

CONCLUDING PERSPECTIVES

Nitrogenase is a fascinating enzyme. It houses not one but two unique iron-sulfur clusters, the FeMo-cofactor and the P-cluster. Neither cluster has been successfully synthesized outside a cellular environment. Due to the complexity of the clusters' structure and their interactions with substrate, electrons, and protons, the mechanism of biological nitrogen fixation has attracted the attention of numerous scientists for the better part of a century but yet remains elusive. Solving the mechanism of biological nitrogen fixation will not only advance the fields of chemistry and biochemistry at a fundamental level and through the invention of new characterization techniques, it also has the potential to significantly impact agriculture by leading to new and cleaner methods of ammonia production for fertilizer.

The work described in this thesis contributes to our understanding of nitrogenase using structural and spectroscopic techniques. Major results include the structural characterization of a 1.08 Å Cp1 structure and its comparison to Av1, analysis of pathways within the MoFe protein, and the characterization of a reversible protonated resting state of the MoFe protein. An underlying principle in all the work described herein is the importance of investigating multiple nitrogenase species that are phylogenetically distinct. While the structure of the cofactors is conserved among all known MoFe proteins, their enzyme activity and structure are not wholly conserved. Understanding the similarities and differences between nitrogenases elucidates the relationship of the structure and function of nitrogenase.

Looking ahead, much work is still needed to achieve a conclusive and complete understanding of the mechanism of biological nitrogen fixation. New technology and creative approaches will undoubtedly get the job done, and I look forward to seeing the results!

References

1. Lineweaver, H., Burk, D. & Deming, W. E. The Dissociation Constant of Nitrogen-Nitrogenase in *Azotobacter*. *J. Am. Chem. Soc.* **56**, 225–230 (1934).
2. Burk, D., Lineweaver, H. & Horner, C. K. The Specific Influence of Acidity on the Mechanism of Nitrogen Fixation by *Azotobacter*. *J. Bacteriol.* **27**, 325–340 (1934).
3. Smil, V. *Enriching the Earth: Fritz Haber, Carl Bosch, and the Transformation of World Food Production*. (The MIT Press, 2001).
4. Ritter, S. K. The Haber-Bosch Reaction: An Early Chemical Impact On Sustainability. *C&EN* **86**, (2008).
5. Zhang, X. Biogeochemistry: A Plan for Efficient use of Nitrogen Fertilizers. *Nature* **543**, 322–323 (2017).
6. Loxton, S. The Haber Process Nitrogen Fertilizer from the Air. *The Compost Gardener* (2008). Available at: <http://www.the-compost-gardener.com/haber-process.html>. (Accessed: 3rd August 2017)
7. Anderson, J. S., Rittle, J. & Peters, J. C. Catalytic Conversion of Nitrogen to Ammonia by an Iron Model Complex. *Nature* **501**, 84–87 (2013).
8. Rittle, J. & Peters, J. C. An Fe-N₂ Complex that Generates Hydrazine and Ammonia via Fe=NNH₂: Demonstrating a Hybrid Distal-to-Alternating Pathway for N₂ Reduction. *J. Am. Chem. Soc.* **138**, 4243–4248 (2016).
9. Creutz, S. E. & Peters, J. C. Catalytic Reduction of N₂ to NH₃ by an Fe–N₂ Complex Featuring a C-Atom Anchor. *J. Am. Chem. Soc.* **136**, 1105–1115 (2014).
10. Rittle, J. & Peters, J. C. Fe–N₂/CO Complexes that Model a Possible Role for the Interstitial C Atom of FeMo-cofactor (FeMoco). *Proc. Natl. Acad. Sci. U. S. A.* **110**, 15898–15903 (2013).
11. Withers, N. Nitrogenase: Carbon in the Middle. *Nat. Chem.* **4**, 68 (2012).
12. Ribbe, M. W., Hu, Y., Hodgson, K. O. & Hedman, B. Biosynthesis of Nitrogenase Metalloclusters. *Chem. Rev.* **114**, 4063–4080 (2014).
13. Oldroyd, G. E. D. & Dixon, R. Biotechnological Solutions to the Nitrogen Problem. *Curr. Opin. Biotechnol.* **26**, 19–24 (2014).
14. Beatty, P. H. & Good, A. G. Future Prospects for Cereals that Fix Nitrogen. *Science* **333**, 416–417 (2011).
15. Temme, K., Zhao, D. & Voigt, C. A. Refactoring the Nitrogen Fixation Gene Cluster from *Klebsiella oxytoca*. *Proc. Natl. Acad. Sci.* **109**, 7085–7090 (2012).
16. Song, W. J. & Tezcan, F. A. A Designed Supramolecular Protein Assembly with *in vivo*

- Enzymatic Activity. *Science* **346**, 1525–1528 (2014).
17. Sontz, P. A., Song, W. J. & Tezcan, F. A. Interfacial Metal Coordination in Engineered Protein and Peptide Assemblies. *Curr. Opin. Chem. Biol.* **19**, 42–49 (2014).
 18. Song, W. J., Sontz, P. A., Ambroggio, X. I. & Tezcan, F. A. Metals in Protein–Protein Interfaces. *Annu. Rev. Biophys.* **43**, 409–431 (2014).
 19. Rees, D. C. *et al.* in *Molybdenum Enzymes, Cofactors, and Model Systems* **535**, 11–170 (American Chemical Society, 1993).
 20. Woo, D. Crystal Structure of the Nitrogenase Iron Protein from *Clostridium pasteurianum*. (University of California, Los Angeles, 1995).
 21. Kim, J., Woo, D. & Rees, D. C. X-Ray Crystal Structure of the Nitrogenase Molybdenum-Iron Protein from *Clostridium pasteurianum* at 3.0-Å Resolution. *Biochemistry* **32**, 7104–7115 (1993).
 22. Spatzal, T. *et al.* Evidence for Interstitial Carbon in Nitrogenase FeMo Cofactor. *Science* **334**, 940 (2011).
 23. Zhang, L. *et al.* The Sixteenth Iron in the Nitrogenase MoFe Protein. *Angew. Chemie Int. Ed.* **52**, 10529–10532 (2013).
 24. Howard, J. B., Kechris, K. J., Rees, D. C. & Glazer, A. N. Multiple Amino Acid Sequence Alignment Nitrogenase Component 1: Insights into Phylogenetics and Structure-Function Relationships. *PLoS One* **8**, e72751 (2013).
 25. Seefeldt, L. C., Hoffman, B. M. & Dean, D. R. Mechanism of Mo-Dependent Nitrogenase. *Annu. Rev. Biochem.* **78**, 701 (2009).
 26. Burgess, B. K. & Lowe, D. J. Mechanism of Molybdenum Nitrogenase. *Chem. Rev.* **96**, 2983–3012 (1996).
 27. Schindelin, H., Kisker, C., Schlessman, J. L., Howard, J. B. & Rees, D. C. Structure of ADP-AlF₄⁻-Stabilized Nitrogenase Complex and its Implications for Signal Transduction. *Nature* **387**, 370–376 (1997).
 28. Tezcan, F. A. *et al.* Nitrogenase Complexes: Multiple Docking Sites for a Nucleotide Switch Protein. *Science* **309**, 1377–1380 (2005).
 29. Igarashi, R. Y. & Seefeldt, L. C. Nitrogen Fixation: The Mechanism of the Mo-Dependent Nitrogenase. *Crit. Rev. Biochem. Mol. Biol.* **38**, 351–384 (2003).
 30. Duval, S. *et al.* Electron Transfer Precedes ATP Hydrolysis during Nitrogenase Catalysis. *Proc. Natl. Acad. Sci.* **110**, 16414–16419 (2013).
 31. Hageman, R. V, Orme-Johnson, W. H. & Burris, R. H. Role of Magnesium Adenosine 5'-Triphosphate in the Hydrogen Evolution Reaction Catalyzed by Nitrogenase from *Azotobacter vinelandii*. *Biochemistry* **19**, 2333–2342 (1980).

32. Thorneley, R. N., Lowe, D. J., Eday, R. R. & Miller, R. W. The Coupling of Electron Transfer in Nitrogenase to the Hydrolysis of Magnesium Adenosine Triphosphate. *Biochem. Soc. Trans.* **7**, 633–636 (1979).
33. Hoffman, B. M., Lukoyanov, D., Yang, Z.-Y., Dean, D. R. & Seefeldt, L. C. Mechanism of Nitrogen Fixation by Nitrogenase: The Next Stage. *Chem. Rev.* **114**, 4041–4062 (2014).
34. Lukoyanov, D. *et al.* Identification of a Key Catalytic Intermediate Demonstrates That Nitrogenase is Activated by the Reversible Exchange of N₂ for H₂. *J. Am. Chem. Soc.* **137**, 3610–3615 (2015).
35. Simpson, F. B. & Burris, R. H. A Nitrogen Pressure of 50 Atmospheres does not Prevent Evolution of Hydrogen by Nitrogenase. *Science* **224**, 1095–1097 (1984).
36. Schrock, R. R. Nitrogen Reduction: Molybdenum does it Again. *Nat. Chem.* **3**, 95–96 (2011).
37. Spatzal, T., Perez, K. A., Einsle, O., Howard, J. B. & Rees, D. C. Ligand Binding to the FeMo-Cofactor: Structures of CO-Bound and Reactivated Nitrogenase. *Science* **345**, 1620–1623 (2014).
38. Spatzal, T., Perez, K. A., Howard, J. B. & Rees, D. C. Catalysis-Dependent Selenium Incorporation and Migration in the Nitrogenase Active Site. *Elife* **4**, e11620 (2015).
39. Danyal, K., Dean, D. R., Hoffman, B. M. & Seefeldt, L. C. Electron Transfer within Nitrogenase: Evidence for a Deficit-Spending Mechanism. *Biochemistry* **50**, 9255–9263 (2011).
40. Wilson, P. E., Nyborg, A. C. & Watt, G. D. Duplication and Extension of the Thorneley and Lowe Kinetic Model for *Klebsiella pneumoniae* Nitrogenase Catalysis using a MATHEMATICA Software Platform. *Biophys. Chem.* **91**, 281–304 (2001).
41. Lowe, D. J. & Thorneley, R. N. The Mechanism of *Klebsiella pneumoniae* Nitrogenase Action: Pre-Steady-State Kinetics of H₂ Formation. *Biochem. J.* **224**, 877–886 (1984).
42. Thorneley, R. N. & Lowe, D. J. The Mechanism of *Klebsiella pneumoniae* Nitrogenase Action: Pre-Steady-State Kinetics of an Enzyme-Bound Intermediate in N₂ Reduction and of NH₃ Formation. *Biochem. J.* **224**, 887–894 (1984).
43. Lowe, D. J. & Thorneley, R. N. F. The Mechanism of *Klebsiella pneumoniae* Nitrogenase Action: The Determination of Rate Constants Required for the Simulation of the Kinetics of N₂ Reduction and H₂ Evolution. *Biochem. J.* **224**, 895–901 (1984).
44. Thorneley, R. N. & Lowe, D. J. Nitrogenase of *Klebsiella pneumoniae*. Kinetics of the Dissociation of Oxidized Iron Protein from Molybdenum-Iron Protein: Identification of the Rate-Limiting Step for Substrate Reduction. *Biochem. J.* **215**, 393–403 (1983).
45. Thorneley, R. N. & Lowe, D. J. *Molybdenum Enzymes*. (John Wiley & Sons, 1985).
46. Young, J. P. W. in *Biological Nitrogen Fixation* (eds. Stacey, G., Burris, R. H. & Evans, H. J.) 43–86 (Routledge, Chapman and Hall, 1992).

47. Mallette, M. F., Reece, P. & Dawes, E. A. Culture of *Clostridium pasteurianum* in Defined Medium and Growth as a Function of Sulfate Concentration. *Appl. Microbiol.* **28**, 999–1003 (1974).
48. Rogers, P. & Palosaari, N. *Clostridium acetobutylicum* Mutants that Produce Butyraldehyde and Altered Quantities of Solvents. *Appl. Environ. Microbiol.* **53**, 2761–2766 (1987).
49. Maddox, I. S. *et al.* The cause of ‘Acid-Crash’ and ‘Acidogenic Fermentations’ during the Batch Acetone-Butanol-Ethanol (ABE-) Fermentation Process. *J. Mol. Microbiol. Biotechnol.* **2**, 95–100 (2000).
50. Sabra, W., Groeger, C., Sharma, P. N. & Zeng, A.-P. Improved *n*-Butanol Production by a Non-Acetone Producing *Clostridium pasteurianum* DSMZ 525 in Mixed Substrate Fermentation. *Appl. Microbiol. Biotechnol.* **98**, 4267–4276 (2014).
51. Biebl, H. Fermentation of Glycerol by *Clostridium pasteurianum*: Batch and Continuous Culture Studies. *J. Ind. Microbiol. Biotechnol.* **27**, 18–26 (2001).
52. Bahadur, K. & Saroj, K. K. A Study of the Influence of Hydrogen Ion Concentration of the Culture Media on the Formation of Acetone and Butanol by *Clostridium pasteurianum*, *Clostridium butylicus*, and *Clostridium acetobutylicum* using Sucrose as Substrate. *Jpn. J. Microbiol.* **4**, 341–349 (1960).
53. Daesch, G. & Mortenson, L. E. Effect of Ammonia on the Synthesis and Function of the N₂-Fixing Enzyme System in *Clostridium pasteurianum*. *J. Bacteriol.* **110**, 103–109 (1972).
54. Ljones, T. & Burris, R. H. Nitrogenase: The Reaction between Iron Protein and Bathophenanthrolinedisulfonate as a Probe for Interactions with MgATP. *Biochemistry* **17**, 1866–1872 (1978).
55. Seefeldt, L. C., Yang, Z.-Y., Duval, S. & Dean, D. R. Nitrogenase Reduction of Carbon-Containing Compounds. *BBA-Bioenergetics* **1827**, 1102–1111 (2013).
56. Weston, M. F., Kotake, S. & Davis, L. C. Interaction of Nitrogenase with Nucleotide Analogs of ATP and ADP and the Effect of Metal Ions on ADP Inhibition. *Arch. Biochem. Biophys.* **225**, 809–817 (1983).
57. Kabsch, W. XDS. *Acta Crystallogr. Sect. D Biol. Crystallogr.* **66**, 125–132 (2010).
58. CCP4. The CCP4 Suite: Programs for Protein Crystallography. *Acta Crystallogr. Sect. D Biol. Crystallogr.* **50**, 760–763 (1994).
59. Brunger, A. T. *et al.* Crystallography & NMR System: A New Software Suite for Macromolecular Structure Determination. *Acta. Cryst.* **54**, 905–921 (1998).
60. Murshudov, G. N. *et al.* REFMAC 5 for the Refinement of Macromolecular Crystal Structures. *Acta Crystallogr. Sect. D Biol. Crystallogr.* **67**, 355–367 (2011).
61. Murshudov, G. N., Vagin, A. A. & Dodson, E. J. Refinement of Macromolecular Structures by the Maximum-Likelihood Method. *Acta Crystallogr. Sect. D Biol. Crystallogr.* **53**, 240–

- 255 (1997).
62. Afonine, P. V *et al.* Towards Automated Crystallographic Structure Refinement with Phenix.Refine. *Acta Crystallogr. Sect. D Biol. Crystallogr.* **68**, 352–367 (2012).
 63. The PyMOL Molecular Graphics System, Version 1.8 Schrödinger, LLC.
 64. Zhang, L. L.-M., Morrison, C. N., Kaiser, J. T. & Rees, D. C. Nitrogenase MoFe Protein from *Clostridium pasteurianum* at 1.08 Å Resolution: Comparison with the *Azotobacter vinelandii* MoFe Protein. *Acta Crystallogr. Sect. D Biol. Crystallogr.* **71**, 274–282 (2015).
 65. Kim, J. & Rees, D. C. Structural Models for the Metal Centers in the Nitrogenase Molybdenum-Iron Protein. *Science* **257**, 1677–1682 (1992).
 66. Mayer, S. M., Lawson, D. M., Gormal, C. A., Roe, S. M. & Smith, B. E. New Insights into Structure-Function Relationships in Nitrogenase: A 1.6 Å Resolution X-Ray Crystallographic Study of *Klebsiella pneumoniae* MoFe-Protein. *J. Mol. Biol.* **292**, 871–891 (1999).
 67. Schmid, B. *et al.* Biochemical and Structural Characterization of the Cross-Linked Complex of Nitrogenase: Comparison to the ADP- AlF_4^- -Stabilized Structure. *Biochemistry* **41**, 15557–15565 (2002).
 68. Emerich, D. W. & Burris, R. H. Interactions of Heterologous Nitrogenase Components that Generate Catalytically Inactive Complexes. *Proc. Natl. Acad. Sci. U. S. A.* **73**, 4369–4373 (1976).
 69. Eady, R. R., Smith, B. E., Cook, K. A. & Postgate, J. R. Nitrogenase of *Klebsiella pneumoniae*: Purification and Properties of the Component Proteins. *Biochem. J.* **128**, 655–675 (1972).
 70. Winkler, J. R. & Gray, H. B. Could Tyrosine and Tryptophan serve Multiple Roles in Biological Redox Processes? *Philos. Trans. A. Math. Phys. Eng. Sci.* **373**, 20140178 (2015).
 71. Hu, Y., Fay, A. W., Lee, C. C., Yoshizawa, J. & Ribbe, M. W. Assembly of Nitrogenase MoFe Protein. *Biochemistry* **47**, 3973–3981 (2008).
 72. Lancaster, K. M. *et al.* X-ray Emission Spectroscopy Evidences a Central Carbon in the Nitrogenase Iron-Molybdenum Cofactor. *Science* **334**, 974–977 (2011).
 73. Einsle, O. *et al.* Nitrogenase MoFe-Protein at 1.16 Å Resolution: A Central Ligand in the FeMo-Cofactor. *Science* **297**, 1696–1700 (2002).
 74. Chan, M. K., Kim, J. & Rees, D. C. The Nitrogenase FeMo-Cofactor and P-Cluster Pair: 2.2 Å Resolution Structures. *Science* **260**, 792–794 (1993).
 75. Peters, J. W. *et al.* Redox-Dependent Structural Changes in the Nitrogenase P-Clustes. *Biochemistry* **36**, 1181–1187 (1997).
 76. Emsley, P., Lohkamp, B., Scott, W. G. & Cowtan, K. Features and Development of Coot. *Acta Crystallogr. Sect. D Biol. Crystallogr.* **66**, 486–501 (2010).

77. Chen, V. B. *et al.* MolProbity: All-Atom Structure Validation for Macromolecular Crystallography. *Acta Crystallogr. Sect. D Biol. Crystallogr.* **66**, 12–21 (2010).
78. Morrison, C. N., Hoy, J. A., Zhang, L., Einsle, O. & Rees, D. C. Substrate Pathways in the Nitrogenase MoFe Protein by Experimental Identification of Small Molecule Binding Sites. *Biochemistry* **54**, 2052–2060 (2015).
79. Erisman, J. W., Sutton, M. A., Galloway, J., Klimont, Z. & Winiwarter, W. How a Century of Ammonia Synthesis Changed the World. *Nat. Geosci.* **1**, 636–639 (2008).
80. Kitano, M. *et al.* Ammonia Synthesis using a Stable Electride as an Electron Donor and Reversible Hydrogen Store. *Nat. Chem.* **4**, 934–940 (2012).
81. Hartwig, J. F. *Organotransition Metal Chemistry: From Bonding to Catalysis*. (Science Books, 2010).
82. Rees, D. C. & Howard, J. B. Nitrogenase: Standing at the Crossroads. *Curr. Opin. Chem. Biol.* **4**, 559–566 (2000).
83. Scott, E. E., Gibson, Q. H. & Olson, J. S. Mapping the Pathways for O₂ Entry Into and Exit from Myoglobin. *J. Biol. Chem.* **276**, 5177–5188 (2001).
84. Radding, W. & Phillips, G. N. Kinetic Proofreading by the Cavity System of Myoglobin: Protection from Poisoning. *BioEssays* **26**, 422–433 (2004).
85. Teeter, M. M. Myoglobin Cavities Provide Interior Ligand Pathway. *Protein Sci.* **13**, 313–318 (2004).
86. Schoenborn, B. P., Watson, H. C. & Kendrew, J. C. Binding of Xenon to Sperm Whale Myoglobin. *Nature* **207**, 28–30 (1965).
87. Duff, A. P. *et al.* Using Xenon as a Probe for Dioxygen-binding Sites in Copper Amine Oxidases. *J. Mol. Biol.* **344**, 599–607 (2004).
88. Johnson, B. J. *et al.* Exploring Molecular Oxygen Pathways in *Hansenula polymorpha* Copper-Containing Amine Oxidase. *J. Biol. Chem.* **282**, 17767–17776 (2007).
89. Kallio, J. P., Rouvinen, J., Kruus, K. & Hakulinen, N. Probing the Dioxygen Route in *Melanocarpus albomyces* Laccase with Pressurized Xenon Gas. *Biochemistry* **50**, 4396–4398 (2011).
90. Whittington, D. A., Rosenzweig, A. C., Frederick, C. A. & Lippard, S. J. Xenon and Halogenated Alkanes Track Putative Substrate Binding Cavities in the Soluble Methane Monooxygenase Hydroxylase. *Biochemistry* **40**, 3476–3482 (2001).
91. McCormick, M. S. & Lippard, S. J. Analysis of Substrate Access to Active Sites in Bacterial Multicomponent Monooxygenase Hydroxylases: X-ray Crystal Structure of Xenon-Pressurized Phenol Hydroxylase from *Pseudomonas* sp. OX1. *Biochemistry* **50**, 11058–11069 (2011).
92. Svensson-Ek, M. *et al.* The X-ray Crystal Structures of Wild-type and EQ(I-286) Mutant

- Cytochrome *c* Oxidases from *Rhodobacter sphaeroides*. *J. Mol. Biol.* **321**, 329–339 (2002).
93. Luna, V. M., Chen, Y., Fee, J. A. & Stout, C. D. Crystallographic Studies of Xe and Kr Binding within the Large Internal Cavity of Cytochrome *ba3* from *Thermus thermophilus*: Structural Analysis and Role of Oxygen Transport Channels in the Heme–Cu Oxidases. *Biochemistry* **47**, 4657–4665 (2008).
 94. Luna, V. M., Fee, J. A., Deniz, A. A. & Stout, C. D. Mobility of Xe Atoms within the Oxygen Diffusion Channel of Cytochrome *ba3* Oxidase. *Biochemistry* **51**, 4669–4676 (2012).
 95. Darnault, C. *et al.* Ni-Zn-[Fe₄-S₄] and Ni-Ni-[Fe₄-S₄] Clusters in Closed and Open α -Subunits of Acetyl-CoA Synthase/Carbon Monoxide Dehydrogenase. *Nat. Struct. Mol. Biol.* **10**, 271–279 (2003).
 96. Wentworth, P. *et al.* Antibody Catalysis of the Oxidation of Water. *Science* **293**, 1806–1811 (2001).
 97. Tilton, R. F. *et al.* Computational Studies of the Interaction of Myoglobin and Xenon. *J. Mol. Biol.* **192**, 443–456 (1986).
 98. Tilton, R. F., Singh, U. C., Kuntz, I. D. & Kollman, P. A. Protein-Ligand Dynamics. *J. Mol. Biol.* **199**, 195–211 (1988).
 99. Montet, Y. *et al.* Gas Access to the Active Site of Ni-Fe Hydrogenases Probed by X-Ray Crystallography and Molecular Dynamics. *Nat. Struct. Mol. Biol.* **4**, 523–526 (1997).
 100. Czerminski, R. & Elber, R. Computational Studies of Ligand Diffusion in Globins: *I. Leghemoglobin*. *Proteins Struct. Funct. Bioinforma.* **10**, 70–80 (1991).
 101. Smith, D., Danyal, K., Raugei, S. & Seefeldt, L. C. Substrate Channel in Nitrogenase Revealed by a Molecular Dynamics Approach. *Biochemistry* **53**, 2278–2285 (2014).
 102. Dance, I. A Molecular Pathway for the Egress of Ammonia Produced by Nitrogenase. *Sci. Rep.* **3**, 3237 (2013).
 103. Durrant, M. C. Controlled Protonation of Iron-Molybdenum Cofactor by Nitrogenase: A Structural and Theoretical Analysis. *Biochem. J.* **355**, 569–576 (2001).
 104. Barney, B. M., Yurth, M. G., Dos Santos, P. C., Dean, D. R. & Seefeldt, L. C. A Substrate Channel in the Nitrogenase MoFe Protein. *J. Biol. Inorg. Chem.* **14**, 1015–1022 (2009).
 105. Dance, I. The Controlled Relay of Multiple Protons Required at the Active Site of Nitrogenase. *Dalt. Trans.* **41**, 7647–7659 (2012).
 106. Dance, I. Nitrogenase: A General Hydrogenator of Small Molecules. *Chem. Commun.* **49**, 10893–10907 (2013).
 107. Ringe, D., Petsko, G. A., Kerr, D. E. & Ortiz de Montellano, P. R. Reaction of Myoglobin with Phenylhydrazine: A Molecular Doorstop. *Biochemistry* **23**, 2–4 (1984).
 108. Chovancova, E. *et al.* CAVER 3.0: A Tool for the Analysis of Transport Pathways in

- Dynamic Protein Structures. *PLOS Comput. Biol.* **8**, e1002708 (2012).
109. Hu, Y. & Ribbe, M. W. Nitrogenase Assembly. *Biochim. Biophys. Acta* **1827**, 1112–1122 (2013).
 110. Rupnik, K. *et al.* Nonenzymatic Synthesis of the P-Cluster in the Nitrogenase MoFe Protein: Evidence of the Involvement of All-Ferrous $[\text{Fe}_4\text{S}_4]^0$ Intermediates. *Biochemistry* **53**, 1108–1116 (2014).
 111. Schmid, B. *et al.* Structure of a Cofactor-Deficient Nitrogenase MoFe Protein. *Science* **296**, 352–356 (2002).
 112. Stowell, M. H. B. *et al.* A Simple Device for Studying Macromolecular Crystals under Moderate Gas Pressures (0.1–10 MPa). *J. Appl. Crystallogr.* **29**, 608–613 (1996).
 113. Zondlo, N. J. Aromatic–Proline Interactions: Electronically Tunable CH/ π Interactions. *Acc. Chem. Res.* **46**, 1039–1049 (2013).
 114. Leitgeb, B. & Tóth, G. Aromatic–Aromatic and Proline–Aromatic Interactions in Endomorphin-1 and Endomorphin-2. *Eur. J. Med. Chem.* **40**, 674–686 (2005).
 115. Udvardi, M., Brodie, E. L., Riley, W., Kaeppler, S. & Lynch, J. Impacts of Agricultural Nitrogen on the Environment and Strategies to Reduce these Impacts. *Procedia Environ. Sci.* **29**, 303 (2015).
 116. George, S. J., Ashby, G. A., Wharton, C. W. & Thorneley, R. N. F. Time-Resolved Binding of Carbon Monoxide to Nitrogenase Monitored by Stopped-Flow Infrared Spectroscopy. *J. Am. Chem. Soc.* **119**, 6450–6451 (1997).
 117. Meyer, J. Comparison of Carbon Monoxide, Nitric Oxide, and Nitrite as Inhibitors of the Nitrogenase from *Clostridium pasteurianum*. *Arch. Biochem. Biophys.* **210**, 246–256 (1981).
 118. Rivera-Ortiz, J. M. & Burris, R. H. Interactions among Substrates and Inhibitors of Nitrogenase. *J. Bacteriol.* **123**, 537–545 (1975).
 119. Sattler, M. *Introduction to Biomolecular NMR Spectroscopy*. (2004).
 120. Thiyagarajan, P. *et al.* Design and Expected Performance of a High Resolution Macromolecular Neutron Diffractometer (MaNDi) at the Spallation Neutron Source. *Trans. Am. Crystallogr. Assoc.* **38**, 67–76 (2003).
 121. Hainbuchner, M. & Jericha, E. *Bound Coherent Neutron Scattering Lengths*. (2001).
 122. Wilson, C. C. Towards Large Molecule Crystallography at ISIS. *Trans. Am. Crystallogr. Assoc.* **38**, 14–25 (2003).
 123. Teixeira, S. C. M. *Neutron Crystallography*. (2010).
 124. Schoenborn, B. P. Neutron Diffraction Analysis of Myoglobin. *Nature* **224**, 143–146 (1969).

125. Kossiakoff, A. A. & Spencer, S. A. Neutron Diffraction Identifies His57 as the Catalytic Base in Trypsin. *Nature* **288**, 414–416 (1980).
126. Chen, J. C.-H., Hanson, B. L., Fisher, S. Z., Langan, P. & Kovalevsky, A. Y. Direct Observation of Hydrogen Atom Dynamics and Interactions by Ultrahigh Resolution Neutron Protein Crystallography. *Proc. Natl. Acad. Sci. U. S. A.* **109**, 15301–15306 (2012).
127. Kovalevsky, A. Y. *et al.* Direct Determination of Protonation States of Histidine Residues in a 2 Å Neutron Structure of Deoxy-Human Normal Adult Hemoglobin and Implications for the Bohr Effect. *J. Mol. Biol.* **398**, 276–291 (2010).
128. Howard, J. B. & Rees, D. C. How many Metals does it take to Fix N₂? A Mechanistic Overview of Biological Nitrogen Fixation. *Proc. Natl. Acad. Sci.* **103**, 17088–17093 (2006).
129. Münck, E. *et al.* Nitrogenase: VIII. Mössbauer and EPR spectroscopy. The MoFe Protein Component from *Azotobacter vinelandii*. *BBA - Protein Struct.* **400**, 32–53 (1975).
130. Surerus, K. K. *et al.* Mössbauer and Integer-Spin EPR of the Oxidized P-Clusters of Nitrogenase: P^{OX} is a Non-Kramers System with a nearly Degenerate Ground Doublet. *J. Am. Chem. Soc.* **114**, 8579–8590 (1992).
131. Pierik, A. J., Wassink, H., Haaker, H. & Hagen, W. R. Redox Properties and EPR Spectroscopy of the P-Clusters of *Azotobacter vinelandii* MoFe Protein. *Eur. J. Biochem.* **212**, 51–61 (1993).
132. Fisher, K., Newton, W. E. & Lowe, D. J. Electron Paramagnetic Resonance Analysis of Different *Azotobacter vinelandii* Nitrogenase MoFe-Protein Conformations Generated during Enzyme Turnover: Evidence for $S = 3/2$ Spin States from Reduced MoFe-Protein Intermediates. *Biochemistry* **40**, 3333–3339 (2001).
133. Fisher, K. *et al.* Conformations Generated during Turnover of the *Azotobacter vinelandii* Nitrogenase MoFe Protein and their Relationship to Physiological Function. *J. Inorg. Biochem.* **101**, 1649–56 (2007).
134. Lukoyanov, D. *et al.* A Confirmation of the Quench-Cryoannealing Relaxation Protocol for Identifying Reduction States of Freeze-Trapped Nitrogenase Intermediates. *Inorg. Chem.* **53**, 3688–3693 (2014).
135. Wiig, J. A., Hu, Y. & Ribbe, M. W. Refining the Pathway of Carbide Insertion into the Nitrogenase M-Cluster. *Nat. Commun.* **6**, 8034 (2015).
136. Spatzal, T. *et al.* Nitrogenase FeMoco Investigated by Spatially Resolved Anomalous Dispersion Refinement. *Nat. Commun.* **7**, 10902 (2016).
137. Pham, D. N. & Burgess, B. K. Nitrogenase Reactivity: Effects of pH on Substrate Reduction and Carbon Monoxide Inhibition. *Biochemistry* **32**, 13725–13731 (1993).
138. Ellis, K. J. & Morrison, J. F. Buffers of cConstant Ionic Strength for Studying pH-Dependent Processes. *Methods Enzymol.* **87**, 405–426 (1982).
139. Hayward, S. Peptide-Plane Flipping in Proteins. *Protein Sci.* **10**, 2219–2227 (2001).

140. Fay, A. W., Hu, Y., Schmid, B. & Ribbe, M. W. Molecular Insights into Nitrogenase FeMoco Insertion: The Role of His274 and His451 of MoFe Protein α -Subunit. *J. Inorg. Biochem.* **101**, 1630–1641 (2007).
141. Smith, B. E. *et al.* Exploring the Reactivity of the Isolated Iron-Molybdenum Cofactor of Nitrogenase. *Coord. Chem. Rev.* **185–186**, 669–687 (1999).
142. Yang, K. Y., Haynes, C. A., Spatzal, T., Rees, D. C. & Howard, J. B. Turnover-Dependent Inactivation of the Nitrogenase MoFe-Protein at High pH. *Biochemistry* **53**, 333–343 (2014).
143. Bruice, T. C., Maskiewicz, R. & Job, R. The Acid-Base Properties, Hydrolytic Mechanism, and Susceptibility to O₂ Oxidation of Fe₄S₄(SR)₄⁻² Clusters. *Proc. Natl. Acad. Sci. U. S. A.* **72**, 231–234 (1975).
144. Bates, K., Garrett, B. & Henderson, R. A. Rates of Proton Transfer to Fe–S-Based Clusters: Comparison of Clusters Containing [MFe(μ_2 -S)₂]_n⁺ and [MFe₃(μ_3 -S)₄]_n⁺ (M = Fe, Mo, or W) Cores. *Inorg. Chem.* **46**, 11145–11155 (2007).
145. Chen, K. *et al.* Atomically Defined Mechanism for Proton Transfer to a Buried Redox Centre in a Protein. *Nature* **405**, 814–817 (2000).
146. Morgan, T. V., Mortenson, L. E., McDonald, J. W. & Watt, G. D. Comparison of Redox and EPR Properties of the Molybdenum Iron Proteins of *Clostridium pasteurianum* and *Azotobacter vinelandii* Nitrogenases. *J. Inorg. Biochem.* **33**, 111–120 (1988).
147. Spatzal, T., Einsle, O. & Andrade, S. L. A. Analysis of the Magnetic Properties of Nitrogenase FeMo Cofactor by Single-Crystal EPR Spectroscopy. *Angew. Chemie Int. Ed.* **52**, 10116–10119 (2013).
148. Smith, B. E., Lowe, D. J. & Bray, R. C. Studies by Electron Paramagnetic Resonance on the Catalytic Mechanism of Nitrogenase of *Klebsiella pneumoniae*. *Biochem. J* **135**, 331–341 (1973).
149. Good, N. E. *et al.* Hydrogen Ion Buffers for Biological Research. *Biochemistry* **5**, 467–477 (1966).
150. Stoll, S. & Schweiger, A. EasySpin: A Comprehensive Software Package for Spectral Simulation and Analysis in EPR. *J. Magn. Reson.* **178**, 42–55 (2006).