>7b</sup>



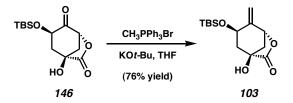
**Keto Lactone 146.** A mixture of TBS lactone **102** (3.72 g, 12.90 mmol), powdered 4Å activated molecular sieves (2.79 g), Celite (2.79 g), pyridinium dichromate (12.13 g, 32.2 mmol), and acetonitrile (185 mL) was heated to 45 °C for 24 h. The reaction was allowed to cool to 23 °C, and then was filtered over a plug of silica gel topped with Celite (EtOAc eluent). The solvent was removed under reduced pressure to afford a brown oil, which was further purified by passage over a plug of silica gel (1:1 hexanes:EtOAc). Evaporating the solvent in vacuo afforded keto lactone **146** (3.35 g, 91% yield) as a pale yellow oil.

Alternate Procedure. Powdered 4Å activated molecular sieves (184.6 g) were agitated and flame-dried under vacuum for approximately 30 min until a fine, powder-

like consistency was obtained. Upon cooling to 23 °C, CH<sub>2</sub>Cl<sub>2</sub> (540 mL) was introduced, and the slurry was cooled to 0 °C. Freshly prepared pyridinium dichromate<sup>49</sup> (148.7 g, 395.3 mmol) was added, and the resulting heterogeneous orange mixture was treated with TBS lactone 102 (70.04 g, 242.8 mmol) portionwise over 4 min. After the addition was complete, the reaction was stirred for 5 min and then freshly distilled AcOH (49.0 mL, 856.0 mmol) was added dropwise over a 20 min period. The reaction temperature was maintained at 0 °C for 15 min after the addition was complete, and the mixture was then stirred at 23 °C. After 10 h, the reaction was judged complete by <sup>1</sup>H NMR. The dark mixture was evenly divided into 3 portions, each of which was filtered over a pad of silica gel (10 cm diameter x 7.5 cm height, EtOAc eluent). The filtrates were combined and evaporated in vacuo to afford a dark liquid, and this residue was further coevaporated with toluene  $(3 \times 150 \text{ mL})$ . The crude product was diluted in a mixture of hexanes:EtOAc (10:1; 250 mL) and filtered over a pad of powdered Na<sub>2</sub>SO<sub>4</sub> to remove insoluble impurities. The filtrate was evaporated, and dried in vacuo, to afford keto lactone 146 (55.27 g, 80% yield) as a brown, waxy solid. This material was used immediately in the next step without further purification. Unstable to TLC conditions; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  4.71 (d, J = 6.6 Hz, 1H), 4.52 (dd, J = 10.3 Hz, 8.9 Hz, 1H), 2.95 (s, 1H), 2.88-2.79 (m, 1H), 2.57-2.47 (m, 1H), 2.39 (d, J = 12.4 Hz, 1H), 2.13 (dd, J = 12.4 Hz, 10.5 Hz, 1H), 0.88 (s, 9H), 0.11 (s, 3H), 0.02 (s, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  202.6, 177.4, 79.0, 72.0, 70.6, 43.2, 42.6, 25.8 (3C), 18.5, -4.6, -5.3; IR (film): 3444 (br), 2931, 2858, 1799, 1753, 1254, 1144, 1111 cm<sup>-1</sup>; HRMS-FAB (m/z):  $[M + H]^+$ calc'd for  $C_{13}H_{23}O_5Si$ , 287.1315; found, 287.1316;  $[\alpha]^{19}_{D}$  -96.47° (*c* 1.0,  $C_6H_6$ ).



*Unstable to TLC conditions*; mp 104-6 °C; <sup>1</sup>H NMR (300 MHz, CD<sub>3</sub>OD): δ 4.46 (d, J = 5.8 Hz, 1H), 3.75 (dd, J = 10.7 Hz, 7.0 Hz, 1H), 2.48-2.31 (comp. m, 2H), 2.10-2.00 (m, 1H), 1.76 (app. t, J = 11.4 Hz, 1H), 0.93 (s, 9H), 0.14 (s, 3H), 0.12 (s, 3H); <sup>13</sup>C NMR (75 MHz, CD<sub>3</sub>OD): δ 179.4, 93.2, 81.8, 73.0, 72.3, 41.5, 40.9, 26.5 (3C), 19.1, -4.3, -4.7; IR (KBr): 3440 (br), 3374 (br), 2929, 2858, 1782, 1256, 1108, 1070 cm<sup>-1</sup>; HRMS-CI (*m/z*): [M + H]<sup>+</sup> calc'd for C<sub>13</sub>H<sub>24</sub>O<sub>6</sub>Si, 304.1342; found, 304.1336; [α]<sup>19</sup><sub>D</sub> -54.29° (*c* 1.0, MeOH).

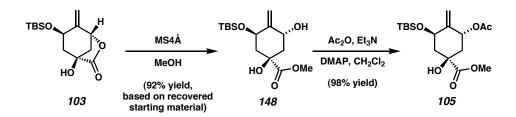


Methylene Lactone 103. To  $CH_3PPh_3Br$  (105 mg, 0.293 mmol) in THF (2.8 mL) at 0 °C was added potassium *t*-butoxide (31.3 mg, 0.279 mmol). The mixture was warmed to 23 °C and stirred for an additional 10 min. Keto lactone 146 (40 mg, 0.140 mmol) in THF (1 mL) was added and stirring was continued at 23 °C for 15 min. The reaction mixture was then refluxed for 2 h and cooled to 23 °C. The solvent was removed under reduced pressure, and the residue was partitioned between Et<sub>2</sub>O (3 mL)

and brine (1.5 mL). The layers were separated, and the aqueous layer was further extracted with  $Et_2O$  (3 x 1 mL). The combined organic layers were washed with brine (1.5 mL), dried by passage over a plug of silica gel ( $Et_2O$  eluent, then 2:1 hexanes:EtOAc eluent), and evaporated under reduced pressure. The crude product was purified by flash chromatography (2:1 hexanes:EtOAc) to afford methylene lactone **103** (30 mg, 76% yield) as a white solid.

Alternate Procedure. To CH<sub>3</sub>PPh<sub>3</sub>Br (82.9 g, 232.1 mmol) in THF (1.10 L) at 23 °C was added potassium t-butoxide (23.8 g, 212.1 mmol) in one portion. The mixture was stirred for 2 h, then cooled to 0 °C. Keto lactone **146** (54.5 g, 190.3 mmol) in THF (240 mL) was added dropwise over a 30 min period. The reaction was allowed to warm slowly to 23 °C over 9 h, then quenched by the addition of ice-cold 15% aq.  $NH_4Cl$  (500 mL). The solvent was evaporated under reduced pressure, and the residue was partitioned between Et<sub>2</sub>O (500 mL) and H<sub>2</sub>O (100 mL). The aqueous phase was extracted with Et<sub>2</sub>O (3 x 250 mL), and the combined organics were washed with H<sub>2</sub>O (100 mL) and brine (100 mL) and dried over MgSO<sub>4</sub>. Evaporation of the solvent afforded a crude yellow oil, which was filtered over a plug of silica gel (4:1 pentane:Et<sub>2</sub>O  $\rightarrow$  3:2 pentane:Et<sub>2</sub>O eluent). After evaporating the solvent in vacuo, the residue was triturated with ice-cold pentane (40 mL). The white solid was filtered and washed with ice-cold pentane (2 x 2 mL). A second crop was collected from the filtrate after concentrating its volume to 15 mL. Drying the collected material in vacuo afforded methylene lactone 103 (22.1 g, 41% yield) as a white solid.  $R_{f}$  0.59 (1:1 hexanes:EtOAc); mp 87-88 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  5.25-5.23 (m, 1H), 5.13-5.10 (m, 1H), 5.07 (d, J = 6.0 Hz, 1H), 4.38-4.29 (m, 1H), 2.85 (s, 1H), 2.67-2.59 (m, 1H), 2.31-2.21 (m, 1H), 2.09 (d, J =

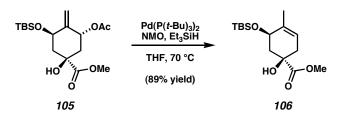
11.5 Hz, 1H), 1.86 (app. t, J = 11.3 Hz, 1H), 0.89 (s, 9H), 0.06 (s, 6H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  178.1, 144.8, 111.0, 79.4, 73.1, 67.1, 44.7, 44.7, 26.0 (3C), 18.5, -4.5, -4.7; IR (film): 3426 (br), 2956, 2931, 2858, 1791, 1254, 1120, 1071 cm<sup>-1</sup>; HRMS-FAB (*m*/*z*): [M + H]<sup>+</sup> calc'd for C<sub>14</sub>H<sub>25</sub>O<sub>4</sub>Si, 285.1522; found, 285.1519; [ $\alpha$ ]<sup>19</sup><sub>D</sub> -101.71° (*c* 1.0, CHCl<sub>3</sub>).



**Methyl Ester 105.** To lactone **103** (420 mg, 1.477 mmol) and activated ovendried 4Å molecular sieves (100 mg) was added MeOH (15 mL). The reaction mixture was stirred at 23 °C for 5.5 h, then filtered over a short plug of Celite (EtOAc eluent). After evaporation of the reaction mixture under reduced pressure, the residue was purified by flash column chromatography (2:1 hexanes:EtOAc eluent) to afford starting material lactone **103** (82 mg, 20% yield) and siloxy diol **148** (345 mg, 74% yield, 92% yield based on recovered starting material), which was used directly in the subsequent reaction.

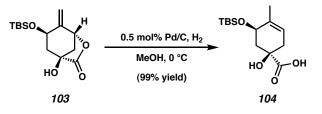
To siloxy diol **148** (80.0 mg, 0.253 mmol) in  $CH_2Cl_2$  (1.5 mL) was added  $Et_3N$  (71 µL, 0.506 mmol), DMAP (3 mg, 0.0253 mmol), followed by  $Ac_2O$  (31 µL, 0.329 mmol). The reaction mixture was stirred at 23 °C for 10 min, quenched with saturated aq. NaHCO<sub>3</sub> (5 mL), and extracted with  $CH_2Cl_2$  (3 x 15 mL). The combined organic layers were filtered over a plug of silica gel ( $CH_2Cl_2$  eluent, then EtOAc eluent) and evaporated under reduced pressure. The crude product was purified by flash

chromatography (3:1 hexanes:EtOAc eluent) to afford methyl ester **105** (89.0 mg, 98% yield) as a colorless oil.  $R_f 0.50$  (1:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  5.90-5.81 (m, 1H), 4.96 (br s, 1H), 4.94 (br s, 1H), 4.91-4.89 (m, 1H), 4.67 (app. t, J = 3.2 Hz, 1H), 3.74 (s, 3H), 2.38 (ddd, J = 12.7, 5.2, 2.2 Hz, 1H), 2.19-2.03 (comp. m, 2H), 2.09 (s, 3H), 1.93 (app. t, J = 12.1 Hz, 1H), 0.87 (s, 9H), 0.09 (s, 3H), 0.08 (s, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  173.7, 169.6, 146.3, 108.5, 76.5, 75.1, 68.0, 52.9, 42.7, 41.2, 25.8 (3C), 21.1, 18.1, -4.6, -5.2; IR (film) 3464 (br), 2954, 2932, 2858, 2888, 1739 (br), 1369, 1233 (br), 1124, 1098, 1072, 1036 cm<sup>-1</sup>; HRMS-FAB (*m/z*): [M + H]<sup>+</sup> calc'd for C<sub>17</sub>H<sub>31</sub>O<sub>6</sub>Si, 359.1890; found, 359.1900; [ $\alpha$ ]<sup>26</sup><sub>D</sub> -26.61° (*c* 1.0, C<sub>6</sub>H<sub>6</sub>).



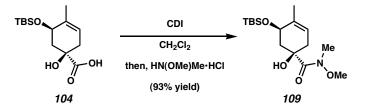
Siloxycyclohexene 106. Methyl ester 105 (94 mg, 0.262 mmol),  $Pd(P(t-Bu_3)_2)$  (40.2 mg, 0.0786 mmol), anhydrous *N*-methylmorpholine *N*-oxide (307 mg, 2.52 mmol), THF (5.2 mL), and freshly distilled Et<sub>3</sub>SiH (1.67 mL, 10.5 mmol) were combined under a glovebox atmosphere. The reaction mixture was immediately removed from the glovebox and placed in a 70 °C oil bath. After 3.5 h, the reaction mixture was cooled to 0 °C, and the volatiles were removed under reduced pressure. Saturated aq. NH<sub>4</sub>Cl (15 mL) was added, and the mixture was extracted with Et<sub>2</sub>O (3 x 25 mL). The combined organic layers were washed with brine (15 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure. The crude product was purified by flash chromatography (5:1 hexanes:EtOAc eluent) to afford siloxycyclohexene 106 (70 mg, 89% yield) as a pale

yellow oil.  $R_f 0.55$  (2:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  5.49-5.42 (m, 1H), 4.62 (s, 1H), 4.18-4.12 (m, 1H), 3.76 (s, 3H), 2.45-2.38 (comp. m, 2H), 2.16-2.10 (comp. m, 2H), 1.79-1.74 (m, 3H), 0.88 (s, 9H), 0.13 (s, 3H), 0.12 (s, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  175.3, 133.7, 120.9, 73.0, 68.7, 52.6, 38.4, 36.9, 25.9 (3C), 21.4, 18.0, -4.3, -4.7; IR (film) 3478 (br), 2955, 2858, 1740, 1451, 1253, 1217, 1111, 1065, 1037 cm<sup>-1</sup>; HRMS-FAB (*m*/*z*): [M + H]<sup>+</sup> calc'd for C<sub>15</sub>H<sub>19</sub>O<sub>4</sub>Si, 301.1835; found, 301.1835; [ $\alpha$ ]<sup>24</sup><sub>D</sub> +77.62° (*c* 0.47, CHCl<sub>3</sub>).

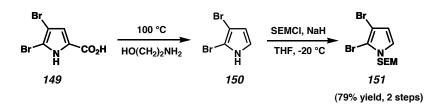


Acid 104. A mixture of methylene lactone 103 (4.0 g, 14.1 mmol) and 10% Pd/C (80 mg, 0.075 mmol) in methanol (120 mL) was cooled to 0 °C. The reaction vessel was evacuated and back-filled with H<sub>2</sub> (3x). After 7 h at 0 °C, the mixture was filtered over a pad of Celite (MeOH eluent), and the solvent was evaporated under reduced pressure to afford a colorless oil. Residual solvent was removed by holding the crude product under vacuum for 10 h, providing acid 104 (4.0 g, 99% yield), which was used immediately without further purification.  $R_f$  0.28 (1:1 hexanes:EtOAc;1% acetic acid); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  5.88 (s, 1H), 5.53-5.48 (m, 1H), 4.16-4.11 (m, 1H), 2.71-2.60 (m, 1H), 2.36-2.22 (m, 1H), 2.18 (dd, *J* = 14.3 Hz, 3.9 Hz, 1H), 2.08-2.01 (m, 1H), 1.79-1.76 (m, 3H), 0.89 (s, 9H), 0.15-0.13 (comp. m, 6H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  176.4, 133.2, 121.1, 73.6, 68.6, 37.9, 35.9, 25.8 (3C), 21.4, 18.0, -4.5, -4.7; IR (film): 3356 (br), 2956,

2931, 2858, 1768 (br), 1718 (br), 1255, 1063 cm<sup>-1</sup>; HRMS-FAB (*m/z*):  $[M + H]^+$  calc'd for C<sub>14</sub>H<sub>27</sub>O<sub>4</sub>Si, 287.1679; found, 287.1675;  $[\alpha]^{19}_{D}$  +37.58° (*c* 1.0, C<sub>6</sub>H<sub>6</sub>).



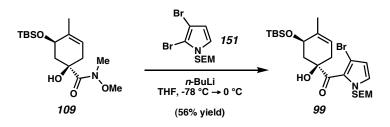
Weinreb Amide 109. To acid 104 (4.0 g, 14.1 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (70 mL) at 23 °C was added 1,1'-carbonyldiimidazole (3.65 g, 22.5 mmol) in equal portions over 15 min. After the final addition, stirring was continued for 10 min, then N,Odimethylhydroxylamine • HCl (3.43 g, 35.16 mmol) was added in one portion. The reaction was allowed to stir at 23 °C for 3 h. Et<sub>2</sub>O was added (50 mL), and the reaction mixture was filtered. The filtrate was evaporated, diluted with Et<sub>2</sub>O (125 mL), washed with 5% aq. citric acid (2 x 50 mL) and brine (50 mL), and dried over MgSO<sub>4</sub>. The crude product was purified by flash chromatography (3:1 hexanes:EtOAc) to afford Weinreb amide **109** (4.29 g, 93% yield) as a colorless oil.  $R_f 0.42$  (2:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  5.43 (m, 1H), 4.72 (s, 1H), 4.17-4.11 (m, 1H), 3.71 (s, 3H), 3.22 (s, 3H), 2.59-2.24 (comp. m, 3H), 2.03 (dd, J = 14.6 Hz, 4.1 Hz, 1H), 1.75-1.71 (m, 3H), 0.86 (s, 9H), 0.11 (s, 3H), 0.09 (s, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, 15/16 C): δ 133.5, 121.5, 74.3, 69.4, 61.2, 38.1, 35.9, 26.0, 25.9 (3C), 21.3, 18.1, -4.3, -4.7; IR (film): 3463 (br), 2956, 2932, 2858, 1655, 1362, 1254 cm<sup>-1</sup>; HRMS-EI (m/z):  $[M + H]^+$  calc'd for  $C_{16}H_{32}NO_4Si$ , 330.2101; found, 330.2085;  $[\alpha]_{D}^{19} + 41.13^{\circ}$  (*c* 1.0, CHCl<sub>3</sub>).



**Dibromopyrrole 151.** A solution of 4,5-dibromopyrrole carboxylic acid (149)<sup>50</sup> (6.05 g, 22.5 mmol) in ethanolamine (36 mL) was heated to 100 °C for 2 h, cooled to 23 °C, and poured into a mixture of  $Et_2O$  (200 mL) and 0.5 N aq. HCl (300 mL). The layers were separated, and the aqueous layer was extracted with  $Et_2O$  (2 x 250 mL). The combined organic layers were washed with brine (200 mL), dried over MgSO<sub>4</sub>, and concentrated to 100 mL. The solution was diluted with hexanes (100 mL), filtered over a plug of silica gel (2:1 hexanes: $Et_2O$ ), and concentrated to 150 mL. THF (100 mL) was added, and the solution was concentrated to 100 mL. This solvent exchange procedure was repeated 2 additional times (2 x 100 mL THF) to afford 2,3-dibromopyrrole (150) as a solution in THF, which was used immediately in the subsequent reaction.

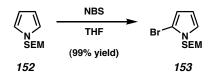
*CAUTION:* Concentrating the above described solutions to dryness or neardryness leads to rapid decomposition of 2,3-dibromopyrrole (**150**).<sup>22</sup>

To 2,3-dibromopyrrole (**150**) in THF at -20 °C was added NaH (60% dispersion in mineral oil, 1.51 g, 37.8 mmol) in 3 equal portions over 3 min. After 10 min at -20°C, SEMCl (4.8 mL, 27.1 mmol) was added dropwise over 1 min. The reaction mixture was allowed to warm to -8 °C over 40 min and was then quenched with saturated aq. NH<sub>4</sub>Cl (30 mL). After warming to 23 °C, the reaction mixture was diluted with Et<sub>2</sub>O (75 mL) and H<sub>2</sub>O (20 mL), and the layers were separated. The aqueous layer was further extracted with Et<sub>2</sub>O (2 x 50 mL). The combined organic layers were washed with brine (50 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure. The crude product was purified by flash chromatography (6:1 hexanes:CH<sub>2</sub>Cl<sub>2</sub>, then 4:1 hexanes:CH<sub>2</sub>Cl<sub>2</sub>) to afford dibromopyrrole **151** (6.25 g, 79% yield) as a yellow oil.  $R_f$  0.17 (6:1 hexanes:CH<sub>2</sub>Cl<sub>2</sub>); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  6.82 (d, *J* = 3.6 Hz, 1H), 6.25 (d, *J* = 3.3 Hz, 1H), 5.21 (s, 2H), 3.48 (t, *J* = 8.1 Hz, 2H), 0.88 (t, *J* = 8.1 Hz, 2H), -0.03 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  123.1, 112.3, 103.7, 99.8, 77.8, 66.2, 17.9, -1.2 (3C); IR (film): 2953, 2896, 1514, 1470, 1279, 1250, 1109, 1084 cm<sup>-1</sup>; HRMS-EI (*m*/*z*): [M + H]<sup>+</sup> calc'd for C<sub>10</sub>H<sub>17</sub>NOSiBr<sub>2</sub>, 352.9446; found, 352.9435.



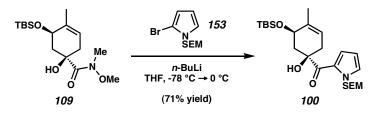
**Bromo Acyl Pyrrole 99.** To dibromopyrrole **151** (6.02 g, 17.06 mmol) in THF (114 mL) at -78 °C was added *n*-BuLi (2.5 M in hexanes, 6.7 mL, 16.8 mmol) dropwise over 1 min. After 10 min at -78 °C, Weinreb amide **109** (1.58 g, 4.80 mmol) in THF (15 mL) was added dropwise over 30 seconds. The reaction vessel was immediately warmed to 0 °C, stirred for 90 min, and cooled to -78 °C. The reaction was quenched with saturated aq. NH<sub>4</sub>Cl (15 mL), then warmed to 23 °C. The volatiles were removed in vacuo, and the residue was partitioned between Et<sub>2</sub>O (75 mL) and H<sub>2</sub>O (30 mL). The layers were separated, and the aqueous layer was further extracted with Et<sub>2</sub>O (2 x 50 mL). The combined organic layers were washed with brine (50 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure. The crude product was purified by flash chromatography (11:9 CH<sub>2</sub>Cl<sub>2</sub>:hexanes) to afford bromo acyl pyrrole **99** (1.47 g, 56%)

yield) as a colorless oil. R<sub>f</sub> 0.29 (11:9 hexanes:CH<sub>2</sub>Cl<sub>2</sub>); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 6.77 (d, J = 2.9 Hz, 1H), 6.20 (d, J = 2.7 Hz, 1H), 5.53-5.47 (m, 1H), 5.35 (d, J = 10.4 Hz, 1H), 5.29 (d, J = 10.4 Hz, 1H), 4.72 (s, 1H), 4.18-4.14 (m, 1H), 3.31 (t, J = 8.2 Hz, 2H), 2.65-2.53 (m, 1H), 2.53-2.41 (m, 1H), 2.32 (dt, J = 14.3 Hz, 1.7 Hz, 1H), 2.15 (dd, J = 14.2 Hz, 4 Hz, 1H), 1.79-1.76 (m, 3H), 0.87 (s, 9H), 0.81 (t, J = 8.2 Hz, 2H), 0.12 (s, 6H), -0.06 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 201.9, 133.2, 129.6, 125.0, 121.6, 112.5, 101.8, 78.9, 78.6, 68.9, 66.2, 38.6, 37.4, 26.0 (3C), 21.5, 18.1, 17.8, -1.2 (3C), -4.1, -4.7; IR (film): 3477 (br), 2953, 1664 (br), 1400, 1253, 1101 cm<sup>-1</sup>; HRMS-EI (*m*/*z*): [M + H]<sup>+</sup> calc'd for C<sub>24</sub>H<sub>43</sub>NO<sub>4</sub>Si<sub>2</sub>Br, 544.1914; found, 544.1903; [α]<sup>19</sup><sub>D</sub> +1.64° (*c* 1.0, CHCl<sub>4</sub>).



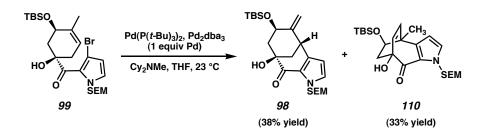
**Bromopyrrole 153.** To SEM pyrrole **152**<sup>14</sup> (1.25 g, 6.33 mmol) in THF (125 mL) at 23 °C was added freshly recrystallized NBS (1.127 g, 6.33 mmol) in one portion. After stirring for 5 min, additional NBS was added (15 mg, 0.084 mmol), and the reaction was immediately judged complete by TLC. The reaction mixture was poured into saturated aq. NaHCO<sub>3</sub> (100 mL) and extracted with Et<sub>2</sub>O (1 x 100 mL, 2 x 50 mL). The combined organic layers were washed with brine (75 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure. The crude product was purified by passage over a plug of silica gel (CH<sub>2</sub>Cl<sub>2</sub> eluent) to afford bromopyrrole **153** (1.73 g, 99% yield) as a pale yellow oil.  $R_f 0.53$  (1:1 CH<sub>2</sub>Cl<sub>2</sub>:hexanes); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  6.83 (app. t, *J* = 2.5 Hz, 1H), 6.18-6.16 (comp. m, 2H), 5.22 (s, 2H), 3.53-3.46 (m, 2H), 0.92-0.85

(m, 2H), -0.03 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 122.9, 111.9, 110.1, 102.0, 76.7, 66.0, 17.9, -1.2 (3C); IR (film): 2953, 2895, 1264, 1249, 1108, 1085 cm<sup>-1</sup>; HRMS-EI (*m/z*): [M + H]<sup>+</sup> calc'd for C<sub>10</sub>H<sub>18</sub>NOSiBr, 275.0341; found, 275.0331.



Acyl Pyrrole 100. To bromopyrrole 153 (1.73 g, 6.26 mmol) in THF (42 mL) at -78 °C was added *n*-BuLi (2.25 M in hexanes, 2.7 mL, 6.16 mmol) dropwise over 1 min. After 10 min at -78 °C, Weinreb amide **109** (655 mg, 1.99 mmol) in THF (5 mL) was added dropwise over 1 min. The reaction vessel was immediately warmed to 0 °C, stirred for 25 min, and cooled to -78 °C. The reaction mixture was quenched with saturated aq. NH<sub>4</sub>Cl (10 mL), then warmed to 23 °C. The volatiles were removed under reduced pressure. The residue was partitioned between Et<sub>2</sub>O (75 mL) and H<sub>2</sub>O (50 mL), and the layers were separated. The aqueous layer was further extracted with  $Et_2O$  (2 x 40 mL). The combined organic layers were washed with brine (50 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure. The crude product was purified by flash chromatography (23:1 hexanes:EtOAc, then 15:1 hexanes:EtOAc) to afford acyl pyrrole **100** (656 mg, 71% yield) as a colorless oil.  $R_f 0.30$  (9:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 7.66 (dd, *J* = 4.0 Hz, 1.7 Hz, 1H), 7.06 (dd, *J* = 2.5 Hz, 1.7 Hz, 1H), 6.19 (dd, J = 4.0 Hz, 2.5 Hz, 1H), 5.71 (d, J = 10.4 Hz, 1H), 5.67 (d, J = 10.0 Hz, 1H), 5.525.47 (m, 1H), 4.90 (s, 1H), 4.19 (app. t, J = 3.1 Hz, 1H), 3.51 (t, J = 8.3 Hz, 2H), 2.52-2.46 (comp. m, 2H), 2.19-2.16 (comp. m, 2H), 1.80-1.78 (m, 3H), 0.92-0.88 (comp. m,

11H), 0.13 (s, 6H), -0.06 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, 22/24 C):  $\delta$  193.7, 133.5, 129.9, 128.0, 123.8, 121.7, 109.0, 78.2, 69.4, 66.3, 38.6, 38.3, 26.0 (3C), 21.5, 18.1, -1.2 (3C), -4.2, -4.7; IR (film): 3476, 2954, 2931, 2859, 1639, 1412, 1310, 1251, 1085 cm<sup>-1</sup>; HRMS-EI (*m*/*z*): [M + H]<sup>+</sup> calc'd for C<sub>24</sub>H<sub>44</sub>NO<sub>4</sub>Si<sub>2</sub>, 466.2809; found, 466.2822; [ $\alpha$ ]<sup>19</sup><sub>D</sub> +34.25° (*c* 1.0, C<sub>6</sub>H<sub>6</sub>).



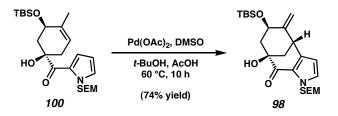
[3.3.1] Bicycle 98. Bromo acyl pyrrole 99 (52.0 mg, 0.0955 mmol),  $Pd_2dba_3$  (21.9 mg, 0.0239 mmol),  $Pd(P(t-Bu)_3)_2$  (24.4 mg, 0.0477 mmol), THF (1.2 mL), and  $Cy_2NMe$  (24.3 µL, 0.115 mmol) were combined under a glovebox atmosphere and stirred at 23 °C for 10 h. The reaction vessel was removed from the glovebox, diluted with 3:1 hexanes:EtOAc (2 mL), and filtered over a plug of silica gel topped with Celite (3:1 hexanes:EtOAc eluent). The solvent was removed under reduced pressure, and the residue was purified by flash chromatography (CH<sub>2</sub>Cl<sub>2</sub>, then 3:1 hexanes:EtOAc). The crude product was further purified by flash chromatography (6:1 hexanes:EtOAc) to afford [3.3.1] bicycle 98 (16.7 mg, 38% yield) and [3.2.2] bicycle 110 (14.4 mg, 33% yield), both as pale yellow oils.

[3.3.1] Bicycle 98: R<sub>f</sub> 0.20 (4:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 7.07 (d, J = 2.7 Hz, 1H), 6.05 (d, J = 2.7 Hz, 1H), 5.71 (d, J = 9.9 Hz, 1H), 5.58 (d, J = 9.9 Hz, 1H), 5.09-5.05 (m, 2H), 4.00 (s, 1H), 3.99-3.90 (m, 1H), 3.84 (app. t, J = 3.0 Hz, 1H),

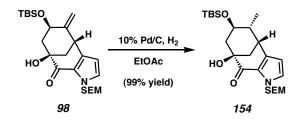
3.55-3.47 (m, 2H), 2.39 (app. dt, J = 7.4 Hz, 3.8 Hz, 1H), 2.13-2.03 (comp. m, 2H), 1.73 (app. t, J = 11.8 Hz, 1H), 0.98-0.76 (comp. m, 11H), -0.04 (s, 9H), -0.11 (s, 6H); <sup>1</sup>H NMR (300 MHz, C<sub>6</sub>D<sub>6</sub>):  $\delta$  6.53 (d, J = 2.5 Hz, 1H), 5.77 (d, J = 2.8 Hz, 1H), 5.55 (d, J = 10.2 Hz, 1H), 5.32 (app. t, J = 1.9 Hz, 1H), 5.26 (d, J = 10.2 Hz, 1H), 5.01-4.97 (m, 1H), 4.29 (s, 1H), 4.27-4.19 (m, 1H), 3.59-3.47 (comp. m, 3H), 2.45-2.31 (comp. m, 2H), 2.16 (dd, J = 12.1 Hz, 3.0 Hz, 1H), 2.07 (app. t, J = 11.8 Hz, 1H), 0.92-0.89 (comp. m, 11H), 0.01 (s, 9H), -0.06 (s, 3H), -0.07 (s, 3H); <sup>13</sup>C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>):  $\delta$  191.5, 149.4, 141.8, 132.0, 125.5, 108.5, 107.4, 76.8, 75.8, 68.4, 66.3, 48.9, 45.5, 40.7, 26.3 (3C), 18.8, 18.2, -0.8 (3C), -4.4, -4.7; IR (film): 3480, 2953, 2858, 1651, 1420, 1318, 1251, 1100, 1077 cm<sup>-1</sup>; HRMS-FAB (*m*/*z*): [M]<sup>+</sup> calc'd for C<sub>24</sub>H<sub>41</sub>NO<sub>4</sub>Si<sub>2</sub>, 463.2574; found, 463.2577; [ $\alpha$ ]<sup>23</sup><sub>D</sub> -275.07° (*c* 1.0, CHCl<sub>3</sub>).

[3.2.2] Bicycle 110:  $R_f 0.42$  (5:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  6.98 (d, J = 2.7 Hz, 1H), 6.15 (d, J = 2.7 Hz, 1H), 6.02 (d, J = 9.3 Hz, 1H), 5.98 (d, J = 8.8 Hz, 1H), 5.69 (d, J = 9.9 Hz, 1H), 5.62 (d, J = 9.9 Hz, 1H), 4.93 (s, 1H), 3.81 (d, J = 7.7 Hz, 1H), 3.50 (t, J = 8.0 Hz, 2H), 2.36 (dd, J = 14.3 Hz, 7.7 Hz, 1H), 1.94 (dd, J = 14.3 Hz, 1.6 Hz, 1H), 1.55 (s, 3H), 0.91-0.83 (comp. m, 11H), 0.02 (s, 3H), 0.01 (s, 3H), -0.07 (s, 9H); <sup>1</sup>H NMR (300 MHz,  $C_6D_6$ ):  $\delta$  6.55 (d, J = 2.7 Hz, 1H), 6.23 (d, J = 8.8 Hz, 1H), 5.96 (d, J = 3.3 Hz, 1H), 5.94 (d, J = 9.2 Hz, 1H), 5.59 (d, J = 10.4 Hz, 1H), 5.40 (d, J = 9.9 Hz, 1H), 5.32 (s, 1H), 3.82-3.75 (m, 1H), 3.46 (t, J = 7.7 Hz, 2H), 2.46 (dd, J = 13.7 Hz, 7.7 Hz, 1H), 2.25 (dd, J = 13.7 Hz, 1G Hz, 1H), 1.52 (s, 3H), 0.92 (s, 9H), 0.82 (t, J = 8.0 Hz, 2H), -0.03 (s, 3H), -0.08 (s, 3H), -0.09 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  188.7, 144.1, 139.4, 134.5, 129.1, 121.8, 107.7, 78.2, 77.8, 73.3, 66.4, 45.7, 45.0, 26.0 (3C),

22.2, 18.2, 18.0, -1.25 (3C), -4.1, -4.6; IR (film): 3432, 2955, 2858, 1645, 1250, 1081 cm<sup>-1</sup>; HRMS-EI (*m/z*):  $[M + H]^+$  calc'd for C<sub>24</sub>H<sub>42</sub>NO<sub>4</sub>Si<sub>2</sub>, 464.2652; found, 464.2665;  $[\alpha]^{19}_{D}$  +19.22° (*c* 1.0, C<sub>6</sub>H<sub>6</sub>).



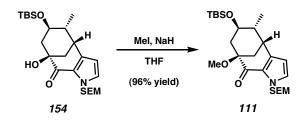
Alternate Procedure. To acyl pyrrole **100** (106.0 mg, 0.227 mmol) was added  $Pd(OAc)_2$  (51.1 mg, 0.227 mmol), DMSO (32.3 µL, 0.455 mmol), *t*-BuOH (18.2 mL), and AcOH (4.5 mL). The mixture was heated to 60 °C for 10 h, cooled to 23 °C, and filtered over a plug of silica gel (3:1 hexanes:EtOAc). The solvent was evaporated, and the residue was again filtered over a plug of silica gel (3:1 hexanes:EtOAc). After removal of solvent in vacuo, the product was purified by flash chromatography on silica gel (6:1 hexanes:EtOAc) to afford [3.3.1] bicycle **98** (78.4 mg, 74% yield) as a pale yellow oil.



**Reduced [3.3.1] Bicycle 154.** [3.3.1] bicycle **98** (360 mg, 0.78 mmol), 10% Pd/C (130 mg, 0.12 mmol), and EtOAc (8 mL) were combined, and the reaction vessel was evacuated and back-filled with  $H_2$  (1 atm). The reaction mixture was stirred under  $H_2$  for 30 min, then filtered over a plug of silica gel topped with Celite (EtOAc eluent) to afford

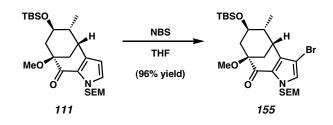
reduced [3.3.1] bicycle **154** as a colorless oil (358 mg, 99% yield).  $R_f$  0.28 (5:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz,  $C_6D_6$ ):  $\delta$  6.55 (d, J = 2.5 Hz, 1H), 5.74 (d, J = 2.5 Hz, 1H), 5.56 (d, J = 10.2 Hz, 1H), 5.30 (d, J = 10.2 Hz, 1H), 4.27 (s, 1H), 3.59-3.45 (m, 2H), 3.19 (ddd, J = 12.9 Hz, 7.7 Hz, 3.3 Hz, 1H), 2.58 (dd, J = 6.5 Hz, 3.2 Hz, 1H), 2.37-2.20 (comp. m, 2H), 2.06-1.90 (comp. m, 2H), 1.63-1.50 (m, 1H), 1.00 (d, J = 6.6 Hz, 3H), 0.94-0.89 (comp. m, 11H), -0.02 (s, 9H), -0.06 (s, 3H), -0.09 (s, 3H); <sup>13</sup>C NMR (75 MHz,  $C_6D_6$ ):  $\delta$  190.8, 140.4, 131.3, 125.2, 110.1, 76.6, 75.6, 71.8, 66.1, 46.8, 44.3, 40.0, 37.3, 25.9 (3C), 18.1, 17.9, 16.5, -1.2 (3C), -4.0, -4.6; IR (film): 3473 (br), 2953, 2931, 2857, 1651, 1420, 1249, 1079 cm<sup>-1</sup>; HRMS-EI (m/z): [M + H]<sup>+</sup> calc'd for  $C_{24}H_{44}NO_4Si_2$ , 466.2809; found, 466.2804; [ $\alpha$ ]<sup>19</sup><sub>D</sub> -166.30° (*c* 1.0,  $C_6H_6$ ).

NOTE: In some instances, trace phosphine contaminants from the Heck reaction (i.e.,  $99 \rightarrow 98$ ) prevented the reduction from occurring. Simply working up the reaction and re-exposing it to the identical reaction conditions (as described above) allowed the reduction to proceed.



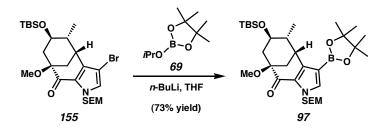
Methyl Ether 111. To reduced [3.3.1] bicycle 154 (358 mg, 0.77 mmol) in THF (7.7 mL) at 23 °C was added NaH (60% dispersion in mineral oil, 123 mg, 3.08 mmol). After stirring for 2 min at 23 °C, MeI was added (335  $\mu$ L, 5.38 mmol). The resulting mixture was stirred for 1 h, cooled to 0 °C, and quenched with saturated aq. NH<sub>4</sub>Cl (4 mL), then warmed to 23 °C. Et<sub>2</sub>O (10 mL) and H<sub>2</sub>O (5 mL) were added, and the layers

were separated. The aqueous layer was further extracted with Et<sub>2</sub>O (2 x 15 mL). The combined organic layers were washed with brine (20 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure. The crude product was purified by flash chromatography (4:1 hexanes:EtOAc) to afford methyl ether **111** (354 mg, 96% yield) as a colorless oil. R<sub>*f*</sub> 0.34 (5:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, C<sub>6</sub>D<sub>6</sub>): δ 6.58 (d, *J* = 2.8 Hz, 1H), 5.78 (d, *J* = 2.5 Hz, 1H), 5.57 (d, *J* = 10.2 Hz, 1H), 5.54 (d, *J* = 10.2 Hz, 1H), 3.65-3.50 (m, 2H), 3.37 (s, 3H), 3.22 (ddd, *J* = 12.9 Hz, 7.9 Hz, 3.1 Hz, 1H), 2.68 (dd, *J* = 6.5 Hz, 3.2 Hz, 1H), 2.59-2.49 (comp. m, 2H), 1.86 (dd, *J* = 12.4 Hz, 11.3 Hz, 1H), 1.72-1.56 (m, 2H), 1.04 (d, *J* = 6.9 Hz, 3H), 0.93-0.85 (comp. m, 11H), -0.02 (s, 9H), -0.07 (s, 3H), -0.10 (s, 3H); <sup>13</sup>C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>, 24/25 C): δ 189.4, 138.3, 130.4, 109.7, 81.9, 76.9, 72.4, 66.2, 51.8, 45.9, 41.3, 41.2, 37.6, 26.4 (3C), 18.5, 18.3, 17.0, -0.9 (3C), -3.6, -4.4; IR (film): 2954, 1657, 1421, 1250, 1085 cm<sup>-1</sup>; HRMS-EI (*m*/*z*): [M + H]<sup>+</sup> calc'd for C<sub>25</sub>H<sub>46</sub>NO<sub>4</sub>Si<sub>2</sub>, 480.2965; found, 480.2970; [α]<sup>19</sup><sub>D</sub> -172.9° (*c* 1.0, C<sub>6</sub>H<sub>6</sub>).



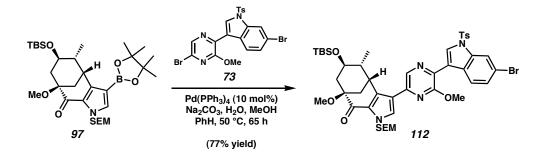
**Bromide 155.** To methyl ether **111** (305 mg, 0.64 mmol) in THF (6 mL) at 0 °C was added freshly recrystallized NBS (147 mg, 0.83 mmol). After stirring for 10 min at 0 °C, the reaction mixture was warmed to 23 °C, and additional NBS (30 mg, 0.17 mmol) was added. After 5 min, the reaction was quenched with saturated aq.  $Na_2S_2O_3$ , diluted with H<sub>2</sub>O (15 mL), and extracted with Et<sub>2</sub>O (3 x 15 mL). The combined organic layers

were washed with brine (15 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure. The crude product was purified by flash chromatography (5:1 hexanes:EtOAc) to afford bromide **155** (340 mg, 96% yield) as a colorless oil. R<sub>f</sub> 0.55 (3:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, C<sub>6</sub>D<sub>6</sub>):  $\delta$  6.57 (s, 1H), 5.46 (d, *J* = 10.2 Hz, 1H), 5.34 (d, *J* = 10.2 Hz, 1H), 3.57-3.41 (m, 2H), 3.32-3.20 (m, 4H), 2.88 (dd, *J* = 6.5 Hz, 3.2 Hz, 1H), 2.46 (ddd, *J* = 12.2 Hz, 5.1 Hz, 2.5 Hz, 1H), 2.28 (app. dt, *J* = 7.4 Hz, 4.0 Hz, 1H), 1.78 (app. t, *J* = 11.8 Hz, 1H), 1.69-1.57 (m, 1H), 1.52 (dd, *J* = 11.8 Hz, 3.0 Hz, 1H), 1.19 (d, *J* = 6.9 Hz, 3H), 0.91-0.80 (comp. m, 11H), -0.05 (s, 9H), -0.09 (s, 3H), -0.12 (s, 3H); <sup>13</sup>C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>):  $\delta$  189.6, 147.2, 137.2, 130.1, 98.4, 81.8, 77.0, 72.1, 66.6, 51.8, 45.8, 42.4, 41.0, 35.9, 26.3 (3C), 18.5, 18.3, 17.8, -0.9 (3C), -3.7, -4.3; IR (film): 2954, 2930, 1664, 1249, 1089 cm<sup>-1</sup>; HRMS-EI (*m*/z): [M + H]<sup>+</sup> - H<sub>2</sub> calc'd for C<sub>25</sub>H<sub>43</sub>NO<sub>4</sub>Si<sub>2</sub>Br, 556.1914; found, 556.1928; [ $\alpha$ ]<sup>19</sup><sub>D</sub> -98.22° (*c* 1.0, C<sub>6</sub>H<sub>6</sub>).



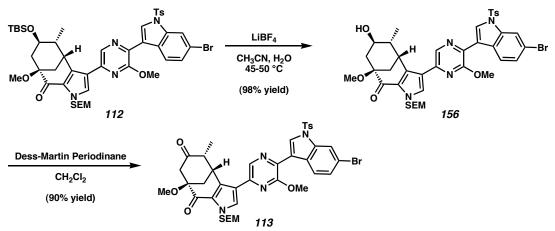
**Boronic Ester 97.** To bromide **155** (116 mg, 0.21 mmol) and 2-isopropoxy-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (**69**) (847  $\mu$ L, 4.15 mmol) in THF (10.4 mL) at -78 °C was added *n*BuLi (2.3 M in hexanes, 1.35 mL, 3.11 mmol) dropwise over 2 min. After stirring for 15 min at -78 °C, the reaction mixture was quenched with saturated aq. NH<sub>4</sub>Cl, warmed to 23 °C, and diluted with H<sub>2</sub>O (10 mL). The mixture was extracted with Et<sub>2</sub>O (3 x 15 mL). The combined organic layers were washed with brine (15 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure. The crude product was purified by

flash chromatography (4:1 hexanes:EtOAc with 0.5% Et<sub>3</sub>N) to afford boronic ester **97** (92 mg, 73% yield) as a white powder, which was used immediately in the next step.  $R_f$  0.50 (3:1 hexanes:EtOAc); mp 143-145 °C; <sup>1</sup>H NMR (300 MHz, C<sub>6</sub>D<sub>6</sub>):  $\delta$  7.42 (s, 1H), 5.55 (d, *J* = 10.1 Hz, 1H), 5.51 (d, *J* = 9.8 Hz, 1H), 3.74-3.68 (m, 1H), 3.60-3.50 (m, 2H), 3.43-3.36 (m, 1H), 3.33 (s, 3H), 2.65-2.53 (comp. m, 2H), 1.91 (app. t, *J* = 11.8 Hz, 1H), 1.89-1.80 (m, 1H), 1.68 (dd, *J* = 11.8 Hz, 2.8 Hz, 1H), 1.34 (d, *J* = 6.6 Hz, 3H), 1.15 (s, 6H), 1.14 (s, 6H), 0.94-0.81 (comp. m, 11H), -0.04 (s, 3H), -0.05 (s, 9H), -0.07 (s, 3H); <sup>13</sup>C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>, 30/31 C):  $\delta$  190.1, 145.1, 139.3, 130.2, 83.5 (2C), 82.0, 77.2, 72.6, 66.5, 51.7, 46.1, 42.0, 41.6, 36.8, 26.4 (3C), 25.4 (2C), 25.2 (2C), 18.5, 18.3, 16.9, -0.9 (3C), -3.6, -4.3; IR (film): 2953, 2931, 2858, 1658, 1543, 1249, 1141, 1085 cm<sup>-1</sup>; HRMS-FAB (*m/z*): [M + H]<sup>+</sup> calc'd for C<sub>31</sub>H<sub>57</sub>BNO<sub>6</sub>Si<sub>2</sub>, 606.3818; found, 606.3805; [ $\alpha$ ]<sup>19</sup><sub>D</sub> -98.84° (*c* 1.0, C<sub>6</sub>H<sub>6</sub>).



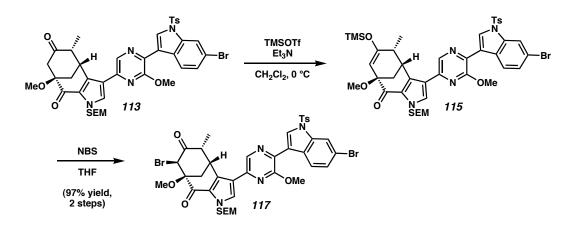
Suzuki Adduct 112. Bromopyrazine 73 (46.5 mg, 0.087 mmol), boronic ester 97 (35 mg, 0.058 mmol), benzene (1.15 mL), methanol (231  $\mu$ L), 2 M aq. Na<sub>2</sub>CO<sub>3</sub> (96  $\mu$ L), and tetrakis(triphenylphosphine)palladium(0) (6.7 mg, 0.0058 mmol) were combined and deoxygenated by sparging with argon for 5 min. The reaction vessel was evacuated, purged with N<sub>2</sub>, sealed, heated to 50 °C for 65 h, cooled to 23 °C, then quenched by the addition of Na<sub>2</sub>SO<sub>4</sub> (200 mg). Following filtration over a pad of silica gel (2:1

hexanes:EtOAc eluent) and evaporation to dryness under reduced pressure, the remaining residue was purified by flash chromatography (3:1 hexanes:EtOAc) to afford Suzuki adduct **112** (41.5 mg, 77% yield) as a yellow oil.  $R_f 0.43$  (2:1 hexanes:EtOAc); <sup>1</sup>H NMR  $(300 \text{ MHz}, \text{CDCl}_3)$ :  $\delta$  8.61 (d, J = 8.5 Hz, 1H), 8.45 (s, 1H), 8.44 (s, 1H), 8.16 (d, J = 1.5 Hz, 1H), 7.80 (d, J = 8.5 Hz, 2H), 7.59 (s, 1H), 7.40 (dd, J = 8.5 Hz, 1.8 Hz, 1H), 7.23 (d, J = 7.9 Hz, 2H), 5.85 (d, J = 10.0 Hz, 1H), 5.78 (d, J = 10.0 Hz, 1H), 4.27-4.21 (m, 1H), 4.19 (s, 3H), 3.72-3.59 (m, 2H), 3.34 (s, 3H), 3.13-3.02 (m, 1H), 2.87-2.77 (m, 1H), 2.32 (s, 3H), 2.22-2.12 (m, 1H), 1.98-1.89 (m, 1H), 1.82-1.72 (m, 1H), 1.67 (app. t, J = 11.7 Hz, 1H), 1.04-0.83 (m, 2H), 0.78 (s, 9H), 0.72 (d, *J* = 6.7 Hz, 3H), -0.02 (s, 9H), -0.09 (s, 3H), -0.16 (s, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub> 44/45 C): δ 190.0, 156.2, 145.7, 143.6, 136.9, 135.7, 135.5, 135.0, 132.7, 130.3 (2C), 130.2, 129.3, 128.8, 128.5, 127.3, 127.1 (2C), 125.3, 120.5, 119.0, 116.9, 116.4, 81.3, 77.2, 71.4, 66.7, 54.3, 51.6, 44.8, 41.8, 40.2, 34.8, 25.9 (3C), 21.8, 18.1, 16.1, -1.1 (3C), -4.0, -4.7; IR (film): 2952, 1660, 1555, 1372, 1372, 1190, 1140, 1089 cm<sup>-1</sup>; HRMS-FAB (m/z): [M]<sup>+</sup> calc'd for  $C_{45}H_{59}N_4O_7Si_2SBr$ , 934.2826; found, 934.2829;  $[\alpha]^{21}_{D}$  +51.73° (*c* 1.0, CHCl<sub>3</sub>).



Ketone 113. Suzuki adduct 112 (113 mg, 0.121 mmol), LiBF<sub>4</sub> (113 mg, 1.21 mmol), acetonitrile (6 mL), and water (600  $\mu$ L) were heated to 45–50 °C. After 9 h, additional LiBF<sub>4</sub> (30 mg, 0.32 mmol) was introduced, and heating was continued. After 6 h, additional LiBF<sub>4</sub> (35 mg, 0.32 mmol) was introduced, and heating was continued for 16 h. The reaction mixture was cooled to 23 °C, quenched with 10% aq. citric acid (10 mL), and extracted with EtOAc (3 x 20 mL). The combined organic layers were dried over MgSO<sub>4</sub> and evaporated under reduced pressure. The crude product was purified by flash chromatography (3:1 EtOAc:hexanes) to yield alcohol **156** (96.9 mg, 98% yield) as a yellow oil, which was used in the subsequent step without further purification.  $R_f = 0.44$  (3:1 EtOAc:hexanes).

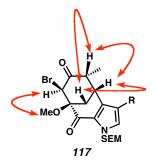
To alcohol **156** (96 mg, 0.117 mmol) in  $CH_2Cl_2$  (2.0 mL) at 23 °C was added Dess-Martin Periodinane (74.3 mg, 0.175 mmol). The mixture was stirred for 3 min, quenched with a solution of saturated aq. NaHCO<sub>3</sub> and saturated aq. Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> (1:1, 5 mL), stirred for 5 min, and extracted with EtOAc (3 x 15 mL). The combined organic layers were washed with brine (15 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure. The crude product was purified by flash chromatography (1:1 hexanes:EtOAc) to yield ketone **113** (86 mg, 90% yield) as a yellow foam.  $R_f = 0.48$  (1:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 8.61 (d, J = 8.5 Hz, 1H), 8.45 (s, 1H), 8.42 (s, 1H), 8.18 (d, J = 1.7 Hz, 1H), 7.81 (d, J = 8.5 Hz, 2H), 7.56 (s, 1H), 7.42 (dd, J =8.7 Hz, 1.8 Hz, 1H), 7.25 (d, J = 7.7 Hz, 2H), 5.77 (d, J = 10.5 Hz, 1H), 5.72 (d, J = 10.2Hz, 1H), 4.62-4.56 (m, 1H), 4.20 (s, 3H), 3.57 (app. dt, J = 8.2 Hz, 1.8 Hz, 2H), 3.43 (s, 3H), 3.14-3.06 (m, 1H), 2.91-2.81 (m, 1H), 2.74 (s, 2H), 2.40 (dd, J = 12.5 Hz, 2.9 Hz, 1H), 2.34 (s, 3H), 0.96-0.88 (m, 2H), 0.78 (d, J = 6.6 Hz, 3H), -0.02 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>, 37/39 C): δ 207.2, 188.0, 156.1, 145.7, 143.2, 136.3, 135.7, 134.9, 132.6, 130.7, 130.3 (2C), 128.8, 128.4, 127.3, 127.1 (2C), 125.4, 120.5, 119.0, 116.8, 116.3, 82.4, 77.1, 66.9, 54.3, 52.2, 52.0, 49.2, 40.2, 35.2, 21.8, 18.1, 12.2, -1.2 (3C); IR (film): 2950, 1716, 1664, 1557, 1373, 1190, 1178, 1090 cm<sup>-1</sup>; HRMS-FAB (*m*/*z*): [M]<sup>+</sup> calc'd for C<sub>39</sub>H<sub>43</sub>N<sub>4</sub>O<sub>7</sub>SiSBr, 818.1805; found, 818.1836; [α]<sup>21</sup><sub>D</sub>+71.61° (*c* 1.0, CHCl<sub>3</sub>).

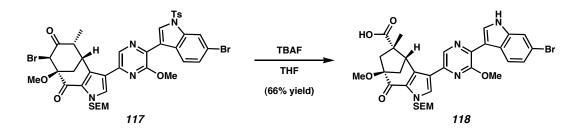


α-Bromoketone 117. To ketone 113 (5.0 mg, 0.0061 mmol) and triethylamine (160 μL, 1.15 mmol) in  $CH_2Cl_2$  (1 mL) at 0 °C was added TMSOTF (70 μL, 0.350 mmol) dropwise over 1 min. The reaction mixture was stirred for 30 min, quenched with saturated aq. NaHCO<sub>3</sub> (2 mL), and extracted with EtOAc (5 x 1 mL). The combined organic layers were washed with brine (1.5 mL) and dried over Na<sub>2</sub>SO<sub>4</sub>. Evaporation of

To crude silvl enol ether product 115 in THF (1.5 mL) at 23 °C was added freshly recrystallized NBS (14 mg, 0.0786 mmol). The reaction mixture was stirred for 1 min, quenched with saturated aq. NaHCO<sub>3</sub> (2 mL), and extracted with EtOAc (5 x 1 mL). The combined organic layers were washed with brine (1.5 mL), dried by passage over a plug of silica gel (EtOAc eluent), and evaporated under reduced pressure to afford the crude product. Purification by preparative thin layer chromatography (1:1 hexanes:EtOAc eluent) afforded  $\alpha$ -bromoketone 117 (5.3 mg, 97% yield, 2 steps) as a colorless oil.  $R_f$ 0.68 (1:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz,  $C_6D_6$ ):  $\delta$  9.01 (d, J = 8.5 Hz, 1H), 8.87 (s, 1H), 8.69 (s, 1H), 8.15 (s, 1H), 7.70 (d, J = 8.3 Hz, 2H), 7.49 (d, J = 8.5 Hz, 1H), 7.10 (s, 1H), 6.40 (d, J = 8.0 Hz, 2H), 5.45 (d, J = 10.2 Hz, 1H), 5.36 (d, J = 10.2 Hz, 1H), 4.75 (s, 1H), 4.14-4.06 (m, 1H), 3.68 (s, 3H), 3.60-3.46 (comp. m, 3H), 3.44 (s, 3H), 2.64-2.55 (m, 1H), 2.52-2.43 (m, 1H), 1.58 (s, 3H), 0.89 (t, J = 8.0 Hz, 2H), 0.78 (d, J =6.6 Hz, 3H), -0.03 (s, 9H); <sup>13</sup>C NMR (125 MHz, C<sub>6</sub>D<sub>6</sub> 38/39 C): δ 202.4, 185.4, 156.6, 145.5, 143.2, 136.9, 136.7, 136.6, 135.8, 133.2, 131.8, 130.5 (2C), 129.8, 129.4, 128.0, 127.3 (2C), 126.5, 121.0, 120.0, 117.7, 117.3, 82.9, 77.3, 67.0, 58.4, 54.1, 53.0, 43.4, 36.7, 35.0, 21.3, 18.4, 12.4, -1.0 (3C); IR (film): 2950, 1719, 1662, 1557, 1374, 1190, 1178, 1141, 1089; HRMS-FAB (m/z):  $[M + H]^+$  calc'd for  $C_{39}H_{43}Br_2N_4O_7SSi$ , 899.0968; found, 899.0952;  $[\alpha]^{27}_{D}$  +10.23° (*c* 0.66, C<sub>6</sub>H<sub>6</sub>).

The relative stereochemistry of  $\alpha$ -bromoketone **117** was determined by NOE experiments. Medium strength NOE interactions were observed as indicated below.<sup>51</sup>

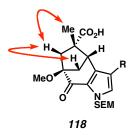


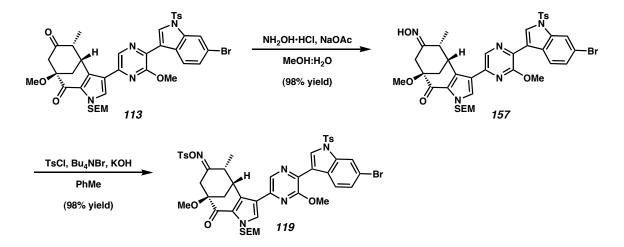


**Favorskii product 118.** To α-bromoketone **117** (3.0 mg, 0.0033 mmol) in THF (1.0 mL) at 23 °C was added TBAF (1.0 M in THF, 20 µL, 0.020 mmol). The reaction mixture was stirred for 15 min, quenched with 10% (*w/v*) aq. citric acid (1 mL), diluted with brine (500 µL), and extracted with EtOAc (5 x 1 mL). The combined organic layers were dried by passage over a plug of silica gel (EtOAc eluent, then 5:1 CH<sub>2</sub>Cl<sub>2</sub>:MeOH eluent) and evaporated under reduced pressure to afford the crude product. Purification by preparative thin layer chromatography (5:1 CH<sub>2</sub>Cl<sub>2</sub>:MeOH eluent) afforded Favorskii product **118** (1.5 mg, 66% yield) as a yellow oil. R<sub>f</sub> 0.53 (5:1 CH<sub>2</sub>Cl<sub>2</sub>:MeOH); <sup>1</sup>H NMR (600 MHz, CD<sub>3</sub>OD): δ 8.61 (d, J = 9.2 Hz, 1H), 8.59 (s, 1H), 8.21 (s, 1H), 7.97 (s, 1H), 7.60 (s, 1H), 7.25 (d, J = 8.2 Hz, 1H), 5.81 (d, J = 10.1 Hz, 1H), 5.76 (d, J = 10.1 Hz, 1H), 4.96 (app. d, J = 3.7 Hz, 1H), 4.24 (s, 3H), 3.65 (m, 2H), 3.42 (s, 3H), 2.98 (d, J = 10.1 Hz, 1H), 4.96 (app. d, J = 3.7 Hz, 1H), 4.24 (s, 3H), 3.65 (m, 2H), 3.42 (s, 3H), 2.98 (d, J = 10.1 Hz, 1H), 4.96 (app. d, J = 3.7 Hz, 1H), 4.24 (s, 3H), 3.65 (m, 2H), 3.42 (s, 3H), 2.98 (d, J = 10.1 Hz, 1H), 4.96 (app. d, J = 3.7 Hz, 1H), 4.24 (s, 3H), 3.65 (m, 2H), 3.42 (s, 3H), 2.98 (d, J = 10.1 Hz, 1H), 4.96 (app. d, J = 3.7 Hz, 1H), 4.24 (s, 3H), 3.65 (m, 2H), 3.42 (s, 3H), 2.98 (d, J = 10.1 Hz, 1H), 4.96 (app. d, J = 3.7 Hz, 1H), 4.24 (s, 3H), 3.65 (m, 2H), 3.42 (s, 3H), 2.98 (d, J = 10.1 Hz, 1H), 4.96 (app. d, J = 3.7 Hz, 1H), 4.24 (s, 3H), 3.65 (m, 2H), 3.42 (s, 3H), 2.98 (d, J = 10.1 Hz, 1H), 4.96 (app. d, J = 3.7 Hz, 1H), 4.24 (s, 3H), 3.65 (m, 2H), 3.42 (s, 3H), 2.98 (d, J = 10.1 Hz, 1H), 4.96 (app. d, J = 3.7 Hz, 1H), 4.24 (s, 3H), 3.65 (m, 2H), 3.42 (s, 3H), 2.98 (d, J = 10.1 Hz, 1H), 4.96 (app. d, J = 3.7 Hz, 1H), 4.24 (s, 3H), 3.65 (m, 2H), 3.42 (s, 3H), 2.98 (d, J = 10.1 Hz, 1H), 4.96 (app. d, J = 3.7 Hz, 1H), 4.24 (s, 3H), 3.65 (m, 2H), 3.42 (s, 3H), 2.98 (d, J = 10.1 Hz, 1H), 4.96 (app. d, J

14.7 Hz, 1H), 2.38 (d, J = 11.0 Hz, 1H), 2.30 (dd, J = 11.0, 4.6 Hz, 1H), 1.63 (d, J = 14.7 Hz, 1H), 1.07 (s, 3H), 0.94-0.88 (m, 2H), -0.02 (s, 9H); <sup>13</sup>C NMR (125 MHz, CD<sub>3</sub>OD):  $\delta$  193.1, 170.2, 157.1, 142.4, 142.3, 139.5, 139.1, 133.1, 131.5, 130.6, 128.4, 126.9, 125.5, 124.4, 121.9, 116.8, 115.3, 112.8, 91.3, 77.8, 67.2, 55.1, 54.1, 46.2, 45.6, 44.7, 30.9, 25.0, 18.8, -1.1 (3C); IR (film): 3288 (br), 2927, 2855, 1711, 1659, 1553, 1535, 1449, 1409, 1367, 1250, 1198, 1093; HRMS-FAB (m/z): [M + H]<sup>+</sup> calc'd for C<sub>32</sub>H<sub>38</sub>BrN<sub>4</sub>O<sub>6</sub>Si, 683.1724; found, 683.1721; [ $\alpha$ ]<sup>23</sup><sub>D</sub> -26.34° (c 0.2, CH<sub>3</sub>OH).

The relative stereochemistry of Favorskii product **118** was determined by NOE experiments. Medium strength NOE interactions were observed as indicated below.<sup>51</sup>



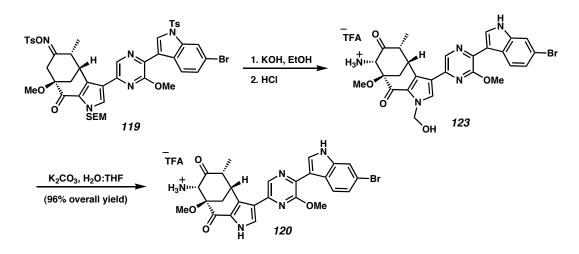


**Tosyl Oxime 119.** To ketone **113** (50.0 mg, 0.061 mmol), NH<sub>2</sub>OH•HCl (85 mg, 1.22 mmol), and NaOAc•3H<sub>2</sub>O (125 mg, 0.915 mmol) was added methanol (2.5 mL),

followed by H<sub>2</sub>O (350 µL), then additional methanol (5 mL). The homogeneous solution was stirred at 23 °C for 8 h, and the solvent was removed under reduced pressure. H<sub>2</sub>O (15 mL) was added, and the resulting mixture was extracted with EtOAc (3 x 15 mL). The combined organic layers were washed with brine (15 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure. The crude product was further purified by filtration over a plug of silica gel (EtOAc eluent) to yield oxime **157** (50.1 mg, 98% yield) as a yellow foam, which was used without purification in the subsequent reaction.  $R_f = 0.46$  (1:1 hexanes:EtOAc).

To a solution of oxime **157** (20.0 mg, 0.0240 mmol), TsCl (14.0 mg, 0.0734 mmol), and Bu<sub>4</sub>NBr (1.0 mg, 0.0031 mmol) in toluene (2.0 mL) at 0 °C was added 50% aq. KOH (310 µL). The reaction mixture was stirred at 0 °C for 2 h, quenched with icecold  $H_2O(1.5 \text{ mL})$  and extracted with ice-cold EtOAc (5 x 1 mL). The combined organic layers were washed with brine (1 mL), dried by passage over a plug of silica gel (EtOAc eluent), and evaporated under reduced pressure. The crude product was purified by flash chromatography (1:1 hexanes: EtOAc) to yield tosyl oxime **119** (23.3 mg, 98% yield) as a yellow foam.  $R_f = 0.48$  (1:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  8.63 (d, J =8.5 Hz, 1H), 8.46 (s, 1H), 8.41 (s, 1H), 8.19 (d, J = 1.4 Hz, 1H), 7.81 (d, J = 8.3 Hz, 2H), 7.65 (d, J = 8.0 Hz, 2H), 7.51 (s, 1H), 7.44 (dd, J = 8.7 Hz, 1.5 Hz, 1H), 7.28-7.19 (comp. m, 4H), 5.87 (d, J = 10.2 Hz, 1H), 5.42 (d, J = 10.2 Hz, 1H), 4.45-4.43 (m, 1H), 4.20 (s, 3H), 3.67-3.53 (comp. m, 3H), 3.38 (s, 3H), 2.98-2.89 (m, 1H), 2.87-2.77 (m, 1H), 2.42 (s, 3H), 2.35 (s, 3H), 2.12 (d, J = 14.0 Hz, 2H), 1.05-0.85 (m, 2H), 0.78 (d, J = 6.6 Hz, 3H), -0.02 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 187.2, 165.8, 156.3, 145.8, 144.8, 143.5, 135.8, 135.7, 135.3, 135.0, 132.9, 132.6, 130.4 (2C), 129.9, 129.4 (2C), 129.1

(2C), 128.9, 128.4, 128.0, 127.5, 127.2 (2C), 125.3, 120.3, 119.2, 116.8, 116.5, 80.8, 77.4, 67.2, 54.4, 52.2, 42.5, 40.3, 36.5, 36.2, 21.9, 21.9, 18.1, 13.7, -1.1 (3C); IR (film): 2946, 1665, 1555, 1373, 1191, 1178, 1140 cm<sup>-1</sup>; HRMS-FAB (m/z): [M]<sup>+</sup> calc'd for C<sub>46</sub>H<sub>50</sub>N<sub>5</sub>O<sub>9</sub>SiS<sub>2</sub>Br, 987.2002; found, 987.2038; [ $\alpha$ ]<sup>20</sup><sub>D</sub> +139.01° (c 1.0, CHCl<sub>3</sub>).

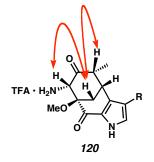


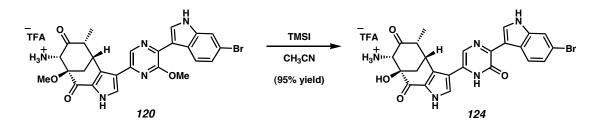
Aminoketone 120. To a stirred solution of tosyl oxime 119 (23.3 mg, 0.0236 mmol) in EtOH (3.5 mL) at 0 °C was added 50% aq. KOH (450 µL) dropwise over 1 min. The reaction mixture was stirred at 0 °C for 3 h, then 6 N aq. HCl (5 mL) was added. The reaction mixture was heated to 60 °C for 10 h, cooled to 23 °C, and purified by reversed-phase filtration through a Sep-Pak column: loaded with water containing 0.1% (*w*/*v*) TFA, washed with 15% acetonitrile:water containing 0.1% (*w*/*v*) TFA to remove salts, then 70% acetonitrile:water containing 0.1% (*w*/*v*) TFA to collect the crude product. The solvents were removed under reduced pressure to afford hemiaminal 123, which was used immediately in the subsequent reaction. Although hemiaminal 123 is typically used in crude form, it has been observed by <sup>1</sup>H NMR. <sup>1</sup>H NMR (600 MHz, CD<sub>3</sub>OD):  $\delta$  8.61 (d, *J* = 8.2 Hz, 1H), 8.52 (s, 1H), 8.24 (s, 1H), 7.94 (s, 1H), 7.60 (s, 1H), 7.25 (d, *J* = 9.2 Hz, 1H), 5.72 (d, *J* = 10.1 Hz, 1H), 5.65 (d, *J* = 10.1 Hz, 1H), 4.85-4.82

(m, 1H), 4.49 (s, 1H), 4.21 (s, 3H), 3.47 (s, 3H), 3.36-3.30 (m, 1H), 3.26 (dd, *J* = 12.8, 2.7 Hz, 1H), 2.61 (dd, *J* = 12.8, 2.7 Hz, 1H), 0.85 (d, *J* = 7.3 Hz, 3H).

Hemiaminal 123 and K<sub>2</sub>CO<sub>3</sub> (60 mg, 0.434 mmol) in THF (2 mL) at 23 °C was added  $H_2O$  (200 µL). The reaction mixture was stirred for 10 min, then purified by reversed-phase filtration through a Sep-Pak column: loaded with water containing 0.1% (w/v) TFA, washed with 10% acetonitrile:water containing 0.1% (w/v) TFA to remove salts, then 70% acetonitrile:water containing 0.1% (w/v) TFA to collect the crude product. After removal of solvents under reduced pressure, the crude material was further purified by reversed-phased HPLC. Concentration under reduced pressure provided aminoketone **120** (15.0 mg, 96% yield) as an orange/red oil. <sup>1</sup>H NMR (300 MHz, CD<sub>3</sub>OD):  $\delta$  8.60 (d, J = 8.5 Hz, 1H), 8.53 (s, 1H), 8.23 (s, 1H), 7.81 (s, 1H), 7.61 (d, J = 1.4 Hz, 1H), 7.25 (dd, J = 8.7 Hz, 1.8 Hz, 1H), 4.82-4.78 (m, 1H), 4.46 (s, 1H),4.21 (s, 3H), 3.47 (s, 3H), 3.41-3.30 (m, 1H), 3.26 (dd, J = 12.9 Hz, 3.9 Hz, 1H), 2.61 (dd, J = 12.9 Hz, 3.0 Hz, 1H), 0.88 (d, J = 6.6 Hz, 3H); <sup>13</sup>C NMR (75 MHz, CD<sub>3</sub>OD, 25/26 C): 8 203.5, 183.3, 156.8, 142.4, 139.9, 139.1, 136.3, 133.4, 130.7, 129.9, 129.6, 126.9, 125.5, 124.5, 123.1, 116.9, 115.4, 112.6, 84.3, 66.0, 54.5, 52.9, 40.4, 36.6, 12.2; IR (film): 3156 (br), 2935, 1674, 1531, 1447, 1409, 1203, 1135 cm<sup>-1</sup>; HRMS-FAB (*m/z*):  $[M + H]^+$  calc'd for  $C_{26}H_{25}N_5O_4Br$ , 550.1090; found, 550.1071;  $[\alpha]^{20}_{D}$  +99.19° (c 0.87, MeOH).

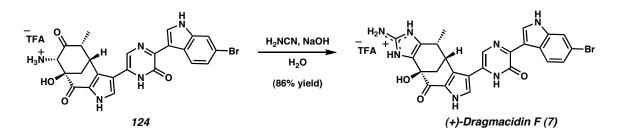
The relative stereochemistry of deprotected aminoketone **120** was determined by NOE experiments. Medium strength NOE interactions were observed as indicated below.<sup>51</sup> Analogous NOE interactions were observed for hemiaminal **123** and deprotected aminoketone **124**.





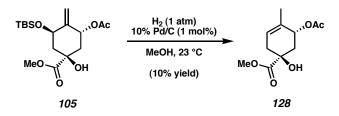
**Deprotected Aminoketone 124.** To a stirred solution of aminoketone **120** (7.5 mg, 0.0113 mmol) in MeCN (1 mL) at 0 °C was added TMSI (500  $\mu$ L, 3.51 mmol) dropwise over 30 sec. The reaction mixture was heated to 60 °C for 48 h, cooled to 0 °C, then transferred dropwise into a chilled solution (0 °C) of saturated aqueous sodium metabisulfite (5 mL). The mixture was diluted with 6 N HCl (15 mL), stirred at 0 °C for 20 min, then purified by reversed-phase filtration through a Sep-Pak column: loaded with water containing 0.1% (*w*/*v*) TFA, washed with 1 N HCl, 10% acetonitrile:water containing 0.1% (*w*/*v*) TFA to remove salts, then 60% acetonitrile:water containing 0.1% (*w*/*v*) TFA to collect the crude product. After removal of solvents under reduced

pressure, the crude material was further purified by reversed-phase HPLC. Concentration under reduced pressure provided deprotected aminoketone **124** (6.8 mg, 95% yield) as an orange/red oil. <sup>1</sup>H NMR (300 MHz, CD<sub>3</sub>OD):  $\delta$  8.69 (s, 1H), 8.59 (d, *J* = 8.5 Hz, 1H), 7.69 (s, 1H), 7.61 (d, *J* = 1.7 Hz, 1H), 7.57 (s, 1H), 7.27 (dd, *J* = 8.5 Hz, 1.7 Hz, 1H), 4.40 (s, 1H), 4.06-3.98 (m, 1H), 3.31-3.21 (m, 1H), 2.87 (dd, *J* = 13.2 Hz, 3.3 Hz, 1H), 2.79 (dd, *J* = 13.1 Hz, 2.9 Hz, 1H), 0.85 (d, *J* = 6.6 Hz, 3H); <sup>13</sup>C NMR (75 MHz, CD<sub>3</sub>OD, 23/24 C):  $\delta$  203.4, 186.0, 157.4, 139.1, 136.3, 132.5, 132.4, 130.2, 130.1, 128.2, 126.7, 126.7, 125.6, 124.9, 117.1, 115.4, 113.6, 79.3, 67.1, 49.6, 45.5, 36.7, 12.3; IR (film): 3164 (br), 2927, 1674, 1451, 1207, 1143 cm<sup>-1</sup>; HRMS-FAB (*m*/*z*): [M + H]<sup>+</sup> calc'd for C<sub>24</sub>H<sub>21</sub>N<sub>5</sub>O<sub>4</sub>Br, 522.0777; found, 522.0783; [ $\alpha$ ]<sup>22</sup><sub>D</sub>+86.88° (*c* 0.33, MeOH).



(+)–Dragmacidin F (7). To deprotected aminoketone 124 (3.6 mg, 0.0056 mmol) and cyanamide (120 mg, 2.86 mmol) in H<sub>2</sub>O (2 mL, degassed by sparging with argon) at 23 °C was added 10% aq. NaOH (80  $\mu$ L). The reaction mixture was heated to 60 °C for 2 h, cooled to 23 °C, then purified by reversed-phase filtration through a Sep-Pak column: loaded with water containing 0.1% (*w/v*) TFA, washed with 10% acetonitrile:water containing 0.1% (*w/v*) TFA to remove salts, then 60% acetonitrile:water containing 0.1% (*w/v*) TFA to collect the crude product. After removal of solvents under reduced pressure, the product was further purified by reversed-phase HPLC. Concentration under reduced pressure afforded (+)–dragmacidin F (7, 3.2 mg,

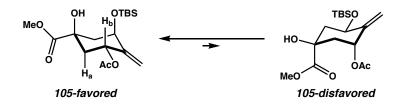
86% yield) as an orange/red oil. <sup>1</sup>H NMR (600 MHz, CD<sub>3</sub>OD): δ 8.69 (s, 1H), 8.59 (d, J = 8.7 Hz, 1H), 7.68 (s, 1H), 7.60 (s, 1H), 7.47 (s, 1H), 7.26 (d, J = 8.7 Hz, 1H), 4.12 (br s, 1H), 3.40-3.34 (m, 1H), 2.73 (dd, J = 12.0 Hz, 2.9 Hz, 1H), 2.45 (d, J = 11.6 Hz, 1H), 0.92 (d, J = 7.0 Hz, 3H); <sup>13</sup>C NMR (125 MHz, CD<sub>3</sub>OD, 22/25 C): δ 188.5, 157.5, 149.6, 139.1, 132.6, 132.4, 128.5, 128.4, 126.7, 126.2, 125.6, 124.9, 124.8, 123.3, 117.1, 115.4, 113.7, 72.8, 45.3, 36.9, 33.3, 15.9; IR (film): 3175 (br), 2925, 1679, 1637, 1205, 1141 cm<sup>-1</sup>; UV (MeOH)  $\lambda_{max}$  283, 389 nm; HRMS-FAB (m/z): [M + H]<sup>+</sup> calc'd for C<sub>25</sub>H<sub>21</sub>N<sub>7</sub>O<sub>3</sub>Br, 546.0889; found, 546.0883; [α]<sup>23</sup><sub>D</sub> +146.21° (*c* 0.45, MeOH).



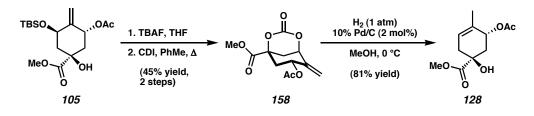
Acetoxycyclohexene 128. A mixture of methyl ester 105 (50.0 mg, 0.140 mmol) and 10% Pd/C (1.5 mg, 0.0014 mmol) in MeOH (1.3 mL) was stirred under an  $H_2$ atmosphere at 23 °C. After 35 min, the reaction mixture was filtered over a Celite plug (MeOH eluent), and the solvent was evaporated in vacuo. <sup>1</sup>H NMR integration showed that acetoxycyclohexene 128 was formed in approximately 10% yield.

Alternate Procedure. A mixture of methyl ester **105** (21.4 mg, 0.06 mmol) and 10% Pd/C (0.3 mg, 0.0003 mmol) in MeOH (1.5 mL) was cooled to 0 °C. The reaction vessel was then evacuated and back-filled with  $H_2$  (4x). After 1 h, the reaction mixture was filtered over a Celite plug (MeOH eluent), and the solvent was evaporated in vacuo. <sup>1</sup>H NMR integration showed that acetoxycyclohexene **128** was formed in approximately 3% yield.

The stable chair conformer of methyl ester **105** was determined using homodecoupling NMR experiments. The coupling constant between H<sub>a</sub> and H<sub>b</sub> was measured as  $J_{ab} = 10.7$  Hz.



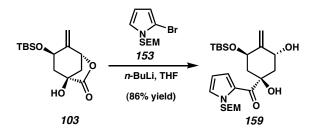
An analytical sample of 105 was prepared via an alternate route as follows:



Acetoxycarbonate 158. To a solution of methyl ester 105 (44.8 mg, 0.12 mmol) in THF (2 mL) was added TBAF (1.0 M in THF, 140 µL, 0.14 mmol). After 3 min of stirring, the reaction was quenched by the addition of saturated aq. NH<sub>4</sub>Cl (2 mL). EtOAc (4 mL) was added, and the phases were partitioned. The aqueous phase was further extracted with EtOAc (2 x 2 mL). The combined organic layers were successively washed with  $H_2O$  (1 mL) and brine (1 mL), and dried over MgSO<sub>4</sub>. The solvent was evaporated in vacuo, and the residue was dissolved in toluene (4 mL). 1,1'carbonyldiimidazole (82.1 mg, 0.51 mmol) was added, and the mixture was heated at reflux for 2 h. After cooling to 23 °C, the crude reaction mixture was directly purified by flash column chromatography (3:2 hexanes:EtOAc eluent) afford to pure

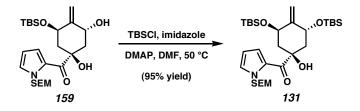
acetoxycarbonate **158** (16.9 mg, 45% yield, 2 steps).  $R_f 0.15$  (1:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  5.70-5.62 (m, 1H), 5.25 (app. d, J = 2.5 Hz, 1H), 5.19 (app. d, J = 2.5 Hz, 1H), 5.16 (dd, J = 4.1, 1.9 Hz, 1H), 3.81 (s, 3H), 2.84 (ddd, J = 13.4, 6.4, 2.7 Hz, 1H), 2.55-2.48 (m, 1H), 2.32-2.26 (m, 1H), 2.12 (s, 3H), 1.96 (dd, J = 13.3, 11.1 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  169.3, 168.3, 146.6, 140.2, 113.7, 81.6, 79.5, 66.4, 53.7, 39.3, 32.7, 20.9; IR (film) 1763 (br), 1230, 1180, 1120 cm<sup>-1</sup>; HRMS-FAB (m/z): [M + H]<sup>+</sup> calc'd for C<sub>12</sub>H<sub>15</sub>O<sub>7</sub>, 271.0818; found, 271.0810; [ $\alpha$ ]<sup>25</sup><sub>D</sub> -154.53° (*c* 1.0, C<sub>6</sub>H<sub>6</sub>).

Acetoxycyclohexene 128. A mixture of acetoxycarbonate 158 (18.5 mg, 0.07 mmol) and 10% Pd/C (1.4 mg, 0.001 mmol) in MeOH (1.3 mL) was cooled to 0 °C. The reaction vessel was then evacuated and back-filled with H<sub>2</sub> (3x). After 1 hr at 0 °C, the reaction mixture was filtered over a Celite plug (MeOH eluent), and the solvent was evaporated in vacuo. The residue was purified by flash chromatography (1:1 EtOAc:hexanes eluent) to afford acetoxycyclohexene 128 (12.6 mg, 81% yield) as a colorless oil.  $R_f$  0.46 (2:1 EtOAc:hexanes); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  5.57-5.48 (comp. m, 2H), 3.77 (s, 3H), 3.06 (br s, 1H), 2.69-2.58 (m, 1H), 2.29-2.20 (m, 1H), 2.16-1.91 (comp. m, 2H), 2.05 (s, 3H), 1.69-1.66 (m, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  176.1, 170.9, 132.7, 122.0, 73.8, 70.7, 53.2, 37.1, 35.3, 21.3, 19.2; IR (film) 3477 (br), 2953, 1736, 1239 cm<sup>-1</sup>; HRMS-FAB (*m*/*z*): [M + H]<sup>+</sup> calc'd for C<sub>11</sub>H<sub>17</sub>O<sub>5</sub>, 229.1076; found 229.1066; [ $\alpha$ ]<sup>25</sup><sub>D</sub> -3.31° (*c* 0.6, CHCl<sub>3</sub>).



Anti-diol 159. To 2-bromo SEM pyrrole (153, 4.66 g, 16.87 mmol) in THF (112 mL) at -78 °C was added *n*-BuLi (2.5 M in hexanes, 6.04 mL, 15.09 mmol) dropwise over 1 min. After 7 min at -78 °C, lactone 103 (1.26 g, 4.44 mmol) in THF (10 mL) was added dropwise over 1 min. The reaction vessel was immediately warmed to -42 °C, stirred for 30 min, and cooled to -78 °C. The reaction mixture was quenched with saturated aq. NH<sub>4</sub>Cl (50 mL), then warmed to 23 °C. The volatiles were removed under reduced pressure. The residue was partitioned between Et<sub>2</sub>O (125 mL) and  $H_2O$  (100 mL), and the layers were separated. The aqueous layer was further extracted with Et<sub>2</sub>O (2 x 125 mL). The combined organic layers were washed with brine (75 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure. The crude product was purified by flash chromatography (4:1 hexanes: EtOAc eluent) to afford anti-diol 159 (1.84 g, 86% yield) as a pale yellow foam.  $R_f 0.48$  (2:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz,  $C_6D_6$ ):  $\delta$  8.11 (dd, J = 4.1, 1.7 Hz, 1H), 6.78 (app. t, J = 2.1 Hz, 1H), 6.15 (dd, J = 4.0, 2.6 Hz, 1H),5.71 (d, J = 9.9 Hz, 1H), 5.58 (d, J = 10.2 Hz, 1H), 5.26 (s, 1H), 5.17 (app. t, J = 1.8 Hz, 1H), 4.92-4.82 (m, 1H), 4.76-4.73 (m, 1H), 4.45 (app. t, J = 3.0 Hz, 1H), 3.47 (t, J = 7.7Hz, 2H), 2.66 (ddd, J = 12.4, 5.2, 2.5 Hz, 1H), 2.39 (dd, J = 14.4, 2.9 Hz, 1H), 2.20 (app. dt, J = 8.7, 4.8 Hz, 1H), 1.92 (app. t, J = 12.0 Hz, 1H), 0.88-0.80 (comp. m, 12H), -0.04 (s, 3H), -0.06 (s, 3H), -0.06 (s, 9H); <sup>13</sup>C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>): δ 192.8, 151.6, 130.5, 128.6, 124.8, 109.3, 108.3, 83.0, 78.5, 76.7, 66.4, 66.2, 48.5, 42.1, 26.1 (3C), 18.4, 18.4, -0.9 (3C), -4.4, -5.1; IR (film): 3456 (br), 2953, 1637, 1406, 1250, 1091 cm<sup>-1</sup>; HRMS-FAB

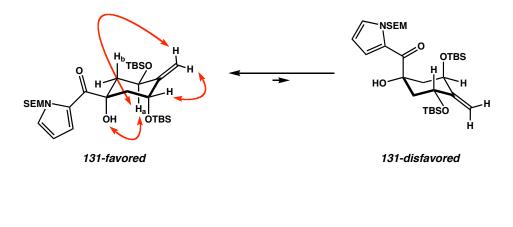
(m/z):  $[M + H]^+$  calc'd for  $C_{24}H_{44}NO_5Si_2$ , 482.2758; found, 482.2751;  $[\alpha]^{28}_{D}$  -21.18° (*c* 1.0,  $C_6H_6$ ).

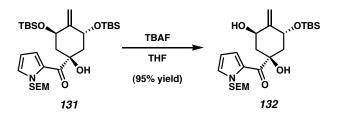


**Bis(silylether)** 131. To a solution of anti-diol 159 (253.1 mg, 0.53 mmol), imidazole (147.1 mg, 2.16 mmol), and DMAP (23.5 mg, 0.19 mmol) in DMF (5.0 mL), was added TBSCl (152.5 mg, 1.01 mmol). The solution was warmed to 50 °C for 70 min, cooled to 0 °C, then quenched by the addition of 10% (w/v) aq. citric acid (10 mL). Et<sub>2</sub>O (40 mL) was added, and the layers were partitioned. The aqueous phase was further extracted with Et<sub>2</sub>O (2 x 30 mL). The combined organic extracts were washed with brine (15 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure. The crude product was purified by flash chromatography (9:1 hexanes:EtOAc eluent) to provide bis(silylether) 131 (296.0 mg, 95% yield) as a colorless oil that solidified under reduced pressure.  $R_f 0.61$  (4:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz,  $C_6D_6$ ):  $\delta$  8.17 (dd, J = 4.0, 1.8 Hz, 1H), 6.76 (dd, J = 2.5, 1.7 Hz, 1H), 6.14 (dd, J = 4.0, 2.6 Hz, 1H), 5.68 (d, J = 9.9 Hz, 1H), 5.62 (d, J = 10.2 Hz, 1H), 5.37 (s, 1H), 5.32 (app. t, J = 2.1 Hz, 1H), 5.22-5.14 (m, 1H), 4.77 (app. t, J = 1.9 Hz, 1H), 4.50 (app. t, J = 3.0 Hz, 1H), 3.47 (t, J = 7.8 Hz, 2H), 2.82 (ddd, J = 12.7, 5.1, 2.6 Hz, 1H), 2.45 (dd, J = 14.6, 2.8 Hz, 1H), 2.27-2.18 (comp. m, 2H), 0.99 (s, 9H), 0.88 (s, 9H), 0.82 (t, J = 7.8 Hz, 2H), 0.17 (s, 3H), 0.14 (s, 3H), 0.00 (s, 3H), -0.04 (s, 3H), -0.07 (s, 9H);  $^{13}$ C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>, 29/30 C):  $\delta$ 192.6, 151.6, 130.4, 124.5, 109.3, 108.6, 83.2, 78.5, 76.8, 67.4, 66.3, 49.3, 42.1, 26.4

(3C), 26.1 (3C), 18.9, 18.4, 18.3, -0.9 (3C), -4.3, -4.4, -4.5, -5.1; IR (film): 3464 (br), 1953, 2929, 1640, 1405, 1309, 1251, 1094 cm<sup>-1</sup>; HRMS-FAB (*m/z*):  $[M + H]^+$  calc'd for  $C_{30}H_{58}NO_5Si_3$ , 596.3623; found, 596.3594;  $[\alpha]^{27}_{D}$  -7.16° (*c* 1.0,  $C_6H_6$ ).

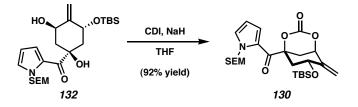
The stable chair conformer of bis(silylether) **131** was determined using a combination of NOESY-1D, gCOSY, and homodecoupling NMR experiments. Medium strength NOE interactions were observed as indicated below.<sup>51</sup> The coupling constant between H<sub>a</sub> and H<sub>b</sub> was measured as  $J_{ab} = 11.0$  Hz.





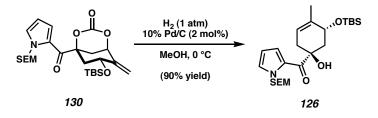
Syn-diol 132. To bis(silylether) 131 (113.9 mg, 0.19 mmol) in THF (10.0 mL) was added TBAF (1.0 M in THF, 195  $\mu$ L, 0.20 mmol) in a dropwise fashion over 1 min. The reaction mixture was stirred for 2 min, quenched with saturated aq. NH<sub>4</sub>Cl (15 mL), then poured into EtOAc (40 mL). The layers were partitioned, and the aqueous layer was further extracted with EtOAc (2 x 40 mL). The combined organic extracts were successively washed with H<sub>2</sub>O (15 mL) and brine (15 mL), dried over MgSO<sub>4</sub>, and

evaporated under reduced pressure. The residue was purified by flash chromatography (7:1 hexanes:EtOAc eluent) to furnish *syn* diol **132** (87.5 mg, 95% yield) as a pale yellow oil.  $R_f 0.29$  (4:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz,  $C_6D_6$ ):  $\delta$  7.09 (dd, J = 4.1, 1.4 Hz, 1H), 6.63 (dd, J = 2.3, 1.5 Hz, 1H), 5.89 (dd, J = 4.1, 2.5 Hz, 1H), 5.51-5.39 (comp. m, 4H), 5.27-5.19 (m, 1H), 5.01 (app. t, J = 2.1 Hz, 1H), 4.52-4.46 (m, 1H), 3.86 (d, J = 8.0 Hz, 1H), 3.37 (t, J = 7.7 Hz, 2H), 2.45-2.23 (comp. m, 3H), 2.04 (app. dt, J = 8.4, 4.9 Hz, 1H), 0.99 (s, 9H), 0.79 (t, J = 7.8 Hz, 2H), 0.14 (s, 3H), 0.11 (s, 3H), -0.09 (s, 9H); <sup>13</sup>C NMR (75 MHz,  $C_6D_6$ ):  $\delta$  191.6, 152.9, 131.4, 126.4, 124.0, 109.8, 108.5, 81.2, 78.8, 74.7, 67.4, 66.6, 49.0, 43.3, 26.4 (3C), 18.9, 18.3, -1.0 (3C), -4.5, -4.5; IR (film): 3363 (br), 2954, 1631, 1410, 1314, 1250, 1101 (br) cm<sup>-1</sup>; HRMS-FAB (m/z): [M + H]<sup>+</sup> calc'd for  $C_{24}H_{44}NO_5Si_2, 482.2758$ ; found, 482.2780; [ $\alpha$ ]<sup>27</sup> - 27.06° (*c* 1.0,  $C_6H_6$ ).



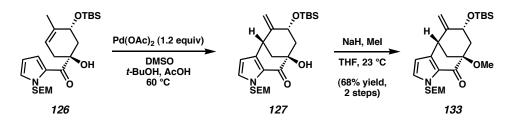
**Carbonate 130.** To *syn* diol **132** (68.2 mg, 0.14 mmol) and 1,1'carbonyldiimidazole (37.0 mg, 0.23 mmol) in THF (2.6 mL) was added NaH (60% dispersion in mineral oil, 21.9 mg, 0.55 mmol) in one portion. The reaction was stirred for 20 min at 23 °C, then quenched by addition of saturated aq. NH<sub>4</sub>Cl (20 mL). The reaction mixture was poured into EtOAc (30 mL), the layers were partitioned, and the aqueous layer was further extracted with EtOAc (2 x 30 mL). The combined organic extracts were successively washed with H<sub>2</sub>O (10 mL) and brine (10 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure. Purification of the residue by flash

chromatography (6:1 hexanes:EtOAc eluent) afforded carbonate **130** (65.8 mg, 92% yield) as a colorless oil.  $R_f 0.29$  (4:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz,  $C_6D_6$ ):  $\delta$  7.91 (dd, J = 4.1, 1.7 Hz, 1H), 6.68 (dd, J = 2.8, 1.7 Hz, 1H), 6.02 (dd, J = 4.3, 2.6 Hz, 1H), 5.51 (d, J = 9.9 Hz, 1H), 5.43 (d, J = 9.9 Hz, 1H), 5.24 (app. t, J = 1.9 Hz, 1H), 4.84-4.75 (m, 1H), 4.69 (app. t, J = 1.8 Hz, 1H), 4.46 (dd, J = 3.9, 1.9 Hz, 1H), 3.39 (t, J = 7.7 Hz, 2H), 2.78 (ddd, J = 13.5, 6.1, 2.5 Hz, 1H), 2.12-1.98 (comp. m, 2H), 1.92-1.85 (m, 1H), 0.86 (s, 9H), 0.81 (t, J = 7.8 Hz, 2H), -0.07--0.08 (comp. m, 12H), -0.10 (s, 3H); <sup>13</sup>C NMR (75 MHz,  $C_6D_6$ ):  $\delta$  185.9, 147.2, 146.4, 132.1, 126.7, 125.0, 112.2, 110.3, 87.9, 80.3, 78.8, 66.8, 66.5, 46.1, 33.7, 26.2 (3C), 18.6, 18.3, -1.0 (3C), -4.7, -5.0; IR (film): 2954, 1764, 1641, 1413, 1354, 1251, 1173, 1089 cm<sup>-1</sup>; HRMS-FAB (m/z): [M + H]<sup>+</sup> calc'd for  $C_{25}H_{42}NO_6Si_2, 508.2551$ ; found, 508.2560; [ $\alpha$ ]<sup>27</sup><sub>D</sub>-54.78° (c 1.0,  $C_6H_6$ ).



**Pyrrolocyclohexene 126.** A mixture of carbonate **130** (40.0 mg, 0.08 mmol) and 10% Pd/C (1.7 mg, 0.002 mmol) in MeOH (1.0 mL) was cooled to 0 °C. The reaction vessel was then evacuated and back-filled with H<sub>2</sub> (3x). After 1.75 hr at 0 °C, the reaction mixture was filtered over a Celite plug (MeOH eluent), and the solvent was evaporated in vacuo. The residue was purified by flash chromatography (9:1 hexanes:EtOAc eluent) to afford pyrrolocyclohexene **126** (33.1 mg, 90% yield) as a colorless oil.  $R_f$  0.53 (4:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz,  $C_6D_6$ ):  $\delta$  6.94 (dd, J = 4.1, 1.4 Hz, 1H), 6.64 (dd, J = 2.6, 1.5 Hz, 1H), 5.89 (dd, J = 4.0, 2.6 Hz, 1H), 5.54 (d, J

= 10.2 Hz, 1H), 5.45 (d, *J* = 10.2 Hz, 1H), 5.39-5.33 (m, 1H), 4.87-4.78 (m, 1H), 4.78 (s, 1H), 3.40 (t, *J* = 7.8 Hz, 2H), 2.97-2.85 (m, 1H), 2.48 (dd, *J* = 12.5, 9.8 Hz, 1H), 2.34-2.26 (m, 1H), 2.21-2.08 (m, 1H), 1.95-1.90 (m, 3H), 0.96 (s, 9H), 0.81 (t, *J* = 7.8 Hz, 2H), 0.06 (s, 3H), 0.03 (s, 3H), -0.08 (s, 9H); <sup>13</sup>C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>): δ 193.8, 138.5, 131.0, 126.4, 123.1, 120.1, 109.7, 78.8, 78.2, 69.6, 66.5, 44.7, 38.9, 26.4 (3C), 20.6, 18.6, 18.3, -1.0 (3C), -3.8, -4.5; IR (film): 3431 (br), 2954, 1634, 1414, 1250, 1089 (br) cm<sup>-1</sup>; HRMS-FAB (*m*/*z*): [M + H]<sup>+</sup> calc'd for C<sub>24</sub>H<sub>44</sub>NO<sub>4</sub>Si<sub>2</sub>, 466.2809; found, 466.2804; [α]<sup>28</sup><sub>D</sub> +26.19° (*c* 1.0, C<sub>6</sub>H<sub>6</sub>).

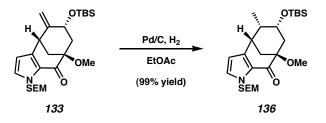


[3.3.1] Bicycle 127. To pyrrolocyclohexene 126 (40.0 mg, 0.0859 mmol) was added Pd(OAc)<sub>2</sub> (23.0 mg, 0.103 mmol), DMSO (14.6  $\mu$ L, 0.206 mmol), *t*-BuOH (6.9 mL), and AcOH (1.7 mL). The mixture was heated to 60 °C for 8 h, cooled to 23 °C, and filtered over a plug of silica gel (2:1 hexanes:EtOAc eluent). The solvent was evaporated, and the product was purified by flash chromatography on silica gel (8:1 hexanes:EtOAc eluent) to afford [3.3.1] bicycle 127 contaminated with a trace amount of pyrrolocyclohexene 126. Although this material was carried on to the subsequent step without further purification, an analytical sample of 127 was obtained by flash chromatography on silica gel (12:1 hexanes:EtOAc eluent) as a colorless oil. R<sub>f</sub> 0.64 (3:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, C<sub>6</sub>D<sub>6</sub>):  $\delta$  6.64 (d, *J* = 2.5 Hz, 1H), 6.25 (d, *J* = 10.2 Hz, 1H), 5.84 (d, *J* = 2.8 Hz, 1H), 5.07 (d, *J* = 9.9 Hz, 1H), 4.79 (br s, 1H), 4.66 (br s,

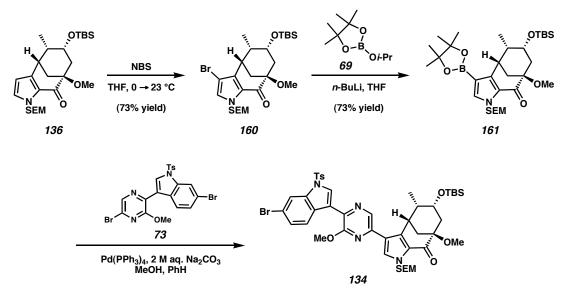
1H), 4.24-4.19 (m, 1H), 4.19 (s, 1H), 3.68-3.51 (m, 2H), 3.43-3.38 (m, 1H), 2.61 (app. dt, J = 7.3, 3.9 Hz, 1H), 2.21-2.10 (m, 2H), 2.06-1.98 (m, 1H), 0.99-0.77 (m, 2H), 0.72 (s, 9H), -0.04 (s, 9H), -0.11 (s, 3H), -0.24 (s, 3H); <sup>13</sup>C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>):  $\delta$  192.0, 148.6, 142.7, 130.5, 126.3, 113.2, 108.3, 77.0, 73.4, 73.0, 66.6, 48.5, 45.5, 40.2, 26.1 (3C), 18.4, 18.3, -1.0 (3C), -4.4, -5.1; IR (film): 3468 (br), 2951, 1648, 1422, 1250, 1094, 1062 cm<sup>-1</sup>; HRMS-FAB (*m*/*z*): [M + H]<sup>+</sup> calc'd for C<sub>24</sub>H<sub>42</sub>NO<sub>4</sub>Si<sub>2</sub>, 464.2652; found, 464.2661; [ $\alpha$ ]<sup>27</sup><sub>D</sub> +319.22° (*c* 1.0, C<sub>6</sub>H<sub>6</sub>).

Methyl Ether 133. The crude mixture of 126 and 127 obtained from the previous step was dissolved in THF (1.5 mL) at 23 °C, and NaH (60% dispersion in mineral oil, 17 mg, 0.429 mmol) was added. After stirring for 1 min at 23 °C, MeI was added (53 µL, 0.859 mmol). The resulting mixture was stirred for 1.5 h, quenched with saturated aq.  $NH_4Cl$  (1.5 mL), and extracted with  $Et_2O$  (4 x 1 mL). The combined organic layers were washed with brine (1 mL), dried by passage over a plug of silica gel (EtOAc eluent), and The crude product was purified by flash evaporated under reduced pressure. chromatography (10:1 hexanes: EtOAc eluent) to afford methyl ether 133 (28.2 mg, 68% yield, 2 steps) as a colorless oil.  $R_t$  0.43 (5:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz,  $C_6D_6$ :  $\delta$  6.62 (d, J = 2.6 Hz, 1H), 6.43 (d, J = 10.3 Hz, 1H), 5.86 (d, J = 2.6 Hz, 1H), 5.06 (d, J = 10.0 Hz, 1H), 4.84 (d, J = 1.5 Hz, 1H), 4.69 (d, J = 1.5 Hz, 1H), 4.29-4.22 (m, 1H), 3.42-3.52 (m, 2H), 3.45 (app. t, J = 2.8 Hz, 1H), 3.39 (s, 3H), 2.79 (app. dt, J =7.4, 3.8 Hz, 1H), 2.49 (app. dt, J = 8.1, 4.4 Hz, 1H), 1.96 (dd, J = 13.8, 4.7 Hz, 1H), 1.70 (dd, J = 11.7, 3.2 Hz, 1H), 0.96-0.82 (m, 2H), 0.73 (s, 9H), -0.06 (s, 9H), -0.11 (s, 3H), -0.06 (s, 9H), -0.11 (s, 3H), -0.06 (s, 9H), -0.01 (s, 9H), -0.0.23 (s, 3H); <sup>13</sup>C NMR (75 MHz,  $C_6D_6$ ):  $\delta$  189.2, 149.2, 140.9, 129.6, 128.9, 112.9, 107.6, 79.0, 77.3, 72.7, 66.6, 51.5, 46.3, 41.7, 39.9, 26.1 (3C), 18.4, 18.4, -1.0 (3C), -4.4,

-5.1; IR (film): 2951, 1661, 1426, 1250, 1113, 1066; HRMS-FAB (m/z): [M + H]<sup>+</sup> calc'd for C<sub>25</sub>H<sub>44</sub>NO<sub>4</sub>Si<sub>2</sub>, 478.2809; found, 478.2815; [ $\alpha$ ]<sup>27</sup><sub>D</sub> +312.37° (c 1.0, C<sub>6</sub>H<sub>6</sub>).



**Reduced Bicycle 136.** Methyl ether **133** (23 mg, 0.0479 mmol), 10% Pd/C (15 mg, 0.014 mmol), and EtOAc (2.5 mL) were combined, and the reaction vessel was evacuated and back-filled with H<sub>2</sub> (1 atm). The reaction mixture was stirred under H<sub>2</sub> for 5 min, then filtered over a plug of silica gel topped with Celite (EtOAc eluent) to afford reduced bicycle **136** as a colorless oil (23 mg, 99% yield). R<sub>f</sub> 0.28 (5:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, C<sub>6</sub>D<sub>6</sub>): 6.64 (d, J = 2.5 Hz, 1H), 6.52 (d, J = 10.2 Hz, 1H) 5.83 (d, J = 2.5 Hz, 1H), 5.05 (d, J = 10.2 Hz, 1H), 3.71-3.51 (comp. m, 3H), 3.42 (s, 3H), 2.78 (app. dt, J = 7.4, 3.9 Hz, 1H), 2.60 (app. q, J = 3.1 Hz, 1H), 2.40 (app. dt, J = 8.1, 4.6 Hz, 1H), 1.81 (dd, J = 13.8, 4.4 Hz, 1H), 1.58 (dd, J = 11.4, 2.9 Hz, 1H), 1.42-1.53 (m, 1H), 0.99-0.81 (m, 2H), 0.87 (d, J = 7.2 Hz, 3H), 0.72 (s, 9H), -0.06 (s, 9H), -0.10 (s, 3H), -0.21 (s, 3H); <sup>13</sup>C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>, 24/25 C):  $\delta$  189.3, 140.3, 129.1, 109.2, 79.2, 77.2, 71.5, 66.5, 51.2, 45.4, 41.9, 38.3, 36.8, 26.1 (3C), 18.4, 18.4, 17.1, -1.0 (3C), -4.4, -5.0; IR (film): 2952, 1660, 1497, 1425, 1251, 1118, 1100, 1042 cm<sup>-1</sup>; HRMS-FAB (*m*/*z*): [M + H]<sup>+</sup> calc'd for C<sub>25</sub>H<sub>46</sub>NO<sub>4</sub>Si<sub>2</sub>, 480.2965; found, 480.2955; [ $\alpha$ ]<sup>25</sup><sub>D</sub> +220.84° (*c* 1.0, C<sub>6</sub>H<sub>6</sub>).

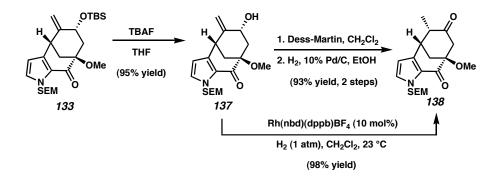


**Pyrazine 134.** To silyl ether **136** (10.0 mg, 0.0208 mmol) in THF (2 mL) at 0 °C was added freshly recrystallized NBS (4.8 mg, 0.0271 mmol) in THF (200  $\mu$ L). After 10 min at 0 °C, the reaction mixture was warmed to 23 °C, stirred for 40 min, then cooled to 0 °C. The reaction was quenched with saturated aq. Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> (1.5 mL), diluted with H<sub>2</sub>O (1 mL), and extracted with EtOAc (5 x 1 mL). The combined organic layers were washed with brine (1 mL), dried by passage over a plug of silica gel (EtOAc eluent), and evaporated under reduced pressure to afford the crude product. Further purification by preparative thin layer chromatography (4:1 hexanes:EtOAc eluent) afforded bromide **160** (8.5 mg, 73% yield) as a colorless oil. R<sub>f</sub> 0.4 (5:1 hexanes:EtOAc).

To bromide **160** (12.7 mg, 0.0227 mmol) and 2-isopropoxy-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (**69**, 190  $\mu$ L, 0.932 mmol) in THF (2.3 mL) at -78 °C was added *n*-BuLi (2.5 M in hexanes, 273  $\mu$ L, 0.682 mmol) dropwise over 1 min. After stirring for 10 min at -78 °C, the reaction mixture was quenched with saturated aq. NH<sub>4</sub>Cl (1.5 mL), warmed to 23 °C, diluted with H<sub>2</sub>O (1 mL), and extracted with EtOAc (5 x 1 mL). The combined organic layers were washed with brine (1 mL), dried by passage over a plug of silica gel (EtOAc eluent), and evaporated under reduced pressure to afford the crude product. Further purification by preparative thin layer chromatography (4:1 hexanes:EtOAc eluent) afforded boronic ester **161** (10.1 mg, 73% yield) as a colorless oil.  $R_f 0.38$  (5:1 hexanes:EtOAc).

A vial charged with bromopyrazine **73** (12.4 mg, 0.0231 mmol), boronic ester **161** (10.0 mg, 0.0165 mmol), and tetrakis(triphenylphosphine)palladium(0) (2.9 mg, 0.00248 mmol) was evacuated and purged with  $N_2$ . Deoxygenated benzene (330  $\mu$ L), deoxygenated methanol (65  $\mu$ L), and deoxygenated 2 M aq. Na<sub>2</sub>CO<sub>3</sub> (28  $\mu$ L) were then added. The reaction vessel was sealed, heated to 50 °C for 82 h, cooled to 23 °C, then quenched by the addition of Na<sub>2</sub>SO<sub>4</sub> (100 mg). Following filtration over a pad of silica gel (1:1 hexanes:EtOAc eluent) and evaporation to dryness under reduced pressure, the residue was purified by preparative thin layer chromatography (2:1 hexanes:EtOAc eluent) to afford pyrazine 134 (4.4 mg, 28% yield) as a yellow foam.  $R_f 0.44$  (2:1 hexanes:EtOAc); <sup>1</sup>H NMR (500 MHz,  $C_6D_6$ ):  $\delta$  9.02 (d, J = 8.8 Hz, 1H), 8.85 (s, 1H), 8.69 (d, J = 2.0 Hz, 1H), 8.35 (s, 1H), 7.71-7.68 (m, 2H), 7.48 (dd, J = 8.8 Hz, 2.0 Hz, 1H), 7.26 (s, 1H), 6.57 (d, J = 10.3 Hz, 1H), 6.40 (d, J = 8.3 Hz, 2H), 5.16 (d, J = 10.3Hz, 1H), 3.92 (d, J = 3.4 Hz, 1H), 3.74 (s, 3H), 3.73-3.58 (comp. m, 3H), 3.45 (s, 3H), 2.91 (app. dt, J = 7.3, 3.6 Hz, 1H), 2.44 (app. t, J = 7.1 Hz, 1H), 1.86 (dd, J = 13.9, 4.2 Hz, 1H), 1.74 (dd, J = 11.7, 2.9 Hz, 1H), 1.64-1.55 (comp. m, 4H), 1.02-0.86 (m, 2H), 0.75 (s, 9H), 0.68 (d, J = 6.8 Hz, 3H), -0.05 (s, 9H), -0.09 (s, 3H), -0.23 (s, 3H);  $^{13}$ C NMR (125 MHz, C<sub>6</sub>D<sub>6</sub>): δ 190.4, 156.7, 145.4, 145.0, 138.6, 136.7, 135.9, 135.8, 133.4, 131.0, 130.4 (2C), 129.5, 129.5, 129.1, 127.9, 127.4 (2C), 126.7, 120.8, 119.8, 118.0, 117.2, 79.0, 77.8, 72.1, 67.1, 53.9, 51.4, 45.4, 41.7, 39.8, 34.6, 26.2 (3C), 21.3, 18.6, 18.5, 17.3,

-1.0 (3C), -4.4, -5.0; IR (film): 2951, 1661, 1556, 1376, 1250, 1178, 1141, 1090, 1011 cm<sup>-1</sup>; HRMS-FAB (*m*/*z*): [M]<sup>+</sup> calc'd for C<sub>45</sub>H<sub>59</sub>BrN<sub>4</sub>O<sub>7</sub>SSi<sub>2</sub>, 934.2826; found, 934.2872;  $[\alpha]^{20}_{D}$  -91.02° (*c* 0.57, C<sub>6</sub>H<sub>6</sub>).



**Ketone 138.** To methyl ether **133** (120 mg, 0.25 mmol) in THF (12.5 mL) was added TBAF (1.0 M in THF, 750 μL, 0.75 mmol). The reaction mixture was stirred for 4 h, quenched with saturated aq. NH<sub>4</sub>Cl (10 mL), diluted with H<sub>2</sub>O (5 mL), and extracted with EtOAc (3 x 25 mL). The combined organic extracts were washed with brine (15 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure. The residue was purified by flash chromatography (1:1 hexanes:EtOAc eluent) to furnish allylic alcohol **137** (86 mg, 95% yield) as a pale yellow oil. R<sub>f</sub> 0.12 (2:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, C<sub>6</sub>D<sub>6</sub>): δ 6.60 (d, J = 2.8 Hz, 1H), 5.81 (d, J = 2.8 Hz, 1H), 5.64 (d, J = 10.2 Hz, 1H), 5.58 (d, J = 10.2 Hz, 1H), 4.80 (d, J = 1.7 Hz, 1H), 4.64 (d, J = 1.7 Hz, 1H), 4.15-4.09 (m, 1H), 3.68-3.59 (m, 2H), 3.42 (t, J = 3.2 Hz, 1H), 3.36 (s, 3H), 2.72 (app. dt, J = 7.4, 3.9 Hz, 1H), 2.58 (app. dt, J = 8.1, 4.9 Hz, 1H), 1.89 (dd, J = 14.2, 5.1 Hz, 1H), 1.65 (dd, J = 11.6, 3.0 Hz, 1H), 0.97-0.88 (m, 2H), 0.59 (d, J = 3.9 Hz, 1H), -0.03 (s, 9H); <sup>13</sup>C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>, 18/19 C): δ 189.4, 149.4, 140.6, 130.4, 113.8, 107.4, 78.9, 76.7, 72.0, 66.2, 51.6, 44.3, 41.1, 39.5, 18.4, -0.9 (3C); IR (film): 3460 (br), 2951, 1659, 1424,

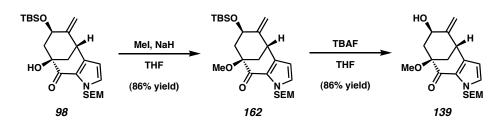
1248, 1111, 1023 cm<sup>-1</sup>; HRMS-FAB (*m/z*):  $[M + H]^+$  calc'd for C<sub>19</sub>H<sub>30</sub>NO<sub>4</sub>Si, 364.1944; found, 364.1942;  $[\alpha]_{-D}^{24} + 330.71^\circ$  (*c* 1.0, C<sub>6</sub>H<sub>6</sub>).

Allylic alcohol **137** (44.0 mg, 0.121 mmol) and freshly prepared Rh(nbd)(dppb)BF<sub>4</sub> (8.6 mg, 0.0121 mmol)<sup>48</sup> were combined under a glovebox atmosphere. The reaction vessel was carefully sealed and removed from the glovebox.  $CH_2Cl_2$  (12.0 mL) was added, and a balloon of H<sub>2</sub> (1 atm) was applied without purging. After 3 h of stirring, the reaction mixture was filtered over a plug of silica gel (CH<sub>2</sub>Cl<sub>2</sub>, then 2:1 hexanes:EtOAc eluent) to afford ketone **138** (43.0 mg, 98% yield) as a colorless oil.

Alternate Procedure. To allylic alcohol **137** (10.6 mg, 0.029 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (1.5 mL) at 23 °C was added Dess-Martin periodinane (50.0 mg, 0.118 mmol). The mixture was stirred for 10 min, quenched with a solution of saturated aq. NaHCO<sub>3</sub> and saturated aq. Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> (1:1, 2 mL), stirred for 10 min, and extracted with EtOAc (4 x 1 mL). The combined organic layers were washed with brine (1 mL), dried by passage over a plug of silica gel (EtOAc eluent), and evaporated under reduced pressure to afford the crude oxidized product, which was used in the subsequent reaction.  $R_f$  0.31 (2:1, hexanes:EtOAc).

A flask containing the crude oxidized product and 10% Pd/C (10 mg, 0.0094 mmol) in EtOH (2.0 mL) at 23 °C was evacuated and back-filled with  $H_2$  (3x). After 20 min, the reaction mixture was filtered over a Celite plug (EtOAc eluent), and the solvent was evaporated in vacuo. The residue was dissolved in EtOAc (2 mL), and then filtered over a short plug of silica gel (EtOAc eluent). After evaporation of solvent under reduced pressure, the crude material was further purified by preparative thin layer

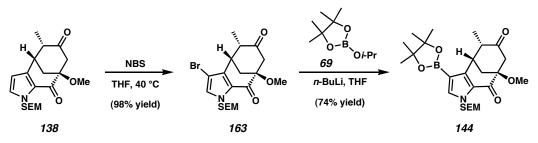
chromatography (2:1 hexanes:EtOAc) to afford ketone **138** (9.9 mg, 93% yield, 2 steps) as a colorless oil.  $R_f 0.30$  (2:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz,  $C_6D_6$ ):  $\delta$  6.53 (d, J = 2.5 Hz, 1H), 5.66 (d, J = 2.5 Hz, 1H), 5.50 (d, J = 10.5 Hz, 1H), 5.36 (d, J = 10.2 Hz, 1H), 3.57-3.38 (m, 2H), 3.34 (s, 3H), 2.98 (dd, J = 14.3, 2.5 Hz, 1H), 2.70-2.64 (m, 1H), 2.57-2.47 (m, 1H), 2.43 (d, J = 14.3 Hz, 1H), 2.11-1.99 (m, 1H), 1.69 (dd, J = 12.2, 2.6 Hz, 1H), 0.95 (d, J = 6.6 Hz, 3H), 0.84 (t, J = 8.0 Hz, 2H), -0.03 (s, 9H); <sup>13</sup>C NMR (125 MHz,  $C_6D_6$ ):  $\delta$  205.7, 187.9, 137.5, 131.1, 126.6, 109.7, 82.9, 76.8, 66.4, 52.7, 52.3, 48.1, 41.0, 37.7, 18.3, 13.0, -1.0 (3C); IR (film): 2952, 2931, 1716, 1660, 1421, 1123, 1097, 1076 cm<sup>-1</sup>; HRMS-FAB (m/z): [M + H]<sup>+</sup> - H<sub>2</sub> calc'd for C<sub>19</sub>H<sub>28</sub>NO<sub>4</sub>Si, 362.1788; found, 362.1778; [ $\alpha$ ]<sup>27</sup><sub>D</sub> +163.23° (*c* 1.0,  $C_6H_6$ ).



Allylic Alcohol 139. To [3.3.1] bicycle 98 (45 mg, 0.097 mmol) in THF (3 mL) at 23 °C was added NaH (60% dispersion in mineral oil, 40 mg, 1.0 mmol). After stirring for 2 min at 23 °C, MeI was added (335  $\mu$ L, 1.6 mmol). The resulting mixture was stirred for 1 h, and then quenched with saturated aq. NH<sub>4</sub>Cl (3 mL). EtOAc (3 mL) and H<sub>2</sub>O (3 mL) were added, and the layers were separated. The aqueous layer was further extracted with EtOAc (3 x 4 mL). The combined organic layers were washed with brine (4 mL), dried by passage over a plug of SiO<sub>2</sub> (EtOAc eluent), and evaporated under reduced pressure. The crude product was purified by flash chromatography (7:1

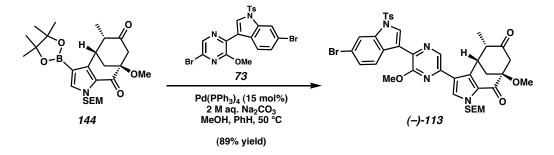
hexanes:EtOAc) to afford methyl ether **162** (40 mg, 86% yield) as a colorless oil.  $R_f 0.38$  (5:1 hexanes:EtOAc).

To methyl ether 162 (19 mg, 0.0396 mmol) in THF (1.5 mL) was added TBAF (1.0 M in THF, 75 µL, 0.075 mmol). The reaction mixture was stirred for 45 min, quenched with saturated aq. NH<sub>4</sub>Cl (1 mL), diluted with H<sub>2</sub>O (1 mL), and extracted with EtOAc (5 x 1 mL). The combined organic extracts were washed with brine (1 mL), dried by passage over a plug of  $SiO_2$  (EtOAc eluent), and evaporated under reduced pressure. The residue was purified by flash chromatography (1:1 hexanes:EtOAc eluent) to furnish allylic alcohol 139 (12.4 mg, 86% yield) as a pale yellow oil.  $R_f 0.16 (2:1)$ hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz,  $C_6D_6$ ):  $\delta$  6.57 (d, J = 2.4 Hz, 1H), 5.82 (d, J = 2.4Hz, 1H), 5.58 (d, J = 10.1 Hz, 1H), 5.49 (d, J = 10.1 Hz, 1H), 5.23-5.18 (m, 1H), 5.00-4.95 (m, 1H), 4.03-3.90 (m, 1H), 3.60-3.51 (comp. m, 3H), 3.32 (s, 3H), 2.61-2.50 (comp. m, 2H), 1.73-1.61 (comp. m, 2H), 1.15 (br s, 1H), 0.87 (t, J = 8.0 Hz, 2H), -0.07(s, 9H); <sup>13</sup>C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>, 18/19 C): δ 189.2, 150.1, 139.9, 131.0, 107.9, 106.6, 81.4, 77.0, 67.0, 66.5, 52.0, 46.3, 41.7, 40.9, 18.3, -1.0 (3C); IR (film): 3452 (br), 2951, 1653, 1420, 1250, 1125, 1076 cm<sup>-1</sup>; HRMS-EI (m/z): [M]<sup>+</sup> calc'd for C<sub>19</sub>H<sub>29</sub>NO<sub>4</sub>Si, 363.1866; found, 363.1857;  $[\alpha]_{D}^{23}$  -410.29° (*c* 1.0, C<sub>6</sub>H<sub>6</sub>).

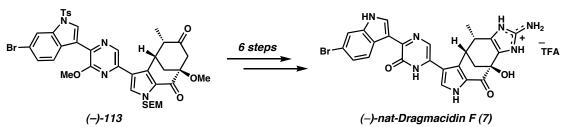


**Boronic Ester 144.** A flask wrapped in aluminum foil at 23 °C was charged with ketone **138** (25 mg, 0.0689 mmol), THF (5 mL), and freshly recrystallized NBS (37.5 mg, 0.211 mmol). The reaction vessel was placed in a 40 °C oil bath, stirred for 15 min, then cooled to 0 °C. The reaction was quenched with saturated aq. Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> (10 mL), diluted with H<sub>2</sub>O (5 mL), and extracted with Et<sub>2</sub>O (3 x 20 mL). The combined organic layers were washed with brine (15 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure to afford the crude product. Further purification by flash column chromatography (3:1 hexanes:EtOAc eluent) afforded bromide **163** (29.9 mg, 98% yield) as a colorless oil. R<sub>t</sub> 0.45 (2:1 hexanes:EtOAc).

To bromide **163** (27 mg, 0.061 mmol) and 2-isopropoxy-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (**69**, 510  $\mu$ L, 2.5 mmol) in THF (7 mL) at -78 °C was added *n*-BuLi (2.5 M in hexanes, 730  $\mu$ L, 0.183 mmol) dropwise over 3 min. After stirring for an additional 10 min at -78 °C, the reaction mixture was quenched with saturated aq. NH<sub>4</sub>Cl (7 mL), warmed to 23 °C, diluted with H<sub>2</sub>O (10 mL), and extracted with EtOAc (3 x 20 mL). The combined organic layers were washed with brine (15 mL), dried over MgSO<sub>4</sub> and evaporated under reduced pressure to afford the crude product. Further purification by flash column chromatography (3:1 hexanes:EtOAc eluent) afforded boronic ester **144** (22 mg, 74% yield) as a colorless oil. R<sub>f</sub> 0.42 (2:1 hexanes:EtOAc); <sup>1</sup>H NMR (300 MHz, C<sub>6</sub>D<sub>6</sub>):  $\delta$  7.37 (s, 1H), 5.46 (d, *J* = 10.2 Hz, 1H), 5.33 (d, *J* = 10.2 Hz, 1H), 3.77-3.72 (m, 1H), 3.49-3.38 (m, 2H), 3.31 (s, 3H), 3.03 (dd, J = 14.0, 2.8 Hz, 1H), 2.61-2.53 (m, 1H), 2.47 (d, J = 13.8 Hz, 1H), 2.36-2.25 (m, 1H), 1.78 (dd, J = 12.4, 3.0 Hz, 1H), 1.24 (d, J = 6.6 Hz, 3H), 1.12 (s, 12H), 0.84-0.77 (m, 2H), -0.05 (s, 9H); <sup>13</sup>C NMR (125 MHz, C<sub>6</sub>D<sub>6</sub>, 23/25 C):  $\delta$  206.4, 188.3, 144.6, 140.0, 83.6 (2C), 83.1, 77.1, 66.5, 52.9, 52.3, 49.0, 41.4, 37.1, 25.3 (2C), 25.2 (2C), 18.3, 13.0, -0.9 (3C); IR (film) 2977, 2951, 1718, 1664, 1543, 1399, 1322, 1263, 1145, 1092, 1074; HRMS-FAB (m/z): [M + H]<sup>+</sup> calc'd for C<sub>25</sub>H<sub>41</sub>NO<sub>6</sub>SiB, 490.2796; found, 490.2800; [ $\alpha$ ]<sup>29</sup><sub>D</sub> +50.77° (c 0.4, C<sub>6</sub>H<sub>6</sub>).



Pyrazine (-)-113. A vial charged with bromopyrazine 73 (29.6 mg, 0.055 mmol), boronic (18 0.0368 ester 144 mg, mmol), and tetrakis(triphenylphosphine)palladium(0) (6.4 mg, 0.0055 mmol) was evacuated and purged with N<sub>2</sub>. Deoxygenated benzene (735  $\mu$ L), deoxygenated methanol (150  $\mu$ L), and deoxygenated 2 M aq. Na<sub>2</sub>CO<sub>3</sub> (61  $\mu$ L) were then added. The reaction vessel was sealed, heated to 50 °C for 72 h, cooled to 23 °C, then quenched by the addition of Na<sub>2</sub>SO<sub>4</sub> (200 Following filtration over a pad of silica gel (3:1 EtOAc:hexanes eluent) and mg). evaporation to dryness under reduced pressure, the residue was purified by flash column chromatography (2:1  $\rightarrow$  1:1 hexanes:EtOAc eluent) to afford pyrazine (-)-113 (26.8 mg, 89% yield) as a yellow foam. R<sub>f</sub>, <sup>1</sup>H NMR, <sup>13</sup>C NMR, HRMS, and IR characterization data for (+)-113 are reported earlier in this section.  $[\alpha]^{27}_{D}$  -72.92° (*c* 1.0, CHCl<sub>3</sub>).



(–)-**Dragmacidin F** (7). Pyrazine (–)-**113** was converted to (–)-dragmacidin F (7) by methods described earlier in this section. <sup>1</sup>H NMR, <sup>13</sup>C NMR, HRMS, and IR characterization data for (+)-7 are also reported above.  $[\alpha]_{D}^{29}$ –148.33° (*c* 0.20, MeOH). For comparison, natural (–)-dragmacidin F (7):  $[\alpha]_{D}^{25}$ –159° (*c* 0.40, MeOH).<sup>1b</sup>

## 3.7 Notes and References

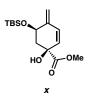
- (1) (a) Wright, A. E.; Pomponi, S. A.; Jacobs, R. S. PCT Int. Appl. WO 9942092
  August 26, 1999. (b) Cutignano, A.; Bifulco, G.; Bruno, I.; Casapullo, A.; Gomez-Paloma, L.; Riccio, R. *Tetrahedron* 2000, *56*, 3743-3748.
- (2) Portions of this work have been published, see: (a) Garg, N. K.; Caspi, D. D.;
  Stoltz, B. M. J. Am. Chem. Soc. 2004, 126, 9552-9553. (b) Garg, N. K.; Caspi, D. D.;
  Stoltz, B. M. J. Am. Chem. Soc. 2005, 127, in press.
- (3) For recent reviews of the Heck reaction, see: (a) Dounay, A. B.; Overman, L. E. *Chem. Rev.* 2003, 103, 2945-2963. (b) Beletskaya, I. P.; Cheprakov, A. V. *Chem. Rev.* 2000, 100, 3009-3066. (c) Amatore, C.; Jutand, A. J. Organomet. Chem. 1999, 576, 254-278.
- (4) For related examples of Pd-mediated carbocyclizations in natural product synthesis, see: (a) Baran, P. S.; Corey, E. J. J. Am. Chem. Soc. 2002, 124, 7904-7905. (b) Williams, R. M.; Cao, J.; Tsujishima, H.; Cox, R. J. J. Am. Chem. Soc. 2003, 125, 12172-12178.
- (5) (a) Stoltz, B. M. *Chem. Lett.* 2004, *33*, 362-367. (b) Trend, R. M.; Ramtohul, Y. K.; Ferreira, E. M.; Stoltz, B. M. *Angew. Chem., Int. Ed.* 2003, *42*, 2892-2895. (c)

Ferreira, E. M.; Stoltz, B. M. J. Am. Chem. Soc. 2003, 125, 9578-9579. d) Zhang,
H.; Ferreira, E. M.; Stoltz, B. M. Angew. Chem., Int. Ed. 2004, 43, 6144-6148.

- (6) For reviews and examples regarding the use of (-)-quinic acid in natural product synthesis, see: (a) Barco, A.; Benetti, S.; De Risi, C.; Marchetti, P.; Pollini, G. P.; Zanirato, V. *Tetrahedron: Asymmetry* 1997, *8*, 3515-3545. (b) Huang, P.-Q. *Youji Huaxue* 1999, *19*, 364-373. (c) Hanessian, S.; Pan, J.; Carnell, A.; Bouchard, H.; Lesage, L. *J. Org. Chem.* 1997, *62*, 465-473. (d) Hanessian, S. In *Total Synthesis of Natural Products: The "Chiron" Approach*, Baldwin, E. J., Ed.; Pergamon Press: Oxford, 1983; pp 206-208.
- (7) (a) Philippe, M.; Sepulchre, A. M.; Gero, S. D.; Loibner, H.; Streicher, W.; Stutz,
  P. J. Antibiot. 1982, 35, 1507-1512. (b) Manthey, M. K.; González-Bello, C.;
  Abell, C. J. Chem. Soc., Perkin Trans. 1 1997, 625-628.
- (8) For a review, see: Tsuji, J.; Mandai, T. Synthesis 1996, 1-24.
- (9) Greene, T. W.; Wuts, P. G. M. Protective Groups in Organic Synthesis, 3<sup>rd</sup> Ed.;
   Wiley-Interscience: New York, 1999.
- (10) Despite its widespread use in modern cross-coupling chemistry, to the best of our knowledge, this is the first report of  $Pd(P(t-Bu)_3)_2$  being used for a  $\pi$ -allyl

palladium substitution reaction. For recent examples of  $Pd(P(t-Bu)_3)_2$  in crosscoupling reactions, see: Hills, I. D.; Fu, G. C. *J. Am. Chem. Soc.* **2004**, *126*, 13178-13179 and references therein.

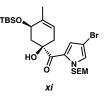
- (11) For the use of NMO as an additive in Stille couplings, see: Han, X.; Stoltz, B. M.;Corey, E. J. J. Am. Chem. Soc. 1999, 121, 7600-7605.
- (12) Under certain conditions, we were able to produce **108** as a mixture of diastereomers. However, upon exposure of **108** to Pd/C and  $H_2$  in MeOH, no reaction took place.
- (13) Treatment of 105 under a variety of standard homogeneous π-allyl reduction conditions<sup>8</sup> led to the formation of 106, 107, and x, all of which presumably arise from loss of OAc<sup>-</sup>. In stark contrast, exposure of 105 to heterogeneous reductive isomerization conditions did not produce any of these compounds (see Scheme 3.4.2).



(14) For the discovery and use of SEM pyrrole, see: (a) Edwards, M. P.; Ley, S. V.;
Lister, S. G.; Palmer, B. D. J. Chem. Soc., Chem. Commun. 1983, 630-633. (b)

Muchowski, J. M.; Solas, D. R. J. Org. Chem. 1984, 49, 203-205. (c) Edwards, M.
P.; Ley, S. V.; Lister, S. G.; Palmer, B. D.; Williams, D. J. J. Org. Chem. 1984, 49, 3503-3516. (d) Edwards, M. P.; Doherty, A. M.; Ley, S. V.; Organ, H. M. Tetrahedron 1986, 42, 3723-3729.

(15) During the conversion of 109 to 99, pyrrole xi was formed as a byproduct.



- (16) Littke, A. F.; Fu, G. C. J. Am. Chem. Soc. 2001, 123, 6989-7000.
- (17) By conducting reactions in THF- $d_8$ , it was possible to monitor Heck reactions by <sup>1</sup>H NMR.
- (18) DMSO has commonly been employed in oxidative Pd(II) chemistry. See: (a) Larock, R. C.; Hightower, T. R. J. Org. Chem. 1993, 58, 5298-5300. (b) Van Benthem, R. A. T. M.; Hiemstra, H.; Michels, J. J.; Speckamp, W. N. J. Chem. Soc., Chem. Commun. 1994, 357-359. (c) Rönn, M.; Bäckvall, J.-E.; Andersson, P. G. Tetrahedron Lett. 1995, 36, 7749-7752. (d) Chen, M. S.; White, M. C. J. Am. Chem. Soc. 2004, 126, 1346-1347. (e) Stahl, S. S. Angew. Chem., Int. Ed. 2004, 43, 3400-3420. See also references therein.

- (19) Gilow, H. M.; Hong, Y. H.; Millirons, P. L.; Snyder, R. C.; Casteel, W. J., Jr. J. *Heterocycl. Chem.* 1986, 23, 1475-1480.
- (20) Reactions conducted in the presence of acetic acid-*d* led to deuterium incorporation in the pyrrole ring of both **100** and **98**, mostly at C(4).
- (21) The instability of pyrroles to oxidants is well known. See: (a) Ciamician, G.; Silber,
  P. *Chem. Ber.* 1912, 45, 1842-1845. (b) Bernheim, F.; Morgan, J. E. *Nature* 1939, 144, 290. (c) Chierici, L.; Gardini, G. P. *Tetrahedron* 1966, 22, 53-56.
- (22) The Heck route required the use of 2,3-dibromopyrrole, an extremely unstable compound. For a discussion regarding the instability of bromopyrroles, see: Audebert, P.; Bidan, G. Synthetic Metals 1986, 15, 9-22.
- (23) In preliminary investigations, late-stage chemistry in the presence of a reactive 3° alcohol was unsuccessful.
- (24) (a) Rathore, R.; Kochi, J. K. J. Org. Chem. 1996, 61, 627-639. (b) Elfehail, F. E.;
  Zajac, W. W., Jr. J. Org. Chem. 1981, 46, 5151-5155. (c) Elfehail, F.; Dampawan,
  P.; Zajac, W. Synth. Commun. 1980, 10, 929-932. (d) Fischer, R. H.; Weitz, H. M.
  Synthesis 1980, 261-282.

- (25) Kreiser, W.; Körner, F. Helv. Chim. Acta 1999, 82, 1610-1629.
- (26) For example, upon treatment of bromoketone 117 with NaN<sub>3</sub>, azidoketone xii formed as the major product.



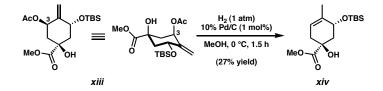
- (27) (a) Favorskii, A. E. J. Russ. Phys. Chem. Soc. 1894, 26, 559. (b) Chenier, P. J. J.
   Chem. Ed. 1978, 55, 286-291.
- (28) Neber, P. W.; Friedolsheim, A. V. Justus Liebigs Ann. Chem. 1926, 449, 109-134.
- (29) (a) For a review, see: O'Brien, C. Chem. Rev. 1964, 64, 81-89. (b) For a recent study involving the Neber rearrangement, see: Ooi, T.; Takahashi, M.; Doda, K.; Maruoka, K. J. Am. Chem. Soc. 2002, 124, 7640-7641.
- (30) (a) Purified by reversed-phase chromatography using trifluoroacetic acid in the eluent. (b) See Section 3.6 for details.

- (31) Derivatives of **119** bearing a free 3° alcohol or a TMS-protected 3° alcohol produced complex mixtures of products when subjected to Neber rearrangement conditions.
- (32) Acid-promoted dimerization of the aminoketone functionalities was not observed.
- (33) (a) Woodward reported a Neber rearrangement during synthetic studies involving lysergic acid. Unfortunately, the Neber rearrangement product could not be further utilized in the synthesis. See: Kornfeld, E. C.; Fornefeld, E. J.; Kline, G. B.; Mann, M. J.; Morrison, D. E.; Jones, R. G.; Woodward, R. B. J. Am. Chem. Soc. 1956, 78, 3087-3114. (b) For the use of the Neber rearrangement in the synthesis of a pharmaceutical substance, see: Chung, J. Y. L.; Ho, G.-J.; Chartrain, M.; Roberge, C.; Zhao, D.; Leazer, J.; Farr, R.; Robbins, M.; Emerson, K.; Mathre, D. J.; McNamara, J. M.; Hughes, D. L.; Grabowski, E. J. J.; Reider, P. J. *Tetrahedron Lett.* 1999, *40*, 6739-6743.
- (34) The intermediacy of azirines in Neber rearrangements is well accepted. These azirines presumably arise from transient nitrenes. See: (a) House, H. O.; Berkowitz, W. F. J. Org. Chem. 1963, 28, 307-311. (b) House, H. O.; Berkowitz, W. F. J. Org. Chem. 1963, 28, 2271-2276.
- (35) Hemiaminal **123** has been isolated and characterized by <sup>1</sup>H NMR.

- (36) Boehm, J. C.; Gleason, J. G.; Pendrak, I.; Sarau, H. M.; Schmidt, D. B.; Foley, J. J.;
   Kingsbury, W. D. J. Med. Chem. 1993, 36, 3333-3340.
- (37) Dragmacidin numbering convention, see reference 1b.
- (38) (a) (+)-Quinic acid ((+)-101) is commercially available in limited quantities from Interbioscreen Ltd. (50 mg/\$305 USD). (b) (+)-Quinic acid ((+)-101) potentially could be prepared via multistep synthesis by applying methods used for the preparation of (-)-quinic acid (101). See: Rapado, L. P.; Bulugahapitiya, V.; Renaud, P. *Helv. Chim. Acta* 2000, *83*, 1625-1632 and references therein.
- (39) Surprisingly, despite its widespread use in natural product synthesis and its near symmetry, (-)-quinic acid (101) has rarely been used in an enantiodivergent manner. For examples, see: (a) Ulibarri, G.; Nadler, W.; Skrydstrup, T.; Audrain, H.; Chiaroni, A.; Riche, C.; Grierson, D. S. *J. Org. Chem.* 1995, *60*, 2753-2761.
  (b) Ulibarri, G.; Audrain, H.; Nadler, W.; Lhermitte, H.; Grierson, D. S. *Pure Appl. Chem.* 1996, *68*, 601-604. (c) Barros, M. T.; Maycock, C. D.; Ventura, M. R. *J. Chem. Soc., Perkin Trans. 1* 2001, 166-173.
- (40) If R = R', **125** is considered to be pseudo- $C_2$ -symmetric. Pseudo- $C_2$ -symmetric molecules are those that would be  $C_2$ -symmetric if they did not contain a central chirotopic, nonstereogenic center. For discussions, see: (a) Schreiber, S. L. *Chem.*

Scr. 1987, 27, 563-566. (b) Poss, C. S.; Schreiber, S. L. Acc. Chem. Res. 1994, 27,
9-17. (c) Magnuson, S. R. Tetrahedron 1995, 51, 2167-2213. (d) Eliel, E. L.;
Wilen, S. H.; Mander, L. N. Stereochemistry of Organic Compounds; Wiley-Interscience: New York, 1994.

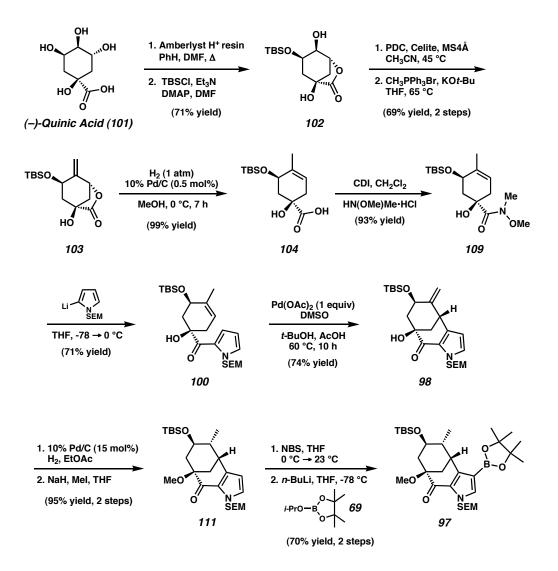
- (41) Olefin 105 was also exposed to the reaction conditions used for the reductive isomerization of 103 to 104. Only trace quantities of 128 were produced under those conditions.<sup>30b,42b</sup>
- (42) (a) Yield determined based on <sup>1</sup>H NMR integration. (b) The material isolated was predominantly a mixture of diastereomeric olefin hydrogenation products.
- (43) The favored conformation of **105** depicted in Scheme 3.4.2 is consistent with NMR studies.<sup>30b</sup>
- (44) Derivatives of **105** bearing C(3)-acetoxy groups (conformationally flexible and good leaving group) were also poor substrates in the reductive isomerization reaction as shown below (**xiii**  $\rightarrow$  **xiv**).<sup>42a</sup> The conformation of **xiii** was established by NMR studies.

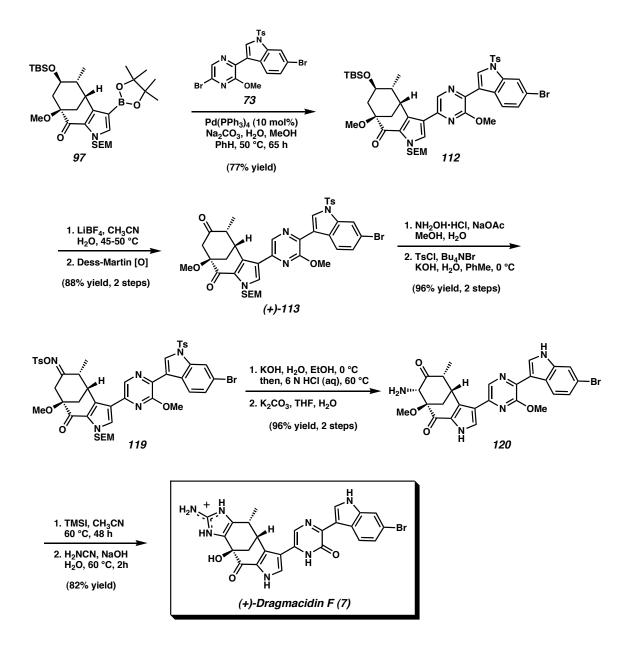


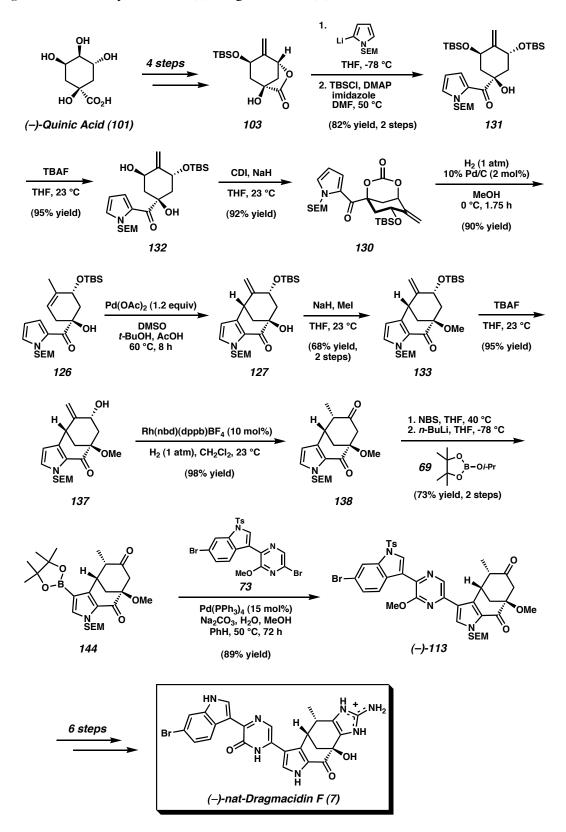
- (45) NMR experiments show that the C(3) silyl ether of 131 is axially disposed.<sup>30b</sup> For similar examples of axial-selective TBS cleavage promoted by TBAF, see: (a) Craig, B. N.; Janssen, M. U.; Wickersham, B. M.; Rabb, D. M.; Chang, P. S.; O'Leary, D. J. J. Org. Chem. 1996, 61, 9610-9613. (b) Meier, R.-M.; Tamm, C. *Helv. Chim. Acta*, 1991, 74, 807-818.
- (46) For a classic example involving the use of conformational analysis to solve stereochemical problems in total synthesis, see: Woodward, R. B.; Bader, F. E.; Bickel, H.; Frey, A. J.; Kierstead, R. W. *Tetrahedron* **1958**, *2*, 1-57.
- (47) Heating reactions above 50 °C led to mixtures of products involving partial and complete cleavage of the SEM and TBS groups.
- (48) (a) Brown, J. M. Angew. Chem., Int. Ed. Engl. 1987, 26, 190-203. (b) Bergens, S. H.; Bosnich, B. J. Am. Chem. Soc. 1991, 113, 958-967.
- (49) Xiao, W. Huaxue Shiji 1992, 14(6), 363-366.
- (50) Bailey, D. M.; Johnson, R. E. J. Med. Chem. 1973, 16, 1300-1302.
- (51) Claridge, T. D. W. In *High-Resolution NMR Techniques in Organic Chemistry*;
   Pergamon: Amsterdam, 1999; pp 320-326.

## **APPENDIX THREE**

Synthetic Summary for (+)- and (–)-Dragmacidin F (7)



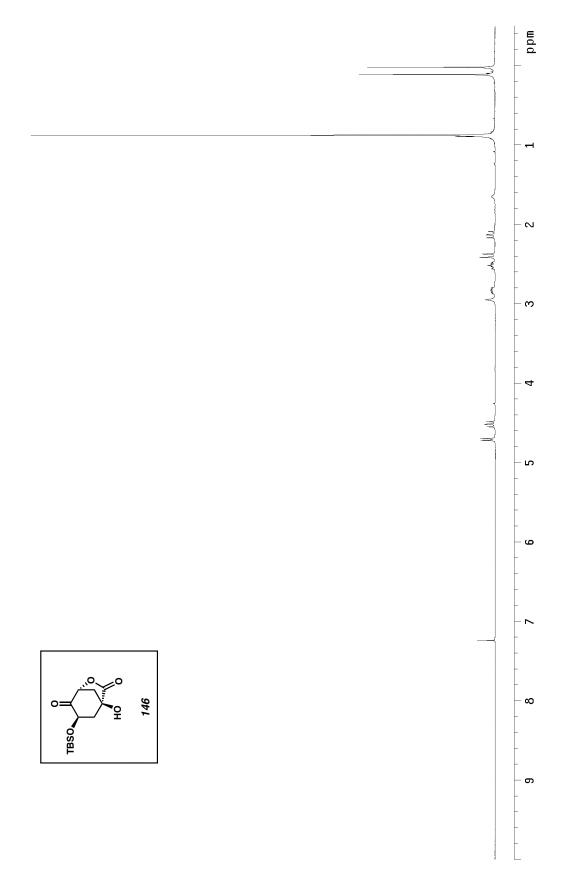




## **APPENDIX FOUR**

Spectra Relevant to Chapter Three:

The Total Synthesis of (+)- and (–)-Dragmacidin F





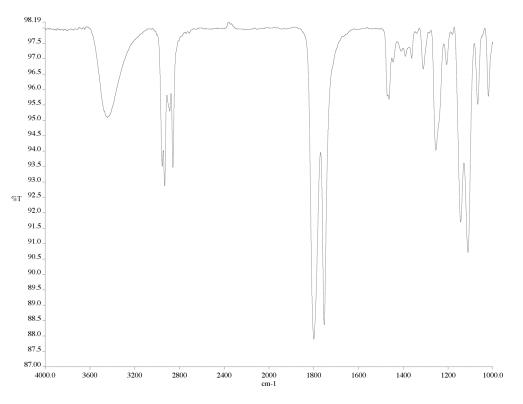
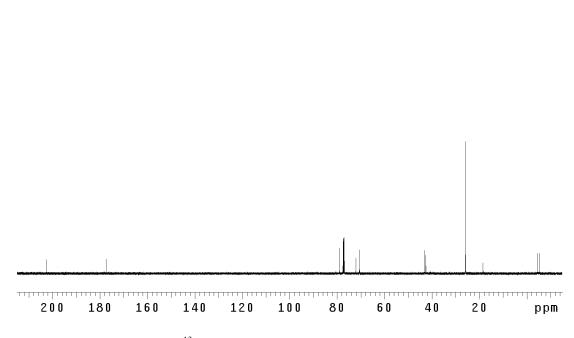
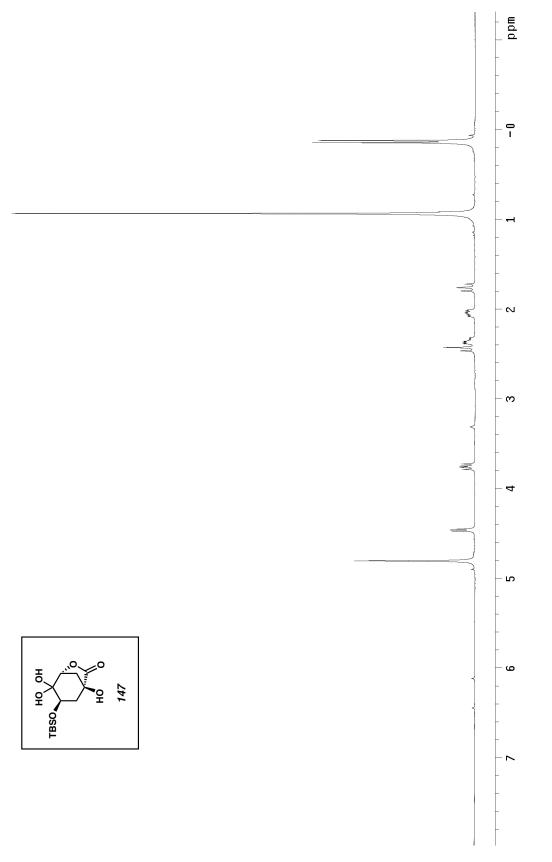


Figure A4.2 Infrared spectrum (thin film/NaCl) of compound 146.



*Figure A4.3* <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) of compound **146**.





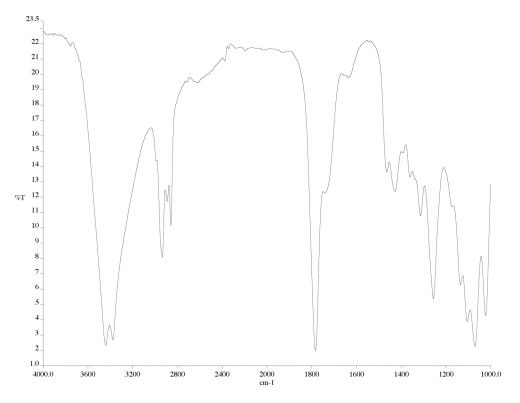
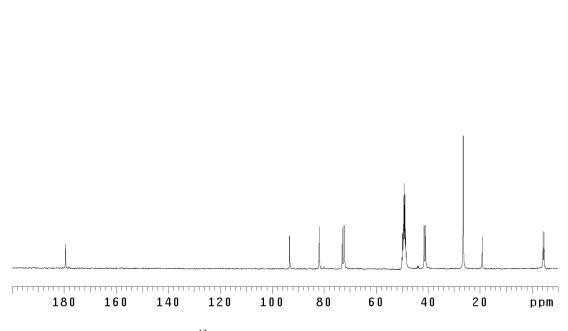


Figure A4.5 Infrared spectrum (thin film/NaCl) of compound 147.



*Figure A4.6*  $^{13}$ C NMR (75 MHz, CD<sub>3</sub>OD) of compound **147**.

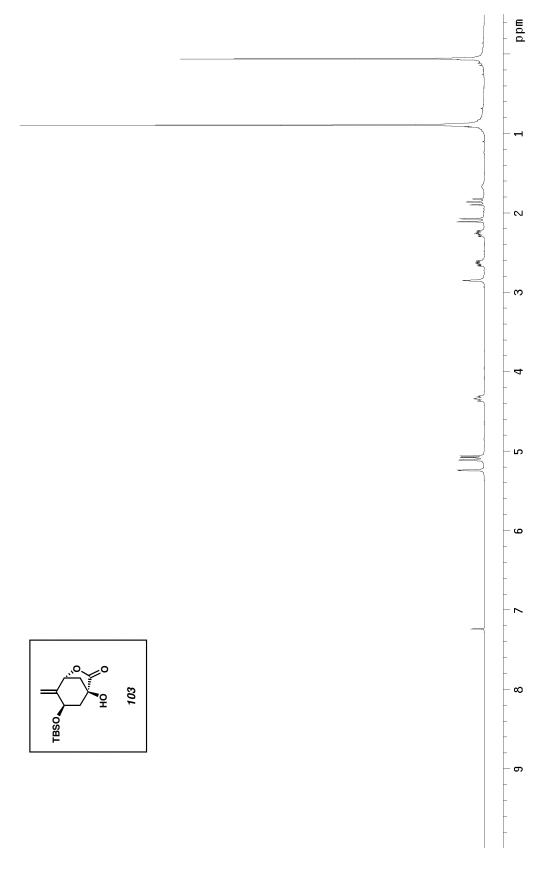


Figure A4.7  $^{1}$ H NMR (300 MHz, CDCl<sub>3</sub>) of compound 103.

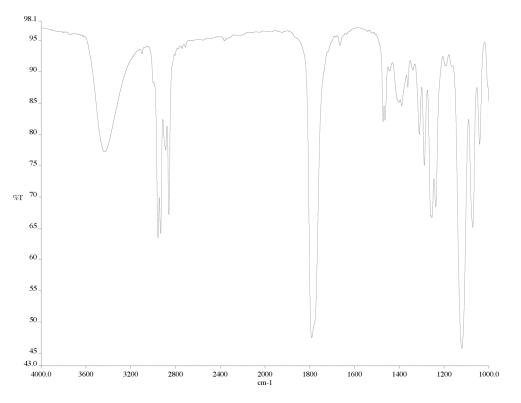
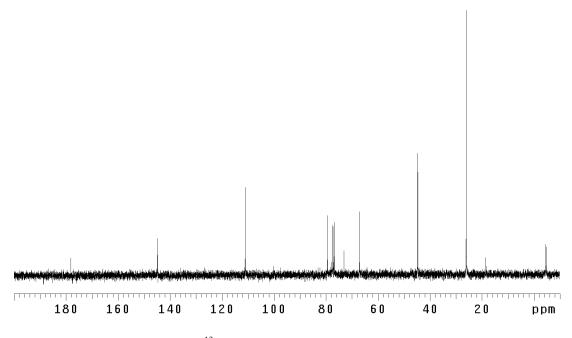
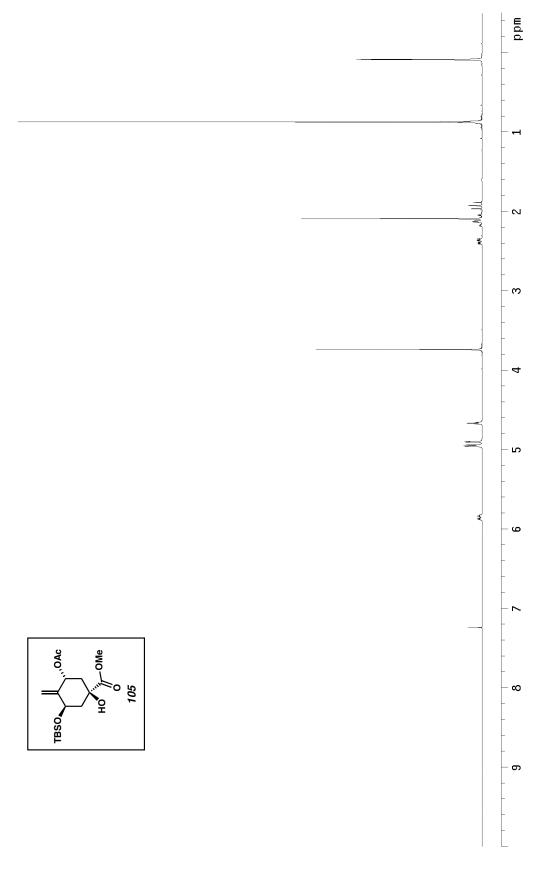


Figure A4.8 Infrared spectrum (thin film/NaCl) of compound 103.



*Figure A4.9* <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) of compound **103**.





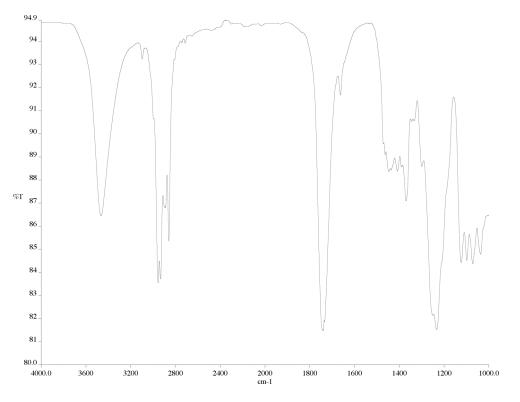
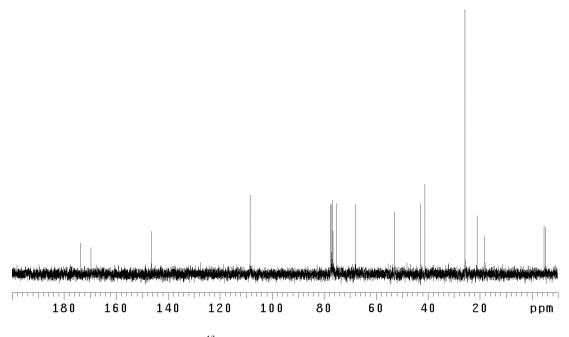
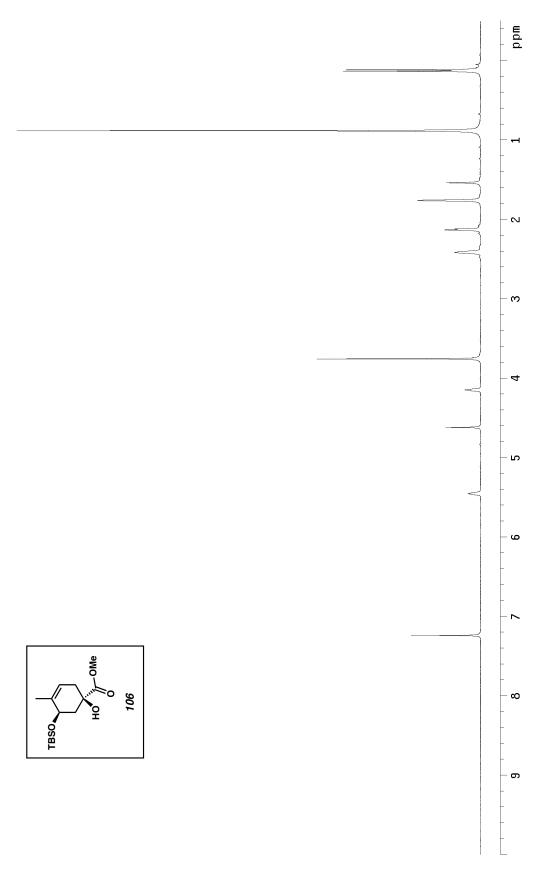


Figure A4.11 Infrared spectrum (thin film/NaCl) of compound 105.



*Figure A4.12* <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) of compound **105**.





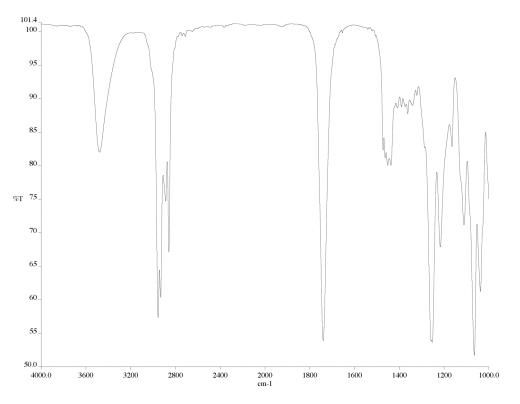
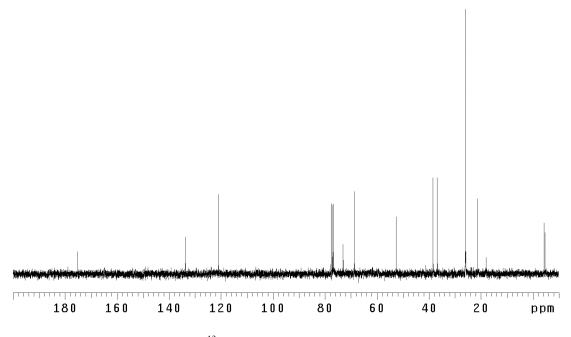
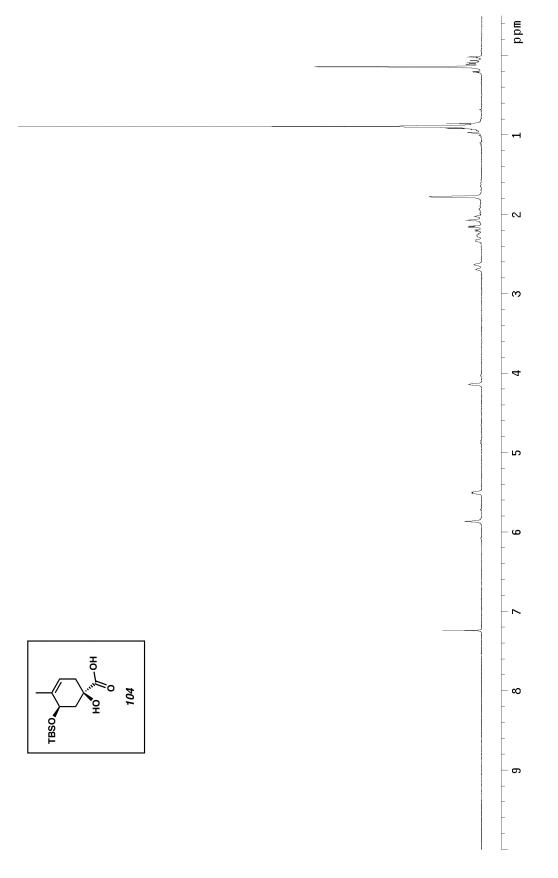


Figure A4.14 Infrared spectrum (thin film/NaCl) of compound 106.



*Figure A4.15* <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) of compound **106**.





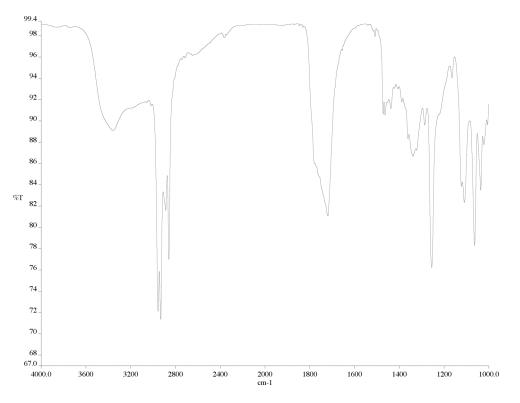
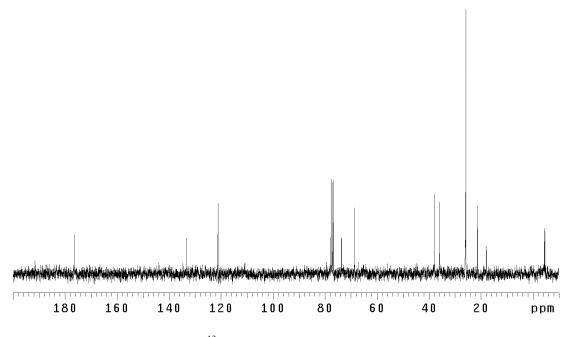
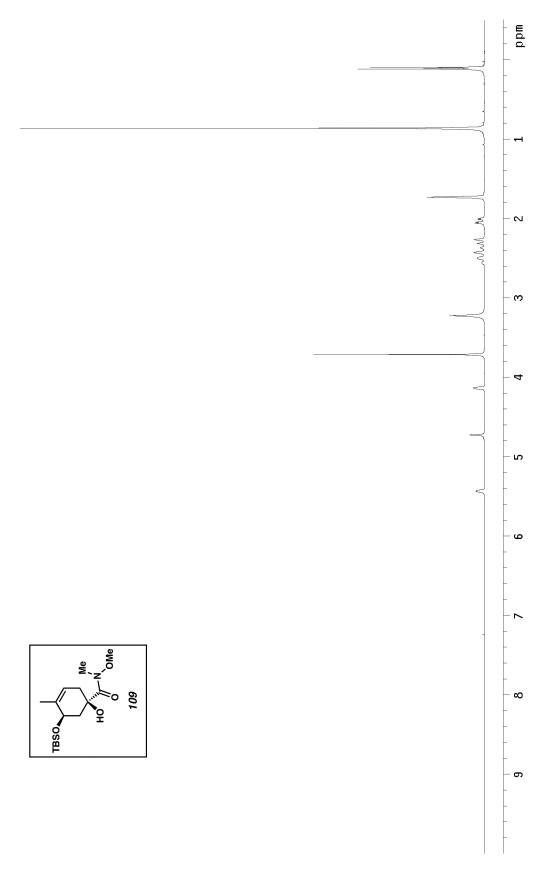


Figure A4.17 Infrared spectrum (thin film/NaCl) of compound 104.



*Figure A4.18* <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) of compound **104**.





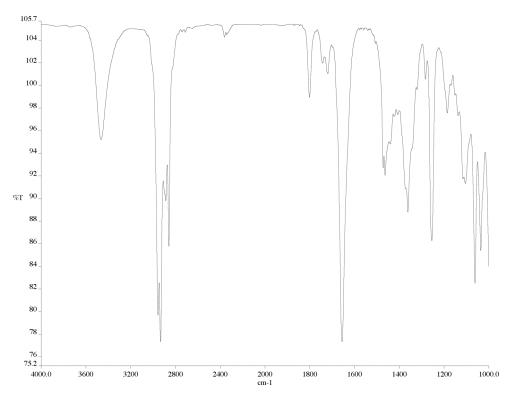
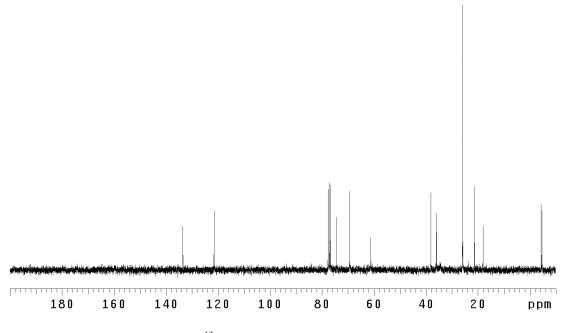
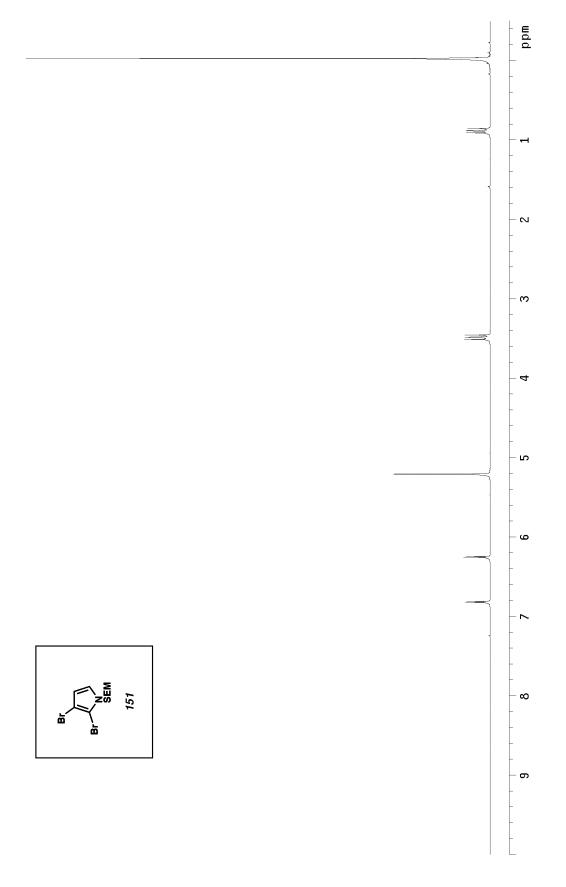


Figure A4.20 Infrared spectrum (thin film/NaCl) of compound 109.



*Figure A4.21*  $^{13}$ C NMR (75 MHz, CDCl<sub>3</sub>) of compound **109**.





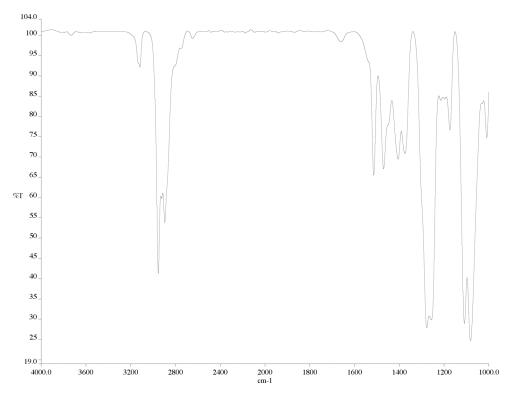
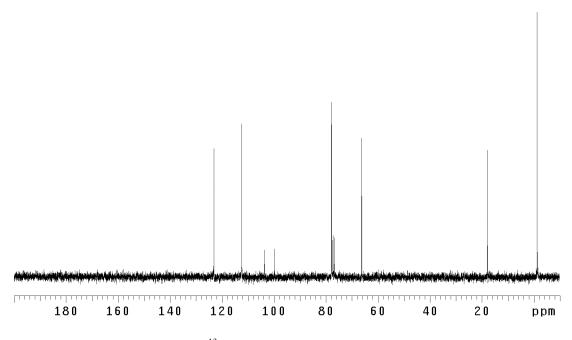
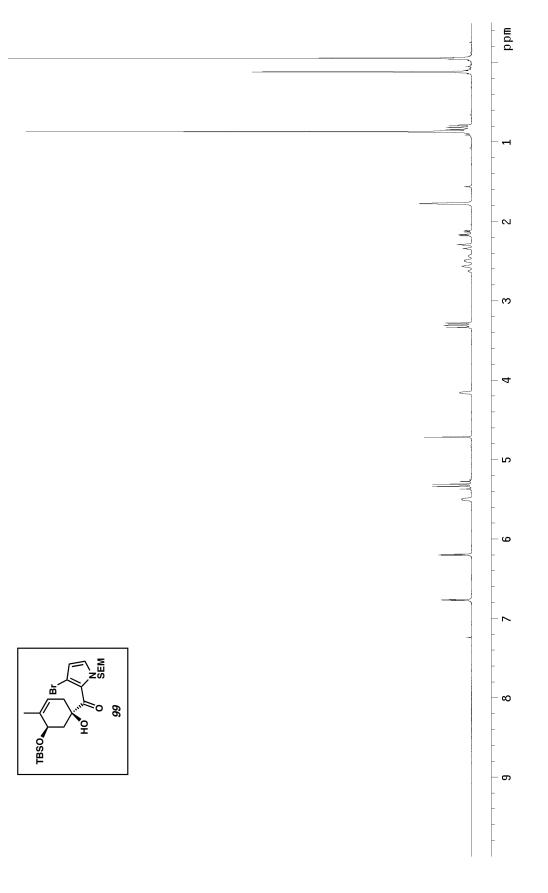


Figure A4.23 Infrared spectrum (thin film/NaCl) of compound 151.



*Figure A4.24* <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) of compound **151**.





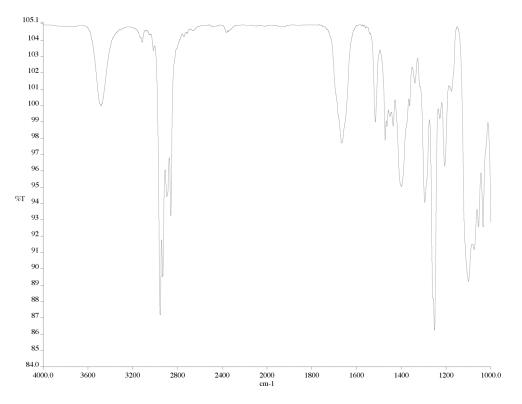
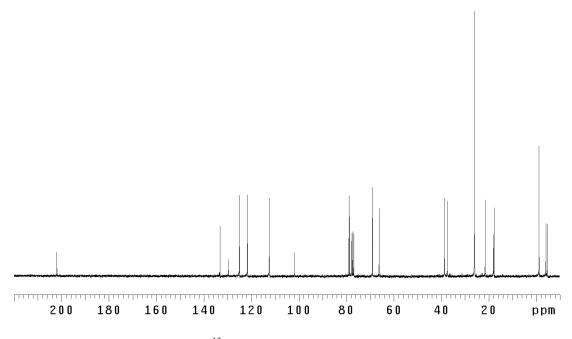
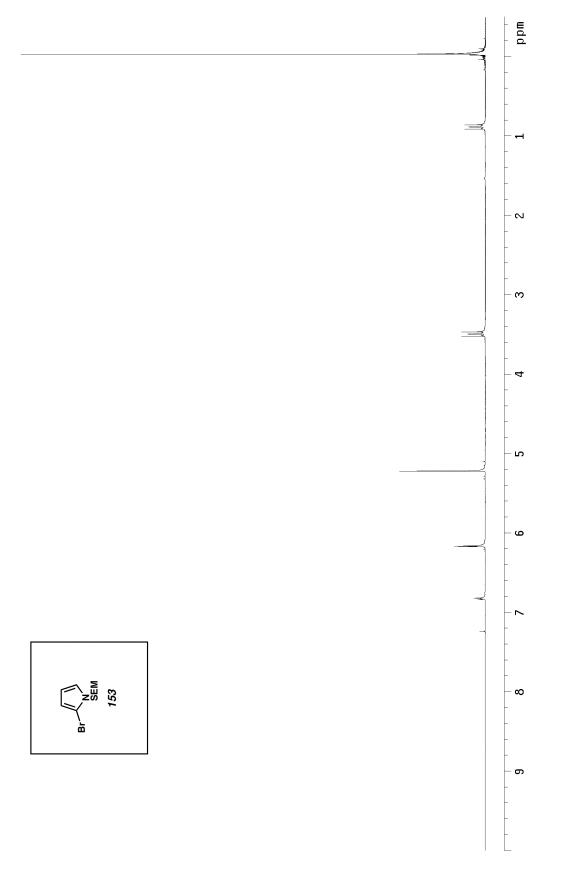


Figure A4.26 Infrared spectrum (thin film/NaCl) of compound 99.



*Figure A4.27*  $^{13}$ C NMR (75 MHz, CDCl<sub>3</sub>) of compound **99**.





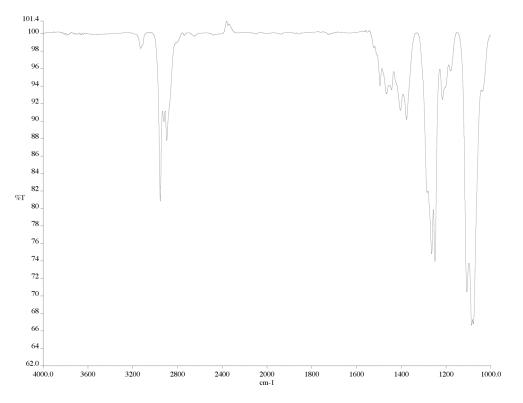
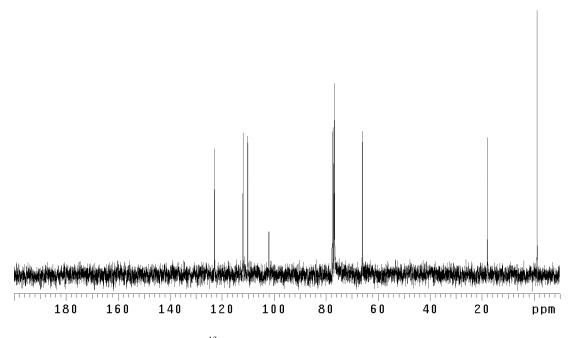
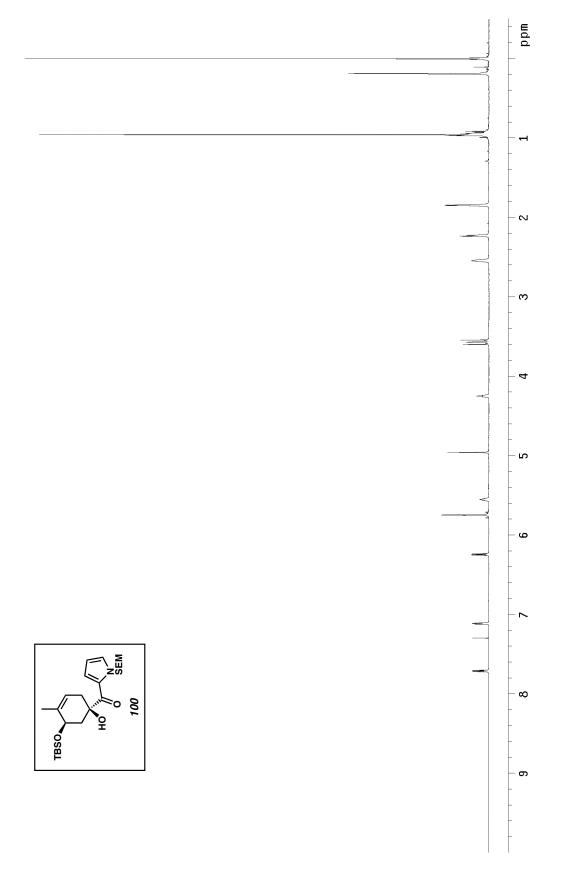


Figure A4.29 Infrared spectrum (thin film/NaCl) of compound 153.



*Figure A4.30* <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) of compound **153**.





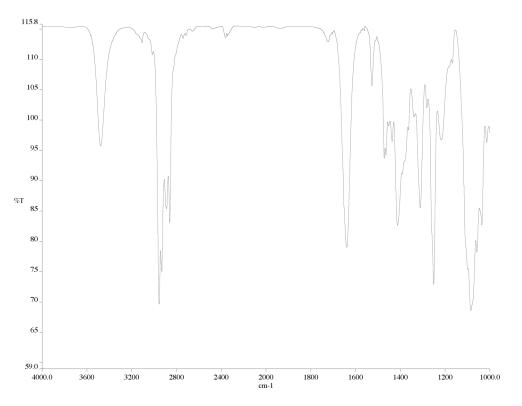
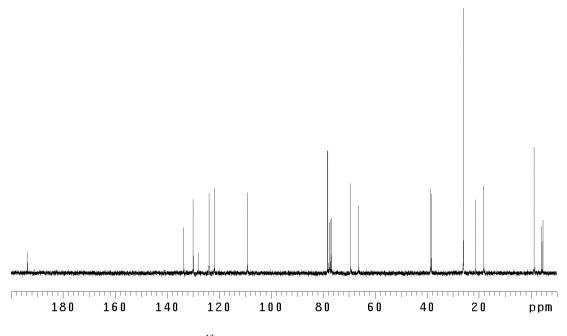
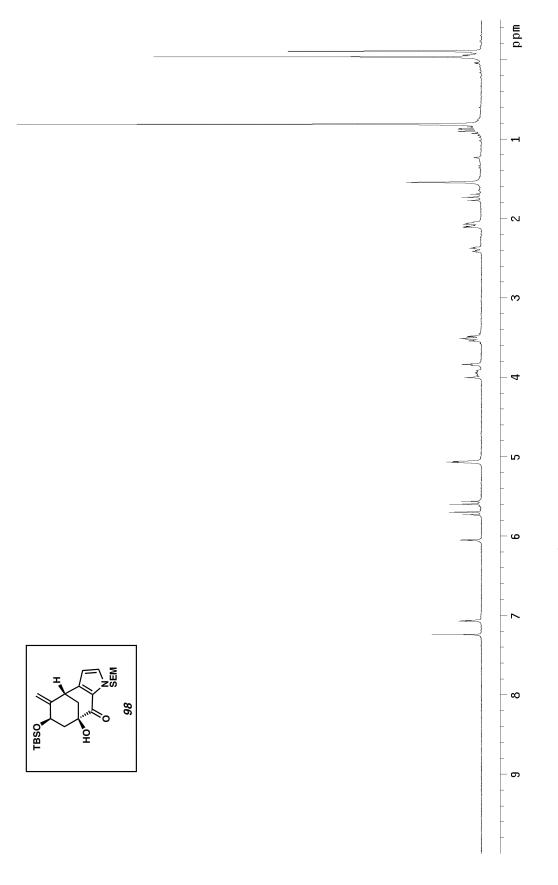


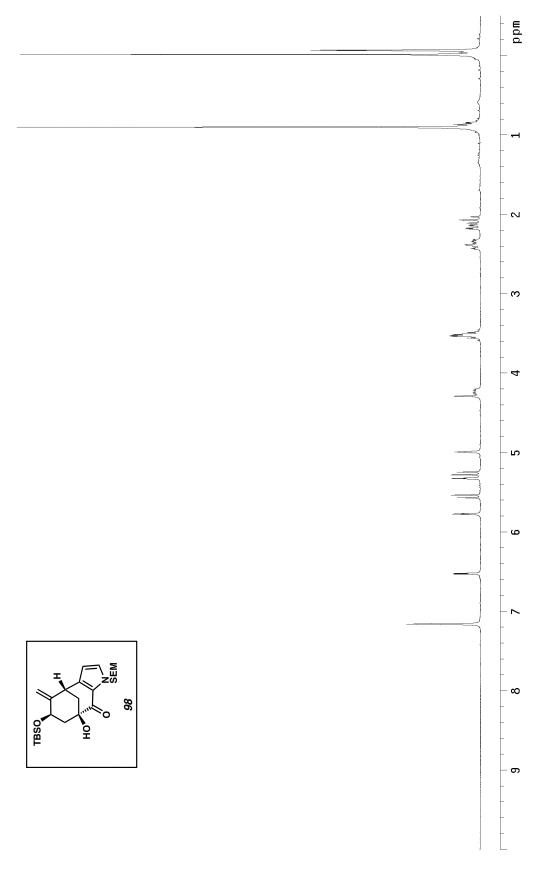
Figure A4.32 Infrared spectrum (thin film/NaCl) of compound 100.



*Figure A4.33*  $^{13}$ C NMR (75 MHz, CDCl<sub>3</sub>) of compound **100**.









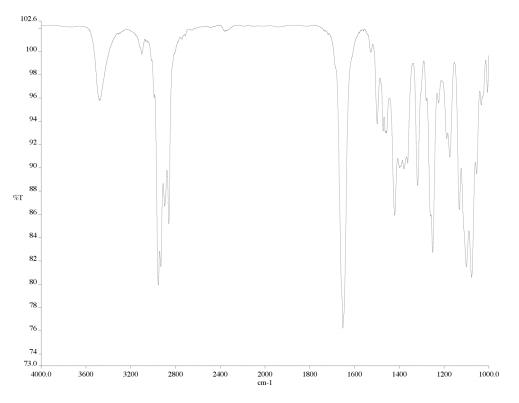
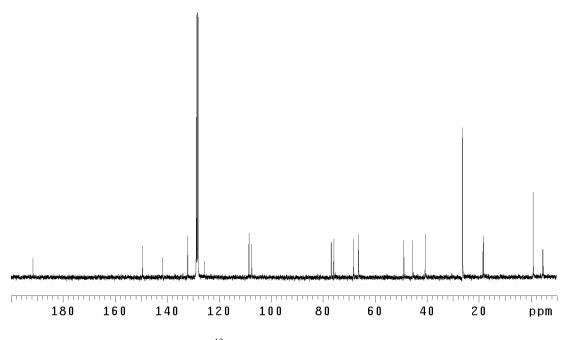
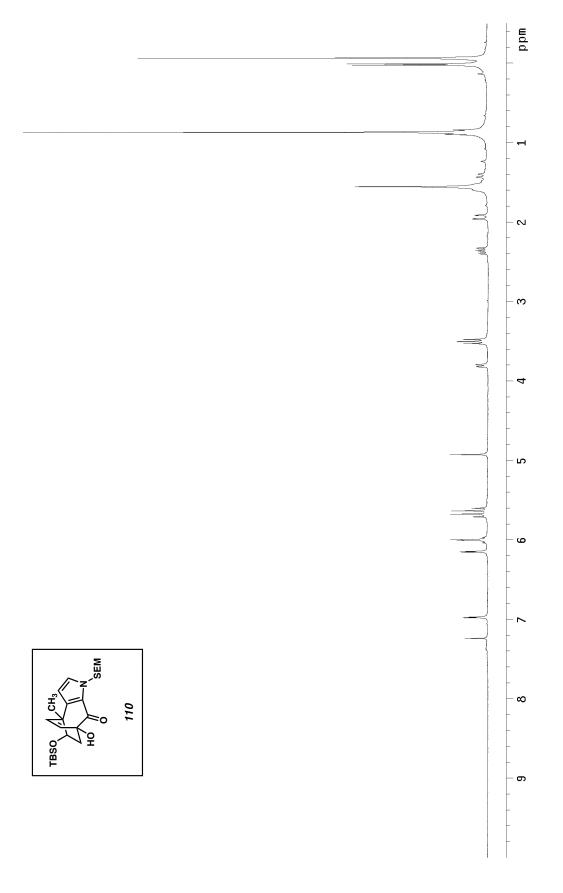


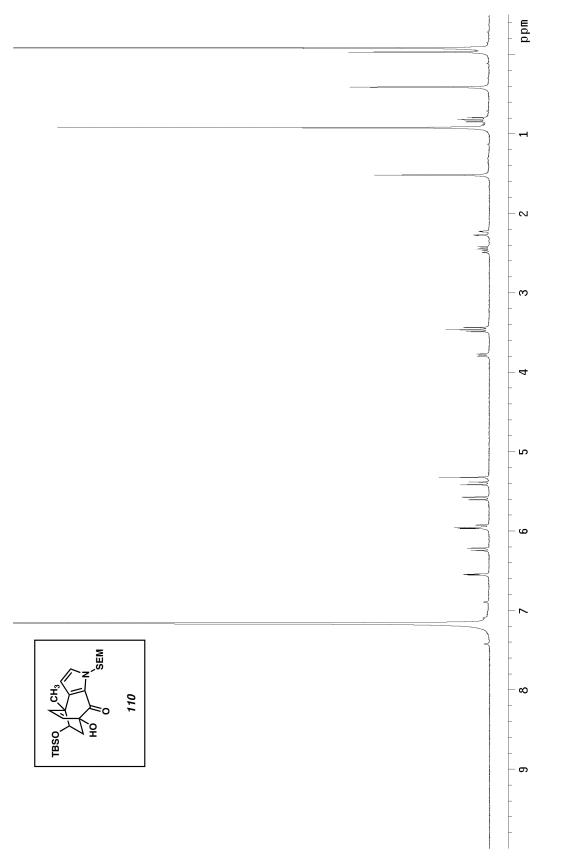
Figure A4.36 Infrared spectrum (thin film/NaCl) of compound 98.



*Figure A4.37*  $^{13}$ C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>) of compound **98**.









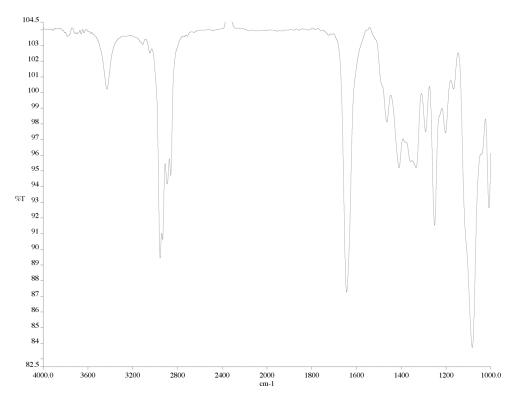
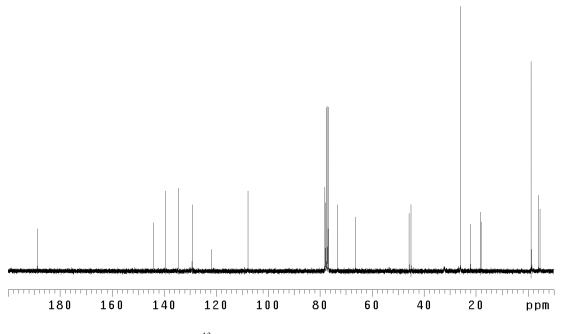
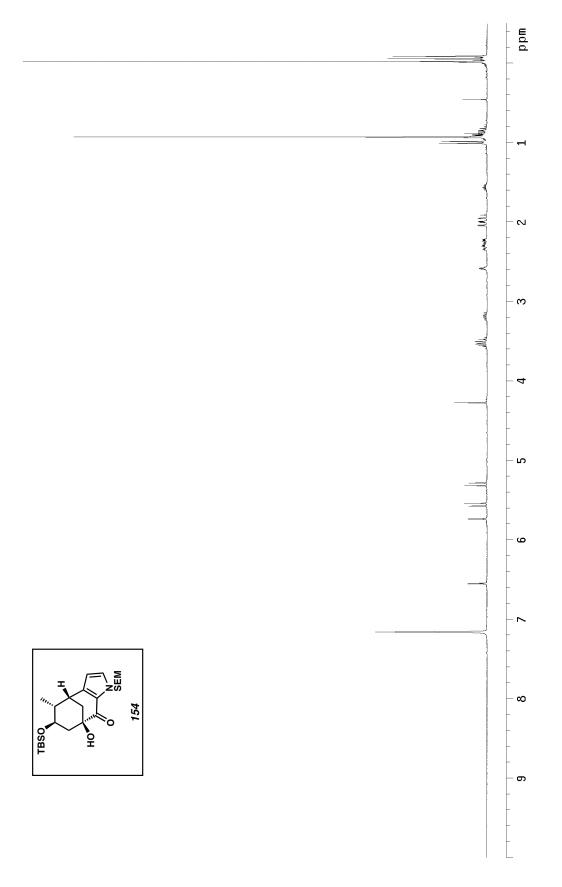


Figure A4.40 Infrared spectrum (thin film/NaCl) of compound 110.



*Figure A4.41*  $^{13}$ C NMR (75 MHz, CDCl<sub>3</sub>) of compound **110**.





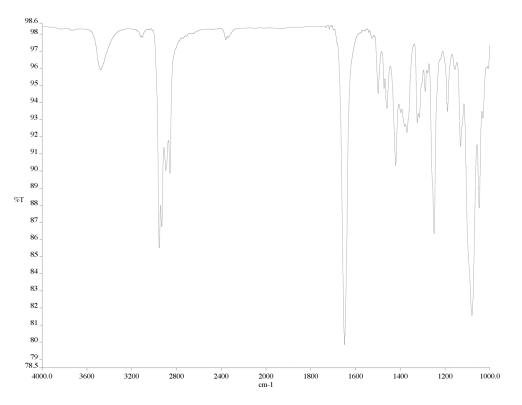
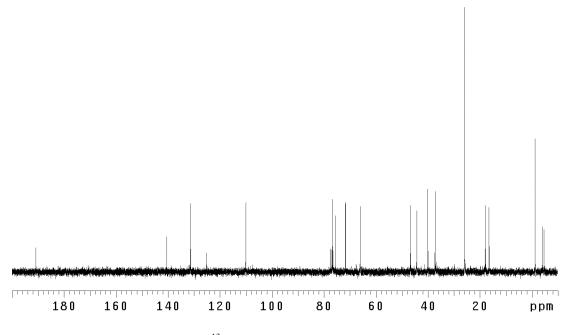
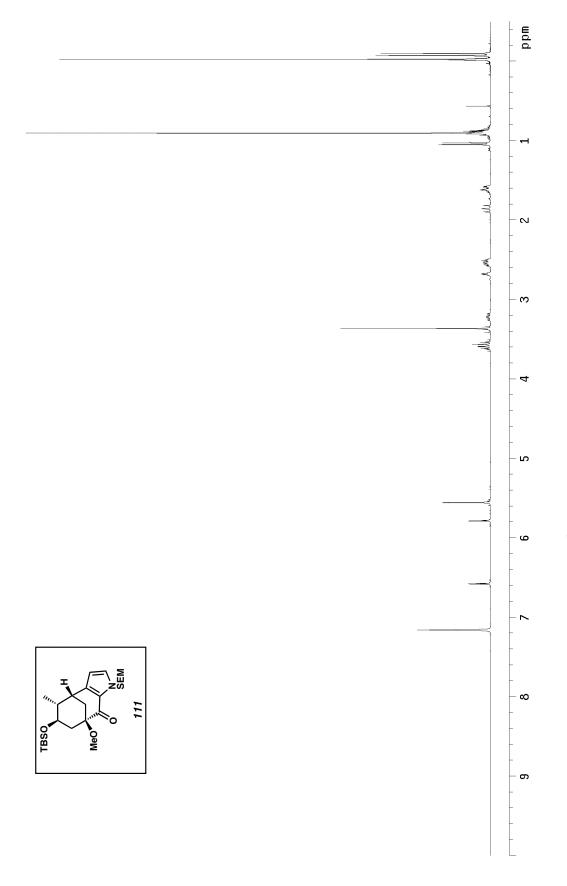


Figure A4.43 Infrared spectrum (thin film/NaCl) of compound 154.



*Figure A4.44*  $^{13}$ C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>) of compound **154**.





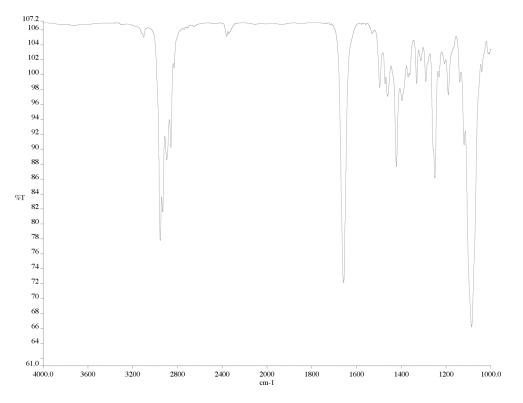
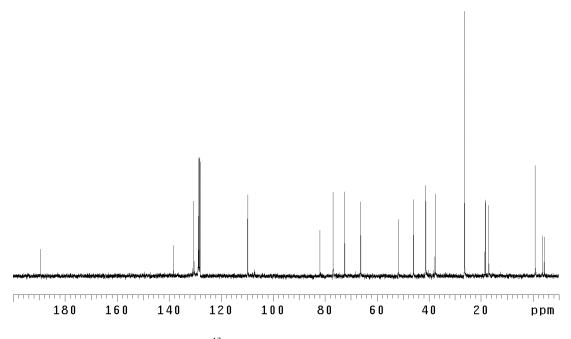
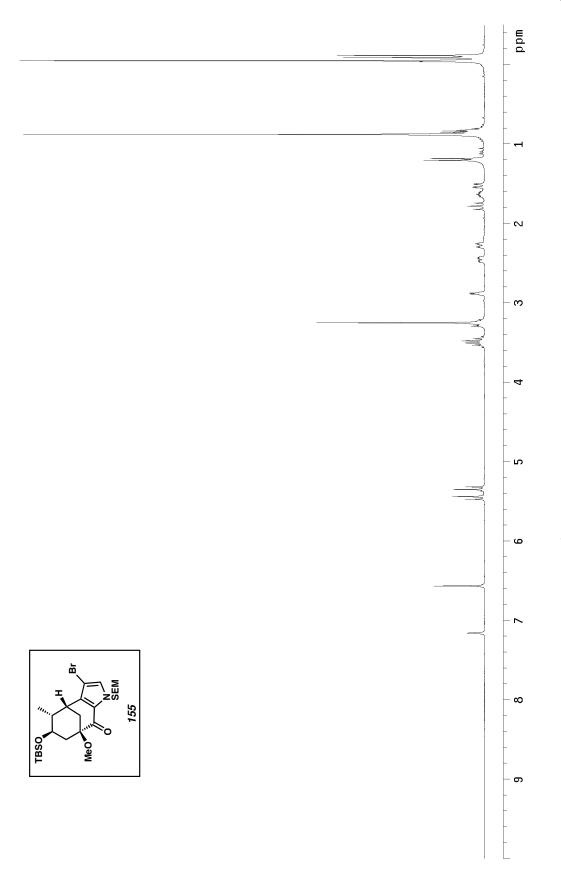


Figure A4.46 Infrared spectrum (thin film/NaCl) of compound 111.



*Figure A4.47*  $^{13}$ C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>) of compound **111**.





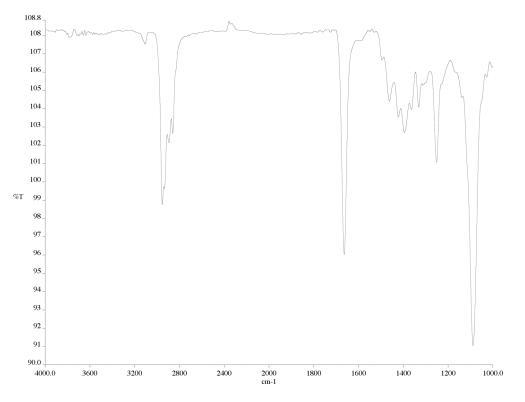
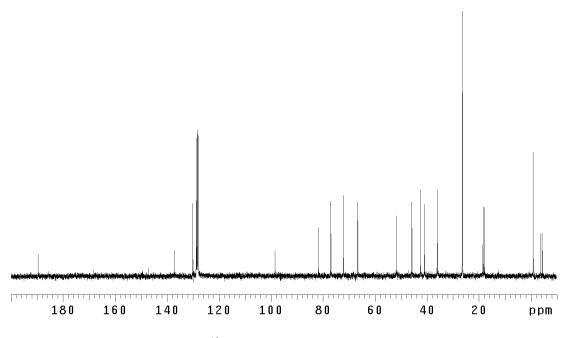
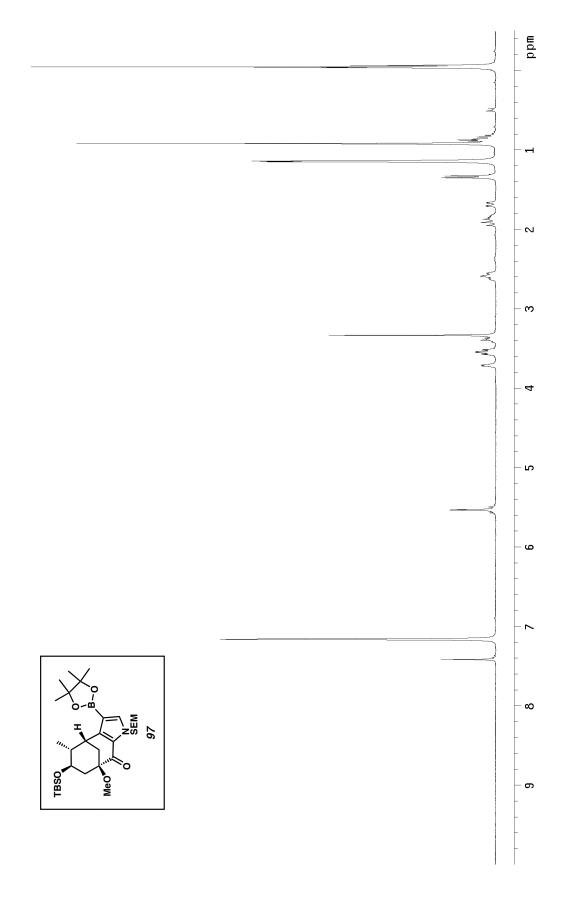


Figure A4.49 Infrared spectrum (thin film/NaCl) of compound 155.



*Figure A4.50*  $^{13}$ C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>) of compound **155**.





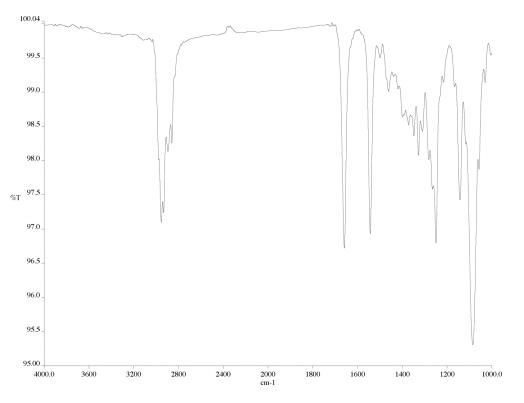
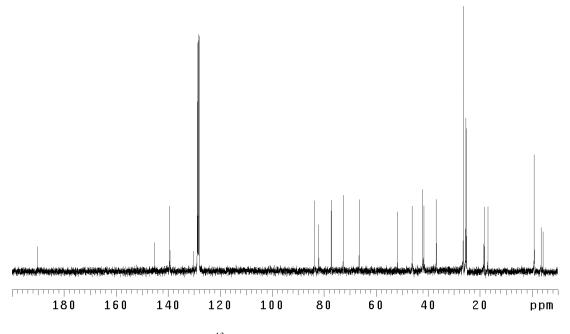
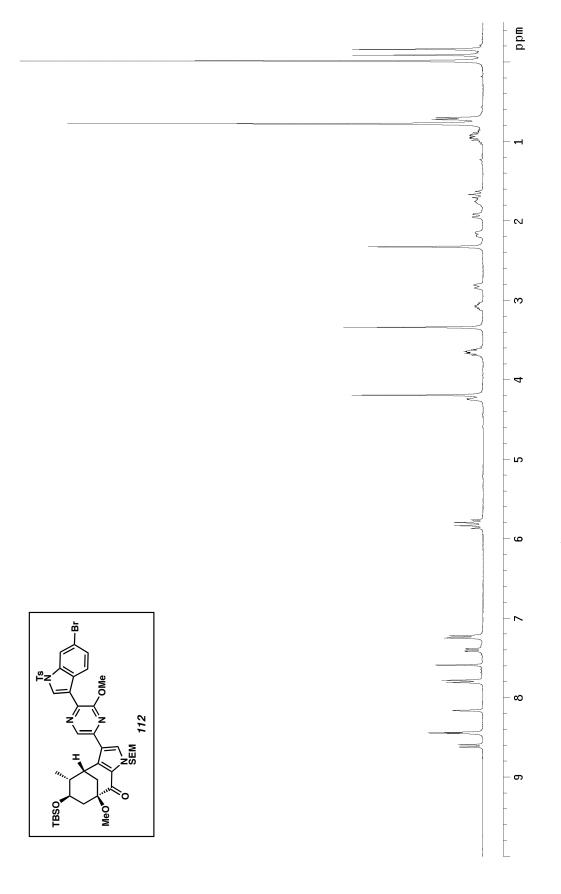


Figure A4.52 Infrared spectrum (thin film/NaCl) of compound 97.



*Figure A4.53*  $^{13}$ C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>) of compound **97**.





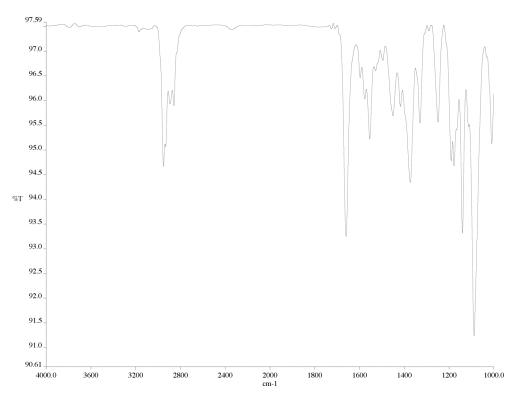
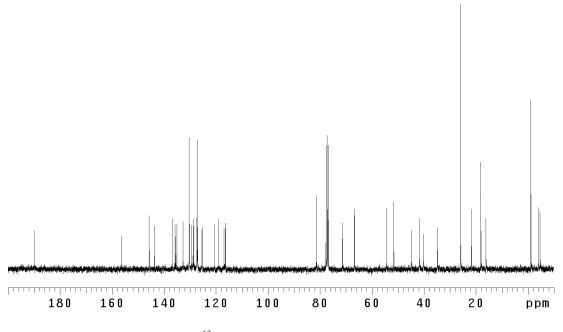
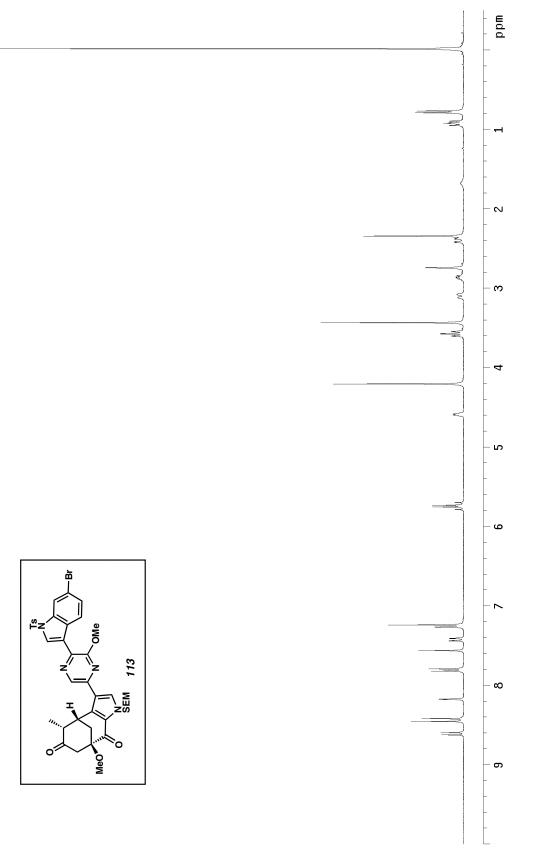


Figure A4.55 Infrared spectrum (thin film/NaCl) of compound 112.



*Figure A4.56* <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) of compound **112**.





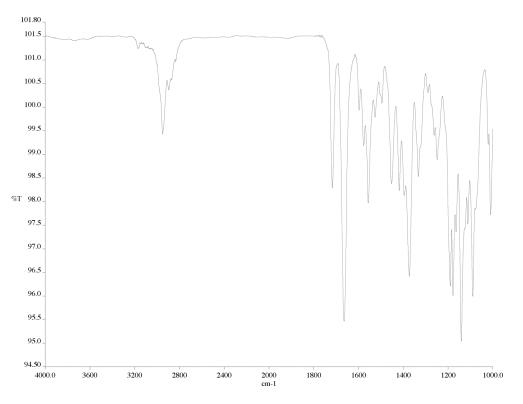
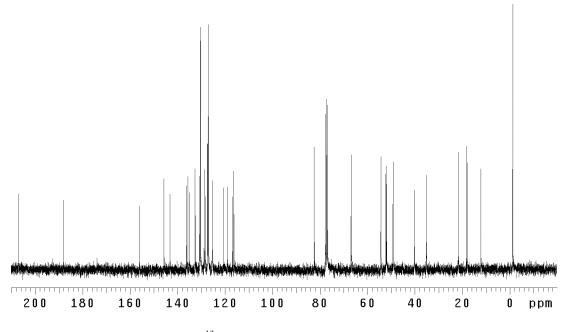
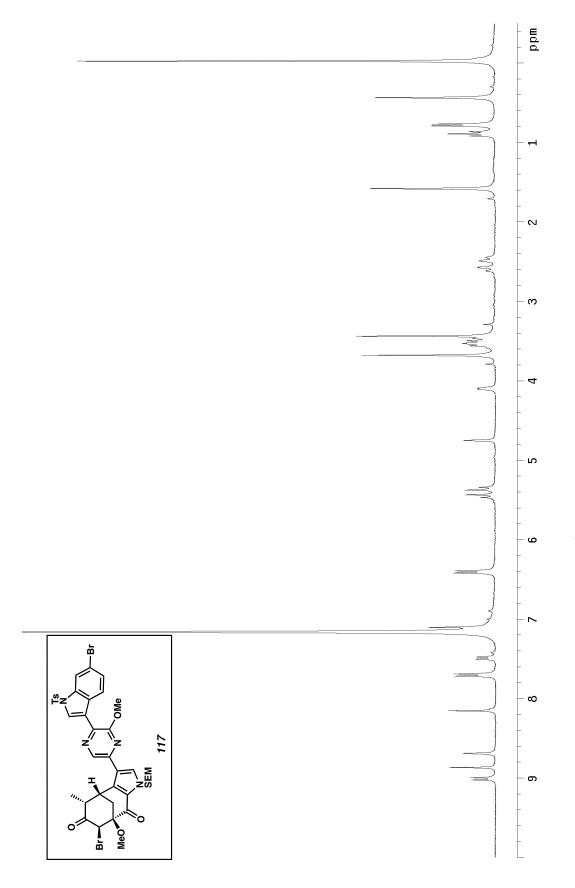


Figure A4.58 Infrared spectrum (thin film/NaCl) of compound 113.



*Figure A4.59* <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) of compound **113**.





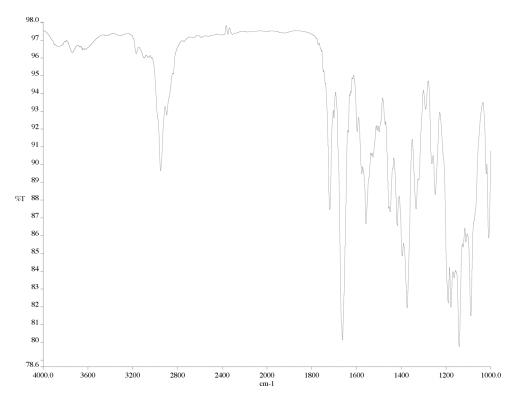
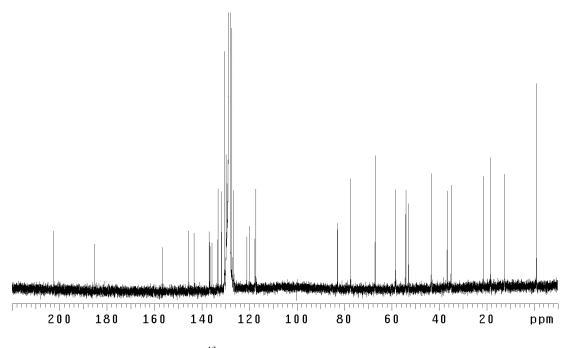
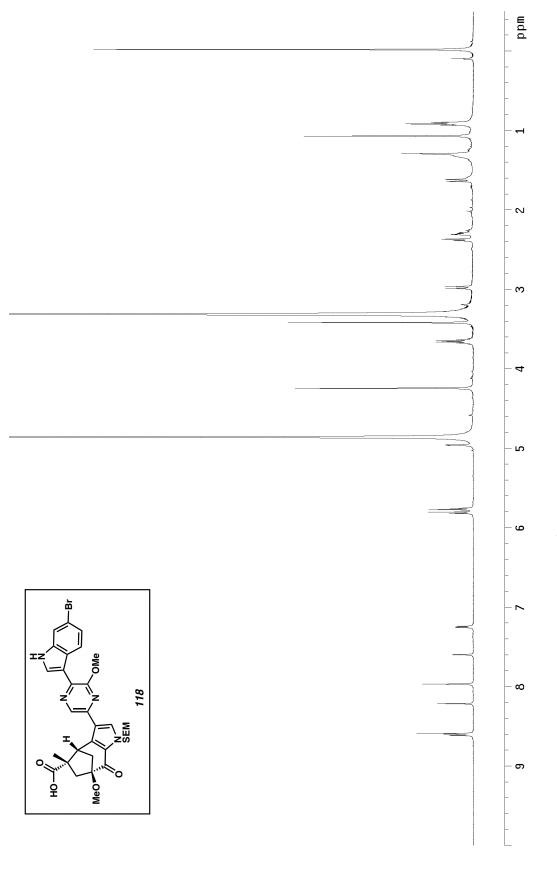


Figure A4.61 Infrared spectrum (thin film/NaCl) of compound 117.



*Figure A4.62*  $^{13}$ C NMR (125 MHz, C<sub>6</sub>D<sub>6</sub>) of compound **117**.





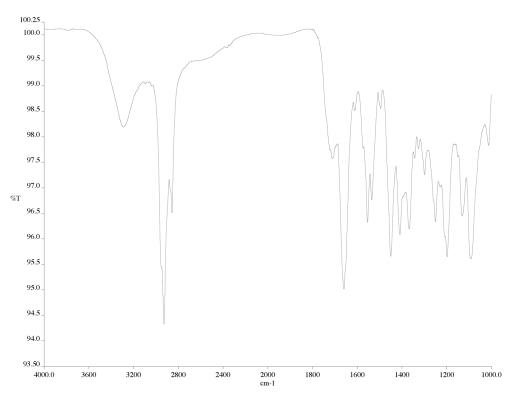
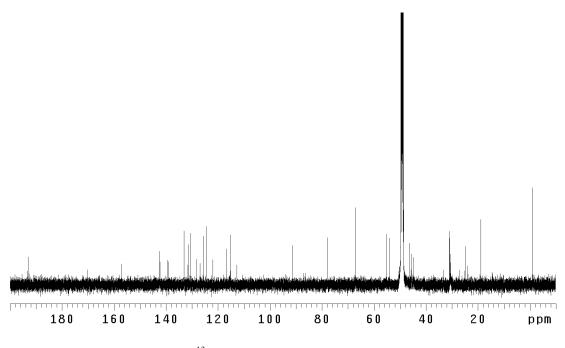
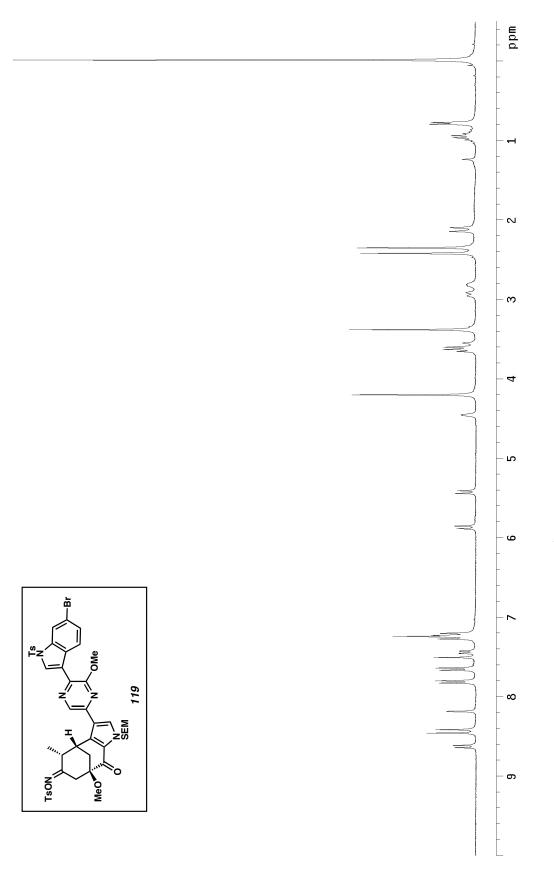


Figure A4.64 Infrared spectrum (thin film/NaCl) of compound 118.



*Figure A4.65*  $^{13}$ C NMR (125 MHz, CD<sub>3</sub>OD) of compound **118**.





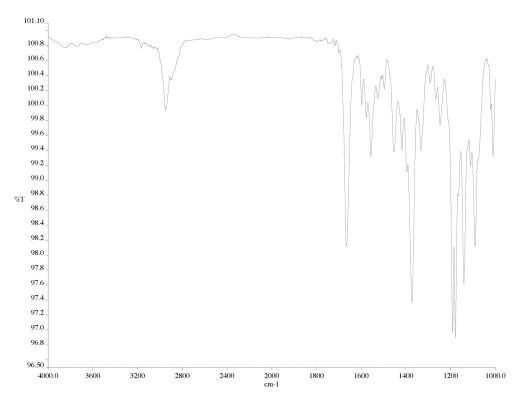
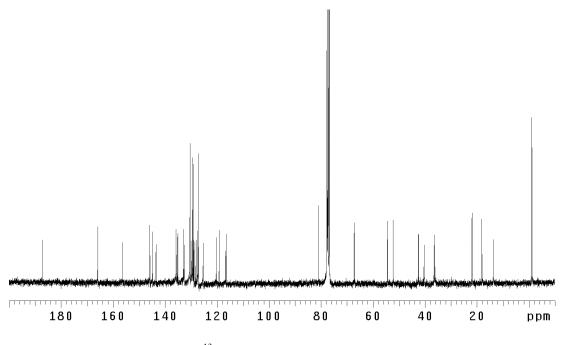
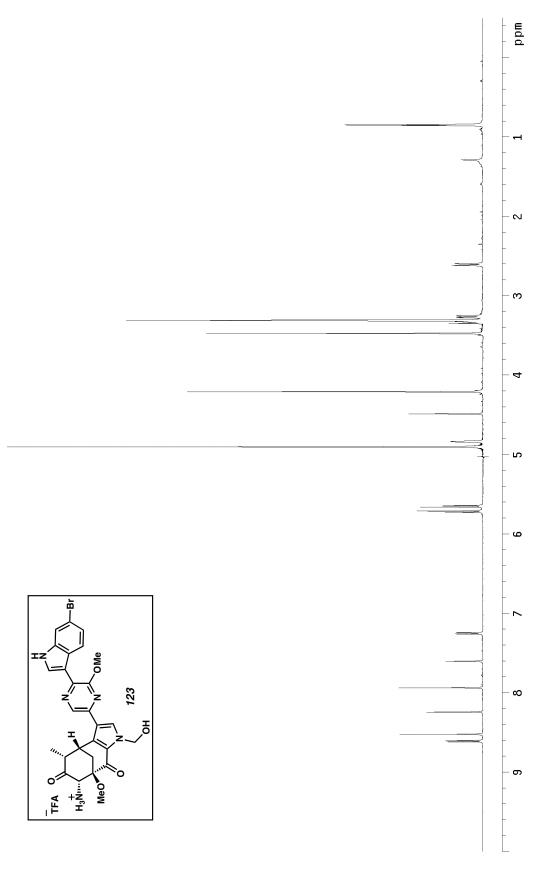


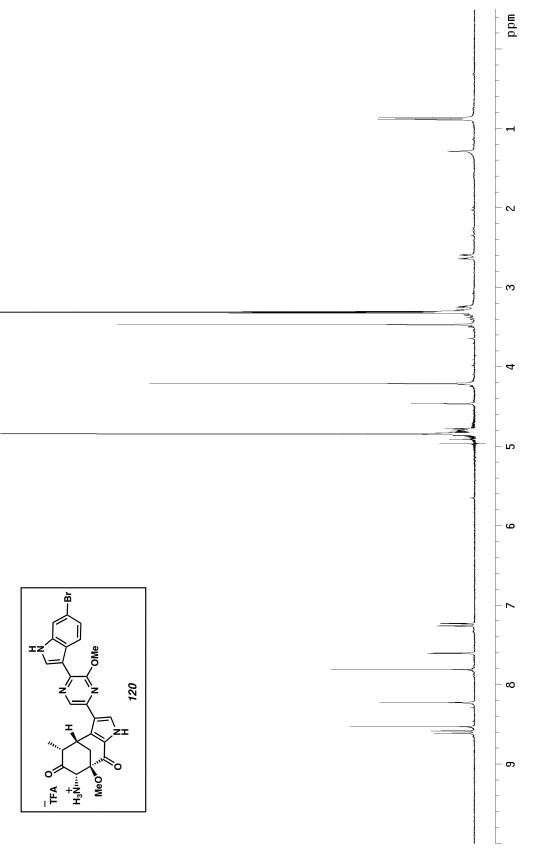
Figure A4.67 Infrared spectrum (thin film/NaCl) of compound 119.



*Figure A4.68* <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) of compound **119**.









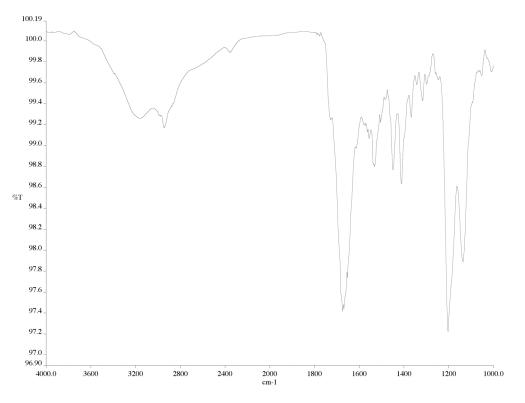
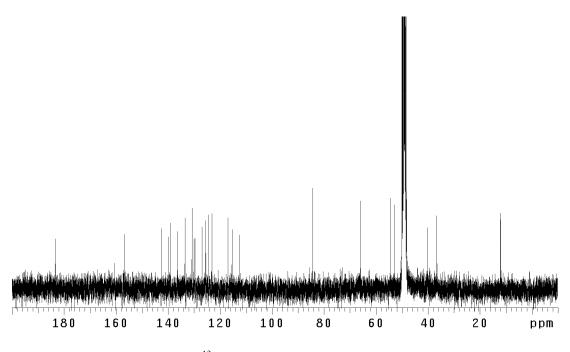
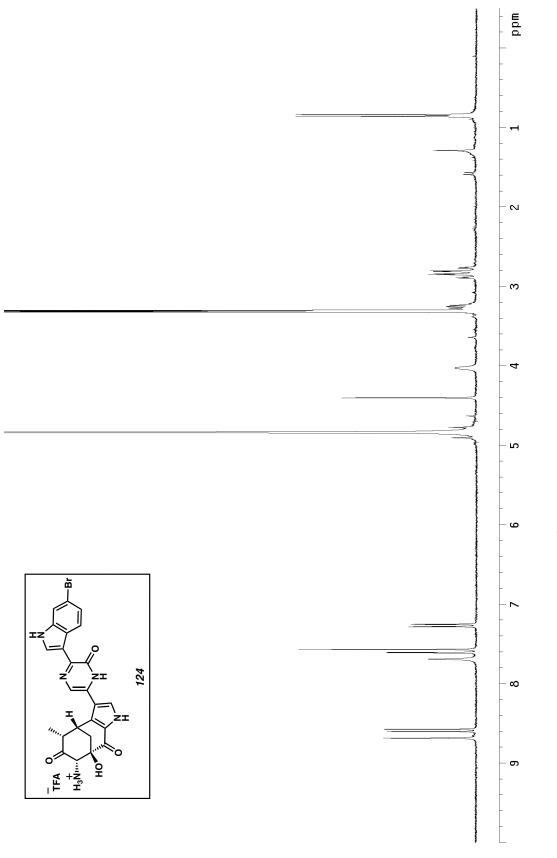


Figure A4.71 Infrared spectrum (thin film/NaCl) of compound 120.



*Figure A4.72*  $^{13}$ C NMR (75 MHz, CD<sub>3</sub>OD) of compound **120**.





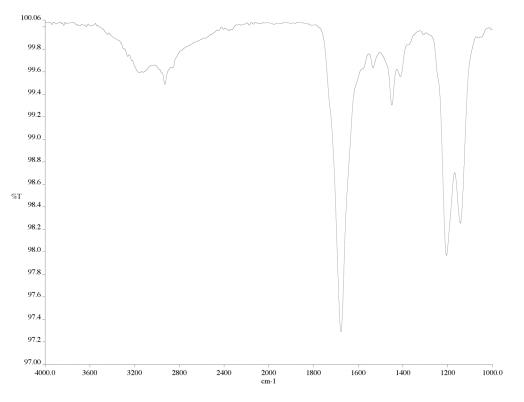
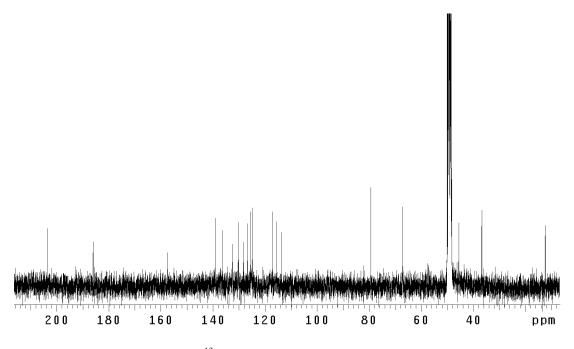
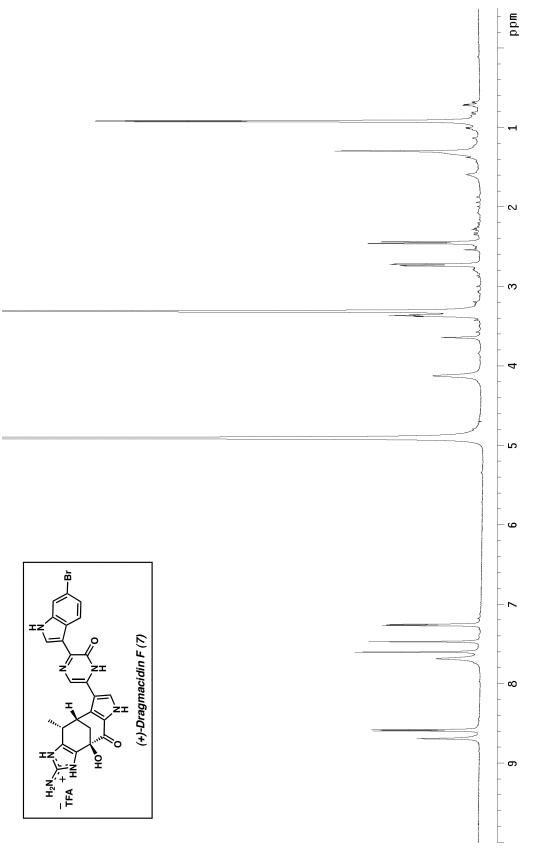


Figure A4.74 Infrared spectrum (thin film/NaCl) of compound 124.



*Figure A4.75* <sup>13</sup>C NMR (75 MHz, CD<sub>3</sub>OD) of compound **124**.





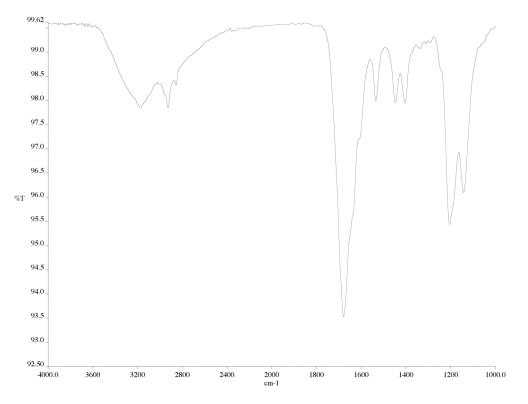
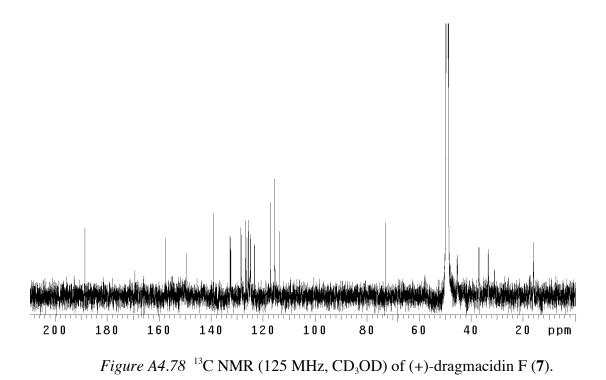
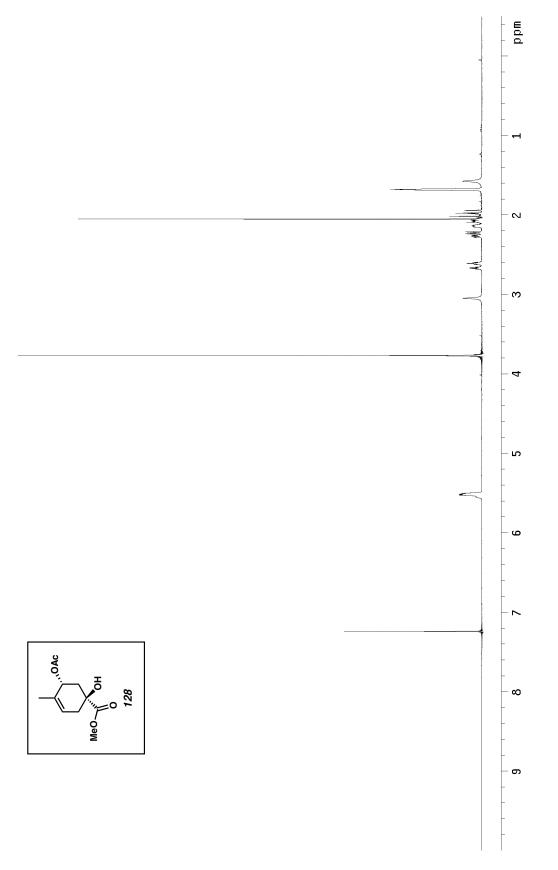


Figure A4.77 Infrared spectrum (thin film/NaCl) of (+)-dragmacidin F (7).







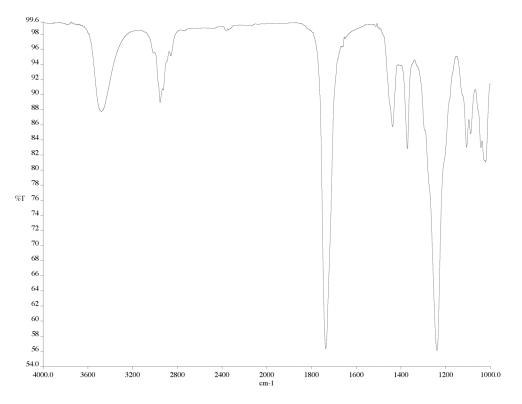
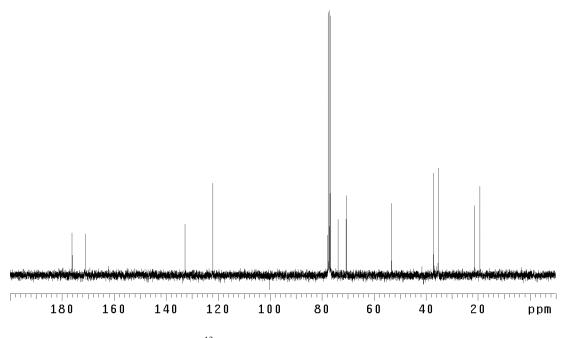
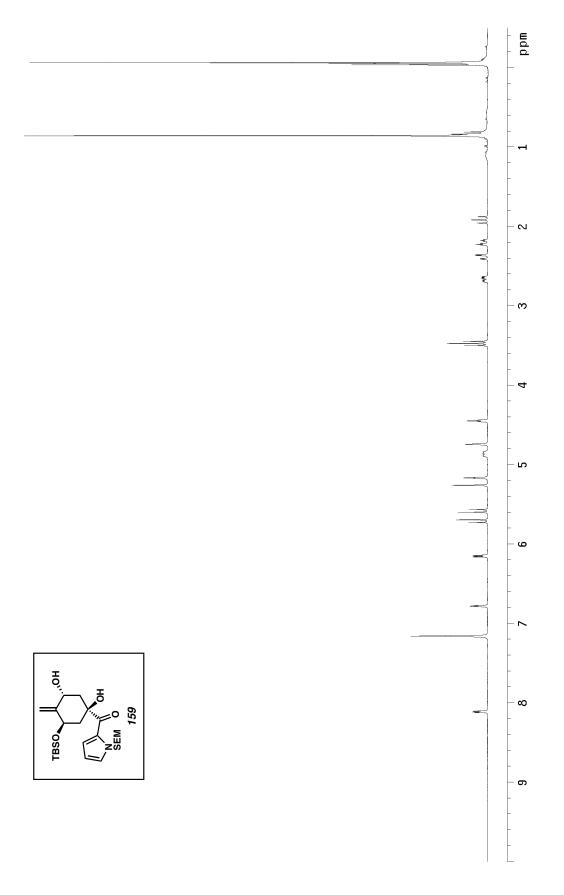


Figure A4.80 Infrared spectrum (thin film/NaCl) of compound 128.



*Figure A4.81* <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) of compound **128**.





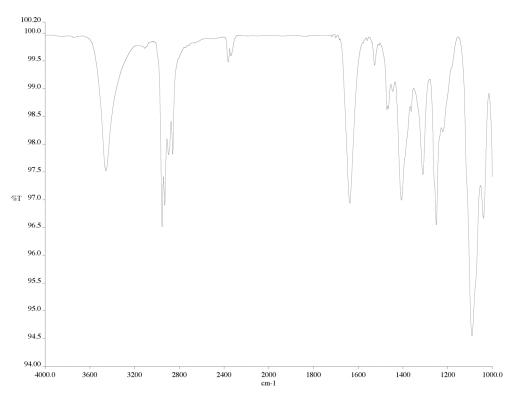
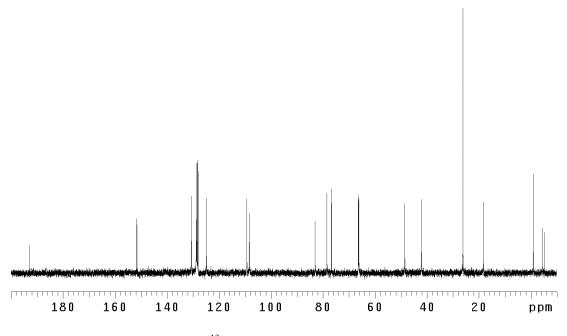
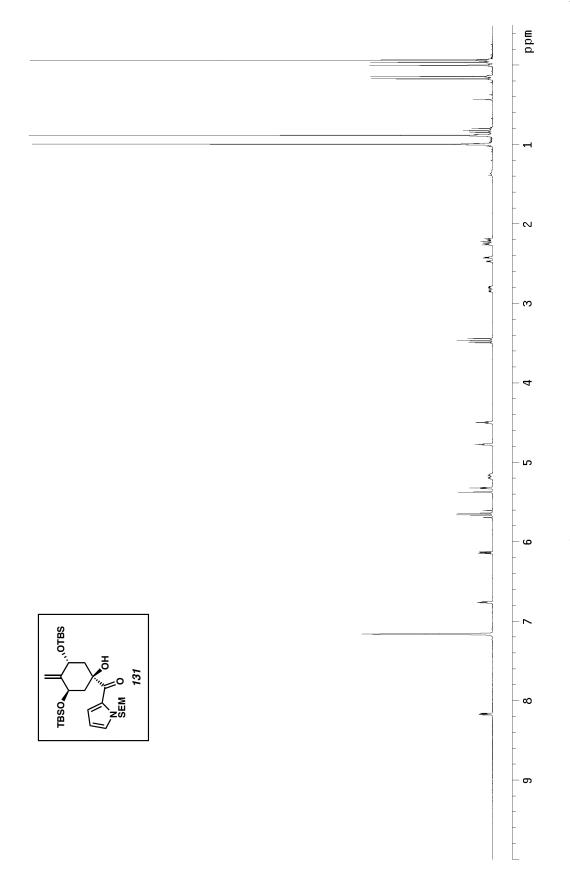


Figure A4.83 Infrared spectrum (thin film/NaCl) of compound 159.



*Figure A4.84*  $^{13}$ C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>) of compound **159**.





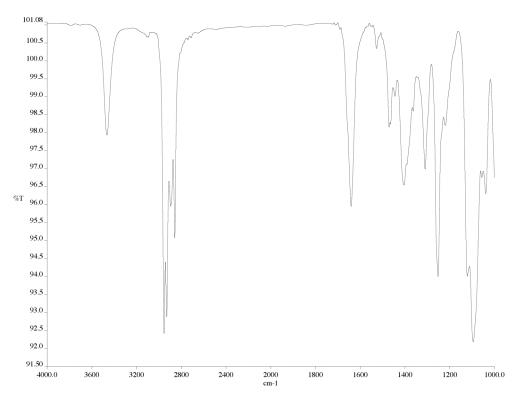
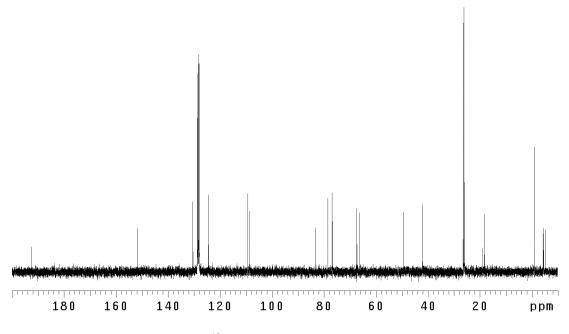
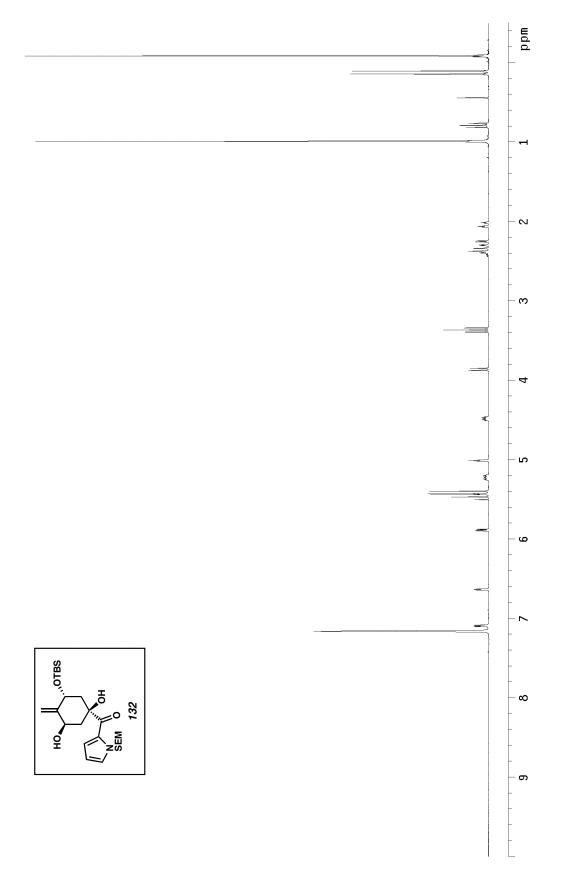


Figure A4.86 Infrared spectrum (thin film/NaCl) of compound 131.



*Figure A4.87*  $^{13}$ C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>) of compound **131**.





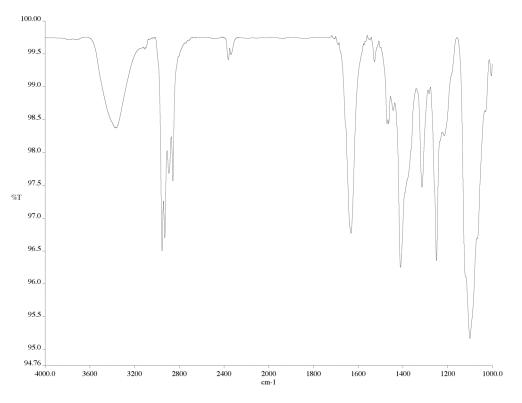
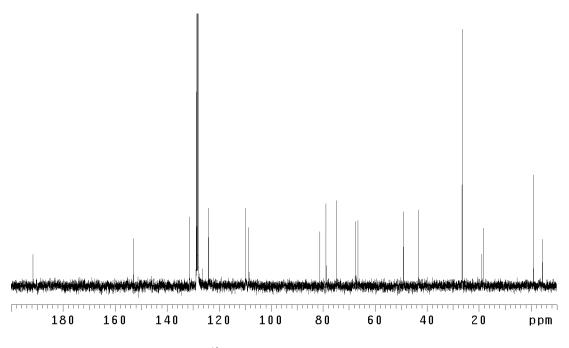
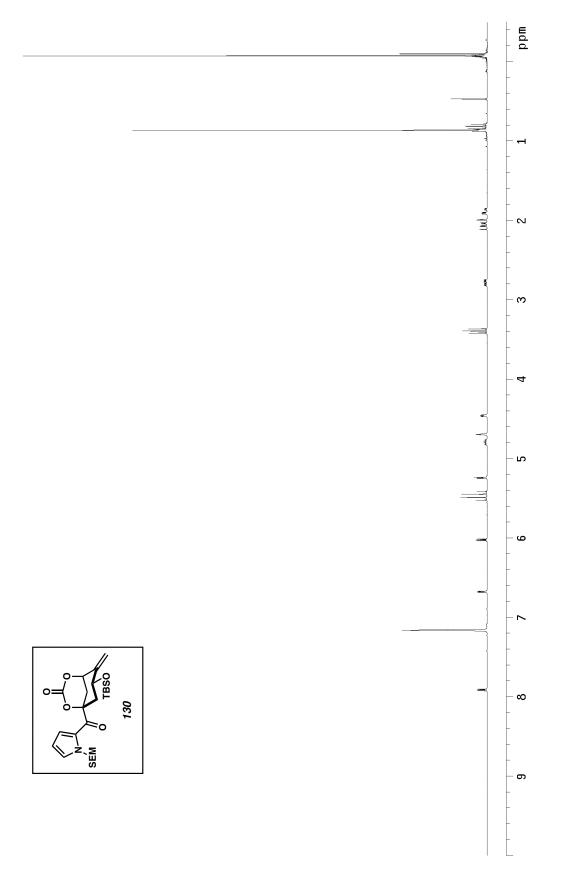


Figure A4.89 Infrared spectrum (thin film/NaCl) of compound 132.



*Figure A4.90*  $^{13}$ C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>) of compound **132**.





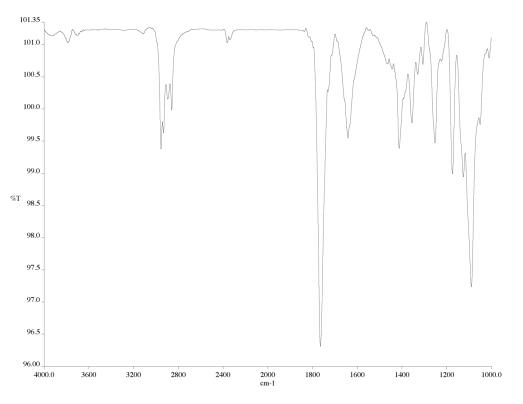
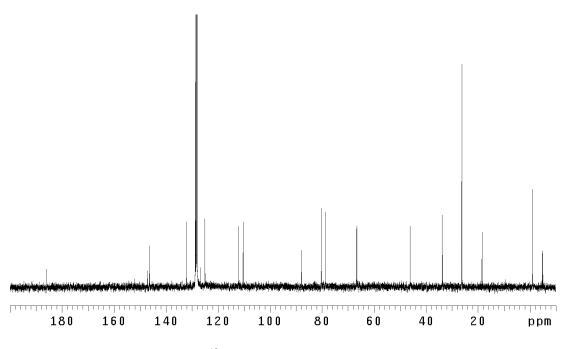
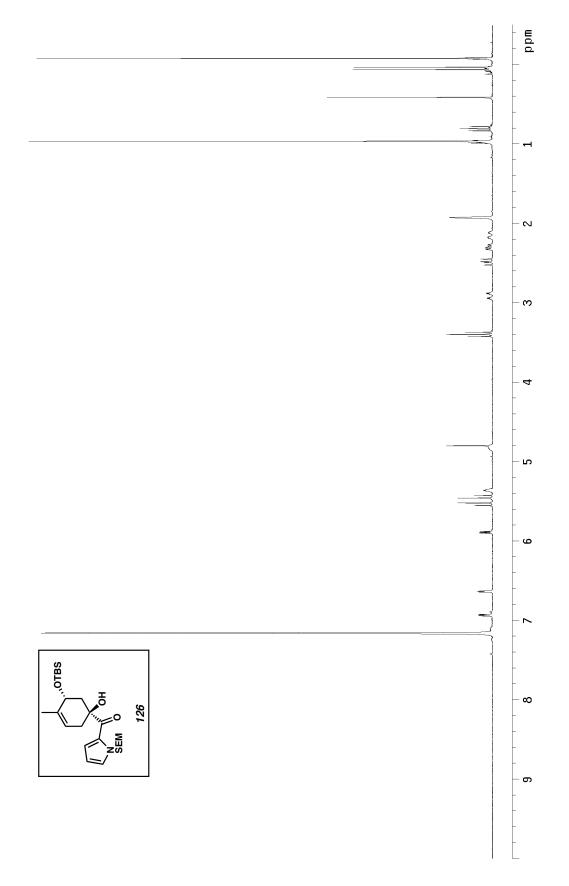


Figure A4.92 Infrared spectrum (thin film/NaCl) of compound 130.



*Figure A4.93*  $^{13}$ C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>) of compound **130**.





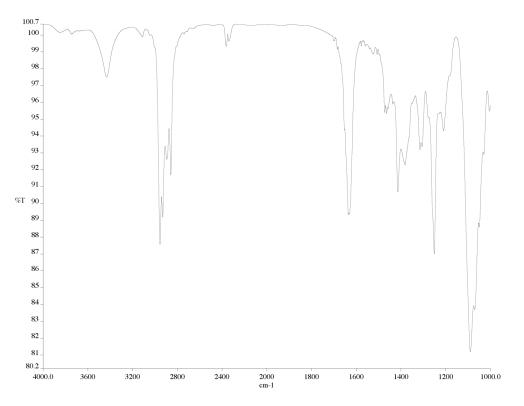
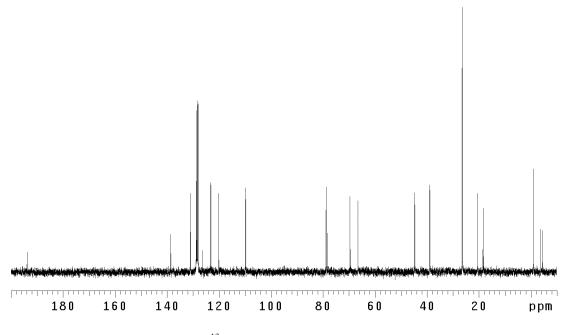
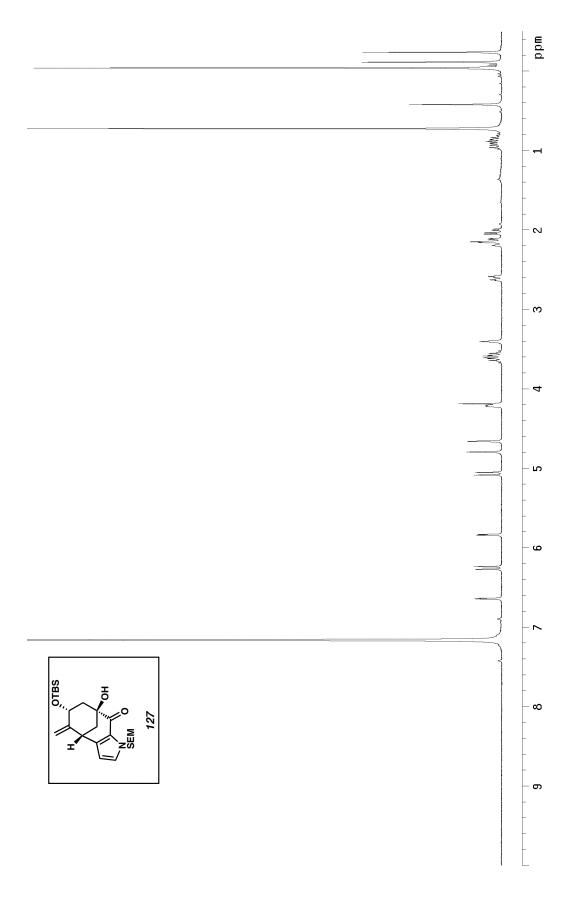


Figure A4.95 Infrared spectrum (thin film/NaCl) of compound 126.



*Figure A4.96*  $^{13}$ C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>) of compound **126**.





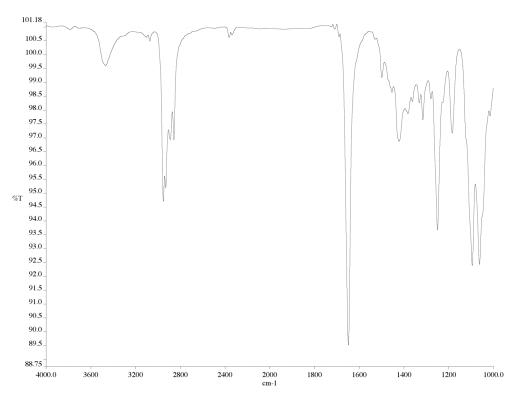
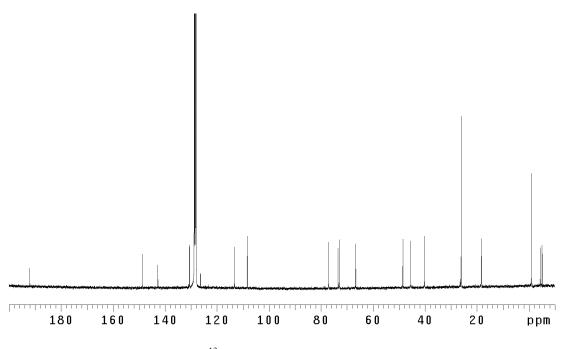
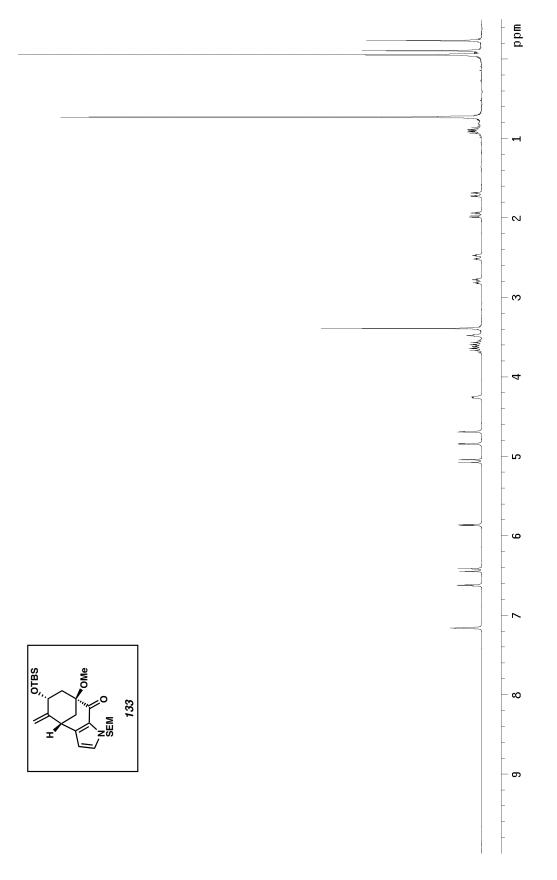


Figure A4.98 Infrared spectrum (thin film/NaCl) of compound 127.



*Figure A4.99*  $^{13}$ C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>) of compound **127**.





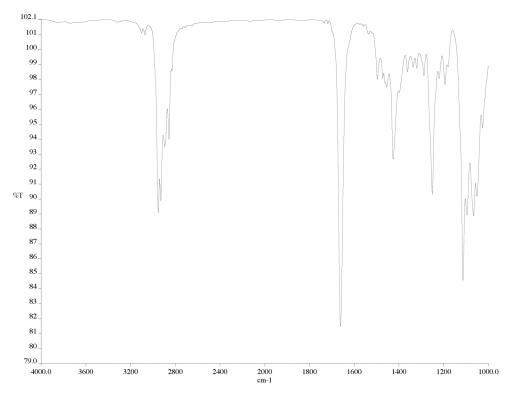
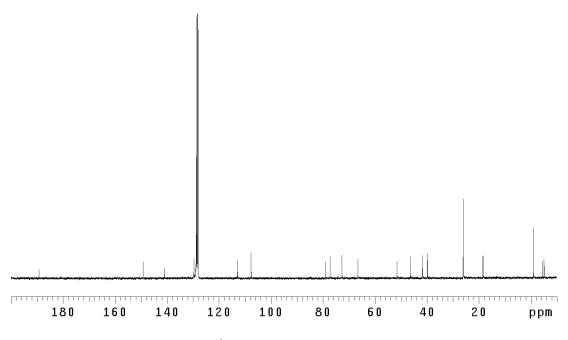
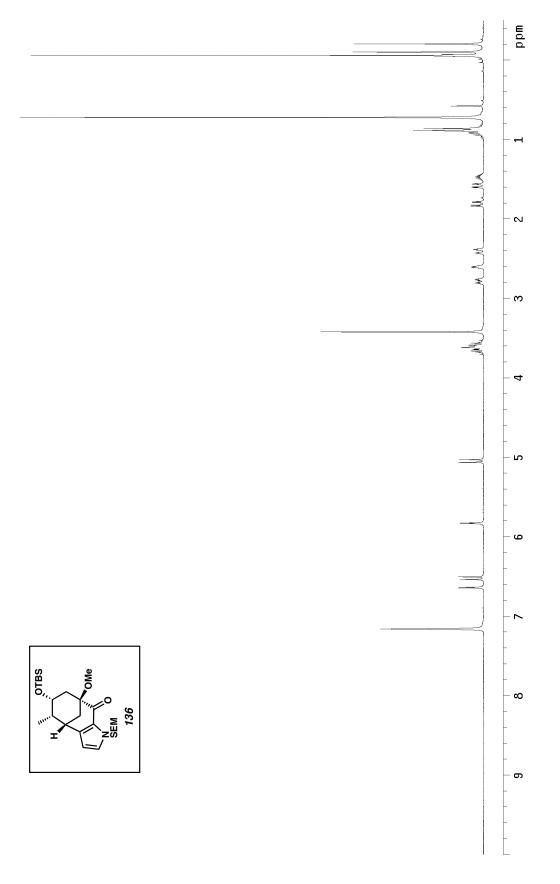


Figure A4.101 Infrared spectrum (thin film/NaCl) of compound 133.



*Figure A4.102*  $^{13}$ C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>) of compound **133**.





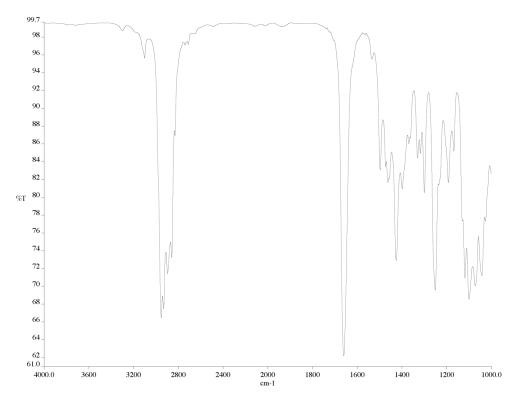
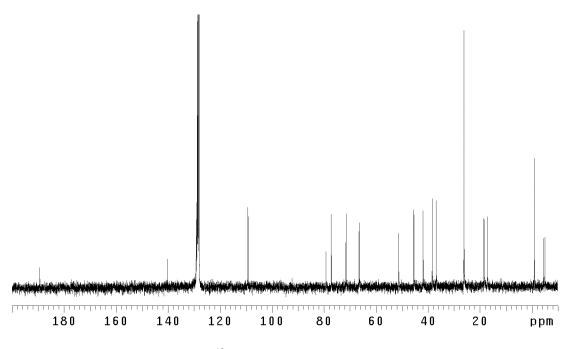
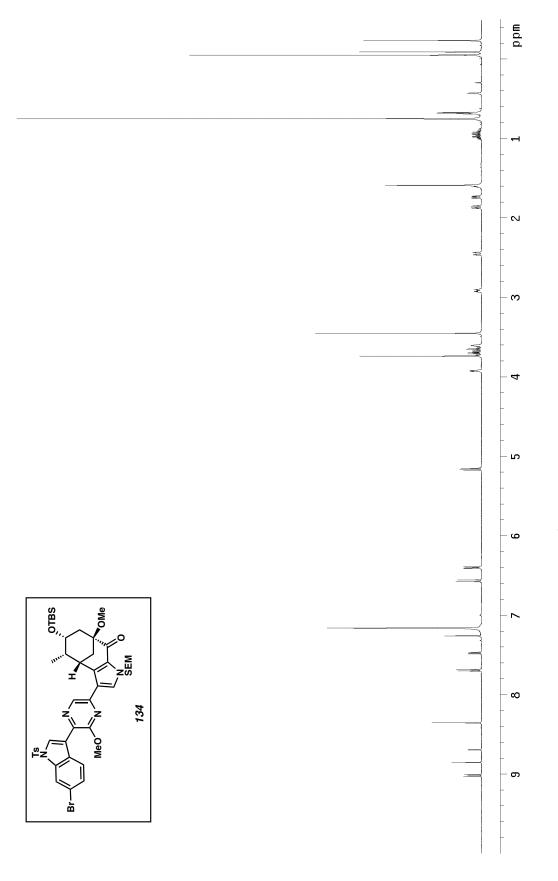


Figure A4.104 Infrared spectrum (thin film/NaCl) of compound 136.



*Figure A4.105*  $^{13}$ C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>) of compound **136**.





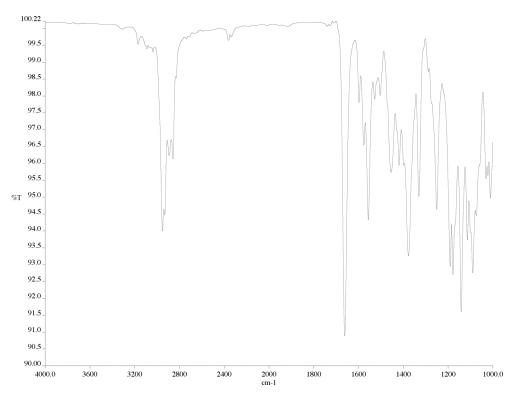
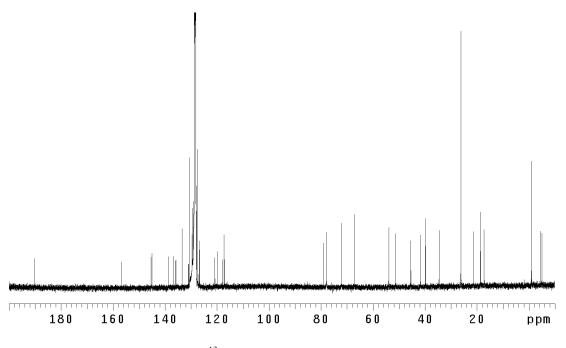
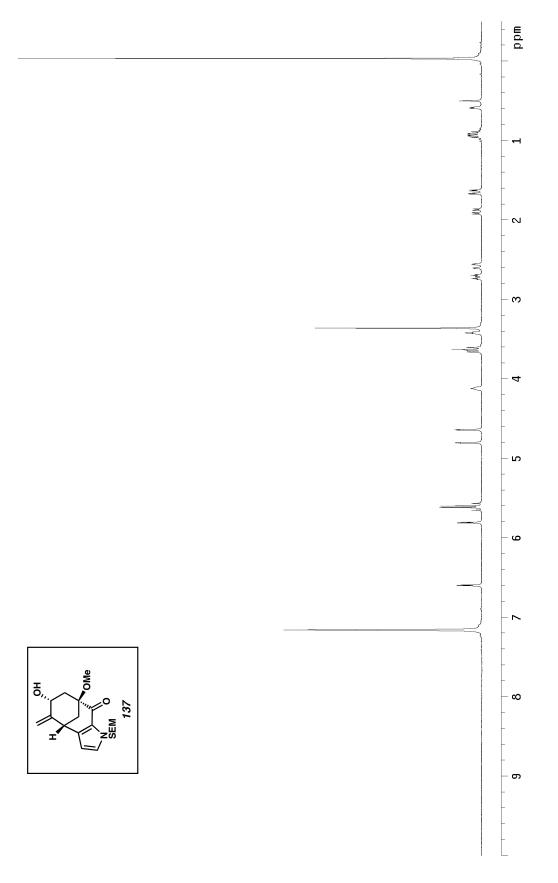


Figure A4.107 Infrared spectrum (thin film/NaCl) of compound 134.



*Figure A4.108*  $^{13}$ C NMR (125 MHz, C<sub>6</sub>D<sub>6</sub>) of compound **134**.





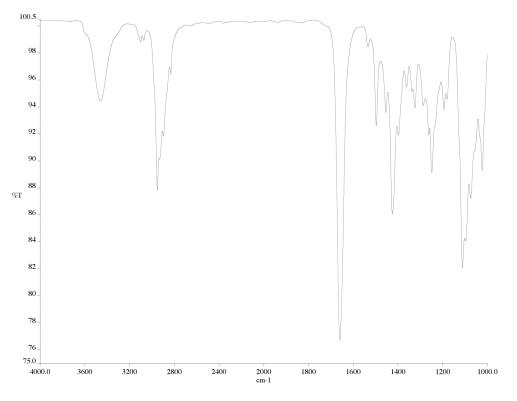
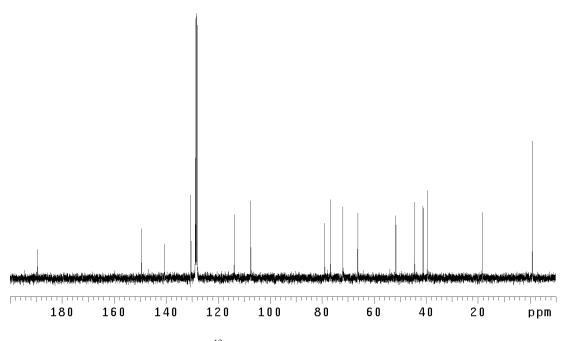
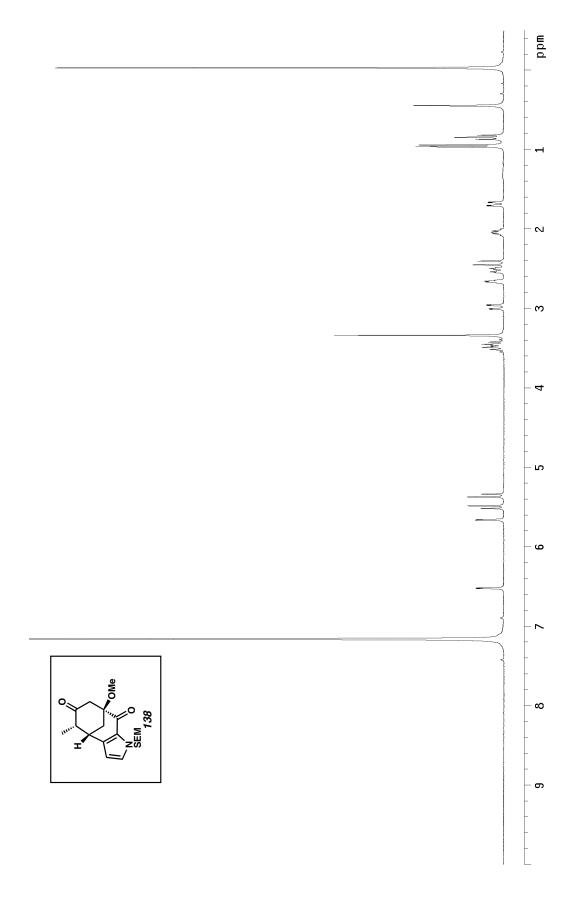


Figure A4.110 Infrared spectrum (thin film/NaCl) of compound 137.



*Figure A4.111* <sup>13</sup>C NMR (75 MHz,  $C_6D_6$ ) of compound **137**.





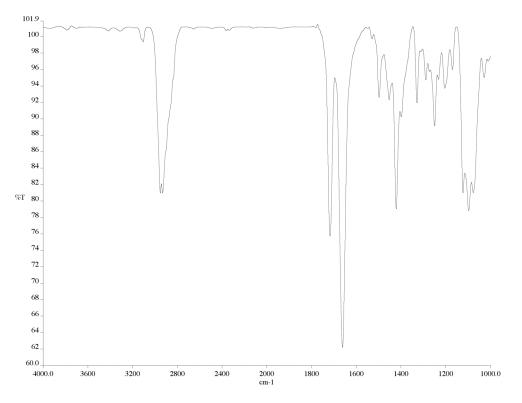
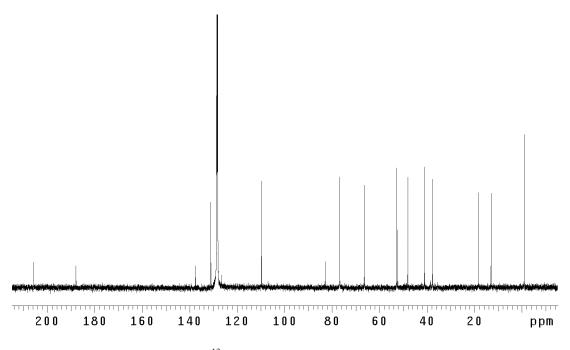
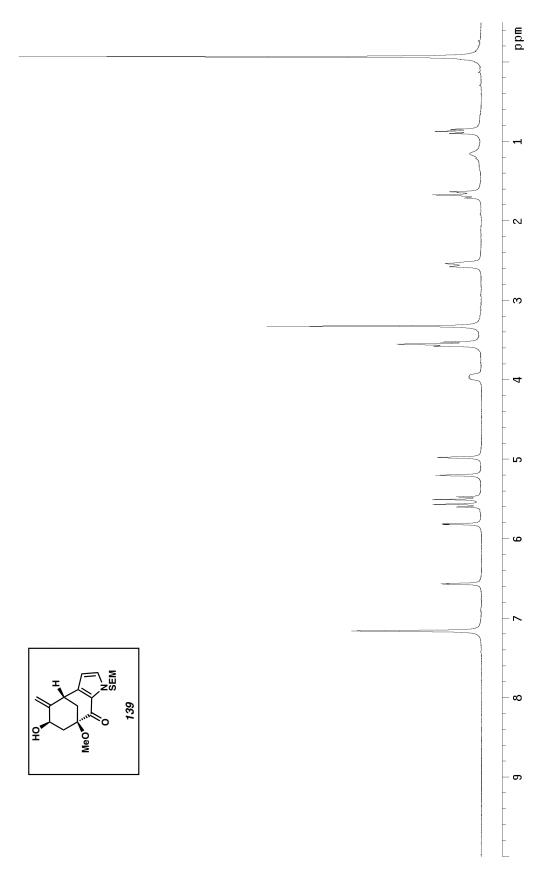


Figure A4.113 Infrared spectrum (thin film/NaCl) of compound 138.



*Figure A4.114*  $^{13}$ C NMR (125 MHz, C<sub>6</sub>D<sub>6</sub>) of compound **138**.





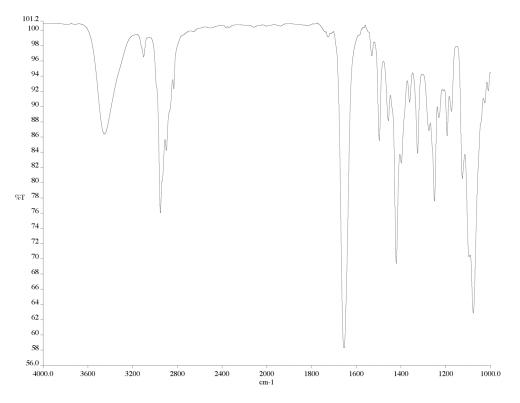
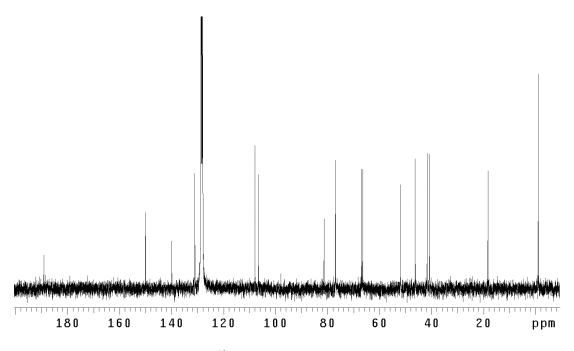
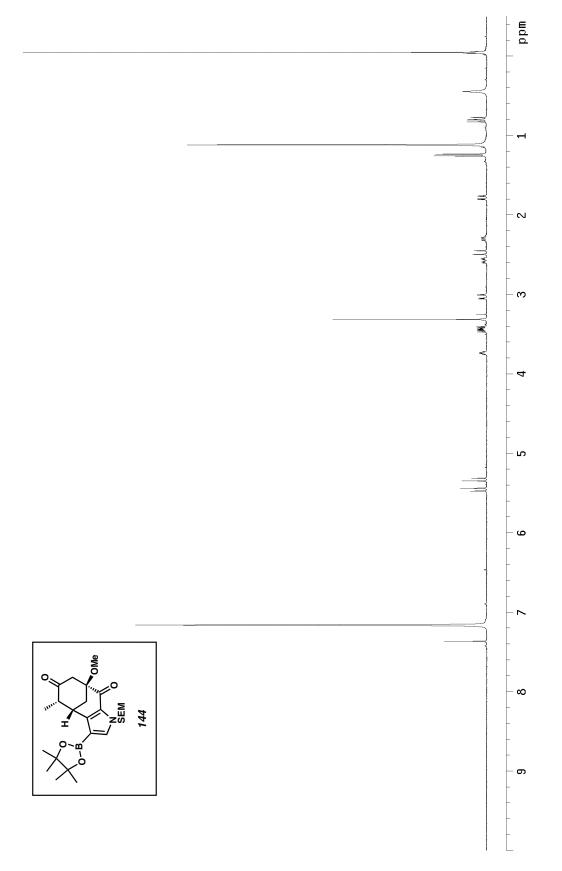


Figure A4.116 Infrared spectrum (thin film/NaCl) of compound 139.



*Figure A4.117*  $^{13}$ C NMR (75 MHz, C<sub>6</sub>D<sub>6</sub>) of compound **139**.





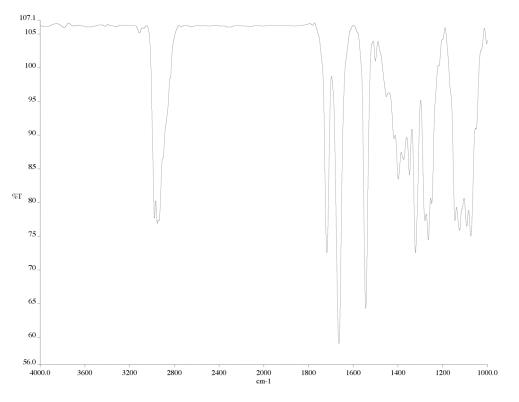
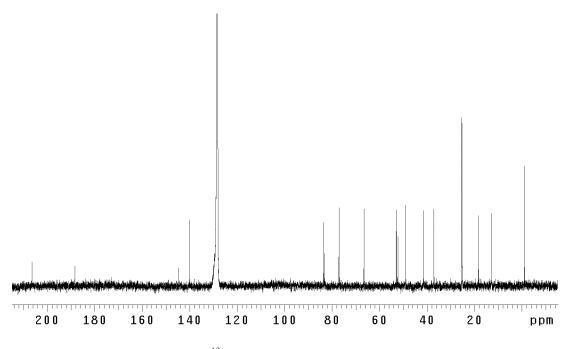
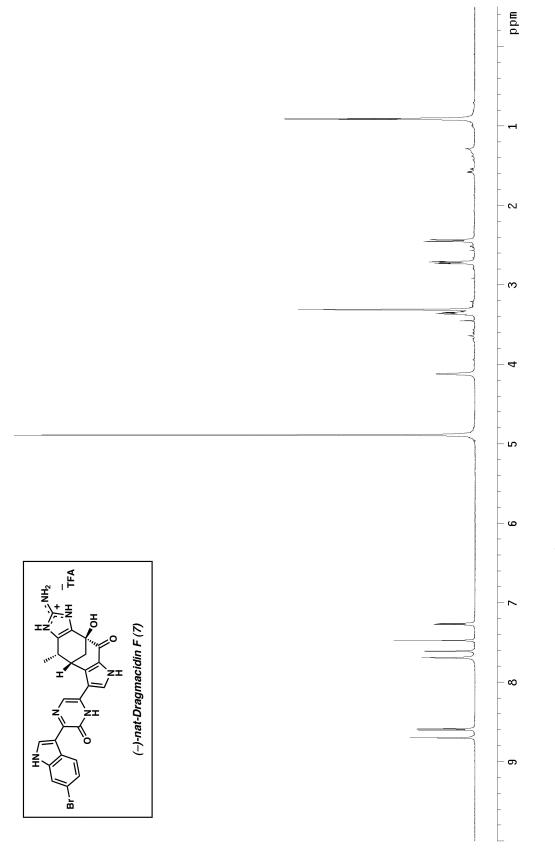


Figure A4.119 Infrared spectrum (thin film/NaCl) of compound 144.



*Figure A4.120*  $^{13}$ C NMR (125 MHz, C<sub>6</sub>D<sub>6</sub>) of compound **144**.





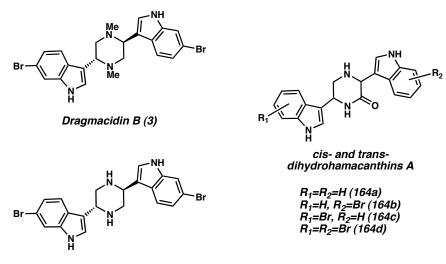
### **APPENDIX FIVE**

# The Formal Total Synthesis of Dragmacidin B, *trans*-Dragmacidin C, and *cis*- and *trans*-Dihydrohamacanthin A

# **A5.1 Introduction**

Having established that halogen-selective Suzuki couplings are a powerful method for constructing the carbon skeleton of the pyrazinone containing dragmacidins, we hypothesized that a similar strategy could be used to access the piperazine dragmacidins and related bis(indole) alkaloids. This appendix section describes the implementation of this approach to achieve the formal total synthesis of dragmacidin B (3),<sup>1</sup> *trans*-dragmacidin C (4),<sup>2</sup> and *cis*- and *trans*-dihydrohamacanthin A  $(164b)^3$  (Figure A5.1.1).<sup>4</sup>

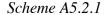


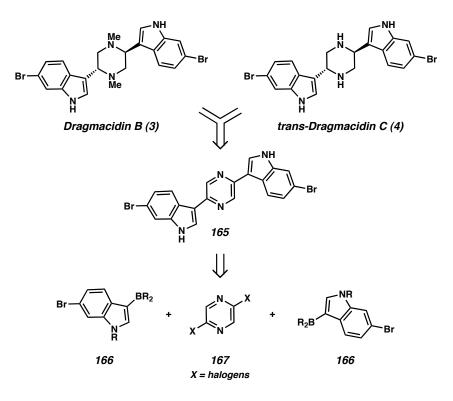


trans-Dragmacidin C (4)

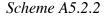
#### A5.2 The Formal Total Synthesis of Dragmacidin B and *trans*-Dragmacidin C

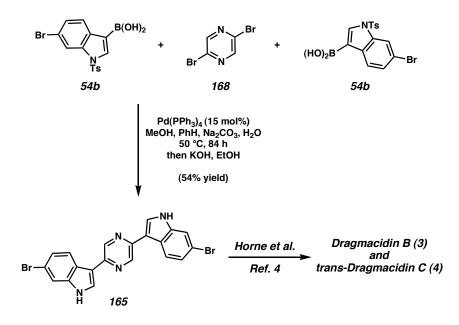
A retrosynthetic strategy for the preparation of dragmacidin B (**3**) and *trans*dragmacidin C (**4**) is shown below in Scheme A5.2.1. Based on conditions reported by Horne, each of the bis(indole) alkaloids can be accessed from unsaturated derivative **165** in a single step.<sup>5</sup> Pyrazine **165**, in turn, would be obtained from two halogen-selective Suzuki cross-coupling reactions of boronic acid **166** with a dihalogenated pyrazine (**167**). We anticipated that the boronic acid fragment employed in our synthesis of dragmacidin D (i.e., **54b**) could be utilized as a surrogate for **166** in order to achieve our current goals. However, the success of our plan would depend highly on the choice of halogens for pyrazine **167**.





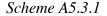
In order to probe the limits of our halogen-selective Suzuki cross-coupling methodology, we chose to use known dibromide  $168^6$  as the critical pyrazine fragment (Scheme A5.2.2). In a one-pot, 4-step transformation, an excess of 6-bromoindolyl-boronic acid (54b) was exposed to dibromopyrazine 168 under our standard cross-coupling conditions. Following quenching with KOH/ethanol, the deprotected pyrazine product (165) was obtained in 54% yield. Notably, although four bromides were introduced in the reaction mixture, only the two pyrazinyl bromides were reactive in the presence of Pd(0) at 50 °C. This rapid synthesis of bis(indole)pyrazine 165 constitutes a formal total synthesis of both dragmacidin B (3) and *trans*-dragmacidin C (4).<sup>5,7,8</sup>

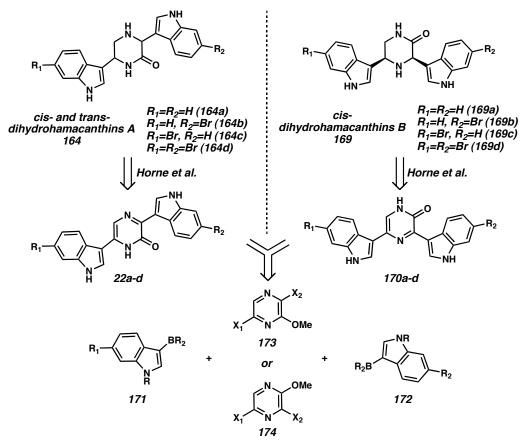




#### A5.3 The Formal Total Synthesis of cis- and trans-Dihydrohamacanthin A

These halogen-selective Suzuki couplings also have great potential for assembling a related family of natural products, the dihydrohamacanthins (**164** and **169**, Scheme A5.3.1). In this scenario, the desired alkaloids (**164** and **169**) would be obtained from their pyrazinone counterparts (**22** and **170**), using the method established by Horne.<sup>9</sup> Intermediates **22** and **170**, in turn, would arise via cross-coupling chemistry using indole fragments **171** and **172**, as well as pyrazine fragments **173** and **174**, in a manner similar to that described above. Halogen-selective cross-couplings will be crucial to prepare all of the halogenation patterns present in this series of natural products (**164a-d**, **169a-d**).

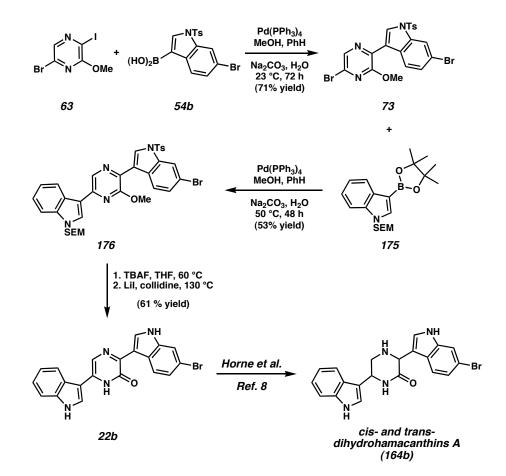




 $X_1, X_2 = halogens$ 

To demonstrate the feasibility of this approach, we prepared one of the dihydrohamacanthin natural products (**164b**, Scheme A5.3.2). In the first Suzuki coupling, dihalopyrazine **63** and bromoindole **54b** were treated with Pd(0) at 23 °C to afford coupled indolylpyrazine **73**, as described in Chapter 2, Section 2.3.4. Dibromide **73**, in turn, was subjected to boronic ester **175** in the presence of Pd(0) at 50 °C to produce bis(indole)pyrazine **176** in 53% yield. In both cases, complete halogen-selectivity was observed. Subsequent removal of all protecting groups furnished pyrazinone **22b**, which has previously been converted to the natural product (**164b**) in a single step.<sup>9,10,11</sup>

Scheme A5.3.2



# **A5.4** Conclusion

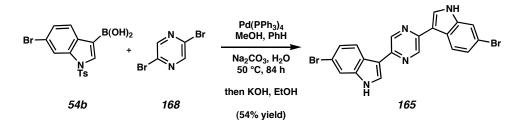
In summary, we have completed the formal total synthesis of dragmacidin B (3) and *trans*-dragmacidin C (4). Our route features a one-pot, 4-step halogen-selective cross-coupling/deprotection sequence to construct the bis(indole) scaffold of our targets. In addition, we have applied this methodology to the formal synthesis of a dihydrohamacanthin natural product (**164b**).

#### **A5.5 Experimental Section**

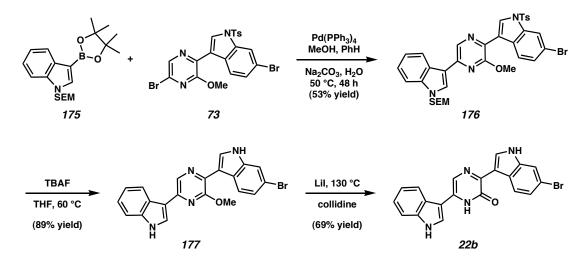
# A5.5.1 Materials and Methods

Unless stated otherwise, reactions were conducted in flame-dried glassware under an atmosphere of nitrogen using anhydrous solvents (either freshly distilled or passed through activated alumina columns). All commercially obtained reagents were used as received. Reaction temperatures were controlled using an IKAmag temperature modulator. Thin-layer chromatography (TLC) was conducted with E. Merck silica gel 60 F254 pre-coated plates (0.25 mm) and visualized using a combination of UV, anisaldehyde, and potassium permanganate staining. ICN silica gel (particle size 0.032-0.063 mm) was used for flash column chromatography.

# **A5.5.2 Preparative Procedures**



Bis(indole)pyrazine 165. A vial charged with dibromopyrazine 168<sup>6</sup> (15.1 mg, 0.0635 mmol), boronic acid 54b (75)0.190 mg, mmol), and tetrakis(triphenylphosphine)palladium(0) (11 mg, 0.0095 mmol) was evacuated and purged with N<sub>2</sub>. Deoxygenated benzene (1.2 mL), deoxygenated methanol (250  $\mu$ L), and deoxygenated 2 M aq. Na<sub>2</sub>CO<sub>3</sub> (105 µL) were added. The reaction mixture was sparged with argon for 3 min. The vial was then sealed, heated to 50 °C for 84 h, and cooled to 23 °C. EtOH (7 mL) and KOH (500 mg) were added. The reaction mixture was heated to 50 °C for 20 h, cooled to 23 °C, then quenched by pouring into 10% (w/v) aq. citric acid (20 mL). EtOAc (30 mL) was added, and the layers were partitioned. The aqueous phase was further extracted with EtOAc (2 x 30 mL). The combined organic layers were washed with brine (15 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure. The crude product was purified by flash chromatography (10:1 CH<sub>2</sub>Cl<sub>2</sub>:MeOH eluent), then further purified by preparative thin layer chromatography (2:1 EtOAc:hexanes eluent) to afford known bis(indole) **165**<sup>5</sup> (16 mg, 54% yield) as a yellow powder.



Pyrazinone 22b. A reaction tube charged with indolylpyrazine  $73^{12}$  (125 mg, 0.335 mmol), boronic (90)0.168 ester 175 mg, mmol), and tetrakis(triphenylphosphine)palladium(0) (35 mg, 0.030 mmol) was evacuated and purged with N<sub>2</sub>. Deoxygenated benzene (3.5 mL), deoxygenated methanol (690  $\mu$ L), and deoxygenated 2 M aq.  $Na_2CO_3$  (180 µL) were then added. The reaction mixture was sparged with argon for 2 min. The tube was then sealed, heated to 50 °C for 48 h, cooled to 23 °C, and quenched by the addition of  $Na_2SO_4$  (200 mg). The reaction mixture was filtered over a plug of SiO<sub>2</sub> (CH<sub>2</sub>Cl<sub>2</sub> eluent), and the solvent was evaporated under

reduced pressure. The crude product was purified by flash chromatography (2:1  $CH_2Cl_2$ :hexanes eluent) to afford bis(indole) **176** (62 mg, 53% yield), which was used immediately in the subsequent reaction.

Bis(indole) **176** (30 mg, 0.043 mmol) was dissolved in 1.0 M TBAF in THF (1 mL, 1 mmol) and heated to 65 °C for 16 h. After cooling to 23 °C, the solvent was removed under reduced pressure, and  $CH_2Cl_2$  (5 mL) was added. The organic layer was washed with  $H_2O$  (2 x 1 mL) and brine (1 mL), concentrated to dryness, then purified by flash chromatography ( $CH_2Cl_2$  eluent) to give bis-*N*-deprotected intermediate **177** (16 mg, 89% yield). A mixture of crude **177** (1.5 mg, 0.0034 mmol), LiI (100 mg, 0.75 mmol), and collidine (1 mL) was heated to 130 °C for 4 days. After cooling to 23 °C, the reaction mixture was diluted with EtOAc (5 mL), washed with  $H_2O$  (3 x 5 mL) and brine (2 mL), then dried by passage over a plug of SiO<sub>2</sub> (EtOAc eluent). The solvent was removed under reduced pressure to afford known pyrazone **22b**<sup>9</sup> (1.0 mg, 69% yield).

#### **A5.6** Notes and References

- (1) Morris, S. A.; Andersen, R. J. *Tetrahedron* **1990**, *46*, 715-720.
- (2) Fahy, E.; Potts, B. C. M.; Faulkner, D. J.; Smith, K. J. Nat. Prod. 1991, 54, 564-569.
- (3) Casapullo, A.; Bifulco, G.; Bruno, I.; Riccio, R. J. Nat. Prod. 2000, 63, 447-451.
- (4) Portions of this work have been described in a communication, see: Garg, N. K.;Stoltz, B. M. *Tetrahedron Lett.* 2005, *46*, 2423-2426.
- (5) Miyake, F. Y.; Yakushijin, K.; Horne, D. A. Org. Lett. 2000, 2, 3185-3187.
- (6) Ellingson, H. J. Am. Chem. Soc. 1949, 71, 2798-2800.
- (7) Treatment of 165 with NaBH<sub>3</sub>CN in formic acid leads to the formation of dragmacidin B (3), while the analogous reaction conducted in acetic acid results in production of *trans*-dragmacidin C (4).<sup>5</sup>
- (8) Sakamoto et al. have shown that natural dragmacidin C is *cis*-fused. See: Kawasaki, T.; Ohno, K.; Enoki, H.; Umemoto, Y.; Sakamoto, M. *Tetrahedon Lett.*2002, 43, 4245-4248.

- (9) Miyake, F. Y.; Yakushijin, K.; Horne, D. A. Org. Lett. 2002, 4, 941-943.
- (10) Reduction of **22b** with NaBH<sub>3</sub>CN leads to the production of **164b**.
- (11) We have also prepared bromopyrazinone 22b by an alternative cyclocondensation route. See Chapter 2, Section 2.2.
- (12) See Chapter 2, Section 2.7.2 for preparative procedure.

#### APPENDIX SIX

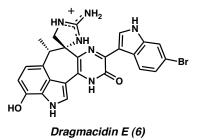
# A Strategy for the Preparation of Dragmacidin E

# A6.1 Background

# A6.1.1 Introduction

To date, dragmacidin E (6) is the only member of the dragmacidin family that has not been synthesized (Figure A6.1.1). In 2000, when the Stoltz laboratory began, we developed a strategy to prepare this complex alkaloid. This appendix section describes our novel approach to the total synthesis of dragmacidin E and highlights our preliminary results involving the synthesis of model systems.<sup>1</sup>

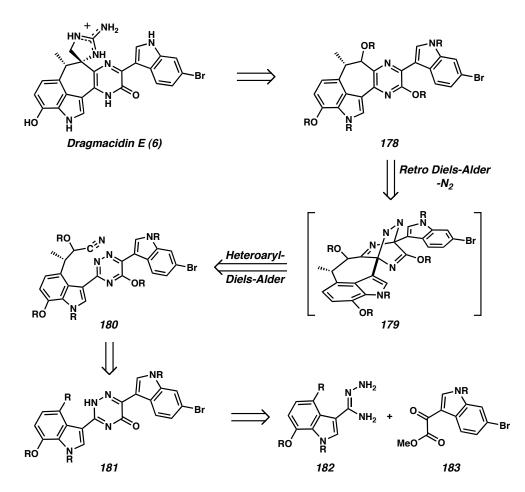
Figure A6.1.1



# A6.1.2 Retrosynthetic Analysis of Dragmacidin E

Our retrosynthetic strategy for the preparation of dragmacidin E ( $\mathbf{6}$ ) is shown below in Scheme A6.1.1. We envisioned that the guanidinium unit could be installed at a late-stage in the synthesis, and the pyrazinone moiety could be masked as a pyrazine. Thus, the natural product ( $\mathbf{6}$ ) was disconnected to bis(indole)pyrazine **178**. The sevenmembered ring of dragmacidin E could then be installed from cyanotriazine 180 via a hetero-aryl Diels-Alder/Retro Diels-Alder sequence with concomitant loss of  $N_2$  (180  $\rightarrow$  $179 \rightarrow 178$ ).<sup>2,3</sup> Aromatic triazine 180 would be obtained from a non-aromatic triazinone (181), which in turn could be prepared via a cyclocondensation reaction of amidrazone 182 and ketoester 183. Although indole-ketoesters are well known in the literature, indole-amidrazones are not. Therefore, our initial goal was to develop a simple synthesis of indole-amidrazones and then utilize those amidrazones to access bis(indole)triazinones.

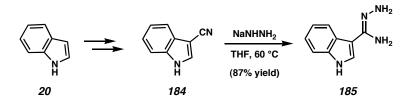
Scheme A6.1.1



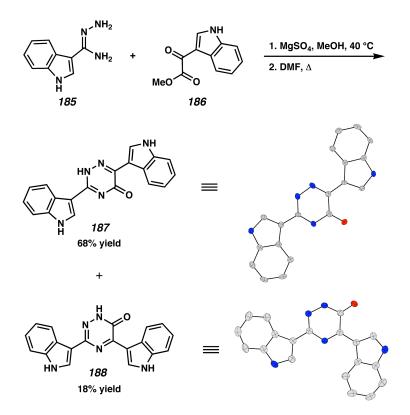
# A6.2 Model Systems: The Facile Synthesis of Bis(indole)-1,2,4-Triazinones

The preparation of unsubstituted indole-amidrazones turned out to be relatively straightforward (Scheme A6.2.1). Beginning from commercially available indole (**20**), we were able to access cyanoindole **184** in three steps using a known protocol.<sup>4</sup> Then, simply treating **184** with sodium hydrazide in refluxing THF afforded the desired amidrazone (**185**) in good yield.<sup>5</sup>

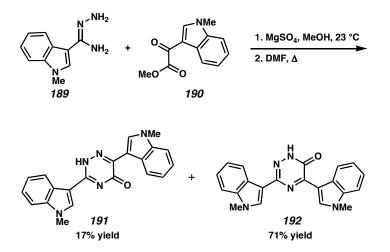
Scheme A6.2.1



In the cyclocondensation reaction, exposure of amidrazone **185** to ketoester **186**<sup>6</sup> in the presence of MgSO<sub>4</sub> in methanol,<sup>5</sup> followed by reflux in DMF, afforded the desired *p*-triazinone product (**187**) in 68% yield (Scheme A6.2.2). *m*-Triazinone **188** was also formed, although in low yield. After separation by silica gel chromatography, the C-C connectivity of each of the triazinone products (**187** and **188**) was determined by single crystal X-ray diffraction studies.<sup>7a,b</sup>

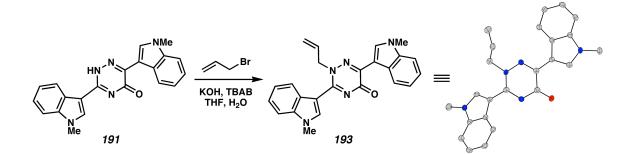


We also prepared the corresponding 1-methylated cyclization starting materials, methylamidrazone **189** and methylketoester **190**<sup>8</sup> (Scheme A6.2.3). When these compounds were reacted under similar conditions to those described above, triazinone formation proceeded readily. However, the product distribution favored mmethyltriazinone **192** over p-methyltriazinone **191**. This reversal in selectivity is presumably due to the electron donating effect of the N-Me group on the ketone functionality of **190**, thereby altering its reactivity.



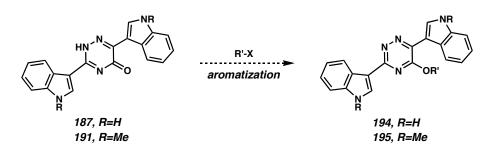
Structural assignments for *N*-methyl derivatives **191** and **192** were made by correlating <sup>1</sup>H NMR and TLC data with data for the corresponding *N*-H compounds (**187** and **188**, respectively). In addition, methyltriazinone **191** was treated with allyl bromide under phase transfer conditions to afford allyl derivative **193** (Scheme A6.2.4). X-ray diffraction analysis of a single crystal revealed the C-C connectivity of allyl species **193** and confirmed that triazinone **191** was *para*-substituted.<sup>7c</sup>

Scheme A6.2.4



While the cyclocondensation strategy described above was effective for the preparation of *para*-substituted bis(indole)triazinones **187** and **191**, initial attempts to aromatize those compounds to their triazine counterparts (**194** and **195**) were met with limited success (Scheme A6.2.5).<sup>9</sup> Although further work in this area has not been carried out, an alternative strategy to directly access bis(indole)triazines, rather than triazinones, would be attractive in order to access substrates suitable for the critical Diels-Alder/Retro Diels-Alder sequence en route to dragmacidin E (**6**).

Scheme A6.2.5

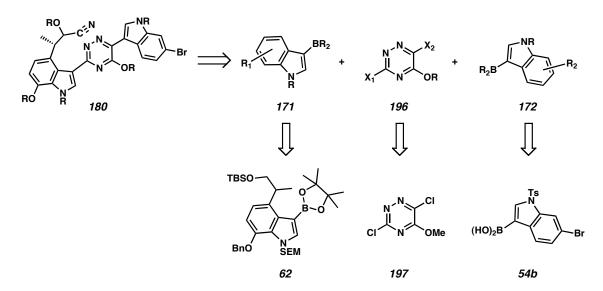


# A6.3 An Alternative Strategy to Access Bis(indole)triazines

Based on our work related to the other dragmacidins (see Chapters 2 and 3), an alternative cross-coupling approach to access aromatic bis(indole)triazines (**180**) can also be envisioned (Scheme A6.3.1). In this scenario, **180** would be obtained by the sequential cross-coupling reactions of metalated indoles (**171** and **172**) and a dihalogenated triazine (**196**). Notably, it may be possible to utilize the same indole fragments (**62** and **54b**) that were employed in the total synthesis of dragmacidin D. The appropriate halogenated triazine (**196**) would likely be discovered after some experimentation; however, known dichloride **197**<sup>10</sup> could serve as a starting point for

optimization. This promising cross-coupling strategy to access dragmacidin E(6) has not yet been explored.

# Scheme A6.3.1



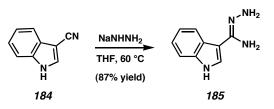
# A6.4 Conclusion

In summary, we have developed a facile method for the synthesis of bis(indole)triazinones involving a cyclocondensation reaction between amidrazone and ketoester functionalities. Although we have only prepared simple model systems thus far, more highly functionalized bis(indole)triazinones could potentially be used as intermediates en route to the total synthesis of dragmacidin E (6). Additionally, in future efforts, it may be possible to access substituted bis(indole)triazines via halogen-selective cross-coupling reactions.

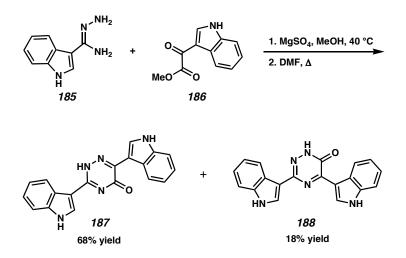
#### A6.5 Experimental Section

# A6.5.1 Materials and Methods

Unless stated otherwise, reactions were conducted in flame-dried glassware under an atmosphere of nitrogen using anhydrous solvents (either freshly distilled or passed through activated alumina columns). All commercially obtained reagents were used as received. Reaction temperatures were controlled using an IKAmag temperature modulator. Thin-layer chromatography (TLC) was conducted with E. Merck silica gel 60 F254 pre-coated plates (0.25 mm) and visualized using a combination of UV, anisaldehyde, and potassium permanganate staining. ICN silica gel (particle size 0.032-0.063 mm) was used for flash column chromatography. <sup>1</sup>H NMR spectra were recorded on a Varian Mercury 300 (at 300 MHz) or a Varian Inova 500 (at 500 MHz) and are reported relative to Me<sub>4</sub>Si ( $\delta$  0.0). Data for <sup>1</sup>H NMR spectra are reported as follows: chemical shift ( $\delta$  ppm), multiplicity, coupling constant (Hz), and integration. <sup>13</sup>C NMR spectra were recorded on a Varian Mercury 300 (at 75 MHz) or a Varian Inova 500 (at 125 MHz) and are reported relative to Me<sub>4</sub>Si ( $\delta$  0.0). Data for <sup>13</sup>C NMR spectra are reported in terms of chemical shift. IR spectra were recorded on a Perkin Elmer Paragon 1000 spectrometer and are reported in frequency of absorption (cm<sup>-1</sup>). High resolution mass spectra were obtained from the California Institute of Technology Mass Spectral Facility. X-ray crystallographic structures were obtained by Mr. Larry M. Henling and Dr. Mike W. Day at the California Institute of Technology Beckman Institute X-Ray Crystallography Laboratory.

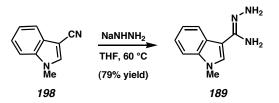


**Amidrazone 185.** To a suspension of NaH (60% dispersion in mineral oil, 167 mg, 4.16 mmol) in Et<sub>2</sub>O (3.5 mL) at 0 °C was added anhydrous hydrazine (131  $\mu$ L, 4.15 mmol). After stirring for 1 h, a solution of cyanoindole **184**<sup>4</sup> (200 mg, 1.39 mmol) in THF (7 mL) was added dropwise over 10 min. The reaction mixture was heated to 60 °C for 6 h, cooled to 23 °C, quenched by the addition of H<sub>2</sub>O (5 mL), and extracted with EtOAc (3 x 20 mL). The combined organic extracts were washed with brine (15 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure. The residue was triturated with Et<sub>2</sub>O (2 mL) and dried under vacuum to afford amidrazone **185** (213 mg, 87% yield), which was used immediately in the subsequent reaction. <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  11.10 (br s, 1H), 8.15 (d, *J* = 7.7 Hz, 1H), 7.69 (s, 1H), 7.33 (d, *J* = 8.1 Hz, 1H), 7.10-7.03 (m, 1H), 7.01-6.93 (m, 1H), 5.48 (br s, 2H), 4.83 (br s, 2H). <sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  145.3, 136.3, 124.8, 123.6, 122.2, 121.2, 119.0, 111.3, 111.1.

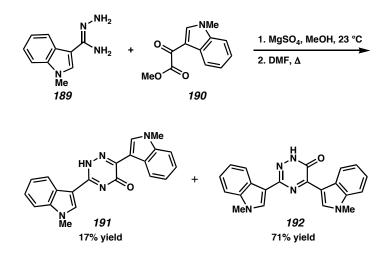


Triazinones 187 and 188. To crude amidrazone 185 (100 mg, 0.568 mmol) and MgSO<sub>4</sub> (171 mg, 1.42 mmol) in MeOH (2 mL) at 23 °C was added a solution of ester **186**<sup>6</sup> (105 mg, 0.516 mmol) in MeOH (5 mL). The reaction mixture was heated to 40 °C for 24 h, then cooled to 23 °C. After removal of solvent under vacuum, DMF (5 mL) was added. The resulting suspension was refluxed for 24 h, then cooled to 23 °C. The solvent was removed under vacuum, and the crude product was purified by flash column chromatography (1:1 hexanes: EtOAc eluent) to afford p-triazinone 187 (115 mg, 68% yield) and *m*-triazinone **188** (30 mg, 18% yield) as yellow solids. For **187**, suitable crystals for X-ray diffraction were grown by the slow diffusion of hexanes into a saturated solution of **187** in 1:1 DMF:MeOH. For **188**, single crystals suitable for X-ray diffraction were obtained by the slow diffusion of hexanes into a saturated solution of **188** in MeOH. *p*-Triazinone **187**:  $R_f 0.28$  (4:1 EtOAc:hexanes); mp >250 °C dec; <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ):  $\delta$  13.66 (br s, 1H), 12.03 (s, 1H), 11.67 (s, 1H), 8.83 (s, 1H), 8.52 (d, J = 7.6 Hz, 1H), 8.50-8.45 (m, 1H), 8.44 (d, J = 2.5 Hz, 1H), 7.55-7.51 (comp. m, 2H), 7.28-7.17 (comp. m, 4H); <sup>13</sup>C NMR (125 MHz, DMSO-d<sub>6</sub>, 16/19 C): δ 136.7, 136.3, 131.6, 129.0, 125.3, 125.2, 122.7, 122.3, 122.1, 121.9, 121.1, 120.5, 112.2, 112.0, 108.1, 106.2. CCDC deposition number 259291; IR (film) 3350, 1520, 1421,

1187 cm<sup>-1</sup>; HRMS-FAB (*m/z*): [M+H]<sup>+</sup> calc'd for C<sub>19</sub>H<sub>14</sub>N<sub>5</sub>O, 328.1198; found, 328.1185. *m*-Triazinone **188**: R<sub>f</sub> 0.61 (4:1 EtOAc:hexanes); mp >250 °C dec; <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  13.12 (s, 1H), 12.13 (s, 1H), 11.54 (s, 1H), 9.12 (d, *J* = 2.9 Hz, 1H), 8.77 (d, *J* = 7.3 Hz, 1H), 8.26 (d, *J* = 7.7 Hz, 1H), 8.14 (d, *J* = 2.6 Hz, 1H), 7.60-7.47 (comp. m, 2H), 7.39-7.26 (comp. m, 2H), 7.25-7.12 (comp. m, 2H). CCDC deposition number 161494.

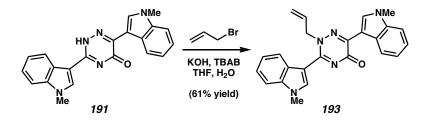


**Methylamidrazone 189**. Methylamidrazone **189** was prepared in a manner analogous to the preparation of **185**. To a suspension of NaH (60% dispersion in mineral oil, 779 mg, 19.48 mmol) in Et<sub>2</sub>O (16.2 mL) at 0 °C was added anhydrous hydrazine (611  $\mu$ L, 19.48 mmol). After stirring for 1 h, a solution of *N*-methyl-3-cyanoindole<sup>11</sup> (**198**, 910 mg, 6.49 mmol) in THF (32.5 mL) was added dropwise over 10 min. The reaction mixture was heated to 60 °C for 6 h, cooled to 23 °C, quenched by the addition of H<sub>2</sub>O (17 mL), and extracted with EtOAc (4 x 25 mL). The combined organic extracts were washed with brine (2 x 25 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure to afford crude amidrazone **189** (880 mg, 79% yield), which was used immediately in the subsequent reaction without further purification. <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  8.16 (d, *J* = 8.0 Hz, 1H), 7.65 (s, 1H), 7.39 (d, *J* = 8.2 Hz, 1H), 7.18-7.11 (m, 1H), 7.07-6.99 (m, 1H), 5.43 (br s, 2H), 4.81 (br s, 2H), 3.76 (s, 3H).



Bis(methyl)triazinones 191 and 192. To crude amidrazone 189 (65 mg, 0.378 mmol) and MgSO<sub>4</sub> (159 mg, 1.32 mmol) in MeOH (2 mL) at 23 °C was added a solution of ester 190<sup>8</sup> (75 mg, 0.343 mmol) in MeOH (3.4 mL). The reaction mixture was stirred at 23 °C for 24 h. After removal of solvent under vacuum, DMF (5 mL) was added. The resulting suspension was refluxed for 24 h, then cooled to 23 °C. The solvent was removed under vacuum, and the crude product was purified by flash chromatography (1:1 hexanes: EtOAc eluent) to afford p-bis(methyl)triazinone **191** (20 mg, 16% yield) as a yellow solid and impure *m*-bis(methyl)triazinone **192**. The crude *m*-triazinone was repurified by flash chromatography (1:1 hexanes: EtOAc eluent) to afford pure 192 (86 mg, 71% yield) as a yellow solid. *p*-Bis(methyl)triazinone **191**:  $R_f$  0.10 (1:1) hexanes:EtOAc); mp >250 °C dec; <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ ):  $\delta$  13.60 (br s, 1H), 8.79 (s, 1H), 8.57-8.49 (comp. m, 2H), 8.34 (s, 1H), 7.60-7.52 (comp. m, 2H), 7.33-7.20 (comp. m, 4H), 3.91 (s, 3H), 3.90 (s, 3H); <sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>, 19/21 C): δ 153.9, 137.2, 136.8, 135.1, 132.5, 125.9, 125.7, 122.6, 122.4, 122.2, 122.1, 121.1, 120.6, 110.5, 110.1, 107.6, 106.3, 33.3, 32.9; IR (film) 3600, 1567, 1539, 1370 cm<sup>-1</sup>; HRMS-FAB (m/z):  $[M + H]^+$  calc'd for  $C_{21}H_{18}N_5O$ , 356.1511; found, 356.1520. m-Bis(methyl)triazinone **192**:  $R_f 0.43$  (1:1 hexanes:EtOAc); mp >250 °C dec; <sup>1</sup>H NMR (300

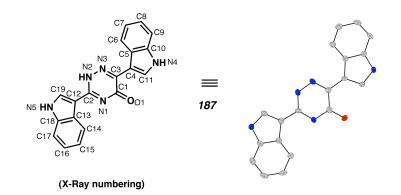
MHz, DMSO- $d_6$ ):  $\delta$  13.15 (br s, 1H), 9.14 (s, 1H), 8.83 (d, J = 7.3 Hz, 1H), 8.28 (d, J = 7.8 Hz, 1H), 8.14 (s, 1H), 7.62 (d, J = 7.0 Hz, 1H), 7.54 (d, J = 7.7 Hz, 1H), 7.47-7.32 (comp. m, 2H), 7.31-7.16 (comp. m, 2H), 3.96 (comp. m, 6H); <sup>13</sup>C NMR (75 MHz, DMSO- $d_6$ , 20/21 C):  $\delta$  157.7, 153.2, 148.0, 140.0, 137.6, 137.2, 131.4, 126.3, 124.9, 123.2, 122.4, 122.1, 121.6, 120.3, 111.2, 110.7, 110.3, 109.2, 33.3, 32.9; IR (film) 3600, 1646, 1465, 1373 cm<sup>-1</sup>; HRMS-FAB (m/z): [M + H]<sup>+</sup> calc'd for C<sub>21</sub>H<sub>18</sub>N<sub>5</sub>O, 356.1511; found, 356.1521.



Allyl triazinone 193. To *p*-triazinone 191 (25 mg, 0.070 mmol) in THF at 23 °C, was added allyl bromide (6.7  $\mu$ L, 0.078 mmol), H<sub>2</sub>O (150  $\mu$ L), powdered KOH (20 mg, 0.35 mmol), and tetrabutylammonium bromide (0.2 mg, 0.0007 mmol) in H<sub>2</sub>O (10  $\mu$ L). The resulting solution was stirred for 24 h, diluted with H<sub>2</sub>O (5 mL), and extracted with EtOAc (3 x 15 mL). The combined organic layers were washed with brine (10 mL), dried over MgSO<sub>4</sub>, and evaporated under reduced pressure to afford allyl triazinone 193 (17 mg, 61% yield). Single crystals suitable for X-ray diffraction were obtained by the slow diffusion of hexanes into a saturated solution of 193 in acetone. <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  8.85 (s, 1H), 8.42 (d, *J* = 7.7 Hz, 1H), 8.14 (d, *J* = 8.1 Hz, 1H), 8.02 (s, 1H), 7.62-7.51 (comp. m, 2H), 7.37-7.18 (comp. m, 4H), 6.30-6.15 (m, 1H), 5.40-5.28 (comp. m, 2H), 5.07 (d, *J* = 5.1 Hz, 2H), 3.92 (s, 3H), 3.91 (s, 3H). <sup>13</sup>C NMR (125 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  160.3, 154.4, 145.6, 136.9, 136.6, 136.0, 133.3, 133.2, 126.4, 125.8, 122.6,

## A6.6 X-Ray Crystallography Reports

#### A6.6.1 X-Ray Crystallographic Report for *p*-Triazinone 187



## Crystal data and structure refinement for 187 (CCDC 259291).

Empirical formula	$C_{19}H_{13}N_5O \bullet 2(C_3H_7NO)$
Formula weight	473.54
Crystallization Solvent	DMF/methanol/hexanes
Crystal Habit	Fragment
Crystal size	$0.26 \text{ x} 0.24 \text{ x} 0.23 \text{ mm}^3$
Crystal color	Colorless

#### **Data Collection**

Type of diffractometer	Bruker SMART 1000			
Wavelength	0.71073 Å MoKα			
Data Collection Temperature	100(2) K			
$\theta$ range for 11276 reflections used in lattice determination	2.77 to 28.01°			
Unit cell dimensions	a = 7.7516(6) Å b = 12.8092(11) Å c = 13.5909(11) Å	$ \begin{aligned} &\alpha = 115.3400(10)^{\circ} \\ &\beta = 90.9730(10)^{\circ} \\ &\gamma = 106.3430(10)^{\circ} \end{aligned} $		
Volume	1155.56(16) Å <sup>3</sup>			
Ζ	2			
Crystal system	Triclinic			
Space group	P1			
Density (calculated)	1.361 Mg/m <sup>3</sup>			

F(000)

Data collection program  $\theta$  range for data collection Completeness to  $\theta = 28.01^{\circ}$ Index ranges Data collection scan type Data reduction program Reflections collected Independent reflections Absorption coefficient Absorption correction Max. and min. transmission 500 Bruker SMART v5.630  $1.68 \text{ to } 28.01^{\circ}$ 93.8 % $-10 \le h \le 10, -16 \le k \le 16, -17 \le 1 \le 17$  $\omega$  scans at 7  $\phi$  settings Bruker SAINT v6.45A 23339  $10181 [R_{int} = 0.0447]$  $0.093 \text{ mm}^{-1}$ None 0.9789 and 0.9761

Structure solution program	Bruker XS v6.12		
Primary solution method	Direct methods		
Secondary solution method	Difference Fourier map		
Hydrogen placement	Difference Fourier map		
Structure refinement program	Bruker XL v6.12		
Refinement method	Full matrix least-squares on F <sup>2</sup>		
Data / restraints / parameters	10181 / 3 / 847		
Treatment of hydrogen atoms	Unrestrained		
Goodness-of-fit on F <sup>2</sup>	1.286		
Final R indices [I> $2\sigma$ (I), 8409 reflections]	R1 = 0.0403, wR2 = 0.0678		
R indices (all data)	R1 = 0.0509, wR2 = 0.0699		
Type of weighting scheme used	Sigma		
Weighting scheme used	$w=1/\sigma^2(\text{Fo}^2)$		
Max shift/error	0.006		
Average shift/error	0.001		
Absolute structure parameter	1.2(7)		
Largest diff. peak and hole	0.283 and -0.236 e.Å <sup>-3</sup>		

## **Special Refinement Details**

Refinement of  $F^2$  against ALL reflections. The weighted R-factor (*w*R) and goodness of fit (S) are based on  $F^2$ , conventional R-factors (R) are based on F, with F set to zero for negative  $F^2$ . The threshold expression of  $F^2 > 2\sigma(F^2)$  is used only for calculating R-factors(gt) etc. and is not relevant to the choice of reflections for refinement. R-factors based on  $F^2$  are statistically about twice as large as those based on F, and R-factors based on ALL data will be even larger.

All esds (except the esd in the dihedral angle between two l.s. planes) are estimated using the full covariance matrix. The cell esds are taken into account individually in the estimation of esds in distances, angles and torsion angles; correlations between esds in cell parameters are only used when they are defined by crystal symmetry. An approximate (isotropic) treatment of cell esds is used for estimating esds involving l.s. planes.

Atomic coordinates ( x 10<sup>4</sup>) and equivalent isotropic displacement parameters (Å<sup>2</sup>x 10<sup>3</sup>) for 187 (CCDC 259291). U(eq) is defined as the trace of the orthogonalized U<sup>ij</sup> tensor.

	x	у	Z	U <sub>eq</sub>
O(1A)	3192(2)	-1559(1)	2826(1)	24(1)
N(1A)	6162(3)	-358(2)	3238(2)	19(1)
N(2A)	6945(3)	1045(2)	2523(2)	20(1)
N(3A)	5230(3)	781(2)	2043(2)	20(1)
N(4A)	-879(3)	-1308(2)	1026(2)	18(1)
N(5A)	12261(3)	1932(2)	4134(2)	19(1)
C(1A)	4396(3)	-704(2)	2747(2)	19(1)
C(2A)	7389(3)	508(2)	3105(2)	15(1)
C(3A)	3952(3)	-75(2)	2149(2)	16(1)
C(4A)	2116(3)	-374(2)	1633(2)	15(1)
C(5A)	1508(3)	155(2)	994(2)	15(1)
C(6A)	2333(4)	1051(2)	674(2)	19(1)
C(7A)	1302(4)	1265(2)	-20(2)	22(1)
C(8A)	-547(4)	601(2)	-399(2)	24(1)
C(9A)	-1405(4)	-265(2)	-81(2)	21(1)
C(10A)	-371(3)	-474(2)	621(2)	17(1)
C(10A) C(11A)	593(3)	-1247(2)	1630(2)	17(1)
C(11A) C(12A)	9284(3)	889(2)	3592(2)	18(1)
C(12A) C(13A)	10029(3)	414(2)	. ,	
	9341(4)		4228(2) 4549(2)	16(1) 10(1)
C(14A)		-515(2) -662(2)	• •	19(1) 24(1)
C(15A)	10492(4)		5222(2)	24(1)
C(16A)	12323(4)	58(2)	5580(2) 5222(2)	23(1)
C(17A)	13055(3)	933(2)	5233(2)	20(1)
C(18A)	11882(3)	1099(2)	4563(2)	17(1)
C(19A)	10709(3)	1797(2)	3556(2)	18(1)
O(1B)	-1098(2)	2912(1)	2152(1)	25(1)
N(1B)	1047(3)	4719(2)	3310(2)	16(1)
N(2B)	1604(3)	6308(2)	2815(2)	19(1)
N(3B)	386(3)	5665(2)	1871(2)	20(1)
N(4B)	-3939(3)	2243(2)	-750(2)	18(1)
N(5B)	5380(3)	8298(2)	5778(2)	19(1)
C(1B)	-217(3)	4002(2)	2370(2)	18(1)
C(2B)	1921(3)	5845(2)	3507(2)	15(1)
C(3B)	-528(3)	4517(2)	1627(2)	16(1)
C(4B)	-1796(3)	3816(2)	624(2)	15(1)
C(5B)	-2163(3)	4183(2)	-215(2)	14(1)
C(6B)	-1504(3)	5244(2)	-341(2)	19(1)
C(7B)	-2213(4)	5250(2)	-1283(2)	23(1)
C(8B)	-3533(4)	4241(2)	-2096(2)	25(1)
C(9B)	-4208(3)	3178(2)	-1997(2)	21(1)
C(10B)	-3498(3)	3167(2)	-1050(2)	18(1)
C(10B) C(11B)	-2924(3)	2622(2)	240(2)	18(1)
C(12B)	3246(3)	6649(2)	4502(2)	17(1)
C(12B) C(13B)	3677(3)	6366(2)	5376(2)	16(1)
C(13B) C(14B)	3097(3)	5356(2)	5597(2)	18(1)
C(14B) C(15B)	3867(4)	5445(2)	6562(2)	25(1)
	5007(4)	5445(2)	0502(2)	23(1)

C(16B)	5211(3)	6513(2)	7325(2)	21(1)
C(17B)	5794(3)	7497(2)	7123(2)	20(1)
C(18B)	5018(3)	7424(2)	6161(2)	16(1)
C(19B)	4315(3)	7825(2)	4797(2)	19(1)
- ( )				
O(1C)	5353(2)	7351(2)	472(2)	28(1)
N(1C)	4737(3)	5870(2)	1035(2)	20(1)
C(1C)	3747(4)	4604(2)	782(2)	24(1)
C(2C)	6150(4)	6563(2)	1997(2)	23(1)
C(3C)	4485(4)	6340(3)	367(2)	26(1)
O(1D)	6049(2)	3243(2)	4744(2)	29(1)
N(1D)	6737(3)	4699(2)	4134(2)	20(1)
C(1D)	7751(4)	5933(3)	4366(2)	24(1)
C(2D)	5339(4)	3990(3)	3159(2)	23(1)
C(3D)	6972(4)	4242(2)	4828(2)	24(1)
O(1E)	3755(2)	9886(2)	8020(2)	28(1)
C(1E)	2070(5)	7586(3)	8921(3)	29(1)
N(1E)	2176(3)	8266(2)	8272(2)	22(1)
C(2E)	769(4)	7800(3)	7344(3)	34(1)
C(3E)	3529(4)	9279(2)	8531(2)	22(1)
O(1F)	7626(2)	625(2)	7005(2)	30(1)
N(1F)	9359(3)	2310(2)	6890(2)	21(1)
C(1F)	9603(5)	3073(3)	6341(3)	31(1)
C(2F)	10667(4)	2687(3)	7848(3)	37(1)
C(3F)	7955(4)	1300(2)	6558(2)	23(1)

0			
Bond lengths [Å]	and angles [°]	for 187 (CC	CDC 259291).

O(1A)-C(1A)	1.268(3) N(4B)-H(4B)	0.94(3)
N(1A)-C(2A)	1.323(3) N(5B)-C(19B)	1.348(3)
N(1A)-C(1A)	1.375(3) N(5B)-C(18B)	1.388(3)
N(2A)-C(2A)	1.345(3) N(5B)-H(5B)	0.97(2)
N(2A)-N(3A)	1.355(3) C(1B)-C(3B)	1.471(3)
N(2A)-H(2A)	1.13(2) C(2B)-C(12B)	1.453(3)
N(3A)-C(3A)	1.316(3) C(3B)-C(4B)	1.428(3)
N(4A)-C(11A)	1.358(3) C(4B)-C(11B)	1.386(3)
N(4A)-C(10A)	1.369(3) C(4B)-C(5B)	1.459(3)
N(4A)-H(4A)	0.78(2) C(5B)-C(6B)	1.396(3)
N(5A)-C(19A)	1.352(3) C(5B)-C(10B)	1.411(3)
N(5A)-C(18A)	1.388(3) C(6B)-C(7B)	1.388(3)
N(5A)-H(5A)	1.05(2) C(6B)-H(6B)	0.97(2)
C(1A)-C(3A)	1.462(3) C(7B)-C(8B)	1.390(4)
C(2A)-C(12A)	1.456(3) C(7B)-H(7B)	1.03(3)
C(3A)-C(4A)	1.446(3) C(8B)-C(9B)	1.379(3)
C(4A)-C(11A)	1.377(3) C(8B)-H(8B)	0.94(2)
C(4A)-C(5A)	1.452(3) C(9B)-C(10B)	1.398(3)
C(5A)-C(6A)	1.392(3) C(9B)-H(9B)	0.98(2)
C(5A)-C(10A)	1.412(3) C(11B)-H(11B)	0.949(15)
C(6A)-C(7A)	1.387(4) C(12B)-C(19B)	1.372(3)
C(6A)-H(6A)	0.99(2) C(12B)-C(13B)	1.442(3)
C(7A)-C(8A)	1.399(4) C(13B)-C(14B)	1.406(3)
C(7A)-H(7A)	1.12(2) C(13B)-C(18B)	1.409(3)
C(8A)-C(9A)	1.366(3) C(14B)-C(15B)	1.377(4)
C(8A)-H(8A)	0.87(2) C(14B)-H(14B)	0.97(2)
C(9A)-C(10A)	1.391(3) C(15B)-C(16B)	1.406(4)
C(9A)-H(9A)	0.92(2) C(15B)-H(15B)	0.97(2)
C(11A)-H(11A)	0.89(2) C(16B)-C(17B)	1.359(3)
C(12A)-C(19A)	1.380(3) C(16B)-H(16B)	1.00(2)
C(12A)-C(13A)	1.442(3) C(17B)-C(18B)	1.385(3)
C(13A)-C(18A)	1.402(3) C(17B)-H(17B)	0.92(3)
C(13A)-C(14A)	1.410(3) C(19B)-H(19B)	1.11(3)
C(14A)-C(15A)	1.371(4) O(1C)-C(3C)	1.224(3)
C(14A)-H(14A)	0.95(2) N(1C)-C(3C)	1.323(3)
C(15A)-C(16A)	1.399(4) N(1C)-C(2C)	1.452(3)
C(15A)-H(15A)	0.919(19) N(1C)-C(1C)	1.467(3)
C(16A)-C(17A)	1.380(3) C(1C)-H(1C1)	1.01(3)
C(16A)-C(17A) C(16A)-H(16A)	1.02(2) C(1C)-H(1C2)	0.95(2)
C(17A)-C(18A)	1.395(3) C(1C)-H(1C3)	1.03(2)
C(17A)-C(18A) C(17A)-H(17A)		
C(17A)-H(17A) C(19A)-H(19A)		0.96(3)
	1.05(2) C(2C)-H(2C2) 1.267(2) C(2C) H(2C2)	1.00(2) 0.99(2)
O(1B)-C(1B)	1.267(3) C(2C)-H(2C3)	
N(1B)-C(2B)	1.313(3) C(3C)-H(3C) 1.372(3) O(1D) C(2D)	1.12(3) 1.222(2)
N(1B)-C(1B) N(2B) N(2B)	1.372(3) $O(1D)-C(3D)$ 1.254(2) $N(1D) C(2D)$	1.233(3)
N(2B)-N(3B)	1.354(3) $N(1D)-C(3D)$	1.340(3)
N(2B)-C(2B)	1.360(3) N(1D)-C(1D)	1.443(3)
N(2B)-H(2B)	1.18(2) N(1D)-C(2D) 1.227(2) C(1D) U(1D1)	1.457(3)
N(3B)-C(3B)	1.327(3) C(1D)-H(1D1)	1.01(2)
N(4B)-C(11B)	1.362(3) C(1D)-H(1D2)	0.93(3)
N(4B)-C(10B)	1.369(3) C(1D)-H(1D3)	1.04(3)

C(2D)-H(2D1)	0.95(3)	C(7A)-C(6A)-H(6A)	120.1(13)
C(2D)-H(2D2)	1.05(3)	C(5A)-C(6A)-H(6A)	120.7(13)
C(2D)-H(2D3)	1.03(3)	C(6A)-C(7A)-C(8A)	121.2(2)
C(3D)-H(3D)	1.01(2)	C(6A)-C(7A)-H(7A)	121.7(12)
O(1E)-C(3E)	1.227(3)	C(8A)-C(7A)-H(7A)	117.1(12)
C(1E)-N(1E)	1.471(3)	C(9A)-C(8A)-C(7A)	121.0(2)
C(1E)-H(1E1)	0.96(3)	C(9A)-C(8A)-H(8A)	121.1(14)
C(1E)-H(1E2)	1.03(2)	C(7A)-C(8A)-H(8A)	117.8(14)
C(1E)-H(1E3)	1.04(2)	C(8A)-C(9A)-C(10A)	117.7(2)
N(1E)-C(3E)	1.319(3)	C(8A)-C(9A)-H(9A)	123.2(12)
N(1E)-C(2E)	1.442(4)	C(10A)-C(9A)-H(9A)	119.1(12)
C(2E)-H(2E1)	1.08(3)	N(4A)-C(10A)-C(9A)	129.2(2)
C(2E)-H(2E2)	1.07(3)	N(4A)-C(10A)-C(5A)	108.0(2)
C(2E)-H(2E3)	1.03(2)	C(9A)-C(10A)-C(5A)	122.7(2)
C(3E)-H(3E)	1.00(2)	N(4A)-C(11A)-C(4A)	110.1(2)
O(1F)-C(3F)	1.230(3)		122.8(13)
N(1F)-C(3F)	1.324(3)		126.9(13)
N(1F)-C(1F)	1.439(3)		106.6(2)
N(1F)-C(2F)	1.449(4)		126.5(2)
C(1F)-H(1F1)	0.99(3)	C(13A)-C(12A)-C(2A)	126.8(2)
C(1F)-H(1F2)	0.96(3)	C(18A)-C(13A)-C(14A)	118.3(2)
C(1F)-H(1F3)	1.00(3)	C(18A)-C(13A)-C(12A)	106.14(19)
C(2F)-H(2F1)	0.92(3)	C(14A)-C(13A)-C(12A)	135.6(2)
C(2F)-H(2F2)	0.91(2)	C(15A)-C(14A)-C(13A)	118.4(2)
C(2F)-H(2F3)	0.97(2)	C(15A)-C(14A)-H(14A)	124.9(14)
C(3F)-H(3F)		) C(13A)-C(14A)-H(14A)	116.3(14)
	<sup>×</sup>	C(14A)-C(15A)-C(16A)	122.5(2)
C(2A)-N(1A)-C(1A)	117.42(19)		115.8(13)
C(2A)-N(2A)-N(3A)	124.1(2)	C(16A)-C(15A)-H(15A)	121.6(13)
C(2A)-N(2A)-H(2A)	123.9(12)	C(17A)-C(16A)-C(15A)	120.3(2)
N(3A)-N(2A)-H(2A)	112.0(12)	C(17A)-C(16A)-H(16A)	117.6(12)
C(3A)-N(3A)-N(2A)	116.6(2)	C(15A)-C(16A)-H(16A)	122.1(12)
C(11A)-N(4A)-C(10A)	109.6(2)	C(16A)-C(17A)-C(18A)	117.4(2)
C(11A)-N(4A)-H(4A)	123.5(19)	C(16A)-C(17A)-H(17A)	125.1(13)
C(10A)-N(4A)-H(4A)	126.5(19)	C(18A)-C(17A)-H(17A)	117.2(14)
C(19A)-N(5A)-C(18A)	108.7(2)	N(5A)-C(18A)-C(17A)	128.7(2)
C(19A)-N(5A)-H(5A)	131.4(14)	N(5A)-C(18A)-C(13A)	108.3(2)
C(18A)-N(5A)-H(5A)	119.9(14)	C(17A)-C(18A)-C(13A)	123.0(2)
O(1A)-C(1A)-N(1A)	119.3(2)	N(5A)-C(19A)-C(12A)	110.2(2)
O(1A)-C(1A)-C(3A)	121.6(2)	N(5A)-C(19A)-H(19A)	121.5(13)
N(1A)-C(1A)-C(3A)	119.1(2)	C(12A)-C(19A)-H(19A)	128.2(13)
N(1A)-C(2A)-N(2A)	122.0(2)	C(2B)-N(1B)-C(1B)	117.96(18)
N(1A)-C(2A)-C(12A)	119.4(2)	N(3B)-N(2B)-C(2B)	123.8(2)
N(2A)-C(2A)-C(12A)	118.5(2)	N(3B)-N(2B)-H(2B)	111.8(10)
N(3A)-C(3A)-C(4A)	117.2(2)	C(2B)-N(2B)-H(2B)	124.4(10)
N(3A)-C(3A)-C(1A)	120.8(2)	C(3B)-N(3B)-N(2B)	116.68(19)
C(4A)-C(3A)-C(1A)	122.0(2)	C(11B)-N(4B)-C(10B)	109.2(2)
C(11A)-C(4A)-C(3A)	126.5(2)	C(11B)-N(4B)-H(4B)	125.0(15)
C(11A)-C(4A)-C(5A)	106.0(2)	C(10B)-N(4B)-H(4B)	124.3(15)
C(3A)-C(4A)-C(5A)	127.4(2)	C(19B)-N(5B)-C(18B)	108.5(2)
C(6A)-C(5A)-C(10A)	118.1(2)	C(19B)-N(5B)-H(5B)	127.4(14)
C(6A)-C(5A)-C(4A)	135.5(2)	C(18B)-N(5B)-H(5B)	124.1(14)
C(10A)-C(5A)-C(4A)	106.27(19)	O(1B)-C(1B)-N(1B)	120.0(2)
C(7A)-C(6A)-C(5A)	119.2(2)	O(1B)-C(1B)-C(3B)	120.9(2)

N(1B)-C(1B)-C(3B)	119.10(19)	C(12B)-C(19B)-H(19B)	125.5(15)
N(1B)-C(2B)-N(2B)	122.1(2)	C(3C)-N(1C)-C(2C)	120.5(2)
N(1B)-C(2B)-C(12B)	120.1(2)	C(3C)-N(1C)-C(1C)	122.0(2)
N(2B)-C(2B)-C(12B)	117.7(2)	C(2C)-N(1C)-C(1C)	117.2(2)
N(3B)-C(3B)-C(4B)	117.5(2)	N(1C)-C(1C)-H(1C1)	105.6(14)
N(3B)-C(3B)-C(1B)	120.4(2)	N(1C)-C(1C)-H(1C2)	106.9(14)
C(4B)-C(3B)-C(1B)	122.1(2)	H(1C1)-C(1C)-H(1C2)	120(2)
C(11B)-C(4B)-C(3B)	126.6(2)	N(1C)-C(1C)-H(1C3)	111.8(12)
C(11B)-C(4B)-C(5B)	105.5(2)	H(1C1)-C(1C)-H(1C3)	102.1(19)
C(3B)-C(4B)-C(5B)	127.9(2)	H(1C2)-C(1C)-H(1C3)	110.1(18)
C(6B)-C(5B)-C(10B)	118.8(2)	N(1C)-C(2C)-H(2C1)	113.8(17)
C(6B)-C(5B)-C(4B)	134.9(2)	N(1C)-C(2C)-H(2C2)	112.0(13)
C(10B)-C(5B)-C(4B)	106.3(2)	H(2C1)-C(2C)-H(2C2)	103(2)
C(7B)-C(6B)-C(5B)	118.1(2)	N(1C)-C(2C)-H(2C3)	108.3(13)
C(7B)-C(6B)-H(6B)	122.7(13)	H(2C1)-C(2C)-H(2C3)	118(2)
C(5B)-C(6B)-H(6B)	119.2(13)	H(2C2)-C(2C)-H(2C3)	100.3(18)
C(6B)-C(7B)-C(8B)	122.3(2)	O(1C)-C(3C)-N(1C)	126.0(3)
C(6B)-C(7B)-H(7B)	117.8(15)	O(1C)-C(3C)-H(3C)	124.0(13)
C(8B)-C(7B)-H(7B)	119.9(15)	N(1C)-C(3C)-H(3C)	109.5(13)
C(9B)-C(8B)-C(7B)	121.0(2)	C(3D)-N(1D)-C(1D)	121.5(2)
C(9B)-C(8B)-H(8B)	120.4(13)	C(3D)-N(1D)-C(2D)	120.5(2)
C(7B)-C(8B)-H(8B)	118.5(13)	C(1D)-N(1D)-C(2D)	117.8(2)
C(8B)-C(9B)-C(10B)	117.0(2)	N(1D)-C(1D)-H(1D1)	112.4(12)
C(8B)-C(9B)-H(9B)	125.5(13)	N(1D)-C(1D)-H(1D2)	113.0(19)
C(10B)-C(9B)-H(9B)	117.4(13)	H(1D1)-C(1D)-H(1D2)	113(2)
N(4B)-C(10B)-C(9B)	128.6(2)	N(1D)-C(1D)-H(1D3)	115.0(16)
N(4B)-C(10B)-C(5B)	108.5(2)	H(1D1)-C(1D)-H(1D3)	102(2)
C(9B)-C(10B)-C(5B)	122.8(2)	H(1D2)-C(1D)-H(1D3)	101(2)
N(4B)-C(11B)-C(4B)	110.5(2)	N(1D)-C(2D)-H(2D1)	110.4(16)
N(4B)-C(11B)-H(11B)	122.1(10)	N(1D)-C(2D)-H(2D2)	104.6(15)
C(4B)-C(11B)-H(11B)	127.4(10)	H(2D1)-C(2D)-H(2D2)	125(2)
C(19B)-C(12B)-C(13B)	106.9(2)	N(1D)-C(2D)-H(2D3)	110.6(14)
C(19B)-C(12B)-C(2B)	126.8(2)	H(2D1)-C(2D)-H(2D3)	97.8(19)
C(13B)-C(12B)-C(2B)	126.4(2)	H(2D2)-C(2D)-H(2D3)	108(2)
C(14B)-C(13B)-C(18B)	118.0(2)	O(1D)-C(3D)-N(1D)	125.5(3)
C(14B)-C(13B)-C(12B)	136.2(2)	O(1D)-C(3D)-H(3D)	121.1(11)
C(18B)-C(13B)-C(12B)	105.7(2)	N(1D)-C(3D)-H(3D)	113.3(11)
C(15B)-C(14B)-C(13B)	118.6(2)	N(1E)-C(1E)-H(1E1)	109.9(14)
C(15B)-C(14B)-H(14B)	118.6(13)	N(1E)-C(1E)-H(1E2)	109.0(11)
C(13B)-C(14B)-H(14B)	122.7(13)	H(1E1)-C(1E)-H(1E2)	109.7(18)
C(14B)-C(15B)-C(16B)	121.8(2)	N(1E)-C(1E)-H(1E3)	107.0(12)
C(14B)-C(15B)-H(15B)	117.8(13)	H(1E1)-C(1E)-H(1E3)	109.0(19)
C(16B)-C(15B)-H(15B)	120.3(12)	H(1E2)-C(1E)-H(1E3)	112.2(17)
C(17B)-C(16B)-C(15B)	120.4(2)	C(3E)-N(1E)-C(2E)	120.6(2)
C(17B)-C(16B)-H(16B)	117.9(12)	C(3E)-N(1E)-C(1E)	120.8(2)
C(15B)-C(16B)-H(16B)	121.5(12)	C(2E)-N(1E)-C(1E)	118.6(2)
C(16B) - C(17B) - C(18B)	118.4(2)	N(1E)-C(2E)-H(2E1)	107.6(13)
C(16B) - C(17B) - H(17B)	125.0(15)	N(1E) - C(2E) - H(2E2) N(1E) - C(2E) - H(2E2)	111.8(18)
C(18B)-C(17B)-H(17B)	116.5(15)	H(1E) C(2E) H(2E2) H(2E1)-C(2E)-H(2E2)	128(2)
C(17B)-C(18B)-N(5B)	129.0(2)	N(1E)-C(2E)-H(2E3)	107.9(13)
C(17B)-C(18B)-C(13B)	122.7(2)	H(2E1)-C(2E)-H(2E3)	107.4(18)
N(5B)-C(18B)-C(13B)	122.7(2) 108.3(2)	H(2E1)-C(2E)-H(2E3) H(2E2)-C(2E)-H(2E3)	91(2)
N(5B)-C(19B)-C(12B)	110.6(2)	O(1E)-C(3E)-N(1E)	125.3(3)
N(5B)-C(19B)-H(19B)	123.7(15)	O(1E)-C(3E)-H(3E)	118.7(13)
	123.1(13)		110.7(13)

$\begin{array}{l} N(1E)-C(3E)-H(3E)\\ C(3F)-N(1F)-C(1F)\\ C(3F)-N(1F)-C(2F)\\ C(1F)-N(1F)-C(2F)\\ N(1F)-C(1F)-H(1F1)\\ N(1F)-C(1F)-H(1F2)\\ H(1F1)-C(1F)-H(1F3)\\ H(1F1)-C(1F)-H(1F3)\\ H(1F1)-C(1F)-H(1F3)\\ H(1F2)-C(1F)-H(1F3)\\ N(1F)-C(2F)-H(2F1)\\ N(1F)-C(2F)-H(2F2)\\ H(2F1)-C(2F)-H(2F2)\\ N(1F)-C(2F)-H(2F3)\\ \end{array}$	$\begin{array}{c} 115.9(13)\\ 121.7(2)\\ 119.8(2)\\ 118.5(2)\\ 111.1(15)\\ 106.9(16)\\ 98(2)\\ 109.4(16)\\ 116(2)\\ 114(2)\\ 116.7(17)\\ 106.1(14)\\ 126(2)\\ 108.5(13)\\ \end{array}$
H(2F1)-C(2F)-H(2F2)	126(2)

Anisotropic displacement parameters (Å<sup>2</sup>x 10<sup>4</sup>) for 187 (CCDC 259291). The anisotropic displacement factor exponent takes the form:  $-2\pi^2$  [  $h^2 a^{*2}U^{11} + ... + 2 h k a^* b^* U^{12}$ ].

	$U^{11}$	U <sup>22</sup>	U <sup>33</sup>	U <sup>23</sup>	U <sup>13</sup>	$U^{12}$
(1A)	227(8)	207(8)	297(8)	156(7)	9(6)	23(7)
(1A)	150(10)	200(10)	197(10)	88(9)	2(8)	44(8)
(2A)	143(10)	209(10)	230(11)	88(9)	19(8)	41(8)
(3A)	179(11)	206(11)	180(11)	63(9)	-10(9)	76(9)
4A)	130(11)	193(11)	204(12)	92(10)	6(9)	13(9)
5A)	181(11)	191(11)	195(11)	91(9)	17(9)	33(9)
1A)	180(13)	226(13)	141(12)	49(10)	26(10)	70(11)
2A)	206(13)	123(11)	139(12)	51(10)	17(10)	91(10)
3A)	181(12)	166(12)	114(12)	47(10)	29(10)	46(10)
4A)	185(13)	153(12)	140(12)	76(10)	25(10)	87(10)
5A)	147(13)	162(12)	131(13)	48(11)	39(10)	72(10)
6A)	185(13)	177(12)	196(13)	79(11)	19(10)	70(10)
7A)	286(13)	187(12)	213(13)	97(11)	39(11)	83(11)
8A)	279(15)	281(14)	203(14)	115(12)	18(12)	167(12)
9A)	148(12)	265(14)	170(13)	61(11)	-27(10)	84(11)
10A)	191(14)	158(13)	152(13)	59(11)	48(11)	72(11)
11A)	188(13)	166(12)	172(13)	95(11)	11(10)	40(10)
11A)	189(13)	151(12)	146(13)	38(11)	11(10) 12(10)	39(10)
12A)	194(14)	131(12) 144(12)	132(13)	38(11)	20(11)	65(10)
13A) 14A)	222(14)	190(13)	171(13)	79(11)	51(11)	78(11)
			201(13)	136(12)	79(12)	151(12)
15A)	379(16)	226(13)				
16A)	305(15)	267(14)	145(13)	61(12)	-11(11)	176(12)
17A)	171(13)	225(13)	178(13)	48(11)	20(11)	81(11)
18A)	185(14)	189(13)	127(13)	43(11)	24(11)	94(11) 72(10)
19A)	201(13)	191(13)	152(13)	69(11)	6(10)	73(10)
1B)	281(9)	189(9)	267(8)	130(7)	-10(6)	15(7)
1B)	147(10)	199(10)	155(11)	85(9)	32(8)	59(8)
(2B)	162(10)	188(10)	167(11)	45(9)	-2(8)	20(8)
3B)	153(11)	256(11)	169(11)	93(10)	-1(9)	55(9)
4B)	149(10)	174(11)	179(10)	57(9)	-14(8)	23(9)
(5B)	226(11)	130(10)	213(11)	82(9)	31(9)	39(9)
1B)	133(12)	234(13)	177(13)	80(11)	33(10)	83(10)
2B)	135(12)	195(12)	131(12)	75(10)	37(9)	68(10)
3B)	146(12)	174(12)	200(13)	100(10)	62(10)	85(10)
4B)	154(13)	171(12)	164(13)	98(11)	53(10)	61(10)
5B)	138(13)	179(13)	124(13)	63(11)	10(10)	68(10)
6B)	155(13)	195(13)	210(14)	91(12)	24(11)	42(11)
7B)	228(14)	261(14)	260(14)	154(12)	69(11)	86(11)
8B)	266(14)	340(15)	217(14)	167(12)	45(12)	151(12)
9B)	186(13)	253(14)	162(13)	53(12)	-5(11)	87(11)
10B)	176(13)	194(13)	214(14)	103(12)	77(11)	97(11)
11B)	193(13)	148(12)	195(13)	84(11)	26(10)	60(10)
12B)	144(13)	207(13)	192(13)	100(11)	40(11)	82(11)
120,			. ,			
13B)	143(13)	177(13)	184(14)	82(12)	00(11)	75(11)
	143(13) 162(13)	177(13) 180(13)	184(14) 235(14)	82(12) 108(12)	68(11) 28(11)	68(11)

C(16B)	214(13)	296(13)	145(13)	113(11)	28(10)	115(11)
C(10B) C(17B)	177(13)	230(13) 231(14)	143(13)	54(11)	28(10) 24(11)	95(11)
C(18B)	152(13)	188(13)	150(13)	62(11)	62(10)	71(11)
C(19B)	157(12)	242(13)	205(13)	134(11)	48(10)	73(10)
0.40	100/10				2 (2)	
O(1C)	190(10)	320(11)	365(12)	236(10)	-8(8)	12(8)
N(1C)	197(12)	220(11)	224(12)	132(10)	40(10)	70(9)
C(1C)	240(15)	188(14)	250(16)	91(13)	44(13)	33(12)
C(2C)	197(15)	208(15)	242(16)	79(13)	14(13)	63(12)
C(3C)	206(15)	294(15)	264(16)	126(14)	-2(12)	56(12)
O(1D)	245(10)	290(10)	333(11)	186(9)	-9(8)	25(8)
N(1D)	175(12)	208(11)	173(12)	62(10)	-1(9)	33(9)
C(1D)	228(16)	254(15)	281(16)	139(14)	71(13)	91(13)
C(2D)	233(15)	259(15)	210(15)	109(13)	11(12)	84(13)
C(3D)	147(14)	286(15)	271(16)	140(14)	16(12)	14(12)
				× /		
O(1E)	300(11)	244(10)	257(10)	102(9)	74(8)	46(8)
C(1E)	381(18)	276(15)	279(16)	139(14)	113(15)	157(13)
N(1E)	185(12)	216(12)	205(12)	76(10)	23(10)	37(10)
C(2E)	260(17)	356(18)	298(18)	91(16)	-42(14)	28(14)
C(3E)	194(14)	212(14)	225(14)	64(12)	62(11)	77(11)
0()		(_ ')	()	- (())	()	( )
O(1F)	325(11)	220(10)	277(10)	92(9)	84(8)	24(8)
N(1F)	200(12)	172(11)	206(12)	58(10)	31(10)	21(9)
C(1F)	346(18)	230(15)	336(18)	121(14)	130(15)	85(13)
C(2F)	231(17)	386(19)	276(18)	23(15)	8(14)	2(14)
C(3F)	226(14)	237(14)	194(14)	42(12)	51(12)	101(12)
	(1 )	207(11)	12 ((1))	12(12)	21(12)	101(12)

	х	У	Z	U <sub>iso</sub>
H(2A)	7960(30)	1770(20)	2381(19)	20(6)
H(4A)	-1870(30)	-1700(20)	990(20)	29(8)
H(5A)	13580(30)	2540(20)	4300(20)	35(7)
H(6A)	3630(30)	1540(20)	951(18)	26(7)
I(7A)	1900(30)	1990(20)	-280(20)	40(8)
I(8A)	-1140(30)	766(19)	-836(17)	16(6)
H(9A)	-2640(30)	-686(17)	-282(16)	9(5)
I(11Å)	520(30)	-1685(18)	1995(16)	7(5)
I(14A)	8070(30)	-930(20)	4350(20)	28(7)
I(15A)	9970(30)	-1226(19)	5467(17)	11(6)
(16A)	13160(30)	-29(19)	6107(18)	27(6)
I(17A)	14380(30)	1530(20)	5480(20)	40(8)
I(19A)	10670(30)	2380(20)	3200(19)	31(7)
	( )	~ /		
H(2B)	2370(30)	7308(18)	2954(16)	3(5)
H(4B)	-4620(30)	1430(20)	-1230(20)	36(8)
I(5B)	6280(30)	9100(20)	6168(19)	30(7)
H(6B)	-600(30)	5950(20)	240(18)	21(6)
I(7B)	-1760(40)	6050(20)	-1360(20)	49(8)
I(8B)	-4010(30)	4313(19)	-2697(18)	23(6)
I(9B)	-5080(30)	2410(20)	-2557(19)	23(6)
I(11B)	-3050(20)	2115(14)	596(13)	-11(4)
I(14B)	2160(30)	4610(20)	5112(19)	18(6)
I(15B)	3430(30)	4747(19)	6714(17)	11(6)
(16B)	5680(30)	6603(18)	8059(17)	19(6)
I(17B)	6720(30)	8200(20)	7560(20)	30(7)
(19B)	4400(40)	8310(30)	4270(20)	71(10)
(101)	2020(40)	4260(20)	(0/20)	41(0)
H(1C1)	2920(40)	4260(20)	60(20)	41(8)
H(1C2)	3250(30)	4620(20)	1421(19)	12(6)
I(1C3)	4590(30) 7100(40)	4070(19)	582(17)	12(6)
I(2C1)	7190(40)	6290(20)	1920(20)	44(9)
(2C2)	5720(30)	6484(19)	2659(19)	18(6)
I(2C3)	6350(30)	7440(20)	2241(18)	16(6)
I(3C)	3480(40)	5630(20)	-380(20)	44(8)
I(1D1)	8810(30)	6310(20)	4984(19)	21(7)
H(1D2)	7010(40)	6420(30)	4470(30)	61(10)
H(1D3)	8390(40)	6030(20)	3730(20)	40(8)
I(2D1)	4910(40)	3160(20)	3020(20)	40(8)
l(2D1)	5850(40)	4290(20)	2580(20)	46(8)
(2D2)	4150(40)	4180(20)	3330(20)	25(7)
I(3D)	8070(30)	4769(18)	5416(16)	3(5)
I(1E1)	3120(30)	7960(20)	9480(20)	26(7)
I(1E2)	2010(30)	6708(18)	8405(16)	10(5)
I(1E3)	920(30)	7620(20)	9295(19)	22(6)
I(2E1)	1120(30)	8390(20)	6950(20)	52(8)

Hydrogen coordinates ( x 10<sup>4</sup>) and isotropic displacement parameters (Å<sup>2</sup>x 10<sup>3</sup>) for 187 (CCDC 259291).

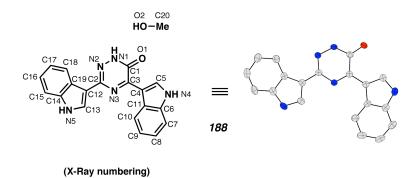
H(2E2)	-540(50)	7400(30)	7500(30)	82(11)
H(2E3)	800(30)	6960(20)	6790(20)	45(7)
H(3E)	4450(30)	9540(20)	9178(19)	22(6)
H(1F1)	10710(40)	3080(20)	5990(20)	38(8)
H(1F2)	10010(40)	3900(30)	6900(20)	54(9)
H(1F3)	8450(40)	2850(20)	5850(20)	46(9)
H(2F1)	10320(40)	3040(20)	8530(20)	58(8)
H(2F2)	11780(30)	2930(20)	7668(18)	19(6)
H(2F3)	10880(30)	1970(20)	7832(18)	34(7)
H(3F)	7190(30)	1099(17)	5830(16)	1(5)

Hydrogen bonds for 187 (CCDC 259291) [Å and °].

D-HA	d(D-H)	d(HA)	d(DA)	<(DHA)
N(2A)-H(2A)O(1B)#1	1.13(2)	1.60(2)	2.700(2)	164(2)
N(4A)-H(4A)O(1C)#2	0.78(2)	2.08(3)	2.836(3)	162(3)
N(5A)-H(5A)O(1D)#1	1.05(2)	1.83(2)	2.842(3)	161(2)
N(2B)-H(2B)O(1A)#3	1.18(2)	1.498(19)	2.656(2)	165.6(17)
N(4B)-H(4B)O(1E)#4	0.94(3)	1.83(3)	2.765(3)	169(2)
N(5B)-H(5B)O(1F)#3	0.97(2)	1.76(2)	2.725(3)	171(2)

Symmetry transformations used to generate equivalent atoms: #1 x+1,y,z #2 x-1,y-1,z #3 x,y+1,z #4 x-1,y-1,z-1

## A6.6.2 X-Ray Crystallographic Report for *m*-Triazinone 188



## Crystal data and structure refinement for 188 (CCDC 161494).

Empirical formula	C20 H17 N5 O2
Formula weight	359.39
Crystallization Solvent	Methanol
Crystal Habit	needle
Crystal size	$0.33 \text{ x} 0.14 \text{ x} 0.09 \text{ mm}^3$
Crystal color	yellow

#### **Data Collection**

Preliminary Photos	none	
Type of diffractometer	CCD	
Wavelength	0.71073 Å MoKα	
Data Collection Temperature	98(2) K	
$\theta$ range for 7767 reflections used in lattice determination	2.20 to 27.35°	
Unit cell dimensions	a = 4.6579(4) Å b = 18.5134(15) Å c = 19.5156(16) Å	$\alpha = 90^{\circ}$ $\beta = 90^{\circ}$ $\gamma = 90^{\circ}$
Volume	1682.9(2) Å <sup>3</sup>	
Ζ	4	
Crystal system	Orthorhombic	
Space group	P2(1)2(1)2(1)	
Density (calculated)	1.418 Mg/m <sup>3</sup>	
F(000)	752	

Data collection program	Bruker SMART
$\theta$ range for data collection	2.09 to 28.53°
Completeness to $\theta = 28.53^{\circ}$	95.9 %
Index ranges	-6<=h<=6, -24<=k<=24, -25<=l<=25
Data collection scan type	phi and omega scans
Data reduction program	Bruker SAINT 6.2
Reflections collected	20360
Independent reflections	4014 ( $R_{int} = 0.0620$ )
Absorption coefficient	0.096 mm <sup>-1</sup>
Absorption correction	None
Structure solution program	SHELXS-97 (Sheldrick, 1990)
Primary solution method	direct
Secondary solution method	difmap
Hydrogen placement	geom
Structure refinement program	SHELXL-97 (Sheldrick, 1997)
Refinement method	Full-matrix least-squares on F <sup>2</sup>
Data / restraints / parameters	4014 / 0 / 312
Treatment of hydrogen atoms	refall
Goodness-of-fit on F <sup>2</sup>	0.903
Final R indices [I> $2\sigma$ (I), 3238 reflections]	R1 = 0.0367, wR2 = 0.0833
R indices (all data)	R1 = 0.0502, $wR2 = 0.0890$
Type of weighting scheme used	calc
Weighting scheme used	calc w=1/[^2^(Fo^2^)+(0.0569P)^2^+0.0000P] where
P=(Fo^2^+2Fc^2^)/3	
Max shift/error	0.010
Average shift/error	0.001
Absolute structure parameter	0.2(12)
Largest diff. peak and hole	0.200 and -0.175 e.Å <sup>-3</sup>

Atomic coordinates ( x 10<sup>4</sup>) and equivalent isotropic displacement parameters (Å<sup>2</sup>x 10<sup>3</sup>) for 188 (CCDC 161494). U(eq) is defined as the trace of the orthogonalized U<sup>ij</sup> tensor.

	X	У	Z	U <sub>eq</sub>
O(1)	8039(3)	8255(1)	10244(1)	29(1)
C(1)	6801(3)	8516(1)	9727(1)	24(1)
C(2)	3843(3)	9014(1)	8628(1)	22(1)
C(3)	7333(3)	9241(1)	9441(1)	22(1)
C(4)	9428(3)	9713(1)	9754(1)	22(1)
C(5)	11233(4)	9558(1)	10295(1)	27(1)
C(6)	12174(4)	10705(1)	10005(1)	29(1)
C(7)	13284(4)	11402(1)	9982(1)	36(1)
C(8)	12196(4)	11856(1)	9489(1)	40(1)
C(9)	10072(4)	11624(1)	9032(1)	39(1)
C(10)	8977(4)	10932(1)	9058(1)	32(1)
C(11)	10029(3)	10457(1)	9559(1)	25(1)
C(12)	2242(3)	9293(1)	8049(1)	24(1)
C(13)	2428(4)	9992(1)	7815(1)	29(1)
C(14)	-771(4)	9450(1)	7138(1)	29(1)
C(15)	-2803(4)	9291(1)	6634(1)	34(1)
C(16)	-3858(4)	8598(1)	6607(1)	38(1)
C(17)	-2948(4)	8074(1)	7078(1)	37(1)
C(18)	-961(4)	8231(1)	7582(1)	30(1)
C(19)	177(3)	8932(1)	7619(1)	25(1)
N(1)	4781(3)	8142(1)	9396(1)	26(1)
N(2)	3216(3)	8360(1)	8843(1)	25(1)
N(3)	5856(3)	9465(1)	8911(1)	22(1)
N(4)	12866(3)	10143(1)	10444(1)	31(1)
N(5)	634(3)	10087(1)	7271(1)	32(1)
C(20)	8632(6)	8407(1)	11949(1)	48(1)
O(2)	6379(4)	8685(1)	11542(1)	54(1)

O(1)-C(1)	1.259(2)	N(3)-C(3)-C(4)	119.81(13)
C(1)-N(1)	1.335(2)	N(3)-C(3)-C(1)	119.89(14)
C(1)-C(3)	1.474(2)	C(4)-C(3)-C(1)	120.29(13)
C(2)-N(2)	1.313(2)	C(5)-C(4)-C(3)	127.56(14)
C(2)-N(3)	1.373(2)	C(5)-C(4)-C(11)	106.19(14)
C(2)-C(12)	1.450(2)	C(3)-C(4)-C(11)	126.25(14)
C(3)-N(3)	1.309(2)	N(4)-C(5)-C(4)	109.93(14)
C(3)-C(4)	1.445(2)	N(4)-C(6)-C(7)	129.08(17)
C(4)-C(5)	1.379(2)	N(4)-C(6)-C(11)	107.66(14)
C(4)-C(11)	1.457(2)	C(7)-C(6)-C(11)	123.26(16)
C(5)-N(4)	1.355(2)	C(8)-C(7)-C(6)	116.94(18)
C(6)-N(4)	1.385(2)	C(7)-C(8)-C(9)	121.24(17)
C(6)-C(7)	1.390(2)	C(10)-C(9)-C(8)	121.55(18)
C(6)-C(11)	1.403(2)	C(9)-C(10)-C(11)	118.56(17)
C(7)-C(8)	1.376(3)	C(6)-C(11)-C(10)	118.44(15)
C(8)-C(9)	1.399(3)	C(6)-C(11)-C(4)	106.52(14)
C(9)-C(10)	1.379(2)	C(10)-C(11)-C(4)	135.04(16)
C(10)-C(11)	1.404(2)	C(13)-C(12)-C(19)	106.59(14)
C(12)-C(13)	1.374(2)	C(13)-C(12)-C(2)	124.22(15)
C(12)-C(19)	1.441(2)	C(19)-C(12)-C(2)	129.18(13)
C(13)-N(5)	1.362(2)	N(5)-C(13)-C(12)	109.97(15)
C(14)-N(5)	1.373(2)	N(5)-C(14)-C(15)	129.58(16)
C(14)-C(15)	1.396(2)	N(5)-C(14)-C(19)	107.96(14)
C(14)-C(19)	1.412(2)	C(15)-C(14)-C(19)	122.46(16)
C(15)-C(16)	1.374(3)	C(16)-C(15)-C(14)	117.76(16)
C(16)-C(17)	1.402(3)	C(15)-C(16)-C(17)	120.89(17)
C(17)-C(18)	1.382(2)	C(18)-C(17)-C(16)	121.47(17)
C(18)-C(19)	1.403(2)	C(17)-C(18)-C(19)	119.00(16)
N(1)-N(2)	1.364(2)	C(18)-C(19)-C(14)	118.41(15)
C(20)-O(2)	1.413(3)	C(18)-C(19)-C(12)	135.30(15)
		C(14)-C(19)-C(12)	106.29(14)
O(1)-C(1)-N(1)	120.81(14)	C(1)-N(1)-N(2)	127.30(13)
O(1)-C(1)-C(3)	125.10(14)	C(2)-N(2)-N(1)	114.03(13)
N(1)-C(1)-C(3)	114.08(13)	C(3)-N(3)-C(2)	118.98(12)
N(2)-C(2)-N(3)	125.72(13)	C(5)-N(4)-C(6)	109.69(15)
N(2)-C(2)-C(12)	117.63(14)	C(13)-N(5)-C(14)	109.19(14)
N(3)-C(2)-C(12)	116.64(13)		

Selected bond lengths [Å] and angles [°] for 188 (CCDC 161494).

# Bond lengths [Å] and angles [°] for 188 (CCDC 161494).

O(1)-C(1)	1.259(2)	N(2)-C(2)-N(3)	125.72(13)
C(1)-N(1)	1.335(2)	N(2)-C(2)-C(12)	117.63(14)
C(1)-C(3)	1.474(2)	N(3)-C(2)-C(12)	116.64(13)
C(2)-N(2)	1.313(2)	N(3)-C(3)-C(4)	119.81(13)
C(2)-N(3)	1.373(2)	N(3)-C(3)-C(1)	119.89(14)
C(2)-C(12)	1.450(2)	C(4)-C(3)-C(1)	120.29(13)
C(3)-N(3)	1.309(2)	C(5)-C(4)-C(3)	127.56(14)
C(3)-C(4)	1.445(2)	C(5)-C(4)-C(11)	106.19(14)
C(4)-C(5)	1.379(2)	C(3)-C(4)-C(11)	126.25(14)
C(4)-C(11)	1.457(2)	N(4)-C(5)-C(4)	109.93(14)
C(5)-N(4)	1.355(2)	N(4)-C(5)-H(5)	120.6(10)
C(5)-H(5)	0.99(2)	C(4)-C(5)-H(5)	129.5(10)
C(6)-N(4)	1.385(2)	N(4)-C(6)-C(7)	129.08(17)
C(6)-C(7)	1.390(2)	N(4)-C(6)-C(11)	107.66(14)
C(6)-C(11)	1.403(2)	C(7)-C(6)-C(11)	123.26(16)
C(7)-C(8)	1.376(3)	C(8)-C(7)-C(6)	116.94(18)
C(7)-H(7)	0.96(2)	C(8)-C(7)-H(7)	122.3(11)
C(8)-C(9)	1.399(3)	C(6)-C(7)-H(7)	120.7(11)
C(8)-H(8)	0.96(2)	C(7)-C(8)-C(9)	121.24(17)
C(9)-C(10)	1.379(2)	C(7)-C(8)-H(8)	119.5(10)
C(9)-H(9)	0.98(2)	C(9)-C(8)-H(8)	119.2(10)
C(10)-C(11)	1.404(2)	C(10)-C(9)-C(8)	121.55(18)
C(10)-H(10)	0.99(2)	C(10)-C(9)-H(9)	118.4(13)
C(12)-C(13)	1.374(2)	C(8)-C(9)-H(9)	120.1(12)
C(12)-C(19)	1.441(2)	C(9)-C(10)-C(11)	118.56(17)
C(13)-N(5)	1.362(2)	C(9)-C(10)-H(10)	122.7(10)
C(13)-H(13)	1.03(2)	C(11)-C(10)-H(10)	118.7(10)
C(14)-N(5)	1.373(2)	C(6)-C(11)-C(10)	118.44(15)
C(14)-C(15)	1.396(2)	C(6)-C(11)-C(4)	106.52(14)
C(14)-C(19)	1.412(2)	C(10)-C(11)-C(4)	135.04(16)
C(15)-C(16)	1.374(3)	C(13)-C(12)-C(19)	106.59(14)
C(15)-H(15)	1.02(2)	C(13)-C(12)-C(2)	124.22(15)
C(16)-C(17)	1.402(3)	C(19)-C(12)-C(2)	129.18(13)
C(16)-H(16)	0.98(2)	N(5)-C(13)-C(12)	109.97(15)
C(17)-C(18)	1.382(2)	N(5)-C(13)-H(13)	120.9(9)
C(17)-H(17)	1.00(2)	C(12)-C(13)-H(13)	129.0(9)
C(18)-C(19)	1.403(2)	N(5)-C(14)-C(15)	129.58(16)
C(18)-H(18)	1.00(2)	N(5)-C(14)-C(19)	107.96(14)
N(1)-N(2)	1.364(2)	C(15)-C(14)-C(19)	122.46(16)
N(1)-H(1N)	0.93(2)	C(16)-C(15)-C(14)	117.76(16)
N(4)-H(4N)	0.87(2)	C(16)-C(15)-H(15)	122.6(11)
N(5)-H(5N)	0.90(2)	C(14)-C(15)-H(15)	119.5(11)
C(20)-O(2)	1.413(3)	C(15)-C(16)-C(17)	120.89(17)
C(20)-H(20A)	0.93(3)	C(15)-C(16)-H(16)	119.1(11)
C(20)-H(20B)	1.02(2)	C(17)-C(16)-H(16)	119.8(11)
C(20)-H(20C)	1.02(2)	C(18)-C(17)-C(16)	121.47(17)
O(2)-H(2O)	0.83(2)	C(18)-C(17)-H(17)	118.8(10)
	0.00(2)	C(16)-C(17)-H(17)	119.7(10)
O(1)-C(1)-N(1)	120.81(14)	C(10) C(17) H(17) C(17) - C(18) - C(19)	119.00(16)
O(1) - C(1) - C(3)	125.10(14)	C(17)-C(18)-H(18)	121.2(11)
N(1)-C(1)-C(3)	114.08(13)	C(19)-C(18)-H(18)	119.8(11)
	11.00(15)		112.0(11)

C(18)-C(19)-C(14) C(18)-C(19)-C(12)	118.41(15) 135.30(15)
C(14)-C(19)-C(12)	106.29(14)
C(1)-N(1)-N(2)	127.30(13)
C(1)-N(1)-H(1N)	119.5(11)
N(2)-N(1)-H(1N)	113.2(11)
C(2)-N(2)-N(1)	114.03(13)
C(3)-N(3)-C(2)	118.98(12)
C(5)-N(4)-C(6)	109.69(15)
C(5)-N(4)-H(4N)	124.2(14)
C(6)-N(4)-H(4N)	125.9(14)
C(13)-N(5)-C(14)	109.19(14)
C(13)-N(5)-H(5N)	130.4(13)
C(14)-N(5)-H(5N)	120.2(13)
O(2)-C(20)-H(20A)	108.3(17)
O(2)-C(20)-H(20B)	105.9(15)
H(20A)-C(20)-H(20B)	108(2)
O(2)-C(20)-H(20C)	106.6(15)
H(20A)-C(20)-H(20C)	118(2)
H(20B)-C(20)-H(20C)	109.4(18)
C(20)-O(2)-H(2O)	103.1(17)

Anisotropic displacement parameters (Å<sup>2</sup>x 10<sup>4</sup>) for 188 (CCDC 161494). The anisotropic displacement factor exponent takes the form:  $-2\pi^2$  [  $h^2 a^{*2}U^{11} + ... + 2h$  k a\* b\* U<sup>12</sup> ].

	U <sup>11</sup>	U <sup>22</sup>	U <sup>33</sup>	U <sup>23</sup>	U <sup>13</sup>	U <sup>12</sup>
O(1)	380(7)	228(5)	264(6)	30(4)	-52(5)	27(5)
C(1)	272(9)	216(7)	226(8)	-6(6)	17(7)	32(6)
C(2)	225(8)	225(7)	215(7)	-11(6)	42(6)	26(6)
C(3)	212(8)	212(7)	224(7)	-14(6)	41(6)	27(6)
C(4)	215(8)	216(7)	243(7)	-22(6)	33(6)	27(6)
C(5)	247(8)	306(8)	248(8)	-7(6)	9(7)	12(7)
C(6)	258(8)	306(8)	295(8)	-57(6)	54(7)	-11(7)
C(7)	284(10)	343(9)	442(11)	-110(8)	65(8)	-83(8)
C(8)	381(10)	264(8)	557(12)	-69(8)	142(10)	-76(8)
C(9)	369(10)	274(8)	517(11)	48(8)	86(9)	8(8)
C(10)	293(9)	267(8)	386(10)	16(7)	16(8)	4(7)
C(11)	220(8)	232(7)	303(8)	-41(6)	55(7)	-1(6)
C(12)	215(8)	263(7)	229(7)	-2(6)	37(6)	20(6)
C(13)	303(9)	295(8)	270(8)	31(6)	16(7)	20(7)
C(14)	237(8)	377(9)	240(8)	15(7)	39(7)	40(7)
C(15)	246(9)	528(11)	248(8)	50(8)	9(7)	70(8)
C(16)	264(9)	598(12)	272(9)	-40(8)	-20(8)	-9(9)
C(17)	326(10)	451(10)	323(9)	-74(8)	15(8)	-50(9)
C(18)	292(9)	359(9)	262(8)	-19(7)	13(7)	14(7)
C(19)	200(8)	317(8)	223(8)	-5(6)	39(6)	34(6)
N(1)	322(8)	195(6)	255(6)	31(5)	-8(6)	-17(6)
N(2)	283(7)	242(6)	227(6)	15(5)	-17(6)	-12(5)
N(3)	221(7)	220(6)	224(6)	3(5)	23(5)	3(5)
N(4)	269(8)	368(8)	287(8)	-40(6)	-14(7)	-30(6)
N(5)	332(8)	337(7)	304(8)	97(6)	-21(6)	45(7)
C(20)	556(15)	440(11)	453(13)	-64(9)	-39(11)	-35(11)
O(2)	617(10)	608(9)	394(8)	-159(7)	12(8)	230(8)

	X	У	Z	U <sub>iso</sub>
H(5)	11440(40)	9105(9)	10561(8)	27(4)
H(7)	14790(50)	11546(10)	10287(9)	44(5)
H(8)	12950(40)	12339(9)	9447(8)	29(4)
H(9)	9280(50)	11960(11)	8692(10)	47(5)
H(10)	7450(40)	10762(9)	8746(8)	27(4)
H(13)	3570(40)	10417(9)	8014(8)	27(4)
H(15)	-3520(50)	9692(10)	6319(10)	45(6)
H(16)	-5350(40)	8481(10)	6275(9)	41(5)
H(17)	-3780(40)	7577(9)	7059(9)	33(5)
H(18)	-340(40)	7859(9)	7920(9)	32(5)
H(1N)	4290(40)	7686(10)	9549(8)	37(5)
H(4N)	14060(50)	10166(11)	10788(11)	52(6)
H(5N)	190(40)	10487(11)	7035(10)	48(6)
H(2O)	6840(50)	8561(12)	11147(13)	59(7)
H(20A)	10300(70)	8664(14)	11848(13)	83(9)
H(20B)	8070(60)	8513(13)	12446(12)	77(8)
H(20C)	8660(60)	7868(14)	11873(11)	72(7)

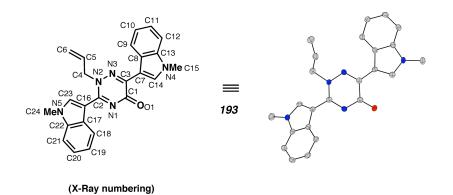
Hydrogen coordinates (  $x~10^4$  ) and isotropic displacement parameters (Å  $^2x~10~^3$  ) for 188 (CCDC 161494).

## Hydrogen bonds for 188 (CCDC 161494) [Å and °].

D-HA	d(D-H)	d(HA)	d(DA)	<(DHA)
N(1)-H(1N)O(1)#1	0.93(2)	1.88(2)	2.800(2)	173(2)
N(5)-H(5N)O(2)#2	0.90(2)	1.95(2)	2.842(2)	171(2)
O(2)-H(2O)O(1)	0.83(2)	1.93(3)	2.765(2)	178(2)

Symmetry transformations used to generate equivalent atoms: #1 x-1/2,-y+3/2,-z+2 #2 -x+1/2,-y+2,z-1/2

## A6.6.3 X-Ray Crystallographic Report for Allyl Triazinone 193



#### Crystal data and structure refinement for 193 (CCDC 259195).

Empirical formula	$C_{24}H_{21}N_5O$
Formula weight	395.46
Crystallization Solvent	hexanes/acetone
Crystal Habit	Fragment
Crystal size	$0.21 \text{ x } 0.11 \text{ x } 0.11 \text{ mm}^3$
Crystal color	Colorless

#### **Data Collection**

Type of diffractometer	Bruker SMART 1000
Wavelength	0.71073 Å MoKα
Data Collection Temperature	100(2) K
$\theta$ range for 6305 reflections used in lattice determination	2.49 to 27.62°
Unit cell dimensions	$  a = 7.8852(8) \text{ Å} \\  b = 15.3322(15) \text{ Å} \\  c = 16.3273(16) \text{ Å} $
Volume	1973.9(3) Å <sup>3</sup>
Z	4
Crystal system	Orthorhombic
Space group	$P2_{1}2_{1}2_{1}$
Density (calculated)	1.331 Mg/m <sup>3</sup>
F(000)	832
Data collection program	Bruker SMART v5.630

 $\theta$  range for data collection 1.82 to 28.45° Completeness to  $\theta = 28.45^{\circ}$ 96.5 %  $-10 \leq \mathbf{h} \leq 10, \, -20 \leq \mathbf{k} \leq 20, \, -21 \leq \mathbf{l} \leq 20$ Index ranges Data collection scan type  $\omega$  scans at 6  $\phi$  settings Data reduction program Bruker SAINT v6.45A Reflections collected 34532 4712 [ $R_{int} = 0.0963$ ] Independent reflections 0.085 mm<sup>-1</sup> Absorption coefficient Absorption correction None 0.9907 and 0.9824 Max. and min. transmission

Structure solution program	Bruker XS v6.12	
Primary solution method	Direct methods	
Secondary solution method	Difference Fourier map	
Hydrogen placement	Difference Fourier map	
Structure refinement program	Bruker XL v6.12	
Refinement method	Full matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	4712 / 0 / 355	
Treatment of hydrogen atoms	Unrestrained	
Goodness-of-fit on F <sup>2</sup>	1.148	
Final R indices [I> $2\sigma$ (I), 3244 reflections]	R1 = 0.0458, wR2 = 0.0560	
R indices (all data)	R1 = 0.0824, wR2 = 0.0596	
Type of weighting scheme used	Sigma	
Weighting scheme used	$w=1/\sigma^2(\text{Fo}^2)$	
Max shift/error	0.000	
Average shift/error	0.000	
Absolute structure parameter	2.8(14)	
Largest diff. peak and hole	0.205 and -0.214 e.Å <sup>-3</sup>	

#### **Special Refinement Details**

Refinement of  $F^2$  against ALL reflections. The weighted R-factor (*w*R) and goodness of fit (S) are based on  $F^2$ , conventional R-factors (R) are based on F, with F set to zero for negative  $F^2$ . The threshold expression of  $F^2 > 2\sigma(F^2)$  is used only for calculating R-factors(gt) etc. and is not relevant to the choice of reflections for refinement. R-factors based on  $F^2$  are statistically about twice as large as those based on F, and R-factors based on ALL data will be even larger.

All esds (except the esd in the dihedral angle between two l.s. planes) are estimated using the full covariance matrix. The cell esds are taken into account individually in the estimation of esds in distances, angles and torsion angles; correlations between esds in cell parameters are only used when they are defined by crystal symmetry. An approximate (isotropic) treatment of cell esds is used for estimating esds involving l.s. planes.

Atomic coordinates ( x 10<sup>4</sup>) and equivalent isotropic displacement parameters (Å<sup>2</sup>x 10<sup>3</sup>) for 193 (CCDC 259195). U(eq) is defined as the trace of the orthogonalized U<sup>ij</sup> tensor.

	Х	у	Z	U <sub>eq</sub>
O(1)	13620(2)	2937(1)	622(1)	25(1)
N(1)	11116(2)	2895(1)	1314(1)	18(1)
N(2)	8961(2)	3837(1)	887(1)	18(1)
N(3)	9849(2)	4135(1)	230(1)	19(1)
N(4)	14226(2)	4330(1)	-1578(1)	19(1)
N(5)	6282(2)	2110(1)	2574(1)	19(1)
C(1)	12133(2)	3195(1)	694(1)	19(1)
C(2)	9552(2)	3205(1)	1384(1)	18(1)
C(3)	11387(2)	3830(1)	125(1)	18(1)
C(4)	7273(3)	4278(1)	968(1)	24(1)
C(5)	7379(3)	5204(1)	713(1)	25(1)
C(6)	6285(3)	5576(2)	212(1)	29(1)
C(7)	12267(2)	4154(1)	-586(1)	17(1)
C(8)	11604(2)	4759(1)	-1185(1)	16(1)
C(9)	10106(2)	5242(1)	-1268(1)	20(1)
C(10)	9911(3)	5772(1)	-1941(1)	21(1)
C(11)	11157(3)	5827(1)	-2547(1)	22(1)
C(12)	12653(3)	5360(1)	-2486(1)	20(1)
C(13)	12856(2)	4843(1)	-1796(1)	18(1)
C(14)	13867(3)	3927(1)	-855(1)	19(1)
C(15)	15833(3)	4284(2)	-2010(2)	26(1)
C(16)	8455(2)	2829(1)	2012(1)	16(1)
C(17)	8953(2)	2551(1)	2817(1)	17(1)
C(18)	10411(3)	2659(1)	3302(1)	20(1)
C(19)	10421(3)	2307(1)	4078(1)	23(1)
C(20)	9031(3)	1841(1)	4384(1)	27(1)
C(21)	7576(3)	1739(1)	3929(1)	24(1)
C(22)	7563(2)	2107(1)	3147(1)	19(1)
C(23)	6824(2)	2544(1)	1896(1)	18(1)
C(24)	4612(3)	1715(2)	2666(2)	27(1)

# Bond lengths [Å] and angles [°] for 193 (CCDC 259195).

O(1)-C(1)	1.243(2) C(23)-H(23)	0.996(15)
N(1)-C(2)	1.327(2) C(24)-H(24A)	1.01(2)
N(1)-C(1)	1.371(2) C(24)-H(24B)	1.05(2)
N(2)-C(2)	1.346(2) C(24)-H(24C)	0.942(18)
N(2)-N(3)	1.361(2)	
N(2)-C(4)	1.499(2) C(2)-N(1)-C(1)	119.13(16)
N(3)-C(3)	1.310(2) C(2)-N(2)-N(3)	122.61(15)
N(4)-C(14)	1.363(2) C(2)-N(2)-C(4)	125.35(16)
N(4)-C(13)	1.382(2) N(3)-N(2)-C(4)	112.03(15)
N(4)-C(15)	1.452(2) C(3)-N(3)-N(2)	117.33(16)
N(5)-C(23)	1.361(2) C(14)-N(4)-C(13)	108.53(16)
N(5)-C(22)	1.378(2) C(14)-N(4)-C(15)	125.45(18)
N(5)-C(24)	1.457(3) C(13)-N(4)-C(15)	125.84(17)
C(1)-C(3)	1.468(2) C(23)-N(5)-C(22)	108.93(16)
C(2)-C(16)	1.460(2) C(23)-N(5)-C(24)	124.85(18)
C(3)-C(7)	1.441(2) C(22)-N(5)-C(24)	126.23(18)
C(4)-C(5)	1.482(3) O(1)-C(1)-N(1)	120.98(18)
C(4)-H(4A)	1.062(19) O(1)-C(1)-C(3)	121.94(19)
C(4)-H(4B)	1.040(19) N(1)-C(1)-C(3)	117.08(16)
C(5)-C(6)	1.319(3) N(1)-C(2)-N(2)	121.85(17)
C(5)-H(5)	1.035(19) N(1)-C(2)-C(16)	117.98(17)
C(6)-H(6A)	1.03(2) N(2)-C(2)-C(16)	120.16(17)
C(6)-H(6B)	1.04(2) N(3)-C(3)-C(7)	115.31(17)
C(7)-C(14)	1.380(3) N(3)-C(3)-C(1)	121.65(18)
C(7)-C(8)	1.445(3) C(7)-C(3)-C(1)	123.03(17)
C(8)-C(9)	1.401(3) C(5)-C(4)-N(2)	110.94(17)
C(8)-C(13)	1.409(2) C(5)-C(4)-H(4A)	112.2(10)
C(9)-C(10)	1.376(3) N(2)-C(4)-H(4A)	111.4(10)
C(9)-H(9)	0.971(18) C(5)-C(4)-H(4B)	110.6(10)
C(10)-C(11)	1.397(3) N(2)-C(4)-H(4B)	103.3(10)
C(10)-H(10)	0.950(17) H(4A)-C(4)-H(4B)	108.0(14)
C(11)-C(12)	1.383(3) C(6)-C(5)-C(4)	123.5(2)
C(11)-H(11)	0.938(17) C(6)-C(5)-H(5)	118.7(12)
C(12)-C(13)	1.388(3) C(4)-C(5)-H(5)	117.5(12)
C(12)-H(12)	1.031(19) C(5)-C(6)-H(6A)	123.8(11)
C(14)-H(14)	1.02(2) C(5)-C(6)-H(6B)	124.7(12)
C(15)-H(15A)	1.00(2) H(6A)-C(6)-H(6B)	111.3(16)
C(15)-H(15B)	0.977(19) C(14)-C(7)-C(8)	106.12(17)
C(15)-H(15C)	1.01(2) C(14)-C(7)-C(3)	127.52(18)
C(16)-C(23)	1.372(2) C(8)-C(7)-C(3)	126.30(17)
C(16)-C(17)	1.437(2) C(9)-C(8)-C(13)	118.32(19)
C(17)-C(22)	1.399(2) C(9)-C(8)-C(7)	135.20(18)
C(17)-C(18)	1.406(2) C(13)-C(8)-C(7)	106.48(16)
C(18)-C(19)	1.378(3) C(10)-C(9)-C(8)	118.94(19)
C(18)-H(18)	0.979(17) C(10)-C(9)-H(9)	120.8(12)
C(19)-C(20)	1.400(3) C(8)-C(9)-H(9)	120.3(12)
C(19)-H(19)	0.994(16) C(9)-C(10)-C(11)	121.5(2)
C(20)-C(21)	1.377(3) C(9)-C(10)-H(10)	117.5(10)
C(20)-H(20)	0.900(19) C(11)-C(10)-H(10)	120.9(10)
C(21)-C(22)	1.394(3) C(12)-C(11)-C(10)	121.1(2)
C(21)-H(21)	1.016(19) C(12)-C(11)-H(11)	120.9(12)

C(10)-C(11)-H(11)	117.8(12)
C(11)-C(12)-C(13)	116.94(19)
C(11)-C(12)-H(12)	123.7(11)
C(13)-C(12)-H(12)	119.4(11)
N(4)-C(13)-C(12)	128.58(18)
N(4)-C(13)-C(8)	108.33(17)
C(12)-C(13)-C(8)	123.09(18)
N(4)-C(14)-C(7)	110.52(18)
N(4)-C(14)-H(14)	122.0(11)
C(7)-C(14)-H(14)	127.4(11)
N(4)-C(15)-H(15A)	107.6(12)
N(4)-C(15)-H(15B)	109.0(11)
H(15A)-C(15)-H(15B)	110.0(16)
N(4)-C(15)-H(15C)	111.5(12)
H(15A)-C(15)-H(15C)	112.1(17)
H(15R) - C(15) - H(15C)	106.6(16)
C(23)-C(16)-C(17)	106.75(18)
C(23)-C(16)-C(2)	125.73(18)
C(17)-C(16)-C(2)	126.73(17)
C(22)-C(17)-C(18)	118.71(17)
C(22)-C(17)-C(16)	106.48(17)
C(18)-C(17)-C(16)	134.76(19)
C(19)-C(18)-C(17)	118.5(2)
C(19)-C(18)-H(18)	120.9(10)
C(17)-C(18)-H(18)	120.6(10)
C(18)-C(19)-C(20)	121.6(2)
C(18)-C(19)-H(19)	118.5(10)
C(10)-C(19)-H(19) C(20)-C(19)-H(19)	119.8(10)
C(20)-C(19)-II(19) C(21)-C(20)-C(19)	121.2(2)
	116.5(13)
C(21)-C(20)-H(20)	
C(19)-C(20)-H(20)	122.1(13)
C(20)-C(21)-C(22)	117.0(2)
C(20)-C(21)-H(21)	122.4(11)
C(22)-C(21)-H(21)	120.6(11)
N(5)-C(22)-C(21)	128.93(19)
N(5)-C(22)-C(17)	108.09(16)
C(21)-C(22)-C(17)	122.98(19)
N(5)-C(23)-C(16)	109.74(18)
N(5)-C(23)-H(23)	119.2(9)
C(16)-C(23)-H(23)	130.8(9)
N(5)-C(24)-H(24A)	106.0(12)
N(5)-C(24)-H(24B)	110.4(11)
H(24A)-C(24)-H(24B)	110.2(15)
N(5)-C(24)-H(24C)	110.2(13)
H(24A)-C(24)-H(24C)	112.0(16)
H(24B)-C(24)-H(24C)	107.9(16)

Anisotropic displacement parameters (Å<sup>2</sup>x 10<sup>4</sup>) for 193 (CCDC 259195). The anisotropic displacement factor exponent takes the form:  $-2\pi^2$  [  $h^2 a^{*2}U^{11} + ... + 2h$  k  $a^* b^* U^{12}$ ].

	U <sup>11</sup>	U <sup>22</sup>	U <sup>33</sup>	U <sup>23</sup>	$U^{13}$	$U^{12}$
O(1)	166(8)	326(9)	248(8)	46(7)	23(7)	52(7)
N(1)	176(9)	198(10)	171(9)	14(8)	-9(8)	-10(8)
N(2)	150(9)	195(10)	200(9)	21(8)	33(8)	42(8)
N(3)	189(10)	232(10)	159(9)	10(8)	14(8)	13(8)
N(4)	142(9)	229(10)	210(10)	9(8)	47(8)	3(8)
N(5)	130(9)	209(10)	224(10)	30(8)	11(8)	-19(8)
C(1)	192(12)	199(12)	173(11)	-26(10)	-36(10)	-10(10)
C(2)	186(12)	167(12)	177(12)	-16(9)	-37(9)	-5(9)
C(3)	181(11)	179(11)	164(11)	-33(9)	-13(9)	-16(10)
C(4)	209(13)	258(13)	257(13)	9(11)	25(11)	43(11)
C(5)	243(13)	262(13)	235(12)	-25(11)	65(11)	35(11)
C(6)	275(14)	265(14)	315(13)	-17(12)	-21(12)	41(12)
C(7)	171(11)	147(11)	182(11)	-4(9)	-19(9)	9(9)
C(8)	152(11)	171(11)	164(11)	-9(9)	-1(9)	-35(9)
C(9)	176(12)	224(12)	190(12)	-4(10)	23(10)	5(10)
C(10)	164(12)	225(12)	243(12)	30(10)	1(10)	15(10)
C(11)	249(12)	206(12)	217(12)	62(11)	-19(11)	-17(11)
C(12)	204(12)	209(12)	197(12)	12(10)	35(11)	-41(10)
C(13)	180(11)	163(11)	191(11)	-9(10)	22(10)	-15(9)
C(14)	190(12)	193(12)	190(12)	-10(10)	-18(10)	-11(10)
C(15)	192(13)	296(15)	285(14)	29(12)	72(11)	5(12)
C(16)	165(11)	151(11)	173(11)	-1(9)	-12(9)	14(9)
C(17)	163(11)	165(11)	180(11)	1(9)	9(9)	28(9)
C(18)	178(12)	183(12)	235(12)	17(10)	18(10)	28(10)
C(19)	176(12)	288(13)	238(12)	3(10)	-67(10)	36(10)
C(20)	303(14)	328(14)	174(12)	67(11)	34(12)	68(11)
C(21)	238(13)	240(13)	239(13)	38(10)	42(11)	38(11)
C(22)	173(11)	196(11)	201(11)	4(10)	21(10)	47(10)
C(23)	216(12)	187(12)	146(11)	-14(10)	-6(10)	70(10)
C(24)	178(13)	255(15)	388(16)	44(13)	11(12)	-43(11)

	х	У	Z	U <sub>iso</sub>
H(4A)	6310(20)	3933(12)	647(11)	35(6)
H(4B)	7010(20)	4231(12)	1591(12)	32(6)
H(5)	8290(20)	5589(13)	994(12)	49(7)
H(6A)	5310(20)	5246(13)	-77(12)	42(7)
H(6B)	6340(30)	6223(14)	20(12)	49(7)
H(9)	9210(20)	5190(12)	-862(11)	32(6)
H(10)	8910(20)	6114(11)	-1973(9)	12(5)
H(11)	10980(20)	6218(12)	-2980(10)	20(5)
H(12)	13600(20)	5380(13)	-2920(12)	41(6)
H(14)	14700(20)	3500(12)	-585(11)	39(6)
H(15A)	16560(30)	3843(14)	-1721(12)	49(7)
H(15B)	16380(20)	4857(13)	-1993(11)	28(6)
H(15C)	15670(30)	4135(14)	-2610(13)	44(7)
H(18)	11410(20)	2962(11)	3085(10)	16(5)
H(19)	11480(20)	2351(11)	4409(10)	23(5)
H(20)	9000(20)	1636(12)	4900(12)	32(7)
H(21)	6550(20)	1414(12)	4144(12)	31(6)
H(23)	6075(19)	2570(11)	1404(10)	12(5)
H(24A)	3910(30)	1938(13)	2193(12)	49(7)
H(24B)	4060(30)	1902(13)	3224(13)	49(7)
H(24C)	4700(20)	1102(12)	2664(10)	19(6)

Hydrogen coordinates (  $x~10^4$  ) and isotropic displacement parameters (Å  $^2x~10~^3$  ) for 193 (CCDC 259195).

#### A6.7 Notes and References

- Portions of this work have been described in a communication, see: Garg, N. K.; Stoltz, B. M. *Tetrahedron Lett.* 2005, 46, 1997-2000.
- (2) (a) Lipinska, T.; Branowska, D.; Rykowki, A. Chem. Heterocycl. Compd. 1999, 35, 334-342. (b) Taylor, E. C.; Pont, J. L.; Warner, J. C. Tetrahedron 1987, 43, 5159-5168. (c) Taylor, E. C.; French, L. G. Tetrahedron Lett. 1986, 27, 1967-1970.
- (3) For the use of similar strategies in natural product synthesis, see: (a) Boger, D. L.;
  Baldino, C. M. J. Am. Chem. Soc. 1993, 115, 11418-11425. (b) Wasserman, H.
  H.; DeSimon, R. W.; Boger, D. L.; Baldino, C. M. J. Am. Chem. Soc. 1993, 115, 8457-8458. (c) Boger, D. L.; Coleman, R. S. J. Am. Chem. Soc. 1987, 109, 2717-2727.
- (4) Shaw, K. N. F.; McMillan, A.; Gudmundson, A. G.; Armstrong, M. D. J. Org. Chem. 1958, 23, 1171-1178.
- (5) Li, J.-H.; Snyder, J. K. J. Org. Chem. 1993, 58, 516-519.
- (6) Faul, M. M.; Winneroski, L. L.; Krumrich, C. A. J. Org. Chem. 1999, 64, 2465-2470.

- (7) Molecular structures are shown with 50% probablility ellipsoids, and hydrogen atoms have been omitted for clarity. Crystallographic data have been deposited at the CCDC, 12 Union Road, Cambridge CB2 1EZ, UK, and copies can be obtained on request, free of charge, by quoting the publication citation and the deposition number; *p*-triazinone **187**: 259291; *m*-triazinone **188**: 161494; allyl triazinone **193**: 259195.
- (8) Faul, M. M.; Winneroski, L. L.; Krumrich, C. A. J. Org. Chem. 1998, 63, 6053-6058.
- (9) Attempted aromatization/protection of 187 or 191 resulted primarily in N-protection, rather than O-protection.
- (10) Sanemitsu, Y.; Nakayama, Y.; Tanabe, Y.; Matsumoto, H.; Hashimoto, S. Agric.
   *Biol. Chem.* 1990, 54, 3367-3369.
- (11) Pletnov, A. A.; Tian, Q.; Larock, R. C. J. Org. Chem. 2002, 67, 9276-9287.

#### **APPENDIX SEVEN**

#### **Notebook Cross-Reference**

The following notebook cross-reference has been included to facilitate access to the original spectroscopic data obtained for the compounds presented in this thesis. For each compound, both hardcopy and electronic characterization folders have been created that contain copies of the original <sup>1</sup>H NMR, <sup>13</sup>C NMR, and IR spectra. All notebooks and spectral data are stored in the Stoltz archives.

Compound	<sup>1</sup> H NMR	<sup>13</sup> C NMR	IR
52	NKGIV-73	NKGIV-73	NKGIV-73
53	NKGIV-45	NKGIV-45	NKGIV-45
22	NKGXI-73	NKGXI-73	NKGXIII-109
63	RSVI-205	RSVI-205	RSVI-205
91	NKGV-247	NKGV-247	NKGV-247
65	RSVI-279		
67	NKGVII-85	NKGVII-85	NKGVII-85
68	NKGVII-49	NKGVII-49	NKGVII-49
70	NKGVII-53	NKGVII-53	NKGVII-53
72	NKGVII-191	NKGVII-191	NKGVII-191
92	NKGVII-211	NKGVII-211	NKGVII-211
93	NKGVII-213	NKGVII-213	NKGVII-213
62	NKGVII-243		
73	NKGVII-193	NKGVII-193	NKGVII-193
74	NKGVII-217	NKGVII-217	NKGVII-217

#### Table A7.1 Compounds Appearing in Chapter 2:

#### The Total Synthesis of Dragmacidin D

Compound	<sup>1</sup> H NMR	<sup>13</sup> C NMR	IR
75	RSVII-161	RSVII-161	RSVII-161
76	RSVII-201	RSVII-201	RSVII-201
77	RSVII-246	RSVII-246	RSVII-246
78	NKGVII-275	NKGVII-275	NKGVII-275
80	NKGVIII-241		
81	NKGVIII-273		
82	NKGX-243	NKGX-243	NKGX-243
83	NKGX-259	NKGX-259	NKGX-259
84	NKGX-241	NKGX-241	NKGX-241
5	NKGXXIII-221	NKGXI-37B	NKGX-253
86	RSX-107	RSX-107	RSX-157
88	RSX-167	RSX-167	RSX-167

## Table A7.2 Compounds Appearing in Chapter 3:

The Total Synthesis of (+)- and (-)-Dragmacio	lin	1			]		ŀ	Ĩ	i	ſ	ĺ		]	1			L	1	1	]	1	]	1	]	1	1	}	ł	ł	1	}	1	1	)	Ì	l	l	l	l	l	Ì	1	1	]	1	1	1	]	1	1	]	1	1	1	1	1	1	1	1	1	1	]	]	1	1	1	1	C	ľ	r	r	1	1	1	1	1	1	r	C	ľ	ľ	I	]	]	]	J	J	Ĺ	í	j	I.	l	l	1	(	(	(	(	(	ĺ	i	í	i	j	•	(	(	ł	ć	l	1	1	ľ	1	)	g	l	ĉ	•	r	ł	)	)	]		]		).	)			(	(		d	(	l	n	J	ł	a	ć	ļ		-	)
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Compound	<sup>1</sup> H NMR	<sup>13</sup> C NMR	IR
146	DDCIV-223	DDCIII-107	DDCIV-223
147	NKGXVI-105	NKGXVI-105	NKGXVI-105
103	DDCIII-121	DDCIII-121	DDCIII-121
105	DDCVIII-65	DDCVIII-65	DDCVIII-65
106	NKGXXIII-75	NKGXIX-133	NKGXXIII-75
104	NKGXIX-237	NKGXIX-131	NKGXIX-131
109	NKGXV-123	NKGXIX-103	NKGXIX-103
151	DDCIV-221	DDCIV-221	DDCIV-221
99	NKGXIX-107	NKGXIX-107	NKGXIX-107
153	DDCIV-217	DDCIV-217	DDCIV-217
100	DDCIV-61	DDCIV-61	NKGXVIII-63
98	NKGXIV-301P3 & NKGXXI-101	NKGXXI-101	NKGXIX-112

Compound	<sup>1</sup> H NMR	<sup>13</sup> C NMR	IR
110	NKGXVI-295P1	NKGXVI-295P1	NKGXVI-295P1
154	NKGXIX-118	NKGXIX-118	NKGXIX-118
111	NKGXIX-119	NKGXIX-119	NKGXIX-119
155	NKGXIX-121	NKGXIX-121	NKGXIX-121
97	NKGXIX-123	NKGXIX-123	NKGXIX-123
112	NKGXIX-135	NKGXIX-135	NKGXIX-135
113	NKGXIX-143	NKGXIX-143	NKGXIX-143
117	NKGXXII-289	NKGXXII-289	NKGXXII-289
118	NKGXVIII-139	NKGXXIII-43	NKGXXIII-43
119	NKGXIX-139	NKGXIX-139	NKGXIX-139
123	NKGXIX-37		
120	NKGXIX-151	NKGXIX-151	NKGXIX-151
124	NKGXIX-163	NKGXIX-163	NKGXIX-155
(+)-7	NKGXIX-227B	NKGXIX-227	NKGXIX-227
128	DDCVIII-143	DDCVIII-143	DDCVIII-143
159	NKGXXII-53	NKGXXII-53	NKGXXII-53
131	DDCVII-201	DDCVII-195	DDCVII-195
132	DDCVII-207	DDCVII-227	DDCVII-207
130	DDCVII-217	DDCVII-213	DDCVII-213
126	DDCVIII-99	NKGXXII-81	NKGXXII-81
127	NKGXXII-116	NKGXXII-116	NKGXXII-116
133	NKGXXII-131	NKGXXII-131	NKGXXII-131
136	NKGXXIII-53	NKGXXIII-53	NKGXXIII-53
134	NKGXXIII-157	NKGXXIII-157	NKGXXIII-157
137	NKGXXIII-57	NKGXXIII-57	NKGXXIII-57
138	NKGXXIII-35	NKGXXII-209	NKGXXII-209
139	NKGXXIII-91	NKGXXIII-91	NKGXXIII-91
144	NKGXXII-213	NKGXXII-213	NKGXXII-213
(–)-7	NKGXXIII-179		

## **COMPREHENSIVE BIBLIOGRAPHY**

Amat, M.; Hadida, S.; Sathyanarayana, S.; Bosch, J. J. Org. Chem. 1994, 59, 10-11.

Amatore, C.; Jutand, A. J. Organomet. Chem. 1999, 576, 254-278.

Aoki, S.; Ye, Y.; Higuchi, K.; Takashima, A.; Tanaka, Y.; Kitagawa, I.; Kobayashi, M. Chem. Pharm. Bull. 2001, 49, 1372-1374.

Arndt, F. Org. Synth., 1943, Coll. Vol. 2, 165.

Audebert, P.; Bidan, G. Synthetic Metals 1986, 15, 9-22.

Auwers, K. Justus Liebigs Ann. Chem. 1907, 357, 85-94.

Ayer, W. A.; Craw, P. A.; Ma, Y. T.; Miao, S. Tetrahedron 1992, 48, 2919-2924.

Ayerbe, M.; Arrieta, A.; Cossio, F. J. Org. Chem. 1998, 63, 1795-1805.

Aygün, A.; Pindur, U. Curr. Med. Chem. 2003, 10, 1113-1127.

Bailey, D. M.; Johnson, R. E. J. Med. Chem. 1973, 16, 1300-1302.

Baran, P. S.; Corey, E. J. J. Am. Chem. Soc. 2002, 124, 7904-7905.

Barco, A.; Benetti, S.; De Risi, C.; Marchetti, P.; Pollini, G. P.; Zanirato, V. *Tetrahedron: Asymmetry* **1997**, *8*, 3515-3545.

Barlin, G. B. Aust. J. Chem. 1983, 36, 983-992.

Barros, M. T.; Maycock, C. D.; Ventura, M. R. J. Chem. Soc., Perkin Trans. 1 2001, 166-173.

Bartik, K.; Braekman, J.-C.; Daloze, D.; Stoller, C.; Huysecom, J.; Vendevyver, G.; Ottinger, R. *Can. J. Chem.* **1987**, *65*, 2118-2121.

Bartoli, G.; Palmieri, G.; Bosco, M.; Dalpozzo, R. Tetrahedron Lett. 1989, 30, 2129-2132.

Batcho, A. D.; Leimgruber, W. Org. Synth. 1985, 63, 214-225.

Beletskaya, I. P.; Cheprakov, A. V. Chem. Rev. 2000, 100, 3009-3066.

Belletete, M.; Beaupre, S.; Bouchard, J.; Blondin, P.; Leclerc, M.; Durocher, G. J. Phys. Chem. B 2000, 104, 9118-9125.

Bergens, S. H.; Bosnich, B. J. Am. Chem. Soc. 1991, 113, 958-967.

Bernheim, F.; Morgan, J. E. Nature 1939, 144, 290.

Blunt, J. W.; Copp, B. R.; Munro, M. H. G.; Northcote, P. T.; Prinsep, M. R. Nat. Prod. Rep. 2004, 21, 1-49.

Boehm, J. C.; Gleason, J. G.; Pendrak, I.; Sarau, H. M.; Schmidt, D. B.; Foley, J. J.; Kingsbury, W. D. *J. Med. Chem.* **1993**, *36*, 3333-3340.

Boger, D. L.; Coleman, R. S. J. Am. Chem. Soc. 1987, 109, 2717-2727.

Boger, D. L.; Baldino, C. M. J. Am. Chem. Soc. 1993, 115, 11418-11425.

Bosco, M.; Dalpozzo, R.; Bartoli, G.; Palmieri, G.; Petrini, M. J. Chem. Soc., Perkin Trans. 2 1991, 5, 657-663.

Brown, J. M. Angew. Chem., Int. Ed. Engl. 1987, 26, 190-203.

Brown, J. M. In *Comprehensive Asymmetric Catalysis*; Jacobsen, E. N.; Pfaltz, A.; Yamamoto, H., Eds.; Springer: Berlin, 1999; Vol. 1, pp 121-195.

Burke, T. R.; Zhang, Z.-Y. Biopolymers 1998, 47, 225-241.

Butler, A.; Carter-Franklin, J. N. Nat. Prod. Rep. 2004, 21, 180-188.

Capon, R. J.; Rooney, F.; Murray, L. M.; Collins, E.; Sim, A. T. R.; Rostas, J. A. P.; Butler, M. S.; Carroll, A. R. J. Nat. Prod. **1998**, 61, 660-662.

Casapullo, A.; Bifulco, G.; Bruno, I.; Riccio, R. J. Nat. Prod. 2000, 63, 447-451.

Chen, M. S.; White, M. C. J. Am. Chem. Soc. 2004, 126, 1346-1347.

Chenier, P. J. J. Chem. Ed. 1978, 55, 286-291.

Chierici, L.; Gardini, G. P. Tetrahedron 1966, 22, 53-56.

Chung, J. Y. L.; Ho, G.-J.; Chartrain, M.; Roberge, C.; Zhao, D.; Leazer, J.; Farr, R.;
Robbins, M.; Emerson, K.; Mathre, D. J.; McNamara, J. M.; Hughes, D. L.; Grabowski,
E. J. J.; Reider, P. J. *Tetrahedron Lett.* **1999**, *40*, 6739-6743.

Ciamician, G.; Silber, P. Chem. Ber. 1912, 45, 1842-1845.

Claridge, T. D. W. In *High-Resolution NMR Techniques in Organic Chemistry*; Pergamon: Amsterdam, 1999; pp 320-326.

Craig, B. N.; Janssen, M. U.; Wickersham, B. M.; Rabb, D. M.; Chang, P. S.; O'Leary,D. J. J. Org. Chem. 1996, 61, 9610-9613.

Cutignano, A.; Bifulco, G.; Bruno, I.; Casapullo, A.; Gomez-Paloma, L.; Riccio, R. *Tetrahedron* **2000**, *56*, 3743-3748.

Dess, D. B.; Martin, J. C. J. Org. Chem. 1983, 48, 4155-4156.

Diederich, F.; Stang, P. J.; Eds.; *Metal-Catalyzed Cross-Coupling Reactions*; Wiley-VCH: Weinheim, 1998.

Dounay, A. B.; Overman, L. E. Chem. Rev. 2003, 103, 2945-2963.

Edwards, M. P.; Ley, S. V.; Lister, S. G.; Palmer, B. D. J. Chem. Soc., Chem. Commun. 1983, 630-633.

Edwards, M. P.; Ley, S. V.; Lister, S. G.; Palmer, B. D.; Williams, D. J. J. Org. Chem. **1984**, 49, 3503-3516.

Edwards, M. P.; Doherty, A. M.; Ley, S. V.; Organ, H. M. *Tetrahedron* **1986**, *42*, 3723-3729.

Elfehail, F.; Dampawan, P.; Zajac, W. Synth. Commun. 1980, 10, 929-932.

Elfehail, F. E.; Zajac, W. W., Jr. J. Org. Chem. 1981, 46, 5151-5155.

Eliel, E. L.; Wilen, S. H.; Mander, L. N. *Stereochemistry of Organic Compounds*; Wiley-Interscience: New York, 1994.

Ellingson, H. J. Am. Chem. Soc. 1949, 71, 2798-2800.

Fahy, E.; Potts, B. C. M.; Faulkner, D. J.; Smith, K. J. Nat. Prod. 1991, 54, 564-569.

Faul, M. M.; Winneroski, L. L.; Krumrich, C. A. J. Org. Chem. 1998, 63, 6053-6058.

Faul, M. M.; Winneroski, L. L.; Krumrich, C. A. J. Org. Chem. 1999, 64, 2465-2470.

Faulkner, D. J. Nat. Prod. Rep. 2002, 19, 1-48.

Favorskii, A. E. J. Russ. Phys. Chem. Soc. 1894, 26, 559.

Ferreira, E. M.; Stoltz, B. M. J. Am. Chem. Soc. 2003, 125, 9578-9579.

Fischer, R. H.; Weitz, H. M. Synthesis 1980, 261-282.

Garg, N. K.; Sarpong, R.; Stoltz, B. M. J. Am. Chem. Soc. 2002, 124, 13179-13184.

Garg, N. K.; Caspi, D. D.; Stoltz, B. M. J. Am. Chem. Soc. 2004, 126, 9552-9553.

Garg, N. K.; Stoltz, B. M. Tetrahedron Lett. 2005, 46, 1997-2000.

Garg, N. K.; Stoltz, B. M. Tetrahedron Lett. 2005, 46, 2423-2426.

Garg, N. K.; Caspi, D. D.; Stoltz, B. M. J. Am. Chem. Soc. 2005, 127, in press.

Geissler, H. In *Transition Metals for Organic Synthesis*; Beller, M.; Bolm, C., Eds.; Wiley-VCH: Weinheim, 1998; Chapter 2.10, p 158.

Gilbert, E. J.; Chisholm, J. D.; Van Vranken, D. L. J. Org. Chem. 1999, 64, 5670-5676.

Gilow, H. M.; Hong, Y. H.; Millirons, P. L.; Snyder, R. C.; Casteel, W. J., Jr. J. *Heterocycl. Chem.* **1986**, *23*, 1475-1480.

Greene, T. W.; Wuts, P. G. M. Protective Groups in Organic Synthesis, 3<sup>rd</sup> Ed.; John Wiley-Interscience: New York, 1999.

Gunasekera, S. P.; McCarthy, P. J.; Kelly-Borges, M. J. Nat. Prod. 1994, 57, 1437-1441.

Gushin, V. V.; Alper, H. Chem. Rev. 1994, 94, 1047-1062.

Han, X.; Stoltz, B. M.; Corey, E. J. J. Am. Chem. Soc. 1999, 121, 7600-7605.

Hanessian, S. In *Total Synthesis of Natural Products: The "Chiron" Approach*, Baldwin,E. J., Ed.; Pergamon Press: Oxford, 1983; pp 206-208.

Hanessian, S.; Pan, J.; Carnell, A.; Bouchard, H.; Lesage, L. J. Org. Chem. 1997, 62, 465-473.

Hashem, M. A.; Sultana, I.; Hai, M. A. Indian J. Chem. Sect. B 1999, 38, 789-794.

Hibino, S.; Choshi, T. Nat. Prod. Rep. 2002, 19, 148-180.

Hills, I. D.; Fu, G. C. J. Am. Chem. Soc. 2004, 126, 13178-13179.

House, H. O.; Berkowitz, W. F. J. Org. Chem. 1963, 28, 307-311.

House, H. O.; Berkowitz, W. F. J. Org. Chem. 1963, 28, 2271-2276.

Howes, P. D.; Cleasby, A.; Evans, D. N.; Feilden, H.; Smith, P. W.; Sollis, S. L.; Taylor,N.; Wonacott, A. J. *Eur. J. Med. Chem.* **1999**, *34*, 225-234.

Huang, P.-Q. Youji Huaxue 1999, 19, 364-373.

Jacobs, R. S.; Pomponi, S.; Gunasekera, S.; Wright, A. PCT Int. Appl. WO 9818466, May 7, 1998.

Jiang, B.; Smallheer, J. M.; Amaral-Ly, C.; Wuonola, M. A. J. Org. Chem. 1994, 59, 6823-6827.

Jiang, B.; Gu, X.-H. Bioorg. Med. Chem. 2000, 8, 363-371.

Jiang, B.; Gu, X.-H. Heterocycles 2000, 53, 1559-1568.

Jin, Z. Nat. Prod. Rep. 2003, 20, 584-605.

Kam, T.-S.; Pang, H.-S.; Lim, T.-M. Org. Biomol. Chem. 2003, 1, 1292-1297.

Kawasaki, T.; Enoki, H.; Matsumura, K.; Ohyama, M.; Inagawa, M.; Sakamoto, M. *Org. Lett.* **2000**, *2*, 3027-3029.

Kawasaki, T.; Ohno, K.; Enoki, H.; Umemoto, Y.; Sakamoto, M. *Tetrahedron Lett.* **2002**, *43*, 4245-4248.

Kerwin, J. F., Jr.; Lancaster, J. R., Jr.; Feldman, P. L. J. Med. Chem. 1995, 38, 4343-4362.

Kharasch, M. S.; Kane, S. S.; Brown, H. C. J. Am. Chem. Soc. 1940, 62, 2242-2243.

Kohmoto, S.; Kashman, Y.; McConnell, O. J.; Rinehart, K. L., Jr.; Wright, A.; Koehn, F. *J. Org. Chem.* **1988**, *53*, 3116-3118.

Kong, Y. C.; Cheng, K. F.; Cambie, R. C.; Waterman, P. G. J. Chem. Soc., Chem. Commun. 1985, 47-48.

Kornfeld, E. C.; Fornefeld, E. J.; Kline, G. B.; Mann, M. J.; Morrison, D. E.; Jones, R.G.; Woodward, R. B. J. Am. Chem. Soc. 1956, 78, 3087-3114.

Kreiser, W.; Körner, F. Helv. Chim. Acta 1999, 82, 1610-1629.

Kuivila, H. G. J. Org. Chem. 1960, 25, 284-285.

Kutney, J. P. Nat. Prod. Rep. 1990, 7, 85-103.

Lancini, G. C.; Lazzari, E.; Sartori, G. J. Antibiot. 1968, 21, 387-392.

Larock, R. C.; Hightower, T. R. J. Org. Chem. 1993, 58, 5298-5300.

Leete, E.; Bjorklund, J. A.; Reineccius, G. A.; Chen, T.-B. Spec. Publ.-R. Soc. Chem. **1992**, 95, 75-95.

Li, J. J.; Gribble, G. W. In *Palladium in Heterocyclic Chemistry: A Guide for the Synthetic Chemist*; Pergamon: Amsterdam, 2000; pp 355-373.

Li, J.-H.; Snyder, J. K. J. Org. Chem. 1993, 58, 516-519.

Li, K.; Du, W.; Que, N. L.; Liu, H. J. Am. Chem. Soc. 1996, 118, 8763-8764.

Lipinska, T.; Branowska, D.; Rykowki, A. Chem. Heterocycl. Compd. 1999, 35, 334-342.

Littke, A. F.; Fu, G. C. J. Am. Chem. Soc. 2001, 123, 6989-7000.

Littke, A. F.; Schwarz, L.; Fu, G. C. J. Am. Chem. Soc. 2002, 124, 6343-6348.

Little, T. L.; Webber, S. E. J. Org. Chem. 1994, 59, 7299-7305.

Longley, R. E.; Isbrucker, R. A.; Wright, A. E. U.S. Patent 6,087,363, July 11, 2000.

Lott, R. S.; Chauhan, V. S.; Virander, S.; Stammer, C. H. J. Chem. Soc., Chem. Commun. 1979, 495-496.

Luzzio, F. A. Tetrahedron 2001, 57, 915-945.

MacDonald, J. C. J. Biol. Chem. 1961, 236, 512-514.

Maehr, H.; Smallheer, J. J. Am. Chem. Soc. 1985, 107, 2943-2945.

Magnuson, S. R. Tetrahedron 1995, 51, 2167-2213.

Maki, S.; Okawa, M.; Matsui, R.; Hirano, T.; Niwa, H. Synlett 2001, 10, 1590-1592.

Manthey, M. K.; González-Bello, C.; Abell, C. J. Chem. Soc., Perkin Trans. 1 1997, 625-628.

Marletta, M. A. J. Med. Chem. 1994, 37, 1899-1907.

McCluskey, A.; Sim. A. T. R.; Sakoff, J. A. J. Med. Chem. 2002, 45, 1151-1175.

McIntire, W. S.; Wemmer, D. E.; Chistoserdov, A.; Lidstrom, M. E. Science 1991, 252, 817-824.

Meier, R.-M.; Tamm, C. Helv. Chim. Acta, 1991, 74, 807-818.

Miyake, F. Y.; Yakushijin, K.; Horne, D. A. Org. Lett. 2000, 2, 3185-3187.

Miyake, F. Y.; Yakushijin, K.; Horne, D. A. Org. Lett. 2002, 4, 941-943.

Miyake, H.; Yamamura, K. Chem. Lett. 1989, 6, 981-984.

Miyaura, N.; Suzuki, A. Chem. Rev. 1995, 95, 2457-2483.

Mocada, S.; Palmer, R. M. J.; Higgs, E. A. Pharmacol. Rev. 1991, 43, 109-142.

Molina, J. A.; Jimenez-Jimenez, F. J.; Orti-Pareja, M.; Navarro, J. A. Drugs Aging 1998, 12, 251-259.

Morris, S. A.; Andersen, R. J. Tetrahedron 1990, 46, 715-720.

Muchowski, J. M.; Solas, D. R. J. Org. Chem. 1984, 49, 203-205.

Murray, L. M.; Lim, T. K.; Hooper, J. N. A.; Capon, R. J. Aust. J. Chem. 1995, 48, 2053-2058.

Neber, P. W.; Friedolsheim, A. Justus Liebigs Ann. Chem. 1926, 449, 109-134.

Nicolaou, K. C.; Webber, S. E. Synthesis 1986, 6, 453-461.

Noble, R. L.; Beer, C. T.; Cutts, J. H. Ann. N.Y. Acad. Sci. 1958, 76, 882-894.

Noyori, R. In Asymmetric Catalysis in Organic Synthesis; Wiley: New York, 1994; pp 16-94.

O'Brien, C. Chem. Rev. 1964, 64, 81-89.

Omura, S.; Iwai, Y.; Hirano, A.; Nakagawa, A.; Awaya, J.; Tsuchiya, H.; Takahashi, Y.; Masuma, R. J. Antibiot. 1977, 30, 275-289.

Ooi, T.; Takahashi, M.; Doda, K.; Maruoka, K. J. Am. Chem. Soc. 2002, 124, 7640-7641.

Philippe, M.; Sepulchre, A. M.; Gero, S. D.; Loibner, H.; Streicher, W.; Stutz, P. J. Antibiot. 1982, 35, 1507-1512.

Piers, E.; Britton, R.; Andersen, R. J. J. Org. Chem. 2000, 65, 530-535.

Pindur, U.; Lemster, T. Curr. Med. Chem. 2001, 8, 1681-1698.

Pletnov, A. A.; Tian, Q.; Larock, R. C. J. Org. Chem. 2002, 67, 9276-9287.

Plieninger, H.; Suhr, K. Chem. Ber. 1956, 89, 270-278.

Poss, C. S.; Schreiber, S. L. Acc. Chem. Res. 1994, 27, 9-17.

Rapado, L. P.; Bulugahapitiya, V.; Renaud, P. Helv. Chim. Acta 2000, 83, 1625-1632.

Rathore, R.; Kochi, J. K. J. Org. Chem. 1996, 61, 627-639.

Rönn, M.; Bäckvall, J.-E.; Andersson, P. G. Tetrahedron Lett. 1995, 36, 7749-7752.

Sakowski, J.; Bohn, M.; Sattler, I.; Dahse, H.; Schlitzer, M. J. Med. Chem. 2001, 44, 2886-2899.

Sanemitsu, Y.; Nakayama, Y.; Tanabe, Y.; Matsumoto, H.; Hashimoto, S. Agric. Biol. Chem. 1990, 54, 3367-3369.

Sasaki, S.; Ehara, T.; Sakata, I.; Fujino, Y.; Harada, N.; Kimura, J.; Nakamura, H.; Maeda, M. *Bioorg. Med. Chem. Lett.* **2001**, *11*, 583-585.

Schmidt, H. H. W.; Walter, U. Cell 1994, 78, 919-925.

Schreiber, S. L. Chem. Scr. 1987, 27, 563-566.

Schumacher, R. W.; Davidson, B. S. Tetrahedron 1999, 55, 935-942.

Shaw, K. N. F.; McMillan, A.; Gudmundson, A. G.; Armstrong, M. D. J. Org. Chem. **1958**, 23, 1171-1178.

Sheppeck, J. E.; Gauss, C.-M.; Chamberlin, A. R. *Bioorg. Med. Chem.* **1997**, *5*, 1739-1750.

Shin, J.; Seo, Y.; Cho, K. W.; Rho, J.-R.; Sim, C. J. J. Nat. Prod. 2000, 62, 647-649.

Stahl, S. S. Angew. Chem., Int. Ed. 2004, 43, 3400-3420.

Stoltz, B. M. Chem. Lett. 2004, 33, 362-367.

Sundberg, R. J., Ed.; Indoles; Academic Press: San Diego, 1996.

Suzuki, A. J. Organomet. Chem. 1999, 576, 147-168.

Takaya, H.; Ohta, T.; Noyori, R. In *Catalytic Asymmetric Synthesis*; Ojima, I., Ed.; VCH Publishers: New York, 1994; pp 1-39.

Taylor, E. C.; French, L. G. Tetrahedron Lett. 1986, 27, 1967-1970.

Taylor, E. C.; Pont, J. L.; Warner, J. C. Tetrahedron 1987, 43, 5159-5168.

Thorns, V.; Hansen, L. M. Exp. Neurol. 1998, 150, 14-20.

Trend, R. M.; Ramtohul, Y. K.; Ferreira, E. M.; Stoltz, B. M. Angew. Chem., Int. Ed. 2003, 42, 2892-2895.

Tsuji, J. In *Transition Metal Reagents and Catalysts*; Wiley: Chichester, U.K., 2000; Chapter 3, p 27.

Tsuji, J.; Mandai, T. Synthesis 1996, 1-24.

Tsujii, S.; Rinehart, K. L.; Gunasekera, S. P.; Kashman, Y.; Cross, S. S.; Lui, M. S.; Pomponi, S. A.; Diaz, M. C. *J. Org. Chem.* **1988**, *53*, 5446-5453.

Turck, A.; Ple, N.; Dognon, D.; Harmoy, C.; Queguiner, G. J. Heterocycl. Chem. 1994, 31, 1449-1454.

Ulhaq, S.; Naylor, M. A.; Chinje, E. C.; Threadgill, M. D.; Stratford, I. J. Anti-Cancer Drug Des. 1997, 12, 61-65.

Ulibarri, G.; Audrain, H.; Nadler, W.; Lhermitte, H.; Grierson, D. S. Pure Appl. Chem. 1996, 68, 601-604.

Ulibarri, G.; Nadler, W.; Skrydstrup, T.; Audrain, H.; Chiaroni, A.; Riche, C.; Grierson,D. S. J. Org. Chem. 1995, 60, 2753-2761.

Van Benthem, R. A. T. M.; Hiemstra, H.; Michels, J. J.; Speckamp, W. N. J. Chem. Soc., Chem. Commun. 1994, 357-359.

Vereshchagin, A. L.; Branskii, O. V.; Semenov, A. A. Chem. Heterocycl. Compd. (Engl. Transl.) 1983, 19, 40-42.

Wasserman, H. H.; DeSimon, R. W.; Boger, D. L.; Baldino, C. M. J. Am. Chem. Soc. 1993, 115, 8457-8458.

Wenkert, E.; Moeller, P. D.; Piettre, S. R.; McPhail, A. T. J. Org. Chem. 1988, 53, 3170-3178.

Whitlock, C. R.; Cava, M. P. Tetrahedron Lett. 1994, 35, 371-374.

Williams, R. M.; Cao, J.; Tsujishima, H.; Cox, R. J. J. Am. Chem. Soc. 2003, 125, 12172-12178.

Woodward, R. B.; Bader, F. E.; Bickel, H.; Frey, A. J.; Kierstead, R. W. *Tetrahedron* **1958**, *2*, 1-57.

Wright, A. E.; Pomponi, S. A.; Cross, S. S.; McCarthy, P. J. Org. Chem. 1992, 57, 4772-4775.

Wright, A. E.; Pomponi, S. A.; Jacobs, R. S. PCT Int. Appl. WO 9942092, August 26, 1999.

Xiao, W. Huaxue Shiji 1992, 14(6), 363-366.

Yang, C.-G.; Huang, H.; Jiang, B. Curr. Org. Chem. 2004, 8, 1691-1720.

Yang, C.-G.; Liu, G.; Jiang, B. J. Org. Chem. 2002, 67, 9392-9396.

Yang, C.-G.; Wang, J.; Jiang, B. Tetrahedron Lett. 2002, 43, 1063-1066.

Yang, C.-G.; Wang, J.; Tang, X.-X.; Jiang, B. *Tetrahedron: Asymmetry* **2002**, *13*, 383-394.

Yaylayan, V. A. J. Agric. Food. Chem. 1996, 44, 2511-2516.

Zhang, H.; Ferreira, E. M.; Stoltz, B. M. Angew. Chem., Int. Ed. 2004, 43, 6144-6148.

Zheng, Q.; Yang, Y.; Martin, A. R. Heterocycles 1994, 37, 1761-1772.

Zhu, Z.; Moore, J. S. J. Org. Chem. 2000, 65, 116-123.

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## **ABOUT THE AUTHOR**

Neil K. Garg was born on December 18, 1978, in White Plains, NY. His immediate family consists of an older brother, Bobby, and two loving parents, Desh and Neena. Neil's childhood years were spent living in Fishkill, NY, a suburb 90 miles north of New York City. He attended John Jay Senior High School and pursued several extracurricular activities including tennis, soccer, and playing the trombone in a jazz band.

After graduating from high school in 1996, Neil began undergraduate studies at New York University. His early years at NYU were spent learning about a broad range of topics including psychology, writing, chemistry, biology, capitalism, and African culture. The sciences were undoubtedly his forte, and he began working in a chemistry research laboratory with Professor Marc Walters during his sophomore year. Neil also grew fond of teaching chemistry and continued both research and teaching until he graduated with a B.S. in chemistry in 2000. During his college years, Neil spent several months living in Strasbourg, France, and conducted research with Professor Mir Wais Hosseini at Université Louis Pasteur. While in France, Neil enjoyed exploring the French countryside on bike and hiking in the Vosges Mountains.

In 2000, Neil moved to Pasadena, CA, to pursue doctoral studies with Professor Brian M. Stoltz at the California Institute of Technology. In 2005, he earned his Ph.D. in chemistry for investigations involving the total synthesis of the dragmacidin alkaloids. Neil will marry his fiancée, Lindsey Bogard, in April 2005, and then begin postdoctoral studies under the direction of Professor Larry E. Overman at UC Irvine soon afterwards.